pISSN 1229-1153 / eISSN 2465-9223 J. Food Hyg. Saf. Vol. 39, No. 4, pp. 353~355 (2024) https://doi.org/10.13103/JFHS.2024.39.4.353

Research Note



Whole-Genome Analysis of Salmonella Enterica subsp. Enterica serovar Gallinarum biovar Gallinarum Strain IJES3-1 Isolated from a Retail Chicken Shell Egg in Korea

Beom Soon Jang, Kun Taek Park*

Department of Biological Sciences, Inje University, Gimhae, Korea

(Received August 19, 2024/Revised August 26, 2024/Accepted August 26, 2024)

ABSTRACT - Salmonella enterica subsp. enterica serovar Gallinarum biovar Gallinarum causes fowl typhoid in poultry. In this study, we isolated Salmonella from a Korean retail chicken shell egg and performed whole-genome sequencing, from which we identified one chromosome (4,659,977-bp) and two plasmids (plasmid_1: 87,506 bp and plasmid_2: 2,331 bp). The isolate serotype was confirmed to be Gallinarum, with a biovar type of Gallinarum, which was finally identified as Salmonella enterica subsp. enterica serovar Gallinarum biovar Gallinarum. Multilocus sequence typing confirmed that the isolate was that of sequence type 78. The antimicrobial resistance gene, aac(6')-laa, was identified on the chromosome, and 166 virulence genes were detected on the chromosome and plasmid 1.

Key words: Salmonella enterica subsp. enterica serovar Gallinarum biovar Gallinarum, Whole-genome sequence, Chicken egg

Salmonella is a rod-shaped, gram-negative, facultative anaerobic bacterium belonging to the *Enterobacteriaceae* family¹). It can be observed in various environments and hosts²). *Salmonella enterica* subsp. *enterica* serovar Gallinarum biovar Gallinarum (*S*. Gallinarum) is adapted to captive birds and primarily affects chickens and turkeys²). Fowl typhoid is a septicemic disease of poultry caused by *S*. Gallinarum²). Although fowl typhoid is controlled or eliminated in numerous developed countries, it remains prevalent in developing countries, including South Africa³). Additionally, the disease causes significant economic losses owing to reduced production and high mortality rates in captive poultry⁴).

S. Gallinarum strain IJES3-1 was isolated from a retail chicken shell egg purchased from a domestic supermarket. Whole-genome sequencing (WGS) was performed using Illumina MiSeq and Nanopore sequencing techniques. For MiSeq sequencing, libraries were prepared using the TruSeq Nano DNA Prep kit (Illumina, San Diego, CA, USA), following the manufacturer's protocol. Libraries were sequenced using the NextSeq P1 600 cycles in a NextSeq 2000 system using 2×300 bp pair-ends. Following

*Correspondence to: Kun Taek Park, Department of Biological Sciences, Inje University, Gimhae 50834, Korea

Tel: +82-55-320-3213, Fax: +82-55-336-7706

E-mail: ktpark@inje.ac.kr

Copyright © The Korean Society of Food Hygiene and Safety. All rights reserved. The Journal of Food Hygiene and Safety is an Open-Access journal distributed under the terms of the Creative Commons Attribution Non-Commercial License(http://creativecommons.org/licenses/by-nc/3.0) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

sequencing, the individual sequence reads were analyzed using FastQC-v.0.11.8. For nanopore sequencing, libraries were prepared using a Ligation Sequencing kit (Oxford Nanopore Technologies, Oxford, UK). Nanopore sequencing data were based on the Guppy v4.2.2. Illumina and nanopore sequencing data were processed and de novo assembled using Unicycler v0.5.0. The genome was annotated using the National Center for Biotechnology Information (NCBI) Prokaryotic Genome Annotation Pipeline⁵⁾. The complete genome sequence of IJES3-1 has been deposited in the NCBI GenBank database under the accession numbers CP157387 (chromosome), CP157388 (plasmid_1), and CP157389 (plasmid_2).

The genome of the *S*. Gallinarum strain IJES3-1 consists of a single chromosome (4,659,977 bp, with 52.20 % GC content), plasmid_1 (87,506 bp, with 53.55 % GC content), and plasmid_2 (2,331 bp, with 47.32 % GC content) (Table 1 and Fig. 1).

Multilocus sequence typing (MLST) analysis using MLST 2.0 (version 2.0.9) confirmed this strain as sequence type (ST) 78⁶). This bacterium was identified as *Salmonella enterica* subsp. *enterica* serovar Gallinarum using SeqSero 1.2 and Kmerfinder 3.2^{7.8}). The biovar type was confirmed as *Salmonella enterica* subsp. *enterica* serovar Gallinarum biovar Gallinarum using in silico polymerase chain reaction (PCR) on Unipro UGENE 50.0, using the primer sequences listed in Table 2⁹). AMRFinderPlus and ResFinder were used to identify antimicrobial resistance (AMR) genes^{10,11}). No antimicrobial

354 Beom Soon Jang and Kun Taek Park

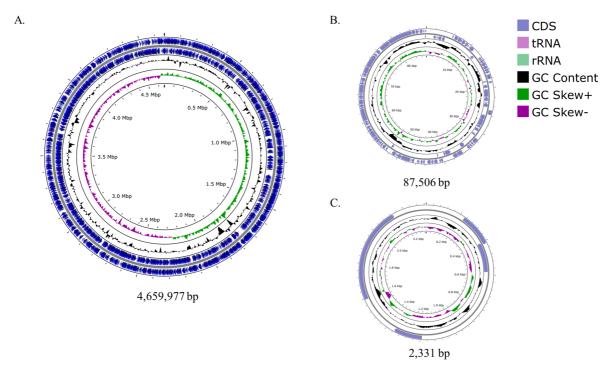


Fig. 1. Complete genome map of *S. gallinarum* strain IJES3-1. (A) Chromosome, (B) plasmid_1, and (C) plasmid_2 sequences are visualized using the Proksee (https://proksee.ca). The outer ring data represents: CDS forward strands, CDS reverse strands, GC content, and GC skew. tRNA (light pink) and rRNA (light green) are located in the same ring as that of the CDS.

Table 1. Genetic characteristics of S. Ga	Illinarum strain IJES3-1
---	--------------------------

Features –	Characteristics			
	Chromosome	Plasmid_1	Plasmid_2	
Genome size	4,659,977 bp	87,506 bp	2,331 bp	
GC content	52.20%	53.55%	47.32%	
No. of CDS	4,400	109	4	
No. of rRNA	22	-	-	
No. of tRNA	77	-	-	
MLST	ST78	-	-	
AMR gene	aac(6')-laa	$N.D^{1)}$	$N.D^{1)}$	
No. of virulence gene	160	6	0	

¹⁾ N.D.: not detected.

Table 2. Confirmation of biovar type using in scilco PCR

Gene		Primer sequence $(5' \rightarrow 3')$	Size	Salmonella		IJES3-1 location	Reference
		Finnel sequence $(5 \rightarrow 5)$	(bp)	$\mathbf{SP}^{1)}$	$SG^{2)}$	(Amplicon size)	Reference
atra	Forward:	TATTTTGCACCACAGCCAGC	131	31 + + 4,476,169-4,476,299 (131bp)	+ +		
stn	Reverse:	CGACCGCGTTATCATCACTG				(131bp)	14)
1137_08605	Forward:	CACTGGAGACTCTGAGGACA	290	+		2,773,759-2,774,048 (290bp)	
	Reverse:	GGGCAGGGAGTCTTGAGATT		Ŧ	Ŧ		
ratA	Forward:	ATTGCTCTCGTCCTGGGTAC	571			1,278,381-1,278,951 (571bp)	
	Reverse:	TACCGATACGCCCAACTACC		/1 -	+		

¹⁾ SP: Salmonella enterica subsp. enterica serovar Gallinarum biovars Pullorum.

²⁾ SG: *Salmonella enterica* subsp. *enterica* serovar Gallinarum biovars Gallinarum.

resistance genes were identified in AMRFinderPlus, but *aac(6')-laa* gene responsible for amikacin, kanamycin, and tobramycin resistance was identified in ResFinder (Table 1)¹²⁾. However, this gene appears to have no clinical significance or evolutionary advantage¹²⁾. Virulence genes were identified using the Virulence Factor Database (VFDB), and 166 genes (160 on the chromosome and 6 on the plasmid_1) were identified (Table 1)¹³⁾.

Acknowledgements

This study was supported by a grant from the Ministry of Food and Drug Safety (22192MFDS021) of the Republic of Korea.

Conflict of interests

The authors declare no potential conflict of interest.

ORCID

Beom Soon Jang https://orcid.org/0000-0003-1657-3105 Kun Taek Park https://orcid.org/0000-0001-6177-0373

References

- Farhat, M., Khayi, S., Berrada, J., Mouahid, M., Ameur, N., El-Adawy, H., Fellahi, S., *Salmonella enterica* serovar Gallinarum biovars Pullorum and Gallinarum in poultry: review of pathogenesis, antibiotic resistance, diagnosis and control in the genomic era. *Antibiotics*, **13**, 23 (2023).
- Lozica, L., Faraguna, S., Artuković, B., Gottstein, Ž., Fowl typhoid outbreak on a commercial turkey farm in Croatia. *Microorganisms*, **12**, 165 (2024).
- Zhou, X., Kang, X., Zhou, K., Yue, M., A global dataset for prevalence of *Salmonella* Gallinarum between 1945 and 2021. *Sci. Data*, 9, 495 (2022).
- Beylefeld, A., Abolnik, C., *Salmonella* Gallinarum strains from outbreaks of fowl typhoid fever in Southern Africa closely related to SG9R vaccines. *Front. Vet. Sci.*, 10, 1191497 (2023).
- Tatusova, T., DiCuccio, M., Badretdin, A., Chetvernin, V., Nawrocki, E.P., Zaslavsky, L., Lomsadze, A., Pruitt, K.D., Borodovsky, M., Ostell, J., NCBI prokaryotic genome annotation pipeline. *Nucleic Acids Res.*, 44, 6614-6624 (2016).
- 6. Larsen, M.V., Cosentino, S., Rasmussen, S., Friis, C., Has-

man, H., Marvig, R.L., Jelsbak, L., Sicheritz-Pontén, T., Ussery, D.W., Aarestrup, F.M., Lund, O., Multilocus sequence typing of total-genome-sequenced bacteria. *J. Clin. Microbiol.*, **50**, 1355-1361 (2012).

- Hasman, H., Saputra, D., Sicheritz-Ponten, T., Lund, O., Svendsen, C.A., Frimodt-Møller, N., Aarestrup, F.M., Rapid whole-genome sequencing for detection and characterization of microorganisms directly from clinical samples. *J. Clin. Microbiol.*, **52**, 139-146 (2014).
- Zhang, S., Yin, Y., Jones, M.B., Zhang, Z., Deatherage Kaiser, B.L., Dinsmore, B.A., Fitzgerald, C., Fields, P.I., Deng, X., *Salmonella* serotype determination utilizing high-throughput genome sequencing data. *J. Clin. Microbiol.*, 53, 1685-1692 (2015).
- Okonechnikov, K., Golosova, O., Fursov, M., The UGENE team, Unipro UGENE: a unified bioinformatics toolkit. *Bio-informatics*, 28, 1166-1167 (2012).
- Bortolaia, V., Kaas, R.S., Ruppe, E., Roberts, M.C., Schwarz, S., Cattoir, V., Philippon, A., Allesoe, R.L., Rebelo, A.R., Florensa, A.F., Fagelhauer, L., Chakraborty, T., Neumann, B., Werner, G., Bender, J.K., Stingl, K., Nguyen, M., Coppens, J., Xavier, B.B., Malhotra-Kumar, S., Westh, H., Pinholt, M., Anjum, M.F., Duggett, N.A., Kempf, I., Nykäsenoja, S., Olkkola, S., Wieczorek, K., Amaro, A., Clemente, L., Mossong, J., Losch, S., Ragimbeau, C., Lund, O., Aarestrup, F.M., ResFinder 4.0 for predictions of phenotypes from genotypes. *J. Antimicrob. Chemother.*, **75**, 3491-3500 (2020).
- Feldgarden, M., Brover, V., Gonzalez-Escalona, N., Frye, J.G., Haendiges, J., Haft, D.H., Hoffmann, M., Pettengill, J.B., Prasad, A.B., Tillman, G.E., Tyson, G.H., Klimke, W., AMRFinderPlus and the reference gene catalog facilitate examination of the genomic links among antimicrobial resistance, stress response, and virulence. *Sci. Rep.*, **11**, 12728 (2021).
- Srednik, M.E., Morningstar-Shaw, B.R., Hicks, J.A., Mackie, T.A., Schlater, L.K., Antimicrobial resistance and genomic characterization of *Salmonella enterica* serovar Senftenberg isolates in production animals from the United States. *Front. Microbiol.*, **13**, 979790 (2022).
- Liu, B., Zheng, D., Zhou, S., Chen, L., Yang, J., VFDB 2022: a general classification scheme for bacterial virulence factors. *Nucleic Acids Res.*, 50, D912-D917 (2022).
- Xiong, D., Song, L., Pan, Z., Jiao, X., Identification and discrimination of *Salmonella enterica* serovar gallinarum biovars pullorum and gallinarum based on a one-step multiplex PCR assay. *Front. Microbiol.*, 9, 1718 (2018).