

RESEARCH ARTICLE

First Report of *Gongronella guangdongensis* Isolated from Soil in Korea

Ally Hassan Wajih, Seung-Yeol Lee, Kallol Das, Leonid N. Ten, Hee-Young Jung*

College of Agriculture and Life Sciences, Kyungpook National University, Daegu 41566, Korea

*Corresponding author: heeyoung@knu.ac.kr

ABSTRACT

A fungal isolate designated KNU16-033 was isolated from a soil sample in Daegu, Korea. White, short, and felt-like aerial mycelia appeared on the surface of colonies of the isolate. Colonies with a smooth texture developed slowly and reached a diameter of 78 mm after 21 days of incubation on potato dextrose agar. This isolate displayed globose, colorless or light yellow sporangiospores, which differed morphologically from the sporangiospores of *Gongronella butleri*, which were hyaline, oval to flattened on one side or almost reniform. Based on these morphological characteristics and phylogenetic analysis using internal transcribed spacer regions, the isolated fungus was identified as *G. guangdongensis* belonging to Cunninghamhamellaceae. To our knowledge, this is the first record of *G. guangdongensis* in Korea.

Keywords: Cunninghamhamellaceae, *Gongronella guangdongensis*, Soil-inhabiting fungi

INTRODUCTION

Many of the fungi in the genus *Gongronella* are economically significant representatives of the class Zygomycetes. The cell walls of Zygomycetes are composed of chitosan [1]. Chitosan is also commercially produced from crustacean shells that are waste materials of food industries. The pin mold *Gongronella butleri* (Lendner) Peyronel & Dal Vesco is a well-known and important *Zygomycetes* fungus used in the production of chitosan. *G. butlei* reportedly produces the highest yield of chitosan [2]. *Gongronella* species have been used in biotechnological applications such as the production of antifungal proteins and enzymes [3-5]. Members of the genus *Gongronella* exclusively inhabit the soil [6-8] and grow relatively slowly between 25°C and 27°C [6]. Recent studies have focused on the genus *Gongronella*, and *G. butleri*, *Gongronella orasabula* and *Gongronella koreana* have been newly reported in Korea [9-11]. In this study, a fungal isolate designated KNU16-033, which was obtained from crop field soil in Korea, was identified as a member of the genus *Gongronella*. Based on the morphological and molecular characteristics, the isolate was identified as *Gongronella guangdongensis*. The isolation and identification of this strain, which is reported for the first time in Korea, is described in this report.

OPEN ACCESS

Kor. J. Mycol. 2018 March, 46(1): 28-33
<https://doi.org/10.4489/KJM.20180004>

pISSN : 0253-651X
eISSN : 2383-5249

Received: February 22, 2018

Revised: February 25, 2018

Accepted: February 25, 2018

© The Korean Society of Mycology



This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/4.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

MATERIALS AND METHODS

Sampling and fungal isolation

Fungal isolate KNU16-033 was isolated from a soil sample collected in Daegu, Korea (N 35°53'41.6", E 128°35'10.1"). The soil sample was collected from the ground, air-dried, and stored in a plastic bag at 4°C until analysis. A conventional dilution plating technique was applied to isolate the fungus [12]. One gram of the soil sample was suspended in 10 mL of sterile distilled water. The suspension was vortexed, diluted, and a defined volume was spread on potato dextrose agar (PDA; Difco, Detroit, MI, USA). To examine the growth rate of fungal colonies, petri dishes containing the inoculated PDA were incubated at 25°C for 3 days. Individual colonies that developed on the agar were purified by subculture on fresh PDA and incubation at 25°C until mycelium development. The pure cultures were preserved on PDA slants at 4°C.

Morphological characterization

For morphological analysis, strain KNU16-033 was grown on PDA at 25°C for 21 days. After incubation colony characteristics such as color, size, and shape were recorded. For the micro observation, samples were observed using a model BX-50 light microscope (Olympus, Tokyo, Japan).

DNA extraction, PCR and sequencing analysis

Genomic DNA of strain KNU16-033 was extracted using the HiGene Genomic DNA Prep Kit (BIOFACT, Daejeon, Korea) following the manufacturer's instructions. To amplify the internal transcribed spacer (ITS) regions, ITS 1F/ITS 4 [13] primer pairs, and the product was purified using ExoSAP-IT (Thermo Fisher Scientific, Waltham, MA, USA) and sequenced (SolGent, Daejeon, Korea). The similarities of obtained sequences were analyzed using the BLAST in NCBI and GENETYX-WIN (ver. 3.2) program. The obtained sequences from KNU16-033 was deposited in NCBI GenBank as accession no. LC372499.

Phylogenetic analysis

To perform the phylogenetic analysis, allied *Gongronella* species were retrieved from GenBank. MEGA 6 software [14] was used for the alignment and the phylogenetic tree constructed by the maximum parsimony method with bootstrap analysis of 1,000 replications.

RESULTS AND DISCUSSION

Gongronella guangdongensis F. Liu, T.T. Liu & L. Cai, Cryptogamie Mycologie 36: 137 (2015)

Morphology of isolate KNU16-033

The KNU16-033 had a slow growth rate, formed white colonies and reached a diameter of 78 mm after 21 days of incubation on PDA at 25°C. Other features included erect and branched sporangiophores, globose sporangium, columellae with a globose apophysis, and one-celled sporangiospores. The morphology of the isolate was compared with previous descriptions of *G. guangdongensis* and *G. butleri* [9, 15]. These species differ morphologically based on sporangiospore characteristics. The sporangiospores of *G. guangdongensis* are globose and colorless or light yellow, while those *G. butleri* sporangiospores are hyaline, and oval to flattened on one side or almost reniform [16]. Taxonomic descriptions and microphotographs of the morphological structures of isolate KNU16-033 are presented in Table 1 and Fig. 1. Colonies having a smooth texture with a white, short, and felt-like aerial mycelium developed slowly and attained a diameter of 78 mm after 21 days of incubation on PDA (Fig. 1A, 1B). On malt extract agar (MEA), growth was slow and the colonies that formed were pale yellow to brown with an irregular margin. The colonies typically reached a diameter of 59 mm after 6 days of incubation (Fig. 2C, 2D). Columellae were hemispherical, spherical or ovoid, smooth, often constricted at the attachment to apophyses, and $2.8\text{--}11.7 \times 2.5\text{--}11.9 \mu\text{m}$ in size (Fig. 1E, 1G). The sporangia were pale to pale mouse grey and always globose with many spores. An apophysis was always present and abortive sporangia were sometimes present. The sporangial wall was thin, smooth, and approximately $15\text{--}22.9 \mu\text{m}$ in diameter (Fig. 1H). Sporangiospores were

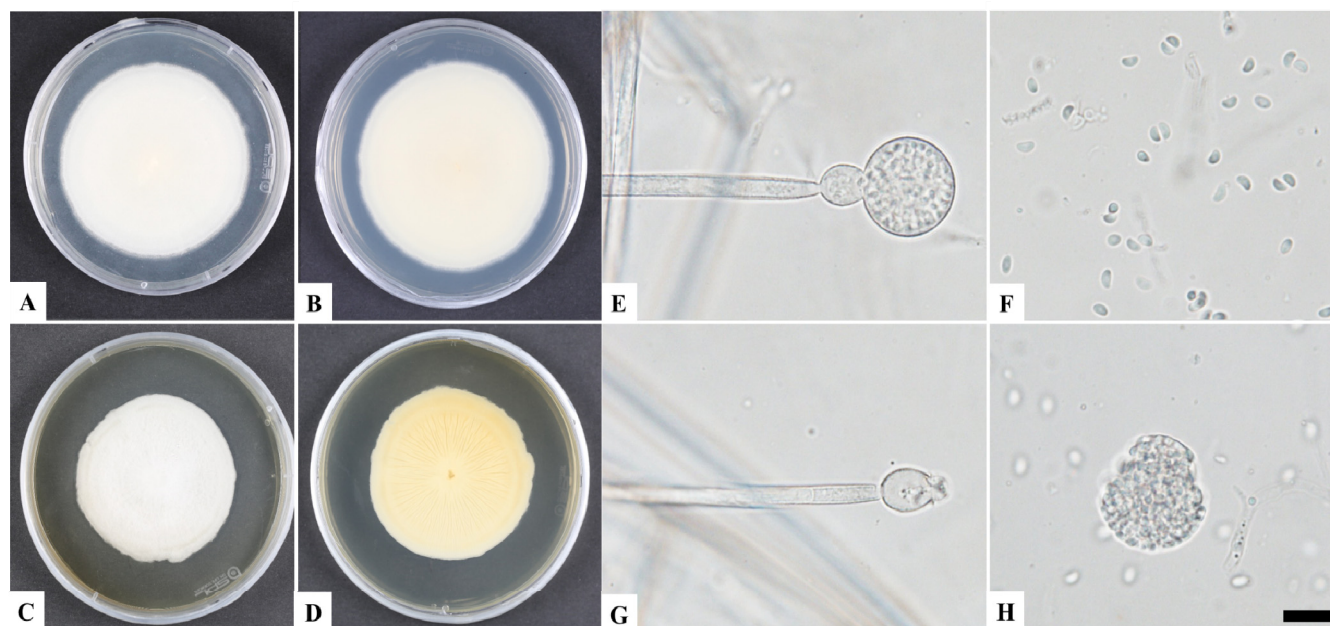


Fig. 1. Cultural and morphological characteristics of *Gongronella guangdongensis* KNU16-033. A, B, colonies on potato dextrose agar; C, D, colonies on malt extract agar; E, columellae and sporangium; F, sporangiospores; G, columellae; H, sporangium (scale bar = 10 μm).

globose, hyaline or light yellow, smooth, and 2.6~3 μm in size (Fig. 1F). Zygospores were not observed. These morphological characteristics of KNU16-033 were similar to those of *G. guangdongensis* and were distinct from those of *G. butleri* (Table 1).

Molecular phylogeny of isolate KNU16-033

After the sequencing analysis, 605 bp was obtained from isolate KNU16-033. As shown in Fig. 2, the KNU16-033 isolate clustered together with the *G. guangdongensis* group with a 99% bootstrap value and was distinct from other *Gongronella* spp. in the phylogenetic tree constructed based on ITS region sequences (Fig. 2). Furthermore, it was revealed that KNU16-033 displayed 99% similarity with *G. guangdongensis* KC462739 that was isolated from soil. In this reason, phylogenetic analysis result support that isolate KNU16-033 was *G. guangdongensis*.

Gongronella species are valuable in the production of chitosan. Chitosan is an important component in the cell walls of Zygomycetes. Some *Gongronella* species produce a large

Table 1. Morphological characteristics of isolate KNU16-033 with reference to *Gongronella guangdongensis* and *Gongronella butleri*

| Characteristics | | KNU16-033 ^a | <i>Gongronella guangdongensis</i> [15] | <i>Gongronella butleri</i> [9] |
|------------------|--------------------|--|---|---|
| Colony | Color | White or pale, colony reverse puff to honey | White or pale, colony reverse puff to honey | White to greyish or smokey brown, pale with brownish zone |
| | Size | 61 mm in diameter after 13 days, and 78 mm in diameter after 21 days of incubation on PDA | 50 mm in diameter after 13 days, and 70 mm after 21 days of incubation on PDA | N/A |
| | Shape | Margin irregular | Margin irregular | N/A |
| | Texture | Smooth | N/A | Woolly to cottony |
| Sporangia | Size (diam.) | 15~22.9 μm | 14~21.5 μm | 7.0~12 \times 5.5~7.7 μm |
| | Shape and position | Pale to pale mouse grey, globose, many spored always with an apophysis, then olivaceous to brown vinaceous with age, always with an apophysis, abortive sporangia sometimes present; sporangial wall thin and smooth | At first pale to pale mouse grey, then olivaceous to brown vinaceous with age, always globose, many spored, always with an apophysis, abortive sporangia sometimes present; sporangial wall thin and smooth | Globose or spherical, about 22.8 μm in diameter, wall smooth and soluble |
| Columellae | Size (diam.) | 2.8~11.7 \times 2.5~11.9 μm | 2.5~12 \times 2~12 μm | N/A |
| | Shape and position | Hemispherical, spherical or ovoid, smooth, often constricted at attachment to apophyses | Hemispherical, spherical or ovoid, smooth, often constricted at attachment to apophyses | N/A |
| Sporangio-spores | Size (diam.) | 2.6~3 μm | 2~3 μm | 11.5~15.5 \times 20.0~24.4 μm |
| | Shape and position | Globose, hyaline or light yellow, smooth | Globose, hyaline or light yellow, smooth | Smooth oval to flattened on one side to reinform |

^aFungal strain studied in this paper.

N/A, not available in previous references.

Sources of descriptions [9, 15].

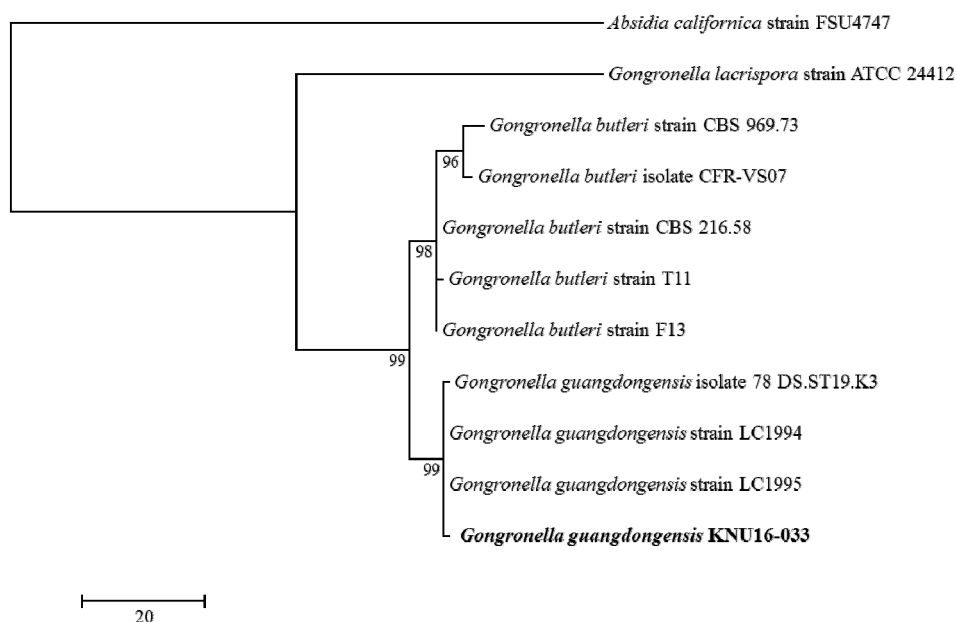


Fig. 2. One of the ten most parsimonious trees generated from maximum parsimony analysis of obtained sequences of the internal transcribed spacer rDNA region. *Absidia californica* strain FSU4747 was an outgroup. The KNU16-033 strain is in bold. The scale bar indicates the number of nucleotide substitutions.

amount of chitosan, which is economically significant in agriculture, medical, biological, and industrial applications [2, 17-19]. The chitosan-producing capability of *G. guangdongensis* will be the subject of future studies. *G. guangdongensis* is uniquely characterized by the presence of globose, hyaline or light yellow sporangiospores. The size and shape of sporangiospores are clearly distinguishing characteristics for other species of *Gongronella*, such as *G. orasabula* and *G. koreana* [10, 11]. To our knowledge, this is the first report of *G. guangdongensis* in Korea.

ACKNOWLEDGEMENTS

This research was supported by a grant from the National Institute of Biological Resources (NIBR), funded by the Ministry of Environment (MOE) of the Republic of Korea for the project on survey and discovery of indigenous fungal species.

REFERENCES

1. Bartniki-Garcia S. Cell wall chemistry, morphogenesis, and taxonomy of fungi. *Annu Rev Microbiol* 1968;22:87-108.
2. Tan SC, Tan TK, Wong SM, Khor E. The chitosan yield of Zygomycetes at their optimum harvesting time. *Carbohydr Polym* 1996;30:239-42.
3. Wang J, Zhou W, Yuan H, Wang Y. Characterization of a novel fungal chitosanase Csn2

- from *Gongronella* sp. JG. Carbohydr Res 2008;343:2583-8.
4. Wei F, Hong Y, Liu J, Yuan J, Fang W, Peng H, Xiao Y. *Gongronella* sp. induces overproduction of laccase in *Panus rudis*. J Basic Microbiol 2010;50:98-103.
 5. Zhou W, Yuan H, Wang J, Yao J. Production, purification and characterization of chitosanase produced by *Gongronella* sp. JG. Lett Appl Microbiol 2008;46:49-54.
 6. Hesseltine CW, Ellis JJ. Notes on Mucorales, especially *Absidia*. Mycologia 1961;53:406-26.
 7. Ho HM, Chen ZC. Morphological study of *Gongronella butleri* (Mucorales) from Taiwan. Taiwaniana 1990;35:259-63.
 8. Upadhyay HP. Soil fungi from north-east and north Brazil-VII: the genus *Gongronella*. Nova Hedwigia 1969;17:65-73.
 9. Babu AG, Kim SW, Adhikari M, Yadav DR, Um YH, Kim C, Lee HB, Lee YS. A new record of *Gongronella butleri* isolated in Korea. Mycobiology 2015;43:166-9.
 10. Li GJ, Hyde KD, Zhao RL, Hongsanan S, Abdel-Aziz FA, Abdel-Wahab MA, Alvarado P, Alves-Silva G, Ammirati JF, Ariyawansa HA, et al. Fungal diversity notes 253-366: taxonomic and phylogenetic contributions to fungal taxa. Fungal Divers 2016;78:1-237.
 11. Ariyawansa HA, Hyde KD, Jayasiri SC, Buyck B, Chethana KW, Dai DQ, Dai YC, Daranagama DA, Jayawardena RS, Lücking R, et al. Fungal diversity notes 111-252: taxonomic and phylogenetic contributions to fungal taxa. Fungal Divers 2015;75:27-274.
 12. Park S, Ten L, Lee SY, Back CG, Lee JJ, Lee HB, Jung HY. New recorded species in three genera of the Sordariomycetes in Korea. Mycobiology 2017;45:64-72.
 13. White TJ, Bruns T, Lee S, Taylor J. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis MA, Gelfand DH, Sninsky JJ, White TJ, editors. PCR protocols: a guide to methods and applications. New York: Academic Press; 1990. p 315-22.
 14. Tamura K, Stecher G, Peterson D, Filipowski A, Kumar S. MEGA 6: Molecular Evolutionary Genetics Analysis version 6.0. Mol Biol Evol 2013;30:2725-9.
 15. Adamčík S, Cai L, Chakraborty D, Chen XH, Cotter HV, Dai DQ, Dai YC, Das K, Deng C, Ghobad-Hejhand M, et al. Fungal biodiversity profiles 1-10. Cryptogam Mycol 2015;36: 121-6.
 16. Hesseltine CW, Ellis JJ. The genus *Absidia*: *Gongronella* and cylindrical-spored species of *Absidia*. Mycologia 1964;56:568-601.
 17. Nge KL, Nwe N, Chandkrachang S, Stevens WF. Chitosan as growth stimulator in orchid tissue culture. Plant Sci 2006;170:1185-90.
 18. Nwe N, Stevens WF. Production of chitin and chitosan and their applications in the medical and biological sector. In: Tamura H, editor. Recent research in biomedical aspects of chitin and chitosan. Thiruvananthapuram: Research Signpost; 2008. p.161-76.
 19. Badawy ME, Rabea EI. A biopolymer chitosan and its derivatives as promising antimicrobial agents against plant pathogens and their applications in crop protection. Int J Carbohydr Chem 2011; Article ID 460381.