RESEARCH ARTICLE

First Report of *Phaeosphaeria chengduensis* Isolated from *Gametis jucunda* in Korea

Soo-Min Hong¹, Seong-Keun Lim¹, Young-Kun Kim¹, Sang Jae Suh^{1,2}, Leonid N. Ten², Seung-Yeol Lee^{1,2}, and Hee-Young Jung^{1,2,*}

¹Department of Plant Medicine, Kyungpook National University, Daegu 41566, Korea ²Institute of Plant Medicine, Kyungpook National University, Daegu 41566, Korea

*Corresponding author: heeyoung@knu.ac.kr

ABSTRACT

A fungal strain, KNUF-4H-A belonging to the genus *Phaeosphaeria* was isolated from the citrus flower chafer (*Gametis jucunda*) in Chungcheongbuk-do, Korea. This strain was further identified as *Phaeosphaeria chengduensis* through phylogenetic analyses based on a concatenated dataset of DNA sequences of internal transcribed spacer (ITS) regions, as well as the small subunit rDNA (SSU), large subunit rDNA (LSU), RNA polymerase II second largest subunit (*RPB2*), and translation elongation factor $1-\alpha$ (*TEF1-* α) genes. The isolate KNUF-4H-A exhibited typical cultural characteristics of *P. chengduensis*, producing colonies that were flattened, greenish-grey at the edges, grey-white at the center, and dark brown on the reverse side. *Phaeosphaeria* species have been reported in various ecosystems, including terrestrial and freshwater environments. The isolation of KNUF-4H-A from the citrus flower chafer provides valuable insights into the habitat diversity of *Phaeosphaeria*. This is the first record of *Phaeosphaeria chengduensis* in Korea.

Keywords: Fungi, Gametis jucunda, Insects, Phaeosphaeria chengduensis, Phylogeny

INTRODUCTION

The family Phaeosphaeriaceae was first established by Barr (1979) with the designation of *Phaeosphaeria* I. Miyake as the generic type of the family [1]. According to the Catalogue of Life (http://www.catalogueoflife. org; COL version 2024.02.24), this family currently comprises more than 170 genera. Notably, these fungi are both ecologically and economically significant due to their ability to adaptively shift between endophytic, pathogenic, or saprobic life strategies depending on environmental conditions [2-7]. When functioning as saprobes, the members of the family Phaeosphaeriaceae facilitate the decomposition of dead plant material. However, this family also harbors several important plant pathogens, such as *Alternaria*, *Bipolaris, Didymella, Leptosphaeria, Neosetophoma, Parastagonospora, Phaeosphaeria, Pyrenophora*, and *Stemphylium* species, which are responsible for severe diseases in economically significant crops [8-12]. The genus *Phaeosphaeria*, belonging to this family, comprises over 200 species listed in the Mycobank (https://www.mycobank.org) and Index Fungorum (http://www.indexfungorum.org) databases. Numerous



OPEN ACCESS

pISSN: 0253-651X eISSN: 2383-5249

Kor. J. Mycol. 2024 June, 52(2): 125-134 https://doi.org/10.4489/kjm.520205

Received: April 22, 2024 Revised: June 12, 2024 Accepted: June 14, 2024

© 2024 THE KOREAN SOCIETY OF MYCOLOGY.



This is an Open Access article distributed

under the terms of the Creative Commons Attribution Non-Commercial License (http: //creativecommons.org/licenses/by-nc/4.0/) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited. species of *Phaeosphaeria* have been recently described, including *P. blodgettiae* Y.P. Tan, Bishop-Hurley, Marney & R.G. Shivas, *P. chengduensis* Wanas. & Maharachch., *P. scalesiae* Crous, *P. sichuanensis* Wanas. & Maharachch., and *P. stonesiae* Y.P. Tan, Sbaraini & E. Lacey, contributing to our knowledge of the diversity of this fungal genus [13-16]. Some species within the genus *Phaeosphaeria* play a crucial role as saprophytes, actively participating in processes such as decomposition and nutrient cycling. They are often associated with decaying plant material, deadwood, or organic debris from monocotyledons. Specifically, the aforementioned *P. chengduensis* is saprobic and was isolated from dead twigs of an unknown deciduous host. However, certain species of *Phaeosphaeria* are recognized as plant pathogens, causing several diseases in various hosts, including crops and forest trees [16].

Due to the remarkable diversity of microbial communities associated with insects, interactions with microbiome members can result in a wide range of effects on the fitness and behavior of insects [17]. Fungi and insects interact reciprocally through a wide array of symbiotic relationships, ranging from instances of parasitism, whereby the fungi gain an advantage by harming the insects, to those where fungi form mutualistic associations with their host [18]. The citrus flower chafer (*Gametis jucunda* Faldermann) is distributed across various regions such as India, Nepal, Tonkin, Taiwan, China, the former Soviet Union, Korea, Japan, and North America [19]. These insects are commonly found on flowers and tree saps across a variety of plant species [20]. Known for their harmful effects, these insects negatively impact citrus fruits, flowers, calyxes, and various other plants [21]. Adults have been observed feeding on Chinese privet (*Ligustrum sinense* Lour.), Wampee [*Clausena lansium* (Lour.) Skeels], and Orange climber [*Toddalia asiatica* (L.) Lam.] flowers in Macau. Additionally, in Hong Kong, they have been found on Gray nicker (*Guilandina bonduc* L.), True rhus (*Rhus chinensis* Mill), and Chinese guger tree (*Schima superba* Gard et Champ var. superba) [19]. Therefore, similar to other insects, *Gametis jucunda* could act as a potential vector for the transmission of microfungi. Nonetheless, conclusive evidence regarding the relationship between *Gametis jucunda* and Phaeosphaeriaceae fungi is currently lacking.

This study focused on investigating less-explored sources with the expectation of discovering previously unreported and novel fungal species. Here, we describe the identification of an insect-associated fungal strain belonging to the genus *Phaeosphaeria*, isolated from the citrus flower chafer (*Gametis jucunda*), using cultural, morphological, and molecular phylogenetic approaches.

MATERIALS AND METHODS

Collection and isolation of the fungal strain

The fungal strain utilized in this study was isolated from specimens of *Gametis jucunda* collected from Chopyeong-myeon, Jincheon-gun, Chungcheongbuk-do (36°49'51.7"N 127°34'08.1"E), South Korea. Following a previously described method, the fungi were isolated using potato dextrose agar (PDA, Difco, Detroit, MI), and incubated at 25°C for 2-3 days [22]. Single colonies were individually transferred onto PDA plates and then incubated at 25°C for 4-5 days. After being transferred onto new PDA plates and

forming colonies, the grey-whitish mycelium was subsequently selected for comprehensive analysis and designated as KNUF-4H-A. The fungal isolate KNUF-4H-A was stored at -80°C in 20% glycerol stock. The isolate, designated as KNUF-4H-A (NIBRFGC000512438), was deposited in the National Institute of Biological Resources (NIBR) as a stock culture.

Morphological characterization

To examine its cultural and morphological characteristics, the isolated fungal strain KNUF-4H-A was cultured on PDA. The characteristics of the colonies, including their color, texture, growth rates, shape, and size, were thoroughly examined after 4 weeks [16]. Additionally, morphological characteristics were observed using a BX-50 microscope (Olympus, Tokyo, Japan).

Molecular analysis

Total genomic DNA was extracted using a commercial extraction kit (HiGene Genomic DNA Prep Kit, Biofact, Daejeon, Korea) according to the manufacturer's instructions. Polymerase chain reaction (PCR) was then performed to amplify the internal transcribed spacer (ITS) regions, using the genomic DNA as the template. The obtained amplicons were then purified using the ExoSAP-IT PCR product cleanup reagent (Thermo Fisher Scientific, Waltham, MA, USA). Recent research has successfully classified 20 species of Phaeosphaeria using the small subunit rDNA (SSU), large subunit rDNA (LSU), translation elongation factor 1- α (*TEF1-\alpha*), and RNA polymerase II second largest subunit (*RPB2*), identifying new species such as P. chengduensis and P. sichuanensis [16]. Five loci were amplified, including the ITS, SSU, LSU, TEF1a, and RPB2 genes using the ITS1F/ITS4 [23,24], NS1/NS4 [24], LROR/LR5 [25], EF1/EF2 [26], and 5F2/7CR [27] primer pairs, respectively. The gene sequences were assembled using SeqMan software (DNASTAR, Madison, WI, USA). Next, gaps and terminal ends in the alignment were edited using BioEdit version 5.0.6 to ensure the accuracy and completeness of the gene sequences. A phylogenetic tree was constructed based on the concatenated sequences of the SSU, LSU, ITS regions, $TEF1-\alpha$, and RPB2 genes via the maximum likelihood method with 1,000 bootstrap replicates using the MEGA 7 software [28]. Genetic divergence between the species was evaluated using Kimura's two-parameter model [29]. To assess evolutionary connections and genetic relatedness among Phaerosphaeria species, sequence alignments were conducted using the National Center for Biotechnology Information (NCBI) Basic Local Alignment Search Tool (BLAST). Upon conducting these BLAST searches, the closest phylogenetic sequences were identified and pairwise sequence similarity values were calculated for each gene, thereby aiding in the determination of genetic connections between Phaerosphaeria species. The gene sequences of closely related phylogenetic relatives were obtained from the NCBI GenBank database (Table 1).

Species	Strain	GenBank accession numbers				
	-	ITS	LSU	SSU	TEF1-α	RPB2
Phaeosphaeria acaciae	KUMCC 20-0214	MW078431	MW078444	MW078482	MW082602	MW192765
Phaeosphaeria ampeli	MFLUCC 18-1641 ^T	NR_165910	MK503808	MK503814	MK503802	-
Phaeosphaeria ampeli	MFLUCC 19-0150	MK503798	MK503809	MK503815	MK503803	-
Phaeosphaeria chengduensis	KUNCC 23-13570	OR206391	OR206410	OR206400	OR195707	OR195716
Phaeosphaeria chengduensis	KUNCC 23-13571 ^T	OR206392	OR206411	OR206401	OR195708	OR195717
Phaeosphaeria chengduensis	KNUF-4H-A	LC802119	LC802127	LC802128	LC802130	LC802129
Phaeosphaeria chiangraina	MFLUCC 13-0231	KM434270	KM434280	KM434289	KM434298	KM434307
Phaeosphaeria chinensis	MFLUCC 19-0217 ^T	MN173212	MN173208	MN173216	-	-
Phaeosphaeria chinensis	KUMCC 19-0161	MN173213	MN173210	MN173217	-	-
Phaerosphaeria oryzae	CBS 110110	MH862850	MH874442	ON408355	ON419509	ON419520
Phaeosphaeria poagena	KUNCC 23-13572	OR206393	OR206412	OR206402	OR195709	OR195718
Phaeosphaeria poagena	KUNCC 23-13573	OR206394	OR206413	OR206403	OR195710	OR195719
Phaeosphaeria sichuanensis	KUNCC 23-13568	OR206389	OR206408	OR206398	OR195705	OR195714
Phaeosphaeria sichuanensis	KUNCC 23-13569 ^T	OR206390	OR206409	OR206399	OR195706	OR195715
Phaeosphaeria thysanolaenicola	MFLUCC 10-0563 ^T	NR_155642	NG_069236	KM434286	KM434295	KM434303
Ophiosphaerella taiwanensis	SICAUCC 23-0003	OR134753	OR134744	OR162596	OR134875	OR424348
Ophiosphaerella taiwanensis	NCYUCC 19-0152 ^T	NR_171874	MT321815	MT321808	-	-

Table 1. List of Phaeosphaeria strains used in the phylogenetic analyses and their GenBank accession numbers

The isolated strain is indicated in bold.

ITS: Internal transcribed spacer regions; LSU: Large subunit rDNA; SSU: Small subunit rDNA; $TEF1-\alpha$: Translation elongation factor 1- α ; RPB2: RNA polymerase II second largest subunit.

RESULTS AND DISCUSSION

Morphological characteristics of the KNUF-4H-A fungal strain

Strain KNUF-4H-A formed circular, convex, flattened, velvety, medium-dense colonies with a greenishgrey edge and a grey-whitish center. The reverse side of the colonies appeared yellowish-brown at the margin and dark brown at the center. Typically, the colonies reached a diameter of 64.4-65.1 mm after 4 weeks of growth at 25°C on PDA medium (Fig. 1A and B). Pycnidial conidiomata were black, globose, and erumpent on the surface of the PDA media after 2 to 4 weeks (Fig. 1C). The conidiophores were borne from the inner hyphal tissue of pycnidia and produced conidia. Conidiophores were reduced to conidiogenous cells, which were hyaline, smooth, and straight to slightly curved (Fig. 1D and E). Strain KNUF-4H-A reproduced asexually through the production of conidia in conidiophores. The chlamydospores appeared hyaline, primarily arranged in chains, smooth, thick-walled, and had a globose to subglobose structure, with a diameter ranging from 4.58 to 13.92 μ m (M=6.47×9.21 μ m, n=25) (Fig. 1F and G). The asexual morphological characteristics of KNUF-4H-A included the formation of yellowish to olive, cylindrical conidia that were either non-septate or one-septate (Fig. 1H). The size of conidia was 3.2-8.1×2.1-3.2 μ m (M=6.2×2.5 μ m, n=50). The cultural characteristics of the isolate on PDA were identical to those of *P. chengduensis* KUNCC 23-13571^T, with colonies appearing flattened, greenish-grey at the edges, greywhite at the center, and dark brown on the reverse side (Table 2). The asexual morphological characteristics of this species have not been reported. *P. poagena* Crous & Quaedvl. was identified as the closest neighbor of *P. chengduensis* according to our phylogenetic analysis and data. Therefore, *P. poagena* was selected as the reference species to conduct a comparative analysis of asexual morphology characteristics [16]. In contrast to KNUF-4H-A, the conidia of *P. poagena* CBS 136771^{T} are solitary, brown, ellipsoidal to subcylindrical with 1-3 septa, slightly constricted at the septa, exhibiting a subobtuse apex, a truncate base, and with a notably longer length on average (Table 2). The differences in color, shape, and size of the conidia distinguish KNUF-4H-A from *P. poagena*, thus providing indirect evidence of its affiliation with *P. chengduensis*.



Fig. 1. Cultural and morphological characteristics of Phaerosphaeria chengduensis KNUF-4H-A. A, B: Front and reverse view of colony grown on potato dextrose agar (PDA) at 25°C for 4 weeks. C: Pycnidia on PDA. D, E: Conidiophores. F, G: Chlamydospores. H: Non-septate and single-septum conidia. Scale bars: C=200 μm, D-H=10 μm.

Characteristics	P. chengduensis	P. chengduensis	P. poagena
	KNUF-4H-A ^a	KUNCC 23-13571 ^{Tb}	CBS 136771 ^{ть}
Cultural	64.4-65.1 mm after 4 weeks, circular,	30 mm after 4 weeks at 25°C, irregular,	40 mm after 4 weeks, irregular, flattened to
characteristics	convex, flattened, velvety, medium dense,	flattened to slightly raised, greenish-grey	slightly raised, various color sectors ranging
	greenish-grey at the edge, grey-whitish at	edge and a grey center, reverse side: dark	from white to creamy orange; reverse side:
	the center; reverse side: yellowish-brown	brown on PDA at 25°C	creamy orange, with occasional dark patches
	at the margin, dark brown at the center, no pigmentation on PDA at 25°C		on PDA at 25°C
Pycnidia	Black, globose, erumpent on the surface of	Undetermined	Black, globose, erumpent, possess a central
	the PDA media after 2 to 4 weeks		ostiole
Chlamydospores	Hyaline, primarily arranged in chains, smooth, thick-walled, globose to subglobose structure, with a diameter ranging from 4.58 to 13.92 μ m (M=6.47×9.21 μ m, n=25)	Undetermined	Undetermined
Conidiophore	Hyaline, smooth, straight to slightly curved	Undetermined	Hyaline, smooth, doliiform
Conidia	Yellowish to olive, cylindrical, non-septate or one-septate, 3.2-8.1×2.1-3.2 μ m (n=50, x=6.2×2.5 μ m)	Undetermined	Solitary, brown, smooth, fusoid, ellipsoidal to subcylindrical, $(1-)$ 3 septate, slightly constricted at the septa, subobtuse apex and a truncate base, $(8-)$ 12-14 $(-16)\times(2.5-)$ 3 (-3.5) um

Table 2. Morphological characteristics of KNUF-4H-A and the reference species Phaeosphaeria chengduensis and Phaeosphaeria poagena

PDA: Potato dextrose agar.

^aFungal strain studied in this research

^bSources of description [16].

Molecular analysis of the fungal strain KNUF-4H-A

The obtained sequence of the ITS regions was 558 bp, exhibiting 100% and 97.2% similarities with those of P. chengduensis KUNCC 23-13570^T and P. poagena CBS 136771, respectively. The SSU gene (993) bp) exhibited 100% similarity with P. musae Sawada LSU 1147, P. chinensis K.K. Zhang, S. Hongsanan, Tennakoon & N. Xie MFLUCC 19-0217, P. oryzae I. Miyake CBS 110110, P. chengduensis OR20640 and P. chengduensis OR206401. For the LSU gene (890 bp), the isolate exhibited 99.8-99.9% similarity with P. sinensis Jayasiri, E.B.G. Jones & K.D. Hyde MFLUCC 18-1552, P. chiangraina Phook. & K.D. Hyde MFLUCC 13-0231, P. thysanolaenicola Phook. & K.D. Hyde MFLUCC 10-0563, and P. chengduensis KUNCC 23-13570^T. For the *RPB2* gene, the obtained sequence was 829 bp and exhibited 97.9% similarity with P. chengduensis KUNCC 23-13570^T. Regarding the TEF1- α gene, the obtained sequence was 858 bp and exhibited 100%, 97.6%, and 97.0% similarities with P. chengduensis KUNCC 23-13570^T, P. cycadis Wanas., Phookamsak & K.D. Hyde KUMCC 18-0161, and P. chinensis MFLUCC 18-1552, respectively. These results demonstrated that none of the five gene sequences alone allowed for the precise identification of strain KNUF-4H-A at the species level. Recently, combined sequences of the ITS region, and the SSU, LSU, TEF1- α , and RPB2 genes were successfully used to classify several new members of the family Phaeosphaeriaceae [16]. The same approach was applied in our study for phylogenetic analysis. In the constructed maximum likelihood (ML) phylogenetic tree, a monophyletic clade composed of the KNUF-4H-A isolate and two *P. chengduensis* strains (KUNCC 23-13571^T and KUNCC 23-13570) with a high bootstrap value of 99%-100% unequivocally indicated that they belong to the same species (Fig. 2). The sequences of RPB2 or TEF1- α and RPB2 genes were not available for the closely related Phaeospaeria

species, namely *P. ampeli* Tennakoon, C.H. Kuo & K.D. Hyde and *P. chinensis*, as the close neighbors of KNUF-4H-A. Therefore, an additional ML phylogenetic tree was constructed using the ITS, LSU, and SSU sequences. Similar to its position in the above-described tree, strain KNUF-4H-A clustered with the two strains of *P. cheugduensis* (Fig. 3). Collectively, the results of our molecular analyses demonstrated that the isolate belongs to *P. cheugduensis*.



Fig. 2. Maximum-likelihood phylogenetic tree of strain KNUF-4H-A based on the concatenated sequences of internal transcribed spacer (ITS) regions, and large subunit rDNA (LSU), small subunit rDNA (SSU), translation elongation factor $1-\alpha$ (*TEF1-\alpha*), and RNA polymerase II second largest subunit (*RPB2*) genes, showing the phylogenetic position of the new isolate among *Phaeosphaeria* species. The numbers above the branches represent the bootstrap values (>70%) obtained for 1,000 replicates. The isolated strain is indicated in bold. *Ophiosphaerella taiwanensis* SICAUCC 23-0003 was used as an outgroup. Bar, 0.01 substitutions per nucleotide position. ^Tindicates type strain.



Fig. 3. Maximum-likelihood phylogenetic tree of strain KNUF-4H-A based on the concatenated sequences of internal transcribed spacer (ITS) regions, as well as the large subunit rDNA (LSU) and small subunit rDNA (SSU) genes, showing the phylogenetic position of the new isolate among *Phaeosphaeria* species. The numbers above the branches represent the bootstrap values (>70%) obtained for 1000 replicates. The isolated strain is shown in bold. *Ophiosphaerella taiwanensis* NCYUCC 19-0152^T was used as an outgroup. Bar, 0.005 substitutions per nucleotide position. ^Tindicates type strain.

The members of the genus Phaeosphaeria are known plant pathogens responsible for inducing various diseases across a wide range of hosts and are commonly found in decaying plant material, deadwood, or organic debris from monocotyledons [16]. Previous studies have identified P. chengduensis as a saprobe, isolated from dead twigs of an unidentified deciduous host. P. poagena is also saprobic, typically found on deceased bamboo, whereas P. sichuanensis was identified on dead leaves of Pandanaceae [16]. In Korea, three species of Penicillium were identified in Korean Muljara (Muljarus japonicas Vuillefroy), European oil beetle (Meloe proscarabaeus Linnaeus), and weevil (Lixus imperessiventris Roelofs) [30], along with three new species of Mucor found in crickets (Gryllus sp.) [31]. Recently, Monochaetia mediana S.Y. Lee & H.Y. Jung was isolated from the hairy long-horned toad beetle (Moechotypa diphysis Pascoe) in Korea [22]. However, to the best of our knowledge, no previous studies had reported on the occurrence of fungal species on the citrus flower chafer (Gametis jucunda). Although members of the genus Phaeosphaeria have been identified in various habitats worldwide [16], the identified strain KNUF-4H-A was isolated from the citrus flower chafer (Gametis jucunda) in Korea. Additionally, sexual morphological characteristics were not observed, but cultural and molecular analyses enabled the identification of strain KNUF-4H-A as P. chengduensis. In conclusion, our findings offer new perspectives into the taxonomic diversity within the genus Phaeospaeria in Korea and provide new insights into the previously undocumented asexual morphology of P. chengduensis KUNCC 23-13571^T. Nevertheless, additional morphological analyses at the species level are required to further validate our findings. Moreover, further research is required to enhance our understanding of the interactions between insects and fungi, along with their classification, etiology, ecology, pathogenicity, and potential activities. To the best of our knowledge, our study constitutes the first record of Phaeosphaeria chengduensis KNUF-4H-A in Korea.

CONFLICT OF INTERESTS

The authors declare that they have no potential conflicts of interest.

ACKNOWLEDGEMENTS

This work was supported by a grant from the National Institute of Biological Resources, funded by the Ministry of Environment of the Republic of Korea [NIBR202333201].

REFERENCES

- 1. Barr ME. A classification of Loculoascomycetes. Mycologia 1979;71:935-57.
- Wong MKM, Hyde KD. Diversity of fungi on six species of Gramineae and one species of Cyperaceae in Hong Kong. Mycol Res 2001;105:1485-91.
- Promputtha I, Lumyong S, Dhanasekaran V, McKenzie EHC, Hyde KD, Jeewon R. A phylogenetic evaluation of whether endophytes become saprotrophs at host senescence. Microb Ecol 2007;53:579-90.

- 4. Promputtha I, Hyde KD, McKenzie EHC, Peberdy JF, Lumyong S. Can leaf degrading enzymes provide evidence that endophytic fungi becoming saprobes? Fungal Divers 2010;41:89-99.
- Ghimire SR, Hyde KD. Fungal endophytes. In: Ghimire SR, Hyde KD, editors. Plant surface microbiology. Berlin, Heidelberg: Springer Berlin Heidelberg; 2004. p. 281-92.
- Szilagyi-Zecchin VJ, Adamoski D, Gomes RR, Hungria M, Ikeda AC, Kava-Cordeiro V, Glienke C, Galli-Terasawa LV. Composition of endophytic fungal community associated with leaves of maize cultivated in south Brazilian field. Acta Microbiol Immunol Hung 2016;63:449-66.
- 7. Yang JW, Yeh YH, Kirschner R. A new endophytic species of *Neostagonospora* (Pleosporales) from the coastal grass *Spinifex littoreus* in Taiwan. Botany 2016;94:593-8.
- 8. Zhang Y, Crous PW, Schoch CL, Hyde KD. Pleosporales. Fungal Divers 2012;53:1-221.
- De Gruyter J, Woudenberg JHC, Aveskamp MM, Verkley GJM, Groenewald JZ, Crous PW. Redisposition of phoma-like anamorphs in Pleosporales. Stud Mycol 2013;75:1-36.
- Quaedvlieg W, Verkley GJM, Shin HD, Barretto RW, Alfenas AC, Swart WJ, Groenewald JZ, Crous PW. Sizing up *Septoria*. Stud Mycol 2013;75:307-90.
- Phookamsak R, Liu JK, McKenzie EHC, Manamgoda DS, Ariyawansa H, Thambugala KM, Dai DQ, Camporesi E, Chukeatirote E, Wijayawardene NN, et al. Revision of Phaeosphaeriaceae. Fungal Divers 2014;68:159-238.
- Tanaka K, Hirayama K, Yonezawa H, Sato G, Toriyabe A, Kudo H, Hashimoto A, Matsumura M, Harada Y, Kurihara Y, et al. Revision of the Massarineae (Pleosporales, Dothideomycetes) Stud Mycol 2015;82:75-136.
- Crous PW, Osieck ER, Shivas RG, Tan YP, Bishop-Hurley SL, Esteve-Raventós F, Larsson E, Luangsa-ard JJ, Pancorbo F, Balashov S, et al. Fungal Planet description sheets: 1478-1549. Pers: Mol Phylogeny Evol Fungi 2023;50:158-310.
- 14. Tan YP, Shivas RG. Index of Australian fungi No. 9. Geneva: Zenodo; 2023. p. 16-7.
- Tan YP, Sbaraini N, Chooi YH, Piggott AM, Foster C, Lacey E. Index of Australian fungi No. 24. Geneva: Zenodo; 2023. p. 10-1.
- Wanasinghe DN, Maharachchikumbura SSN. Exploring the diversity and systematics of Phaeosphaeriaceae: Taxonomic novelties from ecologically diverse habitats and their phylogenetic resolution. J Fungi 2023;9:853.
- Gurung K, Wertheim B, Falcao Salles J. The microbiome of pest insects: It is not just bacteria. Entomol Exp Appl 2019;167:156-70.
- Nicoletti R, Becchimanzi A. Ecological and molecular interactions between insects and fungi. Microorganisms 2022;10:96.
- Perissinotto R, Clennell L. Census of the fruit and flower chafers (Coleoptera, Scarabaeidae, Cetoniinae) of the Macau SAR, China. ZooKeys 2021;1026:17.
- Kishimoto-Yamada K, Kishimoto T, Sakai K, Terayama M, Ota Y, Takakuwa M. Occurrence of non-native species of Cetoniinae (Coleoptera, Insecta) at Port of Tokyo Wild Bird Park, Japan. Japanese J Conserv Ecol 2017;22:159-70.
- 21. Lee DW, Lee KC, Park CG, Choo HY, Kim YS. Scarabs (Coleoptera: Scarabaeidae) in sweet persimmon orchard and effect on sweet persimmon. Korean J Appl Entomol 2002;41:183-9.
- Lim SK, Das K, Hong SM, Suh SJ, Lee SY, Jung HY. Morphological and phylogenetic analyses reveal a new species of genus *Monochaetia* belonging to the family Sporocadaceae in Korea. Mycobiology 2023;51:87-93.

- 23. Gardes M, Brun TD. ITS primers with enhanced specificity for basidiomycetes-application to the identification of mycorrhizae and rusts. Mol Ecol 1993;2:113-8.
- 24. White TJ, Bruns TD, Lee SB, Taylor JW. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. PCR protocols: A guide to methods and applications. In: Innis MA, Gelfand DH, Sninsky JJ, White TJ, editors. San Diego: Academic Press; 1990. p. 315-22.
- 25. Vilgalys R, Hester M. Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. J Bacteriol 1990;172:4238-46.
- 26. O'Donnell K, Sutton DA, Rinaldi MG, Magnon KC, Cox PA, Revankar SG, Sanche S, Geiser DM, Juba JH, van Burik JH, et al. Genetic diversity of human pathogenic members of the *Fusarium oxysporum* complex inferred from multilocus DNA sequence data and amplified fragment length polymorphism analyses: evidence for the recent dispersion of a geographically widespread clonal lineage and nosocomial origin. J Clin Microbiol. 2004;42:5109-20.
- Liu YJ, Whelen S, Hall DB. Phylogenetic relationships among ascomycetes: evidence from an RNA polymerase II subunit. Mol Biol Evol 1999;16:1799-808.
- 28. Kumar S, Stecher G, Tamura K. MEGA7: Molecular evolutionary genetics analysis version 7.0 for bigger datasets. Mol Biol Evol 2016;33:1870-4.
- 29. Kimura M. A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. J Mol Evol 1980;16:111-20.
- Lamsal K, Kim SW, Naeimi S, Adhikari M, Yadav DR, Kim C, Lee HB, Lee YS. Three new records of *Penicillium* species isolated from insect specimens in Korea. Mycobiology 2013;41:116-9.
- 31. Nguyen TTT, Lee HB. Discovery of three new *Mucor* species associated with cricket insects in Korea. J Fungi 2022;8:601.