Physicochemical qualities, antioxidant compounds, and activities of six mini paprika cultivars

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Abstract Paprika is a popular vegetable with high visual appeal and desirable flavor, the health benefits of which are increasingly attracting interest. In this study, the physical qualities, antioxidant content, and activities of six mini paprika cultivars were investigated. Both the edible part (flesh) and the by-products were studied. The average total phenolics and total antioxidant activities were higher in the flesh than in the by-products. The total flavonoids of the flesh and the by-products were 16.41 and 37.80 mg/100 g FW, respectively. “YW glory” and “Raon yellow” flesh had the highest (245.52 mg/100 g FW) and lowest (179.96 mg/100 g FW) total phenolics among the six cultivars, respectively. However, the “RD glory” cultivar showed the highest total phenolic content (232.70 mg/100 g FW) among the by-product samples. The total phenolics in the flesh and by-products were highly correlated to the ABTS radical scavenging activity, with R=0.961 and 0.984, respectively.

Keywords: mini paprika, flavonoids, phenolics, antioxidant, correlation

Introduction

Recently, food consumption patterns have changed as a result of an increase in single-person households and changes in the structure of society and national living standards due to multiple factors (Statistics Korea, 2020; Lee et al., 2019). Led by single-person households, food consumption preference is shifting towards small quantity products with convenience and health (Lee et al., 2015; Cho et al., 2007). The popularity of miniature vegetables has increased and they are conveniently sized while maintaining the nutritional properties of their regular-sized counterparts. This results in more frequent consumption and larger cultivation areas (Hwang, 2019). Paprika (Capsicum annuum L.) is a rich source of phytochemicals, including carotenoid, tocopherol, and phenolic compounds, whose consumption is currently on the rise (Kim et al., 2011; Pérez-Gálvez and Mínguez-Mosquera, 2001).

Paprika is an annual plant that belongs to Solanaceae, Capsicum, and Annuum families (Jung and Hong, 2017). In general, paprika is initially green when not matured, and gradually becomes red as it matures. However, it can take on more colors depending on its variety, including red, orange, and yellow (Luitel and Kang, 2013). The bioactive compound content of paprika is dependent on its maturity, variety, and growing conditions (Hallmann and Rembiatkowska, 2012). Among bioactive compounds, phenolic compounds are strong antioxidants that can protect the human body from reactive oxygen species (ROS). Antioxidants also reduce the levels of ROS related to oxidative stress-related diseases, such as cancer, aging, and inflammatory diseases (Nimse and Pal, 2015). ROS attacks all major biological molecules, including polyunsaturated fatty acid (PUFA), its target on the cell membrane. The oxidative damage of PUFA advances via an autonomous chain reaction, wherein free radicals cause extensive oxidative injury (Poljsak et al., 2013). Antioxidants inhibit this response and delay cell damage. Thus, they are highly regarded as physiologically active substances for the prevention of hypertension, diabetes, carcinogenesis, and aging (Hwang et al., 2020).

Paprika is normally consumed after removing the inedible parts of the vegetable, such as the stalk end, core, and seeds. According to Cho et al. (2007), vegetables comprise 53.1% of food waste, a much higher percentage than fruits (13.7%). Lee et al. (2012) conducted a study on the antioxidant compounds and activities of peels, that is, the inedible parts of fruits, in oranges, golden and green kiwifruits, tangerine, and Korean melon, among others. Similarly, Kabir et al. (2015) studied the antioxidant activities of the inedible parts of cabbage and broccoli. However, there is currently a lack of research on the properties of the inedible parts of paprika. Furthermore, previous studies have mainly focused on the antioxidant compounds and activities of paprika. Šaponjac et al. (2014) investigated the anticancer activity of paprika, while Go et al. (2020) focused on the antibacterial activity of paprika and Hwang et al. (2012) investigated the antioxidative activity of paprika. On the other hand, studies on miniature paprika (or mini paprika) have mainly focused its physicochemical quality with insufficient research on its antioxidant compounds and activities. In this study, six varieties of mini paprika are studied. The six mini paprika cultivars were divided into flesh and by-products, namely core and seeds, to comparatively measure the antioxidant compounds and activities and analyze the correlation among each
acidity was then converted to citric acid %. The pH was measured using drops of 1% phenolphthalein solution. The measured titratable acidity was determined by performing a neutralization titration with 0.1 N NaOH and 3-4 drops of 1% phenolphthalein solution. The measured titratable acidity was then divided into flesh and by-products (core and seeds) (Fig. 1), frozen in a freezer at −196°C, and stored in a freezer at −25°C until analysis of antioxidant compounds and activities.

Materials and Methods

Materials for experiment

Mini paprika cultivars “RD glory”, “OE glory”, and “YW glory” were purchased from Saedanong Co., Ltd. (Hoengseong, Korea), and “Raon red”, “Raon orange”, and “Raon yellow” from Jinju Mini Paprika farm (Jinju, Korea). All cultivars were found at the commercially ripe stage at the time of the experiments. The flesh of the vegetables was analyzed on the day of purchase for physical quality analysis, such as length, color, firmness, pH, acidity, and sugar content. The mini paprika were then divided into flesh and by-products (core and seeds) (Fig. 1), frozen with liquid nitrogen at −196°C, and stored in a freezer at −25°C until analysis of antioxidant compounds and activities.

Color measurement

The color of the flesh was represented as L* value (lightness), a* value (redness), and b* value (yellowness) using a colorimeter (Chroma meter CR-400, Minolta, Tokyo, Japan). For each measurement, 10 samples were measured three times and the mean value was calculated. The Chroma meter was calibrated regularly using the white color plate (Y=87.8, x=0.3156, y=0.3229).

Soluble solid content, firmness, titratable acidity, pH, moisture content, and length

The firmness of the mini paprika flesh was measured using a fruit hardness tester (FHM-1, Demetra Co., Ltd., Tokyo, Japan). The fruit skin of 10 samples was penetrated using a 12Φ×10 mm core probe, and the value was expressed in newtons (N). The soluble solid content, titratable acidity, and pH were measured using a mixture of the flesh homogenized with a blender (JB 3060, Braun Co., Kronberg Germany). The soluble solid content was measured using a digital refractometer (PAL-1, Atago Co., Ltd., Tokyo, Japan). The titratable acidity was determined by mixing 1 g of the mixture and 100 mL of distilled water and performing a neutralization titration with 0.1 N NaOH and 3-4 drops of 1% phenolphthalein solution. The measured titratable acidity was then converted to citric acid %. The pH was measured using a pH meter (Starter300, Ohaus Co., Ltd., Parsippany, NJ, USA). To measure the moisture content of the fruit, samples of the flesh were weighed and stored in an incubator (IB-05G Jeio Tech Co., Ltd., Seoul, Korea) at 70°C. The moisture content was measured and converted into percentages. The length was measured using a digital caliper (1108-150W, INSIZE Co., Ltd., Suzhou, China).

Extraction of samples

Ethanol (80%) was used as the solvent for extracting the flesh and by-products of the mini paprika cultivars. Briefly, 30 g of frozen samples were added to 300 mL of 80% ethanol. This mixture was then homogenized in a blender twice for 3 min and once for 2 min. The homogenized solution was filtered through a decompression filter using a Whatman #2 paper filter (Whatman International Ltd. Kent, UK). Then, the filtered solution was concentrated using a rotary evaporator (N-1000, Eyela, Tokyo, Japan) at 45°C and then stored at −20°C until analysis of antioxidant compounds and activities (Yang et al., 2019).

Total flavonoid content analysis

The total flavonoid content of the extracts from the flesh and by-products was measured by the colorimetric assay (Zulkifli et al., 2020). Briefly, 0.3 mL of 5% NaNO_2 was added to a 15 mL test tube containing 4 mL of distilled water and 1 mL of diluted sample. The mixture was vortexed and left to stand for 5 min at room temperature. Then, 0.3 mL of 10% AlCl_3 was added and the mixture was vortexed and left at room temperature for 6 min. Next, 2 mL of 1 N NaOH and 2.4 mL of distilled water were added to adjust the total volume to 10 mL. The optical density of this solution was measured at a wavelength of 510 nm using a spectrophotometer (Optizen POP, Mecasys, Daejeon, Korea). The standard calibration curve was measured using catechin as a standard, and the total flavonoid content was converted to and expressed in mg catechin equivalents (CE)/100 g fresh weight (FW).

Total phenolic content analysis

The total phenolic content of the flesh and by-product extracts was measured by the Folin-Ciocalteu colorimetric assay (Shin, 2012; Yang et al., 2019). Briefly, 0.2 mL of the Folin-Ciocalteu phenol reagent was added to a 15 mL test tube containing 2.6 mL of distilled water and 0.2 mL of diluted sample. The mixture was vortexed and left to stand at room temperature for 6 min. Next, 2 mL of 7% NaCO_3 was added and the mixture was vortexed and left at room temperature in a dark place for 90 min. The optical density of this solution was measured at a wavelength of 750 nm using a spectrophotometer. The standard calibration curve was measured using gallic acid as a standard, and the total phenolic content was converted to and expressed in mg gallic acid equivalents (GAE)/100 g FW.

DPPH radical scavenging activity

The DPPH radical scavenging activity of the flesh and by-product extracts was measured using the methods described by Hwang et al. (2020) and Sridhar et al. (2019) with some modifications.
The test solution containing 50 µL of diluted sample mixed with 2,950 µL of 0.2 mM DPPH solution was left to react at room temperature in a dark place for 30 min. The optical density of this solution was then measured at a wavelength of 517 nm using a spectrophotometer. The standard calibration curve was measured using vitamin C as a standard, and the antioxidant activity of the extract was converted to and expressed in mg vitamin C equivalents (VCE)/100 g FW.

**ABTS radical scavenging activity**

The ABTS radical scavenging activity of the flesh and by-product extracts was measured using the methods described by Floge et al. (2011) and Sridhar et al. (2019) with some modifications. The test solution containing 20 µL of diluted sample mixed with 980 µL of the ABTS reaction solution was left to react at 37°C for 10 min. The optical density of this solution was then measured at a wavelength of 734 nm using a spectrophotometer. The standard calibration curve was measured using vitamin C as a standard, and the ABTS radical scavenging activity was converted to and expressed in mg VCE/100 g FW.

**Statistical analysis**

For the statistical analysis of each experiment, analysis of variance (ANOVA) was performed using SPSS 20 program (SPSS Inc. Chicago, IL, USA), and Duncan’s multiple range test was used to analyze the significant differences (p<0.05). The correlation between the mean values of each factor was expressed using Pearson’s correlation coefficient.

**Results and Discussion**

**Color**

The color measurement results of the different mini paprika varieties are shown in Table 1. The variety with the highest L* value (lightness) was “YW glory” with 65.43±0.48. The a* value (redness) of the two red varieties was 31.80±0.72 for “RD glory” and 29.18±0.96 for “Raon red”, while “RD glory” significantly higher. The b* value (yellowness) of the two yellow varieties was 42.63±0.97 for “YW glory” and 37.97±0.87 for “Raon yellow”, with “YW glory” significantly higher. A comparative study by Lee et al. (2016) on the characteristics of paprika by variety and color also showed that “RD glory” had higher a* value than “Raon red”, and “YW glory” had a higher b* value than “Raon yellow”.

**Soluble solid content, firmness, titratable acidity, pH, moisture content, and length**

The soluble solid content, firmness, titratable acidity, pH, moisture content, and length of the mini paprika flesh are shown in Table 2. The soluble solid content of the mini paprika cultivars ranged from 7.20 to 8.90%. Kim et al. (2011) reported that the main free sugars of green and red paprika were fructose and glucose. According to Kang et al. (2008), the soluble solid content of larger paprika varieties, namely the “Special” and “Fiesta” varieties, ranged from 5.84 to 7.19%, indicating that the soluble content of the mini paprika cultivars used in this experiment was higher, with a firmness of 4.20-5.22 N and a titratable acidity and pH of 0.52-0.66% and 4.90-5.19, respectively. According to Lee et al. (2017), the titratable acidity of red paprika is 0.8%, which is higher than our findings for mini paprika. However, the pH of 5.10 for the red paprika was similar to our findings for mini paprika, with a moisture content ranging from 85.18% to 90.80%. Jeong et al. (2006) studied the quality of paprika and found a moisture content of 89.80-90.92%, which is similar to our findings for mini paprika. The length of the mini paprika varieties ranged from 77.87 to 83.73 mm, with the “Raon orange” variety significantly longer than the “OE glory” and “YW glory” varieties.

**Table 1. Hunter L*,a*,b* color of mini paprika flesh**

<table>
<thead>
<tr>
<th>General name</th>
<th>Cultivar name</th>
<th>L* (Lightness)</th>
<th>a* (Redness)</th>
<th>b* (Yellowness)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Vinesweet</td>
<td>RD glory</td>
<td>47.33±0.34</td>
<td>31.80±0.72</td>
<td>12.95±0.54</td>
</tr>
<tr>
<td></td>
<td>OE glory</td>
<td>58.36±0.67</td>
<td>26.11±0.78</td>
<td>32.43±1.36</td>
</tr>
<tr>
<td></td>
<td>YW glory</td>
<td>65.43±0.48</td>
<td>13.28±1.08</td>
<td>42.63±0.97</td>
</tr>
<tr>
<td>Raon</td>
<td>Raon red</td>
<td>49.56±0.38</td>
<td>29.18±0.90</td>
<td>17.13±0.89</td>
</tr>
<tr>
<td></td>
<td>Raon orange</td>
<td>56.55±0.45</td>
<td>22.31±0.47</td>
<td>28.41±0.75</td>
</tr>
<tr>
<td></td>
<td>Raon yellow</td>
<td>62.60±0.52</td>
<td>10.74±0.91</td>
<td>37.97±0.87</td>
</tr>
</tbody>
</table>

*RD: Red color, OE: Orange color, YW: Yellow color

**Table 2. Soluble solid contents, firmness, titratable acidities, pH, moisture contents and length of mini paprika flesh**

<table>
<thead>
<tr>
<th>General name</th>
<th>Cultivar name</th>
<th>SSC (% brix)</th>
<th>Firmness (N/12 mmO)</th>
<th>Titratable acidity (%)</th>
<th>pH</th>
<th>SSC/TA ratio</th>
<th>Moisture contents (%)</th>
<th>Length (mm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Vinesweet</td>
<td>RD glory</td>
<td>11.50±0.07</td>
<td>4.20±0.18</td>
<td>0.66±0.02</td>
<td>5.15±0.002</td>
<td>17.42</td>
<td>85.36±0.11</td>
<td>79.72±2.66</td>
</tr>
<tr>
<td></td>
<td>OE glory</td>
<td>10.87±0.02</td>
<td>5.04±0.23</td>
<td>0.54±0.02</td>
<td>5.19±0.01</td>
<td>20.13</td>
<td>85.18±0.12</td>
<td>78.22±3.49</td>
</tr>
<tr>
<td></td>
<td>YW glory</td>
<td>11.07±1.38</td>
<td>5.22±0.29</td>
<td>0.66±0.02</td>
<td>5.15±0.004</td>
<td>16.77</td>
<td>85.42±0.12</td>
<td>77.87±2.26</td>
</tr>
<tr>
<td>Raon</td>
<td>Raon red</td>
<td>11.47±0.06</td>
<td>4.48±0.02</td>
<td>0.66±0.02</td>
<td>5.01±0.002</td>
<td>17.38</td>
<td>88.79±0.15</td>
<td>82.99±2.50</td>
</tr>
<tr>
<td></td>
<td>Raon orange</td>
<td>9.87±0.16</td>
<td>4.35±0.10</td>
<td>0.54±0.02</td>
<td>4.90±0.002</td>
<td>18.28</td>
<td>89.90±0.14</td>
<td>83.73±1.98</td>
</tr>
<tr>
<td></td>
<td>Raon yellow</td>
<td>9.20±0.04</td>
<td>4.78±0.17</td>
<td>0.52±0.01</td>
<td>4.90±0.003</td>
<td>17.69</td>
<td>90.80±0.11</td>
<td>82.15±2.18</td>
</tr>
</tbody>
</table>

*RD: Red color, OE: Orange color, YW: Yellow color
The total flavonoid content of the different mini paprika parts and varieties is shown in Fig. 2A. The by-products (core and seeds) showed a higher flavonoid content than the flesh. In particular, the flavonoid content of the by-products was significantly high in the “OE glory” and “YW glory” varieties, with 48.04±0.45 and 48.41±0.5 mg CE/100 g, respectively, and lowest in the “Raon orange” variety, with 26.84±0.21 mg CE/100 g FW. On the other hand, the flavonoid content of the flesh was significantly high in the “RD glory” and “YW glory” varieties, with 20.64±0.49 and 21.78±0.92 mg CE/100 g FW, respectively, and lowest in the “Raon red” and “Raon orange” varieties, with 12.85±0.33 and 11.70±0.74 mg CE/100 g FW, respectively. Ribes-Moya et al. (2020) studied the flavonoids of paprika and found that the flavonoid content of the paprika varieties “Bola”, “ECU-944”, and “JalapenoM” was 88.54, 57.11, and 65.37 mg/kg FW, respectively, which is lower than our findings for mini paprika. Two flavonoids represent approximately 41% of total flavonoid content in paprika, namely quercetin and luteolin (Anotonio et al., 2018). According to Ribes-Moya et al. (2020), the “Serrano” variety had a higher content of luteolin and quercetin than myricetin, apigenin, and Kaempferol.

Total phenolic content

Paprika is rich in bioactive compounds with health promoting properties, including carotenoid, ascorbic acid, tocopherol, and...
Antioxidant of paprika

Phenolic compounds (Barbosa et al., 2019). The total phenolic content of the mini paprika by part and variety is shown in Fig. 2B. In all varieties except “RD glory”, the flesh showed a significantly higher phenolic content than the by-products. The flesh of paprika is the part that is typically consumed and is the part with the highest phenolic content. The phenolic content of the flesh was highest in the “YW glory” variety, with 245.52±1.94 mg GAE/100 g FW. By contrast, the phenolic content of the by-products was highest in the “RD glory” variety, with 232.70±1.42 mg GAE/100 g FW. According to Kim et al. (2011), the total phenolic content in the flesh of red paprika cultivated in Korea was 731.75 mg CE/100 g DW, which is lower than that of mini paprika in this experiment.

DPPH radical scavenging activity

Using the DPPH method, the antioxidant activity of the mini paprika cultivars was found to be higher in the flesh than the by-products in all the samples (Fig. 3A). In the flesh, the “YW glory” variety showed the highest antioxidant activity, with 211.15±2.45 mg VCE/100 g FW, while the “Raon yellow” variety showed the lowest antioxidant activity, with 146.38±2.19 mg VCE/100 g FW. In the by-products, the “RD glory” and “YW glory” varieties showed the highest antioxidant activity, with 91.23±3.85 and 90.71±1.89 mg VCE/100 g FW, respectively. The changes in DPPH radical scavenging activity

![Fig. 3. DPPH free radical scavenging activities (A) and ABTS free radical scavenging activities (B) of mini paprika by different parts.](image)
radical scavenging activity showed a pattern similar to that of the total phenolic content, but opposite to that of the total flavonoid content. This implies that total phenols contribute more to DPPH radical scavenging activity than total flavonoids.

**ABTS radical scavenging activity**

The ABTS radical scavenging activity of the different mini paprika varieties are shown in Fig. 3B. In general, the antioxidant activity was found to be higher in the flesh than in the by-products. In the flesh, the “YW glory” variety showed the highest antioxidant activity, with 257.38±2.99 mg VCE/100 g FW, while the “Raon yellow” variety showed the lowest antioxidant activity, with 188.77±3.38 mg VCE/100 g FW. In terms of the by-products, the “RD glory” and “YW glory” varieties showed a significantly high antioxidant activity, with 216.15±3.34 and 211.13±2.00 mg VCE/100 g FW, respectively, while the “Raon red”, “Raon orange”, and “Raon yellow” varieties showed significantly low antioxidant activity, with 127.86±2.42, 127.72±1.58, and 136.13±2.00 mg VCE/100 g FW, respectively. In their analysis of the antioxidant activity of green chili between the variety and the antioxidant compounds and activities. Similarly, Park et al. (2019) found that the correlation between the antioxidant compounds and activities of 12 sprout vegetables was very high, with R=0.926 and R=0.988, respectively. Moreover, Kim et al. (2006) found a high correlation between antioxidant compounds and activities were as follows: total flavonoid (R=0.523), total phenolics (R=0.754), DPPH radical scavenging activity (R=0.708), DPPH radical scavenging activity (R=0.964), and ABTS radical scavenging activity (R=0.984). In general, fruits and vegetables show a high correlation between antioxidant compounds and activities (Kim and Shin, 2015; Yun et al., 2018; Hwang et al., 2019). Yoon et al. (2012) found a high correlation (R=0.712) for the antioxidant effect of green chili between the variety and the antioxidant compounds and activities. Moreover, Kim et al. (2006) found a high correlation between antioxidant compounds and activities were as follows: total flavonoid (R=0.523), total phenolics (R=0.922), and ABTS radical scavenging activity (R=0.966).

**Conclusion**

This study measured and comparatively analyzed the color, soluble solid content, firmness, titratable acidity, pH, moisture content, length, antioxidant compound, and antioxidant activity of six varieties of mini paprika: “RD glory”, “OE glory”, “YW glory”, “Raon red”, “Raon orange”, and “Raon yellow”. The total

### Table 3. Pearson correlation among physical qualities, antioxidant compounds, and activities of mini paprika flesh

<table>
<thead>
<tr>
<th></th>
<th>a*</th>
<th>b*</th>
<th>SSC</th>
<th>Firmness</th>
<th>Titratable acidity</th>
<th>pH</th>
<th>Moisture content</th>
<th>Total flavonoids</th>
<th>Total phenolics</th>
<th>DPPH</th>
<th>ABTS</th>
</tr>
</thead>
<tbody>
<tr>
<td>L*</td>
<td>0.891**&lt;1&gt;</td>
<td>0.990**&lt;2&gt;</td>
<td>0.527*</td>
<td>0.692**</td>
<td>0.513*</td>
<td>0.043</td>
<td>0.121</td>
<td>0.077</td>
<td>-0.017</td>
<td>0.000</td>
<td></td>
</tr>
<tr>
<td>a*</td>
<td>-</td>
<td>0.889**</td>
<td>0.673**</td>
<td>0.513*</td>
<td>0.500*</td>
<td>0.257</td>
<td>0.352</td>
<td>0.015</td>
<td>0.243</td>
<td>0.198</td>
<td>0.181</td>
</tr>
<tr>
<td>b*</td>
<td>-</td>
<td>-</td>
<td>0.516*</td>
<td>0.715**</td>
<td>0.519**</td>
<td>0.005</td>
<td>0.076</td>
<td>0.118</td>
<td>0.033</td>
<td>0.024</td>
<td>0.038</td>
</tr>
<tr>
<td>SSC</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>0.094</td>
<td>0.657**</td>
<td>0.656**</td>
<td>0.713**</td>
<td>0.523**</td>
<td>0.708**</td>
<td>0.691**</td>
<td>0.676**</td>
</tr>
<tr>
<td>Firmness</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>0.361</td>
<td>0.407</td>
<td>0.288</td>
<td>0.320</td>
<td>0.191</td>
<td>0.216</td>
<td>0.257</td>
</tr>
<tr>
<td>Titratable acidity</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>0.255</td>
<td>0.296</td>
<td>0.230</td>
<td>0.392</td>
<td>0.323</td>
<td>0.250</td>
</tr>
<tr>
<td>pH</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>0.950**</td>
<td>0.842**</td>
<td>0.847**</td>
<td>0.793**</td>
<td>0.868**</td>
</tr>
<tr>
<td>Moisture content</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>0.854**</td>
<td>-0.901**</td>
<td>-0.937**</td>
<td>-0.977**</td>
</tr>
<tr>
<td>Total flavonoids</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>0.881**</td>
<td>0.853**</td>
<td>0.888**</td>
</tr>
<tr>
<td>Total phenolics</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>0.964**</td>
<td>0.961**</td>
</tr>
<tr>
<td>DPPH</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>0.966**</td>
</tr>
</tbody>
</table>

Pearson correlation (R): **significance at p<0.01, *significance at p<0.05. DPPH: 2,2-diphenyl-1-picrylhydrazyl; ABTS: 2,2-azino-bis (3-ethylbenzothiazoline-6-sulfonic acid). L*: lightness, a*: redness, b*: yellowness.

### Table 4. Pearson correlation among physical qualities, antioxidant compounds, and activities of mini paprika by-products (core and seeds)

<table>
<thead>
<tr>
<th></th>
<th>Total phenolics</th>
<th>DPPH</th>
<th>ABTS</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total flavonoids</td>
<td>0.843**&lt;1&gt;</td>
<td>0.754**</td>
<td>0.881**</td>
</tr>
<tr>
<td>Total phenolics</td>
<td>-</td>
<td>0.922**</td>
<td>0.984**</td>
</tr>
<tr>
<td>DPPH</td>
<td>-</td>
<td>-</td>
<td>0.953**</td>
</tr>
</tbody>
</table>

Pearson correlation (R): **significance at p<0.01, *significance at p<0.05. DPPH: 2,2-diphenyl-1-picrylhydrazyl; ABTS: 2,2-azino-bis (3-ethylbenzothiazoline-6-sulfonic acid).
flavonoid content was significantly higher in the by-products than the flesh of the mini paprika cultivars. By contrast, the total phenolic content was significantly higher in the flesh than in the by-products. The antioxidant activity was measured by DPPH radical scavenging and ABTS radical scavenging activities, and both were higher in flesh than by-products. Moreover, there was a high correlation among antioxidant compounds and antioxidant activities in flesh and by-products. Compared to regular-size paprika, mini paprika in this study contained similar or higher antioxidant compounds and activities, indicating that they have high utility in terms of nutrition. Furthermore, the total flavonoid of mini paprika was higher in by-products, inedible parts, than flesh, the edible part. This indicates that the by-product parts such as core and seeds can be used as dried tea or dried chip as a snack. They can be also expected to be used for animal feed with high health functionality.

Acknowledgments

We would like to thank Yenu Kim and Minji Joung for helping physical quality analysis.

References

Jung HA, Hong JY. Change in quality characteristics of yellow paprika according to drying methods. Korean J. Food Preserv. 24: 1079-1087 (2017)
Lee KI, Hwang YJ, Ban HJ, Lim SJ. Impact of the growth of single-person households on the food market and policy tasks. Korea Rural Economic Institute, Naju, Korea (2015)

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