



Gut microbiota dysbiosis and its impact on asthma and other lung diseases: potential therapeutic approaches

Young-Chan Kim^{1,*}, Kyoung-Hee Sohn^{2,*}, and Hye-Ryun Kang^{1,3}

¹Division of Allergy and Clinical Immunology, Department of Internal Medicine, Seoul National University Hospital, Seoul; ²Division of Respiratory, Allergy and Critical Care Medicine, Department of Internal Medicine, Kyung Hee University Hospital, Seoul; ³Institute of Allergy and Clinical Immunology, Seoul National University Medical Research Center, Seoul National University College of Medicine, Seoul, Korea

*These authors contributed equally to this manuscript.

The emerging field of gut-lung axis research has revealed a complex interplay between the gut microbiota and respiratory health, particularly in asthma. This review comprehensively explored the intricate relationship between these two systems, focusing on their influence on immune responses, inflammation, and the pathogenesis of respiratory diseases. Recent studies have demonstrated that gut microbiota dysbiosis can contribute to asthma onset and exacerbation, prompting investigations into therapeutic strategies to correct this imbalance. Probiotics and prebiotics, known for their ability to modulate gut microbial compositions, were discussed as potential interventions to restore immune homeostasis. The impact of antibiotics and metabolites, including short-chain fatty acids produced by the gut microbiota, on immune regulation was examined. Fecal microbiota transplantation has shown promise in various diseases, but its role in respiratory disorders is not established. Innovative approaches, including mucus transplants, inhaled probiotics, and microencapsulation strategies, have been proposed as novel therapeutic avenues. Despite challenges, including the sophisticated adaptability of microbial communities and the need for mechanistic clarity, the potential for microbiota-based interventions is considerable. Collaboration between researchers, clinicians, and other experts is essential to unravel the complexities of the gut-lung axis, paving a way for innovative strategies that could transform the management of respiratory diseases.

Keywords: Microbiota; Gastrointestinal microbiome; Lung diseases; Asthma

INTRODUCTION

The human body hosts a diverse microbiota that forms a symbiotic relationship with its host, creating a complex microecosystem. Maintaining a balanced microbiota is crucial as imbalances can lead to aberrant immune responses and inflammation. Specifically, the gut microbiota refers to a diverse community of microorganisms residing in the gastrointestinal tract, including bacteria, fungi, protozoa, archaea, and yeasts [1].

The gut, initially sterile during intrauterine development, undergoes colonization immediately after birth, with signifi-

cant fluctuations in bacterial numbers and species occurring during early life [2]. In adults, the gut contains approximately 10^{14} bacteria, two-thirds of which are specific to an individual. In normal subjects, the gut harbors approximately 500 different species of microorganisms, collectively weighing approximately 1.5 kg [3,4]. Dominant genera of the healthy intestinal microbiome include *Clostridium*, *Faecalibacterium*, *Ruminococcus*, *Roseburia*, *Eubacterium*, *Bifidobacterium*, *Prevotella*, and *Bacteroides* [5].

Given its critical role in maintaining overall health, the gut microbiota has garnered significant attention as a potential therapeutic target. This heightened interest stems from

two main factors. First, technological advancements have enabled comprehensive detection and analysis of intestinal bacterial flora, providing detailed insights into its composition and pathological variations. Second, the composition and diversity of gut microbiota can be influenced by various environmental and host immunological factors [6], making external manipulation relatively feasible through oral intake of probiotics, antibiotics, and other agents.

The significant influence of the gut microbiota extends beyond gastrointestinal health, impacting several aspects of human health, ranging from appetite and energy metabolism to immunity [7]. For instance, the gut microbiota plays a crucial role in fermenting non-digestible substrates, including dietary fibers and endogenous intestinal mucus, which, in turn, support the growth of specific microbes capable of producing short-chain fatty acids (SCFAs). An increasing body of evidence also suggests a close link between the gut microbiota and lung health [8]. An imbalance between the gut and lung microbiomes could potentially contribute to altered immune function and the development of chronic airway diseases. However, whether this imbalance is the cause or effect of the disease remains uncertain.

This review aimed to elucidate the significant connection between gut microbiota dysbiosis and lung diseases, focusing in particular on asthma. Furthermore, we explored potential therapeutic approaches for modulating the gut microbiota to mitigate lung inflammation. Through a comprehensive review of this emerging field, we report promising avenues for managing lung diseases through interventions targeting gut microbiota.

GUT MICROBIOME AND PULMONARY DISORDERS

Recent studies have reported an association between gut microbiota changes and lung inflammation [8]. Distant inflammatory responses resulting from a “leaky gut” and microbiome changes may contribute to chronic airway diseases, and vice versa [9]. However, there is a lack of data regarding how lung dysbiosis affects gut dysbiosis in the bidirectional interaction of the lung-gut axis, particularly compared to the reverse interaction, i.e., the effect of gut dysbiosis on lung dysbiosis. An experiment evaluating the impact of type-2 lung inflammation on the lung and gut microbiomes in mice overexpressing lung-specific interleukin

(IL)-13 demonstrated that microenvironmental changes in the lung with profound type-2 inflammation affect not only the lung but also the gut microbiome [10]. Using animal models, researchers have investigated specific gut microbial compositions and their interactions with the host, contributing to our understanding of the processes underlying gut-lung communication [7]. However, there is a need for further large-scale and longitudinal integrative studies on adult asthma, including all members of the gut microbiome, such as the bacteriome, fungiome, and virome, along with their products.

Asthma

Asthma, a prevalent chronic airway disease affecting over 300 million individuals worldwide, holds significant medical importance [11]. Recent research has provided evidence of the involvement of commensal bacteria in asthma through studies conducted on germ-free mice. These studies revealed that germ-free mice exhibited an exaggerated susceptibility to allergic responses, indicating a pivotal role of commensal gut bacteria in regulating immune responses associated with asthma [12,13]. Over the past 15 years, advancements in high-throughput sequencing have provided valuable insights into the significant impact of the gut microbiome on asthma, particularly during the critical period spanning the first 2 years of life [8]. Disruptions in the normal gut microbiota composition during this critical period, attributed to external factors, such as antibiotic exposure, Cesarean-section delivery, shorter breastfeeding duration, and modern residential environments, have been associated with an increased risk of developing asthma and allergic diseases [14,15]. Compositional alterations in gut microbiota during this period, characterized by reduced abundance of the genera *Akkermansia*, *Bifidobacterium*, and *Faecalibacterium*, are potential predictors of asthma and allergic diseases [16]. A Canadian neonatal longitudinal cohort study identified four taxa (*Faecalibacterium*, *Lachnospira*, *Veillonella*, and *Rothia*, together abbreviated as FLVR) exhibiting a protective effect against asthma during the first 100 days of life [17]. Germ-free mice supplemented with FLVR taxa were also found to be protected from allergic airway inflammation, exhibiting increased fecal concentrations of acetate, a SCFA known to protect against asthma. However, there have been fewer studies investigating the gut-lung axis in adult asthma compared to childhood asthma. A pilot study examining the adult asthma phenotype provided compel-

ling evidence of a robust correlation between forced expiratory volume in one second and phylum-level differences in gut bacterial composition. Notably, alterations in the relative abundances of *Bacteroidetes* and *Firmicutes* phyla, such as a lower *Bacteroidetes* to *Firmicutes* ratio, were observed, suggesting a possible association between gut microbial composition and lung function in asthma patients [18].

Several studies have highlighted the significance of specific gut microbes, particularly beneficial probiotics, including the *Lactobacillus* and *Bifidobacterium* genera, in mitigating allergic asthma, both in animal and human studies [19-21]. Oral exposure of adult mice to house dust has demonstrated that *Lactobacillus* enrichment in the intestinal microbiome, resulting in enhanced airway immune defense against allergens and viral infections, was based on the negative regulation of Th2 inflammation by the abundance of *Lactobacillus johnsonii* [19]. In human infant studies, colonization by *Lactobacilli* and *Bifidobacteria* was found to correlate inversely with the risk of allergy [20]. In a murine model of chronic asthma, oral supplementation with specific microbes triggered an anti-inflammatory response. *Bifidobacterium breve* suppressed airway inflammation by inhibiting both neutrophil and eosinophil lung infiltration [22,23]. *Clostridium* species, dominant in the distal small intestine and colon, have been shown to increase the proportion of regulatory T cells (Tregs) in mice [24-27]. For instance, oral administration of *Clostridium leptum* in adult mice has shown a negative correlation with asthma [28]. Oral feeding with *C. leptum* for 2 weeks increased the percentage and total number of Tregs in the spleens and mediastinal lymph nodes, and enhanced IL-10 and transforming growth factor- β 1 production in the lungs. A recent study demonstrated a negative association between asthma severity and gut *Akkermansia muciniphila* in both human and mouse models [29]. These findings suggest that specific gut microbiota species may regulate asthma by modulating airway inflammation.

A proposed mechanism linking gut microbiota to the reduction in allergic airway diseases involves the secretion and disturbance of metabolites that suppress lung inflammation [30,31]. SCFAs, such as butyrate and propionate, are recognized for their pivotal role in the gut-lung axis [32]. The gut microbiota exerts a unique influence on the host's bile acid metabolism by facilitating processes, such as deconjugation and dihydroxylation, leading to primary and secondary bile acid production. Both primary and secondary bile acids activate the farnesoid X receptor and G protein-coupled bile

acid receptor-1 (GPBAR1, also known as TGR5). Secondary bile acids, in particular, significantly impact bacterial translocation and immune regulation through TGR5 [33]. Consequently, gut microbiome dysbiosis can significantly alter the bile acid profiles, thereby influencing asthma pathogenesis through the regulation of both innate and adaptive immunity, particularly in obesity-induced asthma [34].

In vitro and mouse studies have suggested that bile acids could potentially reduce eosinophilic airway inflammation, possibly by promoting Th1 cytokine generation by dendritic cells via the nuclear farnesoid X receptor of these cells [35]. Additionally, an inhibitory effect on the inflammatory unfolded protein response of bronchial epithelial cells has been suggested [36]. A human study reported a significant increase in the levels of taurocholate, a primary bile acid, in children with asthma when compared to healthy individuals [37]. This finding was also replicated among adult individuals with T2-high asthma, with higher levels observed for both taurocholate and glycodeoxycholate [38]. Another possible mechanism that needs further investigation is the gut secretion of soluble IgA. Secretory IgA (SIgA) serves as the first line of defense in protecting the intestinal epithelium from enteric toxins and pathogenic microorganisms [39]. SIgA is known to be induced by either food antigens or intestinal microbiota [40]. Meanwhile, IgA also regulates the composition and metabolic function of gut microbiota [41]. Based on the presence of virulent strains of *Streptococcus pneumoniae* or *Haemophilus influenzae*, these interactions may control the local and systemic SIgA levels by producing IgA-specific proteases, thereby influencing respiratory tract infections [42]. Reduced SIgA levels have been observed in asthma and chronic obstructive pulmonary disease (COPD), possibly affecting disease progression [43].

Current research efforts are focused on the association between gut microbiota and distinct subsets of asthma endotypes. For instance, symptomatic eosinophilic adult asthma patients have exhibited altered gut microbiome compositions [44]. Two recent studies have highlighted the correlation between a dysbiotic gut microbiome and asthma phenotypes. Eosinophilic asthma patients showed reduced *Clostridia*, *Lachnospiraceae*, and *Oscillospiraceae* populations, along with an increased *Prevotella* population in the gut, suggesting an association with bile acid and lipopolysaccharide metabolism [44]. A follow-up conducted at approximately 37.9 days after stepping up the asthma treatment did not reveal any significant difference in gut

bacteria diversity and composition in asthma patients. In another study involving 125 adult asthma patients, gut microbiome data were subjected to clustering analysis, resulting in the identification of three distinct enterotypes [45]. The genus *Prevotella* was found to predominate in the T2-high asthma cluster, characterized by significantly elevated serum periostin, whereas the genera *Clostridium* and *Romboutsia* were prevalent in the T2-low asthma cluster, characterized by a high proportion of serum interferon (IFN)- γ .

COPD

COPD, a progressive inflammatory lung disease characterized by airflow obstruction, ranks as the third leading cause of mortality [46]. Emerging evidence suggests that the gut-lung axis may play a crucial role in COPD pathogenesis [47].

In a study comparing the gut microbiome between COPD patients and healthy controls [48], 16S rRNA gene sequencing revealed significant differences in microbial composition. COPD patients exhibited an increased abundance of *Streptococcus*, *Rothia*, *Romboutsia*, and *Intestinibacter* genera from the family *Peptostreptococcaceae* and *Escherichia* in their guts. Conversely, *Bacteroides*, *Roseburia*, and *Lachnospira* genera from the family *Lachnospiraceae*, as well as several unnamed *Ruminococcaceae* genera, were decreased in COPD patients. Some specific species, such as *Streptococcus sp000187445*, *Streptococcus vestibularis*, and various members of the family *Lachnospiraceae*, were also found to correlate with reduced lung function.

Since smoking is a major risk factor for COPD development, it is essential to consider its impact on the gut microbiome composition. A cohort study reported gut microbiota changes in male smokers [49], with current smokers showing a higher proportion of *Bacteroidetes* and lower proportions of *Firmicutes* and *Proteobacteria* in their gut microbiota compared to those who had never smoked or had quit smoking. Meanwhile, gut microbiota compositions of individuals who had never smoked and former smokers did not show significant differences. These findings suggest that smoking cessation could potentially facilitate the restoration of gut microbiota to a state resembling that of non-smokers.

Various animal models have been used to investigate the role of the microbiome in COPD [50]. Among these, the cigarette smoking (CS) model uniquely sheds light on the potential influence of the gut microbiome on COPD progression. Interestingly, fecal microbiota transplantation

(FMT) from COPD patients (GOLD stage III–IV), exhibiting significantly reduced SCFA levels, into CS murine models aggravated the lung condition [51]. Remarkably, oral administration of antibiotics has demonstrated the capacity to mitigate CS-induced COPD pathogenesis. This intervention yielded a significant reduction in CS-related COPD progression. Within this context, *Parabacteroides goldsteinii* has emerged as a crucial protective species against COPD [52].

A recent Finnish longitudinal cohort study investigated associations between the occurrence of chronic airway disease and the gut microbiome [53]. The study revealed associations between COPD onset and an increase in *Faecalicatena*, *Oscillibacter*, *Lawsonibacter*, *Flavonifractor*, and *Streptomyces*, along with a decrease in *Lachnospira*, *ER4*, *KLE1615*, *Eubacterium*, and *Coproccoccus*. Notably, the gut microbiome exhibited a higher predictive value for incident COPD compared to asthma (area under the curve: 0.780 and 0.593, respectively). Additionally, studies of fecal samples and monitoring for chronic airway diseases in adults have demonstrated that gut dysbiosis precedes respiratory symptoms, indicating its potential role in COPD etiopathogenesis.

Interstitial lung disease

Interstitial lung disease (ILD) refers to a broad spectrum of airway diseases characterized by lung fibrosis and inflammation [54]. Its cause remains elusive, but recent studies have suggested the potential role of the gut and lung microbiome in fibrotic lung disease. The lung microbiome in ILD differs significantly from that of healthy individuals [55,56]. Idiopathic pulmonary fibrosis (IPF) patients exhibit elevated lung bacterial loads, including the genera of *Staphylococcus*, *Streptococcus*, *Campylobacter*, and *Stenotrophomonas*, which may contribute to the exacerbation of IPF [55,56]. Currently, there are no relevant data on the gut microbiome of ILD patients.

In the bleomycin-induced pulmonary fibrosis model, germ-free mice were protected against lung fibrosis morbidity, with increased gut microbial diversity associated with better fibrosis outcomes [57]. Additionally, the relative abundance of some intestinal probiotics, including *Catenibacterium* and *Lactobacillus* (*L. johnsonii* and *L. gasseri*), were significantly reduced, while the relative abundance of *Verrucomicrobiales* and *Enterobacteriales* were significantly increased in the bleomycin mice model.

TARGETING THE GUT MICROBIOTA

The gut-lung axis has emerged as an important area of research in airway diseases, prompting investigations into various therapeutic approaches to target the gut microbiota and its potential influence on lung disorders. It is no longer sufficient to merely explore associations between the gut and/or lung microbiome and disease outcomes; instead, researchers are now focusing on understanding the complex ecological interactions between the microbiota and the host that influence the immune response.

Recent studies have recognized the potential of correcting gut-lung dysbiosis in treating lung disorders. Reviews of preclinical and clinical studies of various interventions, including probiotics/prebiotics, FMT, antibiotics, and metabolites, have reported promising results in modulating the gut microbiota to improve lung health (Table 1) [21,58-70].

However, the mechanisms by which these potential treatment options overcome adaptation and restore the inherent variability in the gut-lung axis are complex and remain poorly understood. The human microbiome is highly dynamic, with its composition changing in response to various factors, including diet, lifestyle, and environmental exposures. Moreover, the gut-lung axis involves intricate interactions between the gut microbiota, immune system, and lung, making it a challenging area to unravel.

Probiotics and prebiotics

Probiotics (i.e., live bacteria administered orally to facilitate intestinal colonization) and prebiotics (i.e., non-viable substrates selectively utilized by host microorganisms to confer health benefits) have been widely used for decades, particularly in infants, to prevent allergic diseases [71]. Probiotic administration may alter the composition of intestinal

Table 1. Potential therapeutic targets for future asthma treatment

Target	Object	Action	Effect
Probiotics	<i>Lactobacillus</i>	Activation FOXP3 receptor in eosinophilic asthma [58] Stimulate dendritic cell maturation [59] Modulation of membrane proteins and modulate key signaling pathways (NF-κB and MAPK) [60]	Reduce GI pathogenic microbiota Reduce collagen deposition
	<i>Bifidobacterium</i>	Increase IL-10 and Foxp3 transcription in lung tissue [21] Augment Foxp3 in blood CD4 ⁺ T cells Promote Th1 and inhibit Th2 immune responses [61]	Suppress Th2 airway inflammation and airway remodeling
	Broncho-Vaxom [®]	Increase the number of regulatory T cells in the airway [62]	Reduce asthma exacerbation in children
Prebiotics	FOS, GOS	Control PI3K gene expression [63]	Induce tolerance in allegro-inflammatory reaction
SCFA	Butyrate	GPR41&43 receptors activation [64] HDAC inhibition [65] T-helper 9 cell inhibition [66]	Stimulation of neutrophils, dendritic cells and macrophage Eosinophilic apoptosis via HDAC inhibition
Antibiotics	Azithromycin	Alter gut microbial composition (increase the proportion of <i>Clostridium</i>) in OVA mice [67] Decline in the alpha diversity and the beneficial <i>Bifidobacterium</i> species in the gut of child asthmatic patients [68]	Reduce Th2 airway inflammation
Inhaler, FMT	Spray microencapsulated microbiome, bacterial trajectory starting from the microbial chamber	Novel approaches for exploring the gut-lung axis [69,70]	

MAPK, mitogen-activated protein kinase; GI, gastrointestinal; IL, interleukin; FOS, fructo-oligosaccharides; GOS, galacto-oligosaccharides; SCFA, short-chain fatty acid; GPR, G-protein coupled receptor; HDAC, histone deacetylase; OVA, ovalbumin; FMT, fecal material transplantation.

microbiota and improve microbial balance in the gut [72]. Well-known probiotic strains, including genera *Lactobacillus* and *Bifidobacterium* spp., ferment oligosaccharides in the colon, leading to SCFA production. Other potential probiotic species from *Clostridium* clusters 4 and 14, including *C. leptum*, *Ruminococcus bromii*, *Faecalibacterium prausnitzii*, *Coccoides*, *Eubacterium rectale*, *Roseburia* spp., and *Butyrivibrio fibrisolvens*, have also shown beneficial effects [73]. These probiotics increased the production of type-1 cytokines, including tumor necrosis factor (TNF)- α and IFN- γ , and decreased the production of type-2 cytokines, including IL-4, in *in vivo* models [74,75]. Additionally, recent studies have suggested a potential role for extracellular vesicles (EVs) derived from *Lactobacillus* and *Lactococcus*. For instance, intranasal administration of EVs isolated from *Lactococcus lactis* in an experimental model of asthma resulted in reduced airway hyper-responsiveness and type-2 inflammation, indicating that EVs may mediate immunomodulatory effects in asthma [76]. These immunomodulatory effects are particularly relevant in allergic diseases, where an imbalance between type-1 and type-2 immune responses is common. However, it is important to note that probiotics may persist in the adult gut for only a few days, necessitating an individualized approach to enhance intestinal adhesion, particularly in adults [77]. Researchers have explored different strategies to improve probiotic colonization in the gut, to maximize their beneficial effects. A study in mice reported that gut inoculation with *L. johnsonii* significantly reduced Th2 inflammatory responses in the lungs [19].

Prebiotics include non-digestible soluble fibers, such as fructo-oligosaccharides, galacto-oligosaccharides, and polysaccharides [69]. These dietary fermentable fibers can influence allergic lung inflammation in mice by altering the gut microbiota, leading to increased circulating SCFA levels [65]. Currently, the combination of probiotics and prebiotics, termed “synbiotics”, is used to enhance their favorable

effects therapeutically and harness the therapeutic potential of the gut microbiota [78].

Antibiotics

Antibiotics are recognized for their ability to induce compositional changes in the intestinal microbiota. Soluble TNF receptors (TNFR1 and TNFR2) were found to be significantly increased in the sputum of T2-low asthma patients [79]. However, long-term azithromycin treatment significantly reduces sputum TNF and TNFR2 concentrations in non-eosinophilic T2-low asthma patients, compared to healthy controls [80]. Azithromycin is considered a potential treatment for airway inflammatory diseases. A recent study demonstrated that azithromycin modified the gut microbial composition and mitigated allergic airway inflammation induced by FMT in a mouse model [67].

The impact of antibiotic use on childhood asthma is a topic of debate. Studies of children aged 2–7 years have demonstrated that macrolide use leads to increased populations of the phyla *Bacteroidetes* and *Proteobacteria*, while decreasing *Actinobacteria* populations, thereby increasing the risk of asthma and overweight [81]. Therefore, the effects of antibiotic administration on the microbiome, along with the optimal dosage and duration of use, remain controversial, necessitating additional longitudinal studies.

Metabolites

The human gut microbiome has several functions, including catalyzing the metabolism of complex carbohydrates and producing SCFAs [82,83]. Additionally, it contributes to tryptophan metabolism [84] and the production of anti-inflammatory lipids [85], which are crucial for maintaining gastrointestinal health. These metabolic products constitute an essential energy source for gastrointestinal epithelial cells.

Among the significant metabolites produced by the gut microbiota, SCFAs generated from microbial fermentation

Table 2. Types of short-chain fatty acids produced by microbiota

SCFA	Producing bacteria
Acetate	<i>Bacteroides</i> , <i>Bifidobacteria</i> , <i>Clostridia</i> , <i>Desulfovibrio</i> , <i>Eubacteria</i> , <i>Fusobacteria</i> , <i>Peptococci</i> , <i>Peptostreptococci</i> , <i>Propionibacteria</i> , <i>Ruminococci</i> , <i>Veillonella</i>
Butyrate	<i>Clostridia</i> , <i>Eubacteria</i> , <i>Fusobacteria</i> , <i>Ruminococci</i>
Propionate	<i>Bacteroides</i> , <i>Clostridia</i> , <i>Propionibacteria</i> , <i>Veillonella</i>

SCFA, short-chain fatty acid.

Data from the article of Ramakrishna. J Gastroenterol Hepatol 2013;28 Suppl 4:9-17 [73].

of dietary fibers, including acetate, butyrate, and propionate, serve as an energy source for colonocytes [86]. Acetate and propionate are present in both small and large intestines, while butyrate is primarily found in the colon. Changes in SCFA levels can indicate host dysbiosis or bacterial infection, making them valuable indicators of bacterial activity influenced by factors such as dysbiosis, diet, lifestyle, and age [87]. The SCFA type produced depends on the bacteria involved in fermentation (Table 2) [73], and the pathways for production of these substrates differ among species.

Despite evidence from murine studies that oral SCFA administration can alleviate allergic inflammation [32,88], its effect on human allergic diseases remains controversial. The PASTURE study reported that children fed a diet composed of yogurt, fish, vegetables, and fruits exhibited increased fecal butyrate levels, along with a decreased risk of sensitization to food and inhalant allergens [32]. Additionally, females with higher gestational fecal SCFA levels were less likely to have offspring with atopic asthma [89]. Histamine, another important gut metabolite, can regulate NLRP6 inflammasome and intestinal IL-18 secretion, thereby influencing the expression of colonic anti-microbial peptides [90]. In another study, supplementation with *Lactobacillus reuteri*, a bacterium producing histidine decarboxylase, resulted in the suppression of colonic inflammation by con-

verting L-histidine to histamine in the gut [91]. However, recent studies have indicated that SCFAs induce the release of TNF- α and IL-6 in lung mesenchymal cells, fibroblasts, and smooth muscle cells, suggesting proinflammatory effects rather than anti-inflammatory effects. This underscores the need for further human studies [92].

FMT

The clinical efficacy of FMT, a technique involving the transfer of fecal matter from a healthy donor to a recipient, has been demonstrated in randomized controlled trials for various conditions, including *Clostridium difficile* infections [93], inflammatory bowel disease [94], obesity [95], type-1 diabetes mellitus [26], and autism spectrum disorder [96]. While FMT has not been established as a therapeutic option for airway diseases due to its high cost and technical complexity, recent research has highlighted its potential impact on lung health. Studies have reported that FMT with feces rich in *Bacteroides fragilis* from adult asthma patients, when transplanted into gnotobiotic mice, induces a Th17 response in murine airways [97]. In animal models of other respiratory diseases, FMT has been reported to alter the immune cell profile, particularly in cases of broad-spectrum antibiotic treatment or germ-free conditions [98,99]. FMT through selective microbiota transplantation using innovative strate-

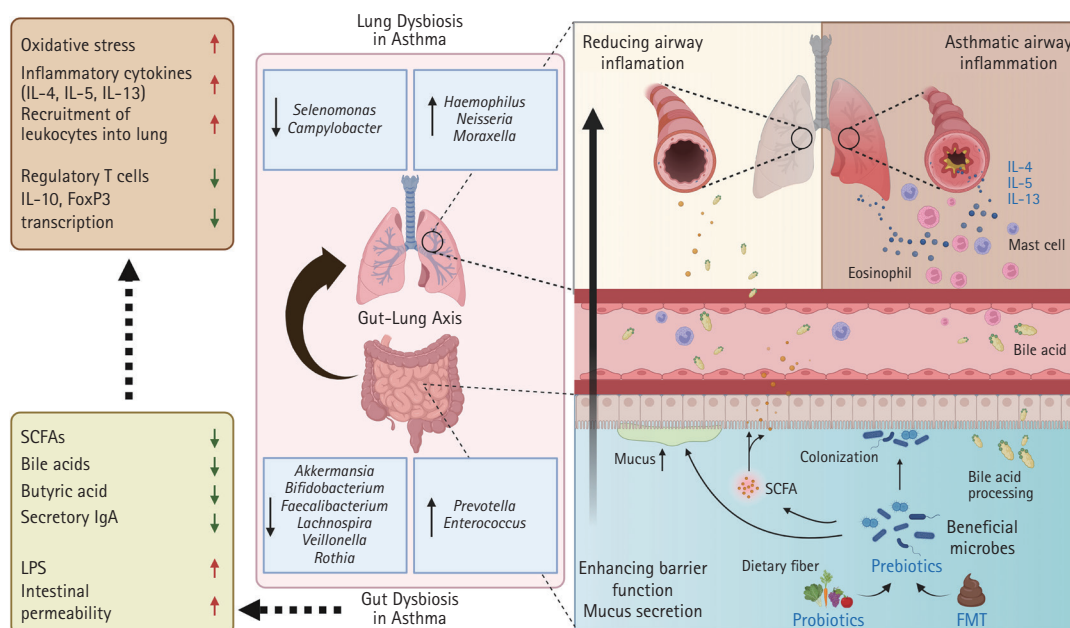


Figure 1. Role of gut microbiome in asthma. IL, interleukin; SCFA, short-chain fatty acid; LPS, lipopolysaccharide; FMT, fecal microbiota transplantation.

gies and ensuring compatibility with the recipient's gut microbiome holds promise for improving the management of chronic airway diseases.

Future therapeutic prospects: inhalational approach

The unique microbiome of lungs has been subjected to extensive scrutiny, revealing its intricate relationships with both pulmonary equilibrium and lung diseases. A previous study analyzing nasal blow samples from asthmatic children suggested that the *Moraxella* cluster poses the highest risk [100]. Regarding phylum-level abundance, dysbiosis characterized by increased *Proteobacteria* or decreased *Bacteroidetes* levels, has been reported in asthmatic airways compared with healthy controls [101]. In a study involving asthmatic patients with varying severities, the genus *Selenomonas* was significantly reduced in asthma, correlating with its severity. Furthermore, intranasal pre-treatment with *Selenomonas* effectively reduced Th2 airway inflammation and airway hyper-responsiveness in an *in vivo* murine asthma model. This suggests the potential for inhalational therapeutic approaches in modulating lung dysbiosis and related immune responses [102].

Within the context of chronic rhinosinusitis, topical probiotic administration as a nasal spray was well-tolerated, although it did not significantly impact symptom severity and the microbiological flora [103]. Another study focusing on topical *Roseomonas mucosa* found a significant reduction in disease severity, the need for topical steroids, and the burden of *Staphylococcus aureus* colonization in pediatric atopic dermatitis patients [104]. Although these studies investigated other allergic diseases, their findings contribute to establishing a potential causal and mechanistic foundation for the encouraging initial results observed with topical microbiome transplantation in the context of allergic airway disease. Theoretically, coating or microencapsulating certain strains and using them in an inhaled form appears to be appropriate in terms of viability and stability. Similarly, the concept of "mucus transplants", analogous to FMT, has been proposed. These emerging concepts highlight the importance of continued research to unravel the complexities of leveraging the gut microbiome as a therapeutic strategy in various respiratory conditions.

PERSPECTIVES

The gut microbiome plays a crucial role in maintaining immune homeostasis throughout the body. Disruptions in its equilibrium can impact lung immune function, potentially leading to respiratory disorders, including asthma. A substantial body of evidence suggests a bidirectional regulatory relationship between intestinal commensals and lung inflammation. Alterations in the intestinal microbial milieu and their metabolites can influence the development and progression of respiratory disorders via immunological pathways. By consolidating recent scientific advancements, this review aimed to clarify the potential link between the gut microbiome and asthma, as well as the gut microbiome's intermediary role in this dynamic, and assess the feasibility of novel therapeutic approaches for modulating gut dysbiosis.

However, understanding the complexity of the microbial landscape is crucial before implementing therapeutic strategies. Studies have reported that approximately two-thirds of the human gut microbiota exhibit individual variations, influenced by factors, such as diet, host genotype, lifestyle, and the use of antibiotics and other medications [105,106]. These variations contribute to the inconsistent gut microbial patterns observed in patients with lung diseases. Additionally, given the heterogeneity of pulmonary diseases, it is essential to consider the interactions of microbiome alterations within these patients [107]. Future research efforts should consider various clinical factors complicating the interpretation of microbial patterns. A comprehensive understanding is essential for developing tailored and effective therapeutic strategies targeting the microbiome. Further mechanistic studies using advanced molecular approaches and longitudinal human studies may enhance our understanding of how changes in gut immune responses induce alterations in respiratory immunity.

REFERNECES

1. Turner PV. The role of the gut microbiota on animal model reproducibility. *Animal Model Exp Med* 2018;1:109-115.
2. Mackie RI, Sghir A, Gaskins HR. Developmental microbial ecology of the neonatal gastrointestinal tract. *Am J Clin Nutr* 1999;69:1035S-1045S.
3. Ley RE, Peterson DA, Gordon JI. Ecological and evolutionary forces shaping microbial diversity in the human intestine.

- Cell 2006;124:837-848.
4. Hooper LV, Littman DR, Macpherson AJ. Interactions between the microbiota and the immune system. *Science* 2012;336:1268-1273.
 5. Thursby E, Juge N. Introduction to the human gut microbiota. *Biochem J* 2017;474:1823-1836.
 6. Lynch SV, Pedersen O. The human intestinal microbiome in health and disease. *N Engl J Med* 2016;375:2369-2379.
 7. Valdes AM, Walter J, Segal E, Spector TD. Role of the gut microbiota in nutrition and health. *BMJ* 2018;361:k2179.
 8. Gensollen T, Iyer SS, Kasper DL, Blumberg RS. How colonization by microbiota in early life shapes the immune system. *Science*. 2016 Apr 29;352(6285):539-44.
 9. Celebi Sozener Z, Ozdel Ozturk B, Cerci P, et al. Epithelial barrier hypothesis: effect of the external exposome on the microbiome and epithelial barriers in allergic disease. *Allergy* 2022;77:1418-1449.
 10. Sohn KH, Baek MG, Choi SM, et al. Alteration of lung and gut microbiota in IL-13-transgenic mice simulating chronic asthma. *J Microbiol Biotechnol* 2020;30:1819-1826.
 11. Brusselle GG, Koppelman GH. Biologic therapies for severe asthma. *N Engl J Med* 2022;386:157-171.
 12. Herbst T, Sichelstiel A, Schär C, et al. Dysregulation of allergic airway inflammation in the absence of microbial colonization. *Am J Respir Crit Care Med* 2011;184:198-205.
 13. Remot A, Descamps D, Noordine ML, et al. Bacteria isolated from lung modulate asthma susceptibility in mice. *ISME J* 2017;11:1061-1074.
 14. van Nimwegen FA, Penders J, Stobberingh EE, et al. Mode and place of delivery, gastrointestinal microbiota, and their influence on asthma and atopy. *J Allergy Clin Immunol* 2011;128:948-955.e1-e3.
 15. Johnson CC, Ownby DR, Alford SH, et al. Antibiotic exposure in early infancy and risk for childhood atopy. *J Allergy Clin Immunol* 2005;115:1218-1224.
 16. Fujimura KE, Sitarik AR, Havstad S, et al. Neonatal gut microbiota associates with childhood multisensitized atopy and T cell differentiation. *Nat Med* 2016;22:1187-1191.
 17. Arrieta MC, Stiemsma LT, Dimitriu PA, et al. Early infancy microbial and metabolic alterations affect risk of childhood asthma. *Sci Transl Med* 2015;7:307ra152.
 18. Begley L, Madapoosi S, Opron K, et al. Gut microbiota relationships to lung function and adult asthma phenotype: a pilot study. *BMJ Open Respir Res* 2018;5:e000324.
 19. Fujimura KE, Demoor T, Rauch M, et al. House dust exposure mediates gut microbiome *Lactobacillus* enrichment and airway immune defense against allergens and virus infection. *Proc Natl Acad Sci U S A* 2014;111:805-810.
 20. Johansson MA, Sjögren YM, Persson JO, Nilsson C, Sverremark-Ekström E. Early colonization with a group of *Lactobacilli* decreases the risk for allergy at five years of age despite allergic heredity. *PLoS One* 2011;6:e23031.
 21. Sagar S, Morgan ME, Chen S, et al. *Bifidobacterium breve* and *Lactobacillus rhamnosus* treatment is as effective as budesonide at reducing inflammation in a murine model for chronic asthma. *Respir Res* 2014;15:46.
 22. Raftis EJ, Delday MI, Cowie P, et al. *Bifidobacterium breve* MRx0004 protects against airway inflammation in a severe asthma model by suppressing both neutrophil and eosinophil lung infiltration. *Sci Rep* 2018;8:12024.
 23. Sagar S, Vos AP, Morgan ME, et al. The combination of *Bifidobacterium breve* with non-digestible oligosaccharides suppresses airway inflammation in a murine model for chronic asthma. *Biochim Biophys Acta* 2014;1842:573-583.
 24. Atarashi K, Tanoue T, Oshima K, et al. T_{reg} induction by a rationally selected mixture of *Clostridia* strains from the human microbiota. *Nature* 2013;500(7461):232-6.
 25. O'Mahony C, Scully P, O'Mahony D, et al. Commensal-induced regulatory T cells mediate protection against pathogen-stimulated NF- κ B activation. *PLoS Pathog* 2008;4:e1000112.
 26. de Groot P, Nikolic T, Pellegrini S, et al. Faecal microbiota transplantation halts progression of human new-onset type 1 diabetes in a randomised controlled trial. *Gut* 2021;70:92-105.
 27. Karimi K, Inman MD, Bienenstock J, Forsythe P. *Lactobacillus reuteri*-induced regulatory T cells protect against an allergic airway response in mice. *Am J Respir Crit Care Med* 2009;179:186-193.
 28. Li YN, Huang F, Liu L, Qiao HM, Li Y, Cheng HJ. Effect of oral feeding with *Clostridium leptum* on regulatory T-cell responses and allergic airway inflammation in mice. *Ann Allergy Asthma Immunol* 2012;109:201-207.
 29. Michalovich D, Rodriguez-Perez N, Smolinska S, et al. Obesity and disease severity magnify disturbed microbiome-immune interactions in asthma patients. *Nat Commun* 2019;10:5711.
 30. Demirci M, Tokman HB, Uysal HK, et al. Reduced *Akkermansia muciniphila* and *Faecalibacterium prausnitzii* levels in the gut microbiota of children with allergic asthma. *Allergol Immunopathol (Madr)* 2019;47:365-371.
 31. Penders J, Thijs C, Vink C, et al. Factors influencing the composition of the intestinal microbiota in early infancy. *Pediatr*

- rics 2006;118:511-521.
32. Roduit C, Frei R, Ferstl R, et al. High levels of butyrate and propionate in early life are associated with protection against atopy. *Allergy* 2019;74:799-809.
 33. Di Vincenzo F, Puca P, Lopetuso LR, et al. Bile acid-related regulation of mucosal inflammation and intestinal motility: from pathogenesis to therapeutic application in IBD and microscopic colitis. *Nutrients* 2022;14:2664.
 34. Joyce SA, Gahan CG. Disease-associated changes in bile acid profiles and links to altered gut microbiota. *Dig Dis* 2017;35:169-177.
 35. Willart MA, van Nimwegen M, Grefhorst A, et al. Ursodeoxycholic acid suppresses eosinophilic airway inflammation by inhibiting the function of dendritic cells through the nuclear farnesoid X receptor. *Allergy* 2012;67:1501-1510.
 36. Nakada EM, Bhakta NR, Korwin-Mihavics BR, et al. Conjugated bile acids attenuate allergen-induced airway inflammation and hyperresponsiveness by inhibiting UPR transducers. *JCI Insight* 2019;4:e98101.
 37. Kelly RS, Sordillo JE, Lasky-Su J, et al. Plasma metabolite profiles in children with current asthma. *Clin Exp Allergy* 2018;48:1297-1304.
 38. Comhair SA, McDunn J, Bennett C, Fettig J, Erzurum SC, Kalhan SC. Metabolomic endotype of asthma. *J Immunol* 2015;195:643-650.
 39. Mantis NJ, Rol N, Corthésy B. Secretory IgA's complex roles in immunity and mucosal homeostasis in the gut. *Mucosal Immunol* 2011;4:603-611.
 40. Hapfelmeier S, Lawson MA, Slack E, et al. Reversible microbial colonization of germ-free mice reveals the dynamics of IgA immune responses. *Science* 2010;328:1705-1709.
 41. Nakajima A, Vogelzang A, Maruya M, et al. IgA regulates the composition and metabolic function of gut microbiota by promoting symbiosis between bacteria. *J Exp Med* 2018;215:2019-2034.
 42. Kilian M, Reinholdt J, Lomholt H, Poulsen K, Frandsen EV. Biological significance of IgA1 proteases in bacterial colonization and pathogenesis: critical evaluation of experimental evidence. *APMIS* 1996;104:321-338.
 43. Wang X, Zhang J, Wu Y, Xu Y, Zheng J. SlgA in various pulmonary diseases. *Eur J Med Res* 2023;28:299.
 44. Gu BH, Choi JP, Park T, et al. Adult asthma with symptomatic eosinophilic inflammation is accompanied by alteration in gut microbiome. *Allergy* 2023;78:1909-1921.
 45. Sohn KH, Choi S, Jung JW, et al. Different inflammatory features of asthma according to gut microbiome enterotype. *Allergy* 2023;78:2997-3001.
 46. Rabe KF, Watz H. Chronic obstructive pulmonary disease. *Lancet* 2017;389:1931-1940.
 47. Quan Y, Yin Z, Chen S, et al. The gut-lung axis: gut microbiota changes associated with pulmonary fibrosis in mouse models induced by bleomycin. *Front Pharmacol* 2022;13:985223.
 48. Bowerman KL, Rehman SF, Vaughan A, et al. Disease-associated gut microbiome and metabolome changes in patients with chronic obstructive pulmonary disease. *Nat Commun* 2020;11:5886.
 49. Lee SH, Yun Y, Kim SJ, et al. Association between cigarette smoking status and composition of gut microbiota: population-based cross-sectional study. *J Clin Med* 2018;7:282.
 50. Tanner L, Single AB. Animal models reflecting chronic obstructive pulmonary disease and related respiratory disorders: translating pre-clinical data into clinical relevance. *J Innate Immun* 2020;12:203-225.
 51. Li N, Dai Z, Wang Z, et al. Gut microbiota dysbiosis contributes to the development of chronic obstructive pulmonary disease. *Respir Res* 2021;22:274.
 52. Lai HC, Lin TL, Chen TW, et al. Gut microbiota modulates COPD pathogenesis: role of anti-inflammatory *Parabacteroides goldsteinii* lipopolysaccharide. *Gut* 2022;71:309-321.
 53. Liu Y, Teo SM, Méric G, et al. The gut microbiome is a significant risk factor for future chronic lung disease. *J Allergy Clin Immunol* 2023;151:943-952.
 54. Molyneaux PL, Cox MJ, Willis-Owen SA, et al. The role of bacteria in the pathogenesis and progression of idiopathic pulmonary fibrosis. *Am J Respir Crit Care Med* 2014;190:906-913.
 55. Molyneaux PL, Cox MJ, Wells AU, et al. Changes in the respiratory microbiome during acute exacerbations of idiopathic pulmonary fibrosis. *Respir Res* 2017;18:29.
 56. Han MK, Zhou Y, Murray S, et al. Lung microbiome and disease progression in idiopathic pulmonary fibrosis: an analysis of the COMET study. *Lancet Respir Med* 2014;2:548-556.
 57. Chioma OS, Mallott EK, Chapman A, et al. Gut microbiota modulates lung fibrosis severity following acute lung injury in mice. *Commun Biol* 2022;5:1401.
 58. Durack J, Kimes NE, Lin DL, et al. Delayed gut microbiota development in high-risk for asthma infants is temporarily modifiable by *Lactobacillus* supplementation. *Nat Commun* 2018;9:707.
 59. Dzidic M, Abrahamsson TR, Artacho A, et al. Aberrant IgA responses to the gut microbiota during infancy precede asthma.

- ma and allergy development. *J Allergy Clin Immunol* 2017; 139:1017-1025.e14.
60. Liu Y, Tran DQ, Rhoads JM. Probiotics in disease prevention and treatment. *J Clin Pharmacol* 2018;58 Suppl 10(Suppl 10):S164-S179.
 61. Wang W, Luo X, Zhang Q, He X, Zhang Z, Wang X. Bifidobacterium infantis relieves allergic asthma in mice by regulating Th1/Th2. *Med Sci Monit* 2020;26:e920583.
 62. Navarro S, Cossalter G, Chiavaroli C, et al. The oral administration of bacterial extracts prevents asthma via the recruitment of regulatory T cells to the airways. *Mucosal Immunol* 2011;4:53-65.
 63. Wu Z, Mehrabi Nasab E, Arora P, Athari SS. Study effect of probiotics and prebiotics on treatment of OVA-LPS-induced of allergic asthma inflammation and pneumonia by regulating the TLR4/NF-kB signaling pathway. *J Transl Med* 2022; 20:130.
 64. Antunes KH, Fachi JL, de Paula R, et al. Microbiota-derived acetate protects against respiratory syncytial virus infection through a GPR43-type 1 interferon response. *Nat Commun* 2019;10:3273.
 65. Trompette A, Gollwitzer ES, Yadava K, et al. Gut microbiota metabolism of dietary fiber influences allergic airway disease and hematopoiesis. *Nat Med* 2014;20:159-166.
 66. Vieira RS, Castoldi A, Basso PJ, Hiyane MI, Câmara NOS, Almeida RR. Butyrate attenuates lung inflammation by negatively modulating Th9 cells. *Front Immunol* 2019;10:67.
 67. Park HK, Choi Y, Lee DH, et al. Altered gut microbiota by azithromycin attenuates airway inflammation in allergic asthma. *J Allergy Clin Immunol* 2020;145:1466-1469.e8.
 68. Wei S, Mortensen MS, Stokholm J, et al. Short- and long-term impacts of azithromycin treatment on the gut microbiota in children: a double-blind, randomized, placebo-controlled trial. *EBioMedicine* 2018;38:265-272.
 69. Trivedi R, Barve K. Gut microbiome a promising target for management of respiratory diseases. *Biochem J* 2020; 477:2679-2696.
 70. Shah P, Fritz JV, Glaab E, et al. A microfluidics-based in vitro model of the gastrointestinal human-microbe interface. *Nat Commun* 2016;7:11535.
 71. Gibson GR, Hutkins R, Sanders ME, et al. Expert consensus document: The International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on the definition and scope of prebiotics. *Nat Rev Gastroenterol Hepatol* 2017;14:491-502.
 72. Williams NT. Probiotics. *Am J Health Syst Pharm* 2010; 67:449-458.
 73. Ramakrishna BS. Role of the gut microbiota in human nutrition and metabolism. *J Gastroenterol Hepatol* 2013;28 Suppl 4:9-17.
 74. Matsusaki T, Takeda S, Takeshita M, et al. Augmentation of T helper type 1 immune response through intestinal immunity in murine cutaneous herpes simplex virus type 1 infection by probiotic *Lactobacillus plantarum* strain 06CC2. *Int Immunopharmacol* 2016;39:320-327.
 75. Won TJ, Kim B, Song DS, et al. Modulation of Th1/Th2 balance by *Lactobacillus* strains isolated from Kimchi via stimulation of macrophage cell line J774A.1 in vitro. *J Food Sci* 2011;76:H55-H61.
 76. Lee DH, Park HK, Lee HR, et al. Immunoregulatory effects of *Lactococcus lactis*-derived extracellular vesicles in allergic asthma. *Clin Transl Allergy* 2022;12:e12138.
 77. Bermudez-Brito M, Plaza-Díaz J, Muñoz-Quezada S, Gómez-Llorente C, Gil A. Probiotic mechanisms of action. *Ann Nutr Metab* 2012;61:160-174.
 78. Fernando WMADB, Flint SH, Ranaweera KKDS, Bamunuarachchi A, Johnson SK, Brennan CS. The potential synergistic behaviour of inter- and intra-genus probiotic combinations in the pattern and rate of short chain fatty acids formation during fibre fermentation. *Int J Food Sci Nutr* 2018;69:144-154.
 79. Niessen NM, Gibson PG, Simpson JL, Scott HA, Baines KJ, Fricker M. Airway monocyte modulation relates to tumour necrosis factor dysregulation in neutrophilic asthma. *ERJ Open Res* 2021;7:00131-2021.
 80. Niessen NM, Gibson PG, Baines KJ, et al. Sputum TNF markers are increased in neutrophilic and severe asthma and are reduced by azithromycin treatment. *Allergy* 2021;76:2090-2101.
 81. Korpela K, Salonen A, Virta LJ, et al. Intestinal microbiome is related to lifetime antibiotic use in Finnish pre-school children. *Nat Commun* 2016;7:10410.
 82. Kelly CJ, Zheng L, Campbell EL, et al. Crosstalk between microbiota-derived short-chain fatty acids and intestinal epithelial HIF augments tissue barrier function. *Cell Host Microbe* 2015;17:662-671.
 83. Furusawa Y, Obata Y, Fukuda S, et al. Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells. *Nature* 2013;504:446-450.
 84. Zelante T, Iannitti RG, Cunha C, et al. Tryptophan catabolites from microbiota engage aryl hydrocarbon receptor and balance mucosal reactivity via interleukin-22. *Immunity* 2013;39:372-385.

85. Viladomiu M, Hontecillas R, Bassaganya-Riera J. Modulation of inflammation and immunity by dietary conjugated linoleic acid. *Eur J Pharmacol* 2016;785:87-95.
86. Sun M, Wu W, Liu Z, Cong Y. Microbiota metabolite short chain fatty acids, GPCR, and inflammatory bowel diseases. *J Gastroenterol* 2017;52:1-8.
87. Wong AC, Levy M. New approaches to microbiome-based therapies. *mSystems* 2019;4:e00122-19.
88. Cait A, Hughes MR, Antignano F, et al. Microbiome-driven allergic lung inflammation is ameliorated by short-chain fatty acids. *Mucosal Immunol* 2018;11:785-795.
89. Lee-Sarwar KA, Kelly RS, Lasky-Su J, et al. Fecal short-chain fatty acids in pregnancy and offspring asthma and allergic outcomes. *J Allergy Clin Immunol Pract* 2020;8:1100-1102.e13.
90. Levy M, Thaiss CA, Zeevi D, et al. Microbiota-modulated metabolites shape the intestinal microenvironment by regulating NLRP6 inflammasome signaling. *cell*. 2015;163:1428-1443.
91. Gao C, Major A, Rendon D, et al. Histamine H2 receptor-mediated suppression of intestinal inflammation by probiotic *Lactobacillus reuteri*. *mBio* 2015;6:e01358-15.
92. Rutting S, Xenaki D, Malouf M, et al. Short-chain fatty acids increase TNF α -induced inflammation in primary human lung mesenchymal cells through the activation of p38 MAPK. *Am J Physiol Lung Cell Mol Physiol* 2019;316:L157-L174.
93. Kelly CR, Khoruts A, Staley C, et al. Effect of fecal microbiota transplantation on recurrence in multiply recurrent *Clostridium difficile* infection: a randomized trial. *Ann Intern Med* 2016;165:609-616.
94. Costello SP, Hughes PA, Waters O, et al. Effect of fecal microbiota transplantation on 8-week remission in patients with ulcerative colitis: a randomized clinical trial. *JAMA* 2019;321:156-164.
95. Leong KSW, Jayasinghe TN, Wilson BC, et al. Effects of fecal microbiome transfer in adolescents with obesity: the gut bugs randomized controlled trial. *JAMA Netw Open* 2020;3:e2030415.
96. Kang DW, Adams JB, Coleman DM, et al. Long-term benefit of Microbiota Transfer Therapy on autism symptoms and gut microbiota. *Sci Rep* 2019;9:5821.
97. Wilson NG, Hernandez-Leyva A, Rosen AL, et al. The gut microbiota of people with asthma influences lung inflammation in gnotobiotic mice. *iScience* 2023;26:105991.
98. Umesaki Y, Okada Y, Matsumoto S, Imaoka A, Setoyama H. Segmented filamentous bacteria are indigenous intestinal bacteria that activate intraepithelial lymphocytes and induce MHC class II molecules and fucosyl asialo GM1 glycolipids on the small intestinal epithelial cells in the ex-germ-free mouse. *Microbiol Immunol* 1995;39:555-562.
99. Ekmekciu I, von Klitzing E, Fiebiger U, et al. Immune responses to broad-spectrum antibiotic treatment and fecal microbiota transplantation in mice. *Front Immunol* 2017;8:397.
100. Zhou Y, Jackson D, Bacharier LB, et al. The upper-airway microbiota and loss of asthma control among asthmatic children. *Nat Commun* 2019;10:5714.
101. Huang YJ. Asthma microbiome studies and the potential for new therapeutic strategies. *Curr Allergy Asthma Rep* 2013;13:453-461.
102. Kim YC, Choi S, Sohn KH, et al. Selenomonas: a marker of asthma severity with the potential therapeutic effect. *Allergy* 2022;77:317-320.
103. Mårtensson A, Abolhalaj M, Lindstedt M, et al. Clinical efficacy of a topical lactic acid bacterial microbiome in chronic rhinosinusitis: a randomized controlled trial. *Laryngoscope Investig Otolaryngol* 2017;2:410-416.
104. Myles IA, Earland NJ, Anderson ED, et al. First-in-human topical microbiome transplantation with *Roseomonas mucosa* for atopic dermatitis. *JCI Insight* 2018;3:e120608.
105. David LA, Maurice CF, Carmody RN, et al. Diet rapidly and reproducibly alters the human gut microbiome. *Nature* 2014;505:559-563.
106. Gupta VK, Paul S, Dutta C. Geography, ethnicity or subsistence-specific variations in human microbiome composition and diversity. *Front Microbiol* 2017;8:1162.
107. Scherzer R, Grayson MH. Heterogeneity and the origins of asthma. *Ann Allergy Asthma Immunol* 2018;121:400-405.

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Correspondence to

Hye-Ryun Kang, M.D., Ph.D.

Division of Allergy and Clinical Immunology, Department of Internal Medicine, Seoul National University Hospital, 101 Daehak-ro, Jongno-gu, Seoul 03080, Korea

Tel: +82-2-2072-0820, Fax: +82-2-742-3291

E-mail: helenmed@snu.ac.kr

<https://orcid.org/0000-0002-2317-4201>

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Credit authorship contributions

Young-Chan Kim: writing - original draft, visualization; Kyoung-Hee Sohn: writing - original draft; Hye-Ryun Kang: conceptualization, writing - review & editing, supervision, project administration, funding acquisition

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