

Review Article



The Role of CD4 T Cell Help in CD8 T Cell Differentiation and Function During Chronic Infection and Cancer

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Conflict of Interest

The authors declare no potential conflicts of interest.

Abbreviations

ACT, adoptive cell therapy; BATF, basic leucine zipper transcription factor, ATF-like; CAR, chimeric Ag receptor; cDC1, type 1 conventional dendritic cell; DC, dendritic

ABSTRACT

CD4 and CD8 T cells are key players in the immune response against both pathogenic infections and cancer. CD4 T cells provide help to CD8 T cells via multiple mechanisms, including licensing dendritic cells (DCs), co-stimulation, and cytokine production. During acute infection and vaccination, CD4 T cell help is important for the development of CD8 T cell memory. However, during chronic viral infection and cancer, CD4 helper T cells are critical for the sustained effector CD8 T cell response, through a variety of mechanisms. In this review, we focus on T cell responses in conditions of chronic Ag stimulation, such as chronic viral infection and cancer. In particular, we address the significant role of CD4 T cell help in promoting effector CD8 T cell responses, emerging techniques that can be utilized to further our understanding of how these interactions may take place in the context of tertiary lymphoid structures, and how this key information can be harnessed for therapeutic utility against cancer.

Keywords: CD4 T cells; CD8 T cells; Chronic infection; Cancer; Immunotherapy

INTRODUCTION

CD4 and CD8 T cells are key players in the immune response against both pathogenic infections and cancer. Oftentimes CD4 T cells are considered to take on a more supportive or “helper” function in CD8 T cell-mediated, as well as humoral, Ab-mediated, immunity. During acute infection and vaccination, CD4 T cell help is important for the development of CD8 T cell memory. However, during chronic viral infection, CD4 T helper cells are critical for the sustained effector CD8 T cell response, through a variety of mechanisms. In this review, we focus on T cell responses in conditions of chronic Ag stimulation, such as chronic viral infection and cancer. In particular, we address the significant role of CD4 T cell help in promoting effector CD8 T cell responses, emerging techniques that can be utilized to further our understanding of how these interactions may take place in the context of tertiary lymphoid structures (TLSs), and how this key information can be harnessed for therapeutic utility against cancer.

cell; GC, germinal center; HPV, human papillomavirus; ICB, immune checkpoint blockade; ICOS, inducible costimulatory; LCMV, lymphocytic choriomeningitis virus; PDAC, pancreatic ductal adenocarcinoma; Tfh, T follicular helper; TIL, tumor-infiltrating lymphocyte; TLS, tertiary lymphoid structure; TME, tumor microenvironment.

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CONCEPTS OF CD4 T CELL HELP

Although it is known that CD4 T cells help CD8 T cells, precisely when, where and by what mechanisms this “help” takes place, continues to be of great scientific interest (1). CD4 T cells provide help via multiple mechanisms that guide the CD8 T cell response during chronic viral infection and cancer. Such mechanisms may include CD4 T cell licensing of dendritic cells (DCs), co-stimulation, and cytokine production.

CD4 T cell help during acute infection and vaccination

One of the critical mechanisms by which CD4 T cells provide help to CD8 T cells is via DC licensing and cross-presentation. Priming of Ag-specific CD8 T cells appears to take place in a two-step process (2). *In vivo* imaging found that upon infection or immunization, CD4 and CD8 T cells undergo their first priming steps independently and in different regions of lymphoid tissues (2-5). During the second step of priming, both Ag-specific CD4 and CD8 T cells interact with the same cross-presenting (6), lymph node-resident XCR1⁺ type 1 conventional dendritic cells (cDC1) (7,8). Cross-presentation takes place when the same Ag-presenting dendritic cell that presents endocytosed Ags on MHC class II surface molecules presents endocytosed Ags via MHC class I molecules, as well (9). Interestingly, a recent study found that CD4 T cells license cDC1 cells, which are necessary for priming both CD8 and CD4 T cells (10). Consistently, deficiency of XCR1⁺ cDC1s results in aberrant memory CD8 T cell formation following viral infection (8). Interaction between CD40L, expressed on CD4 T cells, and CD40 on cDC1 cells signals DCs (11,12) to enhance Ag presentation and expression of costimulatory molecules (13), revealing that CD4 T cells deliver help to CD8 T cells via interaction with DCs (14). Additionally, recent pre-clinical studies have shown the utility of CD40 agonist Abs in combination with immune checkpoint blockade (ICB) as a cancer therapeutic (15-17). In addition to co-stimulation, during this second step of priming, the production of key cytokines such as type 1 IFNs, IL-12, IL-15, and IL-2 by CD4 T cells and cDC1s is also important for driving effector and memory CD8 T cell differentiation and survival (2,18,19).

In the absence of CD4 T cell help, CD8 T cells have a cell-intrinsic deficiency for secondary expansion; thus, help from CD4 T cells is important in generating an optimal CD8 T cell memory response (20-26). In addition, an immunization model found that, without CD4 T cell help, CD8 T cells increase their expression of inhibitory molecules, as a result, developing a transcriptional profile that resembles exhausted CD8 T cells found in chronic infection (27-29). Collectively, these findings substantiate the significant role of helper T cells in supporting and enhancing the CD8 T cell response during immunization or viral infection.

CD4 T cell help during chronic infection

CD4 T cells play a significant role in helping maintain CD8 T cells and their response in many models of chronic infection (30-34). Early studies using genetic knockout or depletion of CD4 T cells during chronic lymphocytic choriomeningitis virus (LCMV) infection suggested that CD4 T cells are critical in sustaining the CD8 T cell response (35,36). Additionally, in the absence of CD4 T cell “help,” CD8 T cells enter a dysfunctional state, losing their cytotoxic capacity for viral control (30,35,36).

The secretion of IL-21 by CD4 T cells was found to be one of the major mechanisms by which CD4 T cells help CD8 T cells maintain their functionality and facilitate viral control during chronic infection (37-39). Consistently, in patients with chronic viral infections, such as HIV and hepatitis C virus, CD4 T cell production of IL-21 often positively correlates with CD8 T

cell function and improved viral control (40-43), suggesting that the IL-21 pathway may hold therapeutic potential.

During viral infection, there are several subsets of CD4 T cells that can produce IL-21, including T follicular helper (Tfh) cells and Th1 cells (25,44,45). Persistent viral infection may promote CD4 T cell differentiation towards Tfh cells (46,47). This differentiation of CD4 T cells towards Tfh cells may be due to their critical role in the germinal center (GC) response (48,49), because resolution of the viremic phase of LCMV Cl13 chronic viral infection critically depends on Ab production (46,47,50,51). In addition to helping CD8 T cells, IL-21 produced by CD4 T cells, specifically Tfh cells, is necessary for GC responses; the absence of IL-21 results in impaired GC maintenance, reduced affinity maturation by B cells, and isotype class switching (52-54). Therefore, CD4 T cell help during chronic infection is critical for both the CD8 T cell-mediated response, as well as the humoral response to control persistent viral infection.

CD4 T cell help affects CD8 T cell heterogeneity

Under chronic Ag stimulation, such as chronic viral infection and cancer, CD8 T cells were suspected to progressively differentiate towards an exhausted state, which is counterproductive in fighting viral infections or preventing tumor growth (55). Due to advancements in biotechnology, identification of CD8 T cell heterogeneity in response to chronic viral infection was defined at single-cell resolution. At least three phenotypically, functionally, and epigenetically distinct CD8 T cell subsets have been identified in response to chronic viral infection. Progenitor Ly108^{hi} TCF-1^{hi} CD8 T cells are a precursor population with self-renewing abilities (56-61). During early infection, progenitor CD8 T cells undergo a bifurcation pathway that can give rise to either terminally differentiated “exhausted” PD-1^{hi} CD8 T cells or, with the help of IL-21 produced by CD4 T cells (62), progenitor cells can give rise to effector CX₃CR1^{hi} CD8 T cells, which maintain their functional capacity to control viral infection (62-68). In particular, the effector CX₃CR1^{hi} CD8 T cell subset exhibits augmented cytolytic function and expression of effector molecules, such as granzyme B (Fig. 1) (62,63,69).

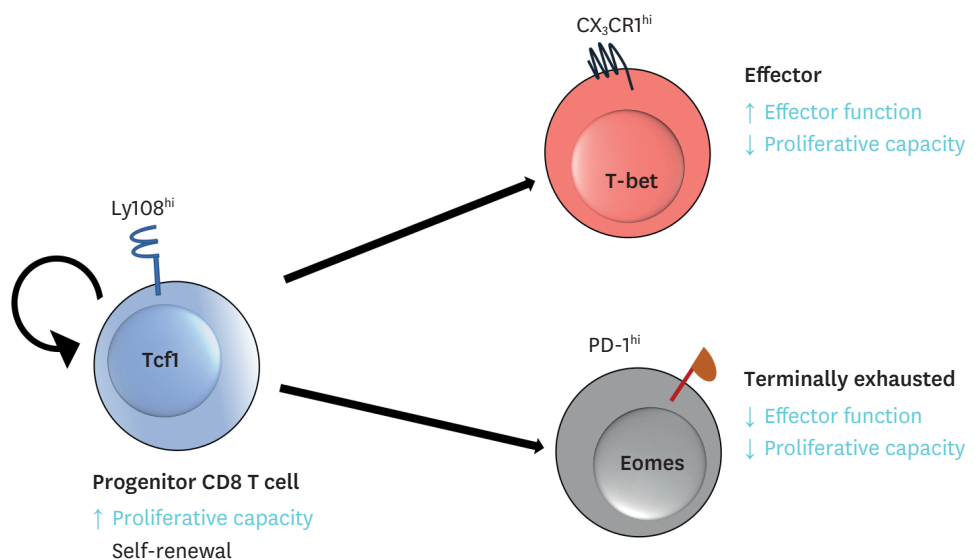


Figure 1. Model of CD8 T cell heterogeneity during chronic antigen stimulation. Progenitor Ly108^{hi} TCF-1^{hi} CD8 T cells maintain a proliferative capacity for self-renewal, while also giving rise to effector CX₃CR1^{hi} CD8 T cells or “exhausted” PD-1^{hi} CD8 T cells.

On the other hand, during cancer, the tumor microenvironment (TME) can be immunosuppressive, with Tregs taking up a significant proportion of CD4 T cells found in some tumors (70-73). Interestingly, in a preclinical melanoma model, CD8 T cells were found to primarily exist in two differentiation states within the TME, either terminally exhausted or progenitor-like (66), which resemble an immunological state similar to “unhelped” CD8 T cells during chronic LCMV infection (62,65). In addition, a recent study of CD8 T cells in human papillomavirus (HPV)-positive head and neck cancer also identified PD1⁺ stem-like CD8 T cells in the TME that resembles progenitor-like cells that contribute to the immune response against tumor cells (74). Our recently published work has shown that when CD4 T cell help is removed via CD4 depletion, the development of CX₃CR1⁺ effector CD8 T cells during chronic LCMV infection is abrogated (62), thus mimicking the TME, where CD4 T cell help is insufficient.

CD4 T cells during cancer

In cancer, CD4 T cell help has been found to promote antitumor CD8 T cell responses, whereas Tregs suppress such responses (71,75,76). Interestingly, expression of MHC class II on the surface of tumor cells, may be associated with improved progression-free survival and overall survival following immunotherapy (77-79). In addition, MHC class II expression on tumor cells was also associated with increased numbers of CD4 and CD8 tumor-infiltrating lymphocytes (TILs), increased TLS formation, and higher expression of IFN- γ , IL-2 and IL-12, and Th1-associated cytokines (80). Some CD4 T cells may even have a direct ability to recognize and kill target cells (81-85). Thus, CD4 T cells may play an important role, both directly and indirectly, in the antitumor response.

Meanwhile, Tregs make up a major component of immune cells found in the immunosuppressive TME (70,71). Tregs may inhibit antitumor activity and are associated with poor prognosis in cancer patients (86,87). Importantly, Tregs maintain their suppressive activity through the production of various immunosuppressive cytokines (IL-10, TGF- β , and IL-35), as well as their surface expression of immunosuppressive receptors (lymphocyte activation gene-3, T-cell immunoglobulin and ITIM domain (TIGIT), CTLA4, and PD-1), which can inhibit effector T cells (88-92). In addition, intra-tumoral Tregs may also interact with dendritic cells to suppress CD80 and CD86 expression, which enhances expression of inhibitory receptor, thus pushing CD8 T cells towards a dysfunctional state (75). Unlike Tregs, helper CD4 T cells are important in maintaining CD8 T cell recruitment, proliferation, and effector function in some models of cancer (93-96). Thus, harnessing the appropriate helper CD4 T cell support may be important in preventing CD8 T cell dysfunction. The following section describes potential applications of CD4 T cell help in enhancing and sustaining effector CD8 T cells.

APPLICATIONS OF CD4 T CELLS

Cancer immunotherapy has made great strides in improving patient outcomes, especially with hematologic malignancies. However, cancer immunotherapy efficacy in treating solid tumors remains limited. A major reason for the setback in treating solid tumors is that TILs often differentiate into dysfunctional states, resembling exhausted T cells that arise during chronic viral infections (97-99). In addition to upregulation of inhibitory molecules such as PD-1, the dysfunctional state of exhaustion in T cells is characterized by diminished effector function, namely decreased cytotoxic activity and reduced secretion of effector molecules, such as

granzyme B and IFN- γ (28,97), resulting in reduced antitumor activity. These dysfunctional TILs are ineffective at killing tumor cells, in part due to epigenetic imprinting that maintains them in an exhausted state of differentiation (100-102). Thus, it is important to understand the cellular and molecular mechanisms that regulate CD8 T cell differentiation in the setting of cancer. Addressing this critical knowledge gap will be essential in developing novel strategies that enhance and restore CD8 T cell effector function within the TME. Applying our current knowledge of CD4 T cell help in generating robust CD8 T cell responses, we will discuss the immunotherapeutic potential of applying these mechanisms of CD4 T cell help against cancer.

ICB

One of the major advances in cancer treatment has been the use of monoclonal Abs to block immune regulatory checkpoints, such as PD-1 and CTLA4 (34,103,104). ICB therapy targets inhibitory receptors, such as PD-1, which are upregulated on the surface of dysfunction tumor-infiltrating T cells (105) in an effort to maintain the T cell response against cancer. Studies have shown that chronic antigenic exposure of T cells results in continuous PD-1 signaling, which epigenetically programs T cell exhaustion (106,107). However, it was recently observed that PD-1 therapy preferentially supports the proliferative burst of a specialized subset of PD-1⁺CXCR5⁺TCF1⁺ CD8 T cells, in a chronic LCMV infection model (60). More recent studies have recapitulated these findings in cancer models, showing that these stem-like, or progenitor, TCF1⁺PD-1⁺ CD8 T cells are preferentially targeted in response to checkpoint blockade immunotherapy (66,108,109). Taken together, these findings suggest that PD-1 therapy supports progenitor CD8 T cells in the tumor, allowing them to continue proliferating and give rise to more effector-like cells in order to control tumors. However, not all patients respond to ICB therapy, and about a third of them relapse (103). Therefore, there has been increased interest in using combination therapies to boost responsiveness to cancer immunotherapies.

Recent studies have shown that the antitumor response to ICB therapy requires the response of both tumor Ag-specific CD8 and CD4 T cells (110). In particular, MHC-II neoantigens are crucial for activating CD4⁺ T cells, which are important for generating functional CD8 T cells in response to checkpoint blockade immunotherapy (110). Interestingly, a study found that enhanced CD4 T cell responses are one of the underlying mechanisms of anti-CTLA-4 blockade (104). Additionally, clinical studies have shown that CD4 T cell populations may determine responsiveness to ICB (111-113). Thus, methods of enhancing the tumor-specific CD4 T helper cell population may provide insight into improving ICB therapy responsiveness.

Adoptive cell therapy (ACT)

ACT entails using tumor-specific cells, typically TILs or genetically engineered chimeric Ag receptor (CAR)-expressing T cells, expanding them *ex vivo*, and then infusing them back into a lymphodepleted patient (114). As previously reported, ACT along with administration of IL-2 can lead to prolonged eradication of tumors in cancer patients that have exhausted other treatment options (115-119). Adoptively transferred TILs typically consist of a mixture of CD4 and CD8 T cells, however most studies in the field of cancer immunotherapy focus on CD8 T cells (120). In a previous study, mutation-specific CD4 T cells from a patient were expanded and adoptively transferred back, resulting in tumor regression (121), which warrants further exploration into the mechanisms of the CD4 T cell response against cancer.

More recently, adoptive cell transfer of autologous, genetically engineered T cells to target tumor Ags has provided a breakthrough treatment for hematological malignancies (122). CAR-T cells are genetically engineered T cell receptors that reprogram T cells to target tumor-

associated Ags (123). Although this cutting-edge technology is efficacious for some patients, 30%–60% of patients relapse after CAR treatment (124). Evidence shows that the cellular composition and immunophenotypes of CAR-T cells is instrumental for therapeutic efficacy (125). In particular, the ratio of CD4 to CD8 T cells may play a role in the antitumor response of CAR-T cells (126). A 2016 clinical trial reported that a 1:1 ratio of CD4 to CD8 T cells during CAR-T manufacturing resulted in high remission rates among B-cell acute lymphoblastic leukemia patients undergoing CAR-T cell therapy (126). This is congruent with preclinical CAR-T therapy studies that found CD4 T cell help induces CD8 T cell memory function, which plays a role in antitumor responses (127). In addition, different subsets of CD4 helper T cells may also affect the effector response of CAR-T therapy. Interestingly, CAR containing the inducible costimulatory (ICOS) intracellular domain redirected CD4 T cells towards a Th17 phenotype with augmented effector function and persistence of CAR-T cells in the circulation (128). Furthermore, CD4 T helper cells play a clinically important role in CAR-T therapy, as observed in a recent study which showed that CD4 CAR-T cells persist in decade-long leukemia, and continue to exhibit functional characteristics of activation, proliferation, and cytotoxicity (129). Thus, CAR CD4 T cells not only facilitate CD8 T cell effector functions but may have the potential for direct cytotoxicity against tumor cells.

CD4 T cells are widely known to assist cytotoxic CD8 T cells (71,130) and help in conferring a cytotoxic T cell effector program (2,131). In addition, ACT of CD4 helper T cells has shown promise in multiple cancer immunotherapy preclinical models (84,132-134), where cells may function as a 'living drug,' by providing costimulatory signals and continued cytokine production (120). Notably, Th17 and Th9 cells may be more effective than Th1 cells in limiting tumor progression (135). Interestingly, IL-21 is a commonly produced cytokine by both Th9 and Th17 T helper cell subsets, which may play an important role in their response to cancer (62,136,137). However, the precise mechanisms by which helper CD4 T cells mediate anti-tumor responses remains under investigation.

MECHANISMS OF CD4 T CELL HELP

IL-21-mediated CD4 T cell help

CD4 T cells are one of the predominant producers of IL-21 (138). Accumulating data reveals that IL-21 signaling on CD8 T cells is vital for their sustained function and control of chronic viral infection (37-39). Additionally, IL-21 treatment of both human and mouse tumor-specific CD8 T cells, resulted in enhanced longevity and anti-tumor activity of CD8 T cells *in vivo* (139-141), supporting the notion that CD4 T cell help via their production of IL-21 may enhance effector CD8 T cell responses. Notably, our pre-clinical studies have shown that adoptive transfer of IL-21-producing CD4 T cell help increases the intra-tumoral effector CX₃CR1⁺ CD8 T cell population, subsequently correlating with reduced tumor burden (62). Thus, harnessing and enhancing CD4 T cell help may hold the answer to combating T cell exhaustion during chronic viral infection and cancer.

Our laboratory has previously shown that IL-21 signaling, through STAT3 induces basic leucine zipper transcription factor, ATF-like (BATF) activation in CD8⁺ T cells resulting in sustained CD8⁺ T cell survival and effector function during chronic viral infection (142), further corroborating the importance of this finding in a preclinical melanoma model (143). BATF cooperatively binds to other transcription factors, which remodel the chromatin landscape to produce changes in chromatin accessibility (144). Through these changes in

the chromatin landscape, BATF promotes the differentiation and function of several types of immune cells, including CD8⁺ T cells (145-152). Most recently, our lab found that BATF is required in maintaining a permissive chromatin structure, which may allow for the transition from progenitor TCF-1⁺ CD8⁺ T cells to effector CX₃CR1⁺ CD8⁺ T cells in a chronic viral infection model (153). Additionally, another study in a preclinical tumor model also showed that BATF may improve CAR T cell antitumor responses by skewing their transcriptional profiles towards a effector phenotype (154). Taken together, these findings support our current knowledge of IL-21-producing CD4 T cell help, via the IL-21-BATF pathway, as being critical for the progenitor to effector differentiation of CD8 T cells, and suggest the potential utility of IL-21⁺ CD4 T cell ACT as a cancer immunotherapeutic.

Cellular localization in chronic infection and cancer

Multiple subsets of CD4 T cells are capable of producing IL-21, however it remained unknown until recently, whether a specific subset may be the primary “helper” of these effector CD8 T cell responses (45). We found that Tfh cells may be the main IL-21 producers critical for sustaining these effector CD8 T cell responses during chronic infection (45,155). Likewise, a recent study by Cui et al. (156) found a correlation of Tfh cell and GC B cell transcriptional signatures in tumors of lung adenocarcinoma patients, which also positively correlated with prolonged survival. Meanwhile, in their preclinical lung adenocarcinoma model, which expresses neoantigens for both T and B cells, they showed that interactions between tumor-specific GC B cells and Tfh cells, along with IL-21 produced by Tfh cells, are necessary for effector CD8 T cell function and tumor control (156). Another study also showed that MHC II-expressing cells may provide niches for maintaining the progenitor subset of CD8 T cells (157). Additionally, the preferential localization of the progenitor subset of CD8 T cells in lymphoid-like stromal areas has also been observed in head and neck cancer (74). Interestingly, our own recent findings, using spatial transcriptomics, support the potential colocalization of B cells, IL-21-producing Tfh cells, and progenitor CD8 T cells, which are the likely responders to IL-21-mediated help (Fig. 2) (155). Taken together, these studies suggest that the organized

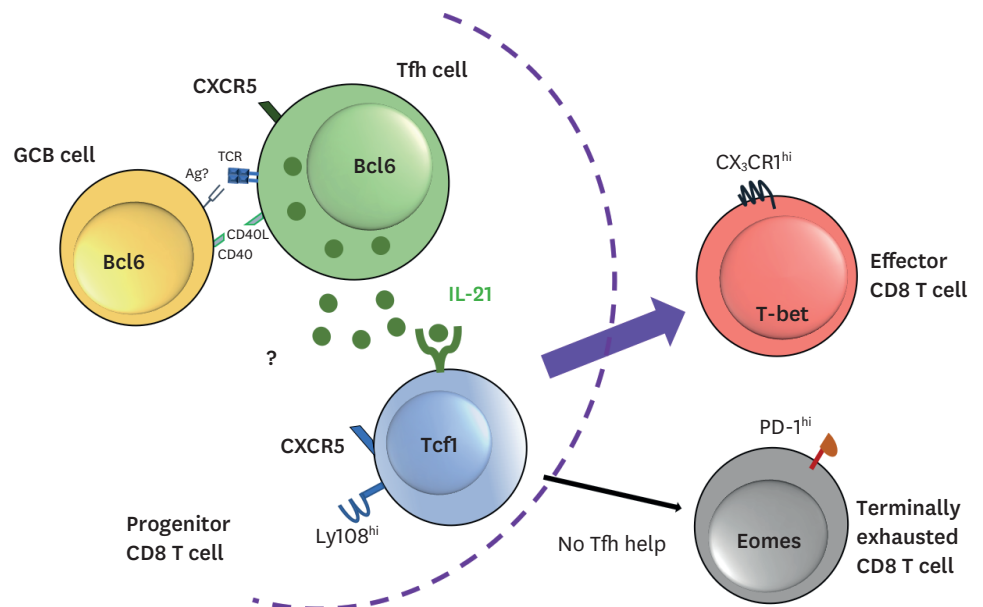


Figure 2. Hypothetical model depicting colocalization of GC B, Tfh, and progenitor CD8 T cells within organized lymphoid structures. GC B cells, and IL-21 producing T follicular helper cell colocalization with progenitor CD8 T cells help facilitate progenitor to effector CD8 T cell differentiation. Importantly, this colocalization may take place within TLSS.

interactions of B cells and Tfh cells with CD8 T cells may be a conserved feature across chronic inflammatory states, such as cancer and chronic infection/inflammation, during which the formation of TLSs may arise (158). In the remainder of this section, we will further discuss TLS in the context of cancer and its correlation with therapeutic efficacy.

It has been known that Tfh cells provide essential help signals for B cells to facilitate GC reactions, isotype class switching, production of high-affinity Abs, and generation of memory B cells and long-lived plasma cells in the context of infection and vaccination (48,49). However, it has become increasingly clear that the presence of Tfh cells (159-161) and B cells (162-170) is correlated with prolonged survival, as well as positive therapeutic responsiveness in patients with a wide variety of cancers (164,165,168). These correlations are particularly strong when B cells and Tfh cells are found in TLSs, which may be important in facilitating their interactions (171-173). A recent study found that in epidermal growth factor receptor-mutant lung cancer patients who have an unfavorable response to anti-PD-1 therapy, there is a disruption in the cooperative interactions between Tfh, B cells, and resident-memory CD8 T cells. This dysregulation may contribute to the reduced effectiveness of immunotherapy in this specific subset of lung cancer patients (174).

TLSs are postnatal ectopic lymphoid formations, consisting of organized aggregates of immune cells that arise in chronically inflamed disease states, such as cancer (158), chronic inflammation/infection (175,176), autoimmune diseases (177-179), and in other immune responses. The presence of TLSs has been reported in multiple types of cancers, including but not limited to breast cancer (161,162), non-small cell lung cancer (163,180,181), head and neck squamous cell carcinoma (160,170,171), colorectal cancer (182,183), gastric cancer (166), ovarian cancer (167,169), and melanoma (164,184). Interestingly, conventional therapies to treat cancer may include corticosteroids, which are often administered alongside chemotherapy or immunotherapy to reduce side effects, resulting in a dampening of the immune response. Corticosteroid administration in cancer patients has been shown to impair TLS formation and maturation, resulting in GC loss (185,186). These findings should be further explored, as steroids, which induce immunosuppression, may play a detrimental role in dampening TLS formation and, in turn, therapeutic efficacy.

Some studies using ICB immunotherapy to treat patients with non-small cell lung cancer (187), or urothelial cancer (185), found an increase in TLSs in responsive or regressing lesions, indicating that TLSs may harbor immune cells that are responsive to ICB therapy. It is important to note that dysfunctional or “exhausted” PD-1^{high} CD8 T cells may also localize within TLSs, near Tfh cells and B cells, and could help predict responsiveness to PD-1 blockade therapy (188). As previously observed, in a chronic LCMV infection model (60) and recapitulated in cancer (66,108,109), blockade of PD-1 may support the proliferative burst of PD-1⁺CXCR5⁺TCF1⁺ CD8 T cells, also recognized as progenitor CD8 T cells. Taken together, this suggests that TLSs may facilitate the environment necessary for progenitor CD8 T cells to respond to ICB therapy by proliferating and giving rise to effector CD8 T cells.

Future studies should be conducted in cancer models, including patient tumor samples, to determine if progenitor CD8 T cells and Tfh cells are found colocalizing within intra-tumoral TLSs, and whether such colocalizations may be associated with enhanced responsiveness of patients to ICB therapy. Furthermore, as TLS presence is associated with patient responsiveness to certain therapeutics, attempts should be made at elucidating methods to initiate or facilitate TLS formation within tumors.

TLSS as prognostic and therapeutic targets

Recent studies have shown evidence of therapeutically-induced TLSS that are associated with tumor control in preclinical models of neuroendocrine pancreatic cancer and breast cancer (158,189,190). In these studies, PD-L1 blockade was used in combination with anti-angiogenic therapies, which resulted in tumor blood vessel transformation into high endothelial venules, TLS formation, enhanced CD8 T cell infiltration and activity, as well as tumor destruction (158,189,190). Additionally, in a preclinical model of colorectal cancer used immunogenic intestinal bacteria to induce an immune response and found that Tfh cells drove TLS formation and tumor control (191). In another study, pancreatic ductal adenocarcinoma (PDAC) patients were therapeutically vaccinated with GVAX, an irradiated allogeneic GM-CSF-secreting PDAC tumor vaccine, and cyclophosphamide (to deplete regulatory T cells), which resulted in a conversion of “nonimmunogenic” PDAC neoplasms, into “immunogenic” neoplasm, with increased TLS formation in the majority of patients (192). These studies suggest potential mechanisms that can be harnessed to induce TLSS in cancer, which may augment ICB and ACT to target “non-responders.”

CONCLUSION

CD4 T cells assist in CD8 T cell effector responses during states of chronic Ag stimulation, such as chronic infection and cancer. Specifically, CD4 T cell help via IL-21 secretion, is critical to facilitating effective CD8 T cell responses in such conditions. Our recent studies discovered Tfh cells as the primary IL-21-producing CD4 T cells that provide this necessary support to CD8 T cells during chronic infection. Additionally, others have further corroborated the importance of IL-21-producing Tfh cells in the cancer model, as well as the necessity of Tfh and GC B cell interactions in effector CD8 T cell responses resulting in tumor control. Interestingly, the presence of TLSS during chronic inflammatory states, including cancer, further suggests the important interplay of these key lymphocytes. Furthermore, there is often a strong, positive correlation between the presence of TLS and anti-cancer therapeutic efficacy, especially when Tfh and B cells are found in TLSS. Therefore, understanding and enhancing CD4 T cell helper mechanisms may harness powerful therapeutic potential against cancer.

REFERENCES

1. Castellino F, Germain RN. Cooperation between CD4⁺ and CD8⁺ T cells: when, where, and how. *Annu Rev Immunol* 2006;24:519-540.
[PUBMED](#) | [CROSSREF](#)
2. Borst J, Ahrends T, Bąbała N, Melief CJ, Kastenmüller W. CD4⁺ T cell help in cancer immunology and immunotherapy. *Nat Rev Immunol* 2018;18:635-647.
[PUBMED](#) | [CROSSREF](#)
3. Bedoui S, Heath WR, Mueller SN. CD4⁺ T-cell help amplifies innate signals for primary CD8⁺ T-cell immunity. *Immunol Rev* 2016;272:52-64.
[PUBMED](#) | [CROSSREF](#)
4. Calabro S, Liu D, Gallman A, Nascimento MS, Yu Z, Zhang TT, Chen P, Zhang B, Xu L, Gowthaman U, et al. Differential intrasplenic migration of dendritic cell subsets tailors adaptive immunity. *Cell Reports* 2016;16:2472-2485.
[PUBMED](#) | [CROSSREF](#)
5. Gerner MY, Casey KA, Kastenmuller W, Germain RN. Dendritic cell and antigen dispersal landscapes regulate T cell immunity. *J Exp Med* 2017;214:3105-3122.
[PUBMED](#) | [CROSSREF](#)

6. Bachem A, Güttler S, Hartung E, Ebstein F, Schaefer M, Tannert A, Salama A, Movassaghi K, Opitz C, Mages HW, et al. Superior antigen cross-presentation and XCR1 expression define human CD11c⁺CD141⁺ cells as homologues of mouse CD8⁺ dendritic cells. *J Exp Med* 2010;207:1273-1281.
[PUBMED](#) | [CROSSREF](#)
7. Hor JL, Whitney PG, Zaid A, Brooks AG, Heath WR, Mueller SN. Spatiotemporally distinct interactions with dendritic cell subsets facilitates CD4⁺ and CD8⁺ T cell activation to localized viral infection. *Immunity* 2015;43:554-565.
[PUBMED](#) | [CROSSREF](#)
8. Eickhoff S, Brewitz A, Gerner MY, Klauschen F, Komander K, Hemmi H, Garbi N, Kaisho T, Germain RN, Kastenmüller W. Robust anti-viral immunity requires multiple distinct T cell-dendritic cell interactions. *Cell* 2015;162:1322-1337.
[PUBMED](#) | [CROSSREF](#)
9. Joffre OP, Segura E, Savina A, Amigorena S. Cross-presentation by dendritic cells. *Nat Rev Immunol* 2012;12:557-569.
[PUBMED](#) | [CROSSREF](#)
10. Ferris ST, Durai V, Wu R, Theisen DJ, Ward JP, Bern MD, Davidson JT 4th, Bagadia P, Liu T, Briseño CG, et al. cDC1 prime and are licensed by CD4⁺ T cells to induce anti-tumour immunity. *Nature* 2020;584:624-629.
[PUBMED](#) | [CROSSREF](#)
11. Bennett SR, Carbone FR, Karamalis F, Flavell RA, Miller JF, Heath WR. Help for cytotoxic-T-cell responses is mediated by CD40 signalling. *Nature* 1998;393:478-480.
[PUBMED](#) | [CROSSREF](#)
12. Schoenberger SP, Toes RE, van der Voort EI, Ofringa R, Melief CJ. T-cell help for cytotoxic T lymphocytes is mediated by CD40-CD40L interactions. *Nature* 1998;393:480-483.
[PUBMED](#) | [CROSSREF](#)
13. Grewal IS, Flavell RA. The role of CD40 ligand in costimulation and T-cell activation. *Immunol Rev* 1996;153:85-106.
[PUBMED](#) | [CROSSREF](#)
14. Ridge JP, Di Rosa F, Matzinger P. A conditioned dendritic cell can be a temporal bridge between a CD4⁺ T-helper and a T-killer cell. *Nature* 1998;393:474-478.
[PUBMED](#) | [CROSSREF](#)
15. Morrison AH, Diamond MS, Hay CA, Byrne KT, Vonderheide RH. Sufficiency of CD40 activation and immune checkpoint blockade for T cell priming and tumor immunity. *Proc Natl Acad Sci U S A* 2020;117:8022-8031.
[PUBMED](#) | [CROSSREF](#)
16. Singh M, Vianden C, Cantwell MJ, Dai Z, Xiao Z, Sharma M, Khong H, Jaiswal AR, Faak F, Hailemichael Y, et al. Intratumoral CD40 activation and checkpoint blockade induces T cell-mediated eradication of melanoma in the brain. *Nat Commun* 2017;8:1447.
[PUBMED](#) | [CROSSREF](#)
17. Djureinovic D, Wang M, Kluger HM. Agonistic CD40 antibodies in cancer treatment. *Cancers (Basel)* 2021;13:1302.
[PUBMED](#) | [CROSSREF](#)
18. Agarwal P, Raghavan A, Nandiwada SL, Curtsinger JM, Bohjanen PR, Mueller DL, Mescher MF. Gene regulation and chromatin remodeling by IL-12 and type I IFN in programming for CD8 T cell effector function and memory. *J Immunol* 2009;183:1695-1704.
[PUBMED](#) | [CROSSREF](#)
19. Pipkin ME, Sacks JA, Cruz-Guilloty F, Lichtenheld MG, Bevan MJ, Rao A. Interleukin-2 and inflammation induce distinct transcriptional programs that promote the differentiation of effector cytolytic T cells. *Immunity* 2010;32:79-90.
[PUBMED](#) | [CROSSREF](#)
20. Janssen EM, Lemmens EE, Wolfe T, Christen U, von Herrath MG, Schoenberger SP. CD4⁺ T cells are required for secondary expansion and memory in CD8⁺ T lymphocytes. *Nature* 2003;421:852-856.
[PUBMED](#) | [CROSSREF](#)
21. Shedlock DJ, Shen H. Requirement for CD4 T cell help in generating functional CD8 T cell memory. *Science* 2003;300:337-339.
[PUBMED](#) | [CROSSREF](#)
22. Sun JC, Bevan MJ. Defective CD8 T cell memory following acute infection without CD4 T cell help. *Science* 2003;300:339-342.
[PUBMED](#) | [CROSSREF](#)
23. Belz GT, Wodarz D, Diaz G, Nowak MA, Doherty PC. Compromised influenza virus-specific CD8⁺-T-cell memory in CD4⁺-T-cell-deficient mice. *J Virol* 2002;76:12388-12393.
[PUBMED](#) | [CROSSREF](#)

24. von Herrath MG, Yokoyama M, Dockter J, Oldstone MB, Whitton JL. CD4-deficient mice have reduced levels of memory cytotoxic T lymphocytes after immunization and show diminished resistance to subsequent virus challenge. *J Virol* 1996;70:1072-1079.
[PUBMED](#) | [CROSSREF](#)
25. Laidlaw BJ, Craft JE, Kaech SM. The multifaceted role of CD4⁺ T cells in CD8⁺ T cell memory. *Nat Rev Immunol* 2016;16:102-111.
[PUBMED](#) | [CROSSREF](#)
26. Williams MA, Tyznik AJ, Bevan MJ. Interleukin-2 signals during priming are required for secondary expansion of CD8⁺ memory T cells. *Nature* 2006;441:890-893.
[PUBMED](#) | [CROSSREF](#)
27. Provine NM, Larocca RA, Aid M, Penaloza-MacMaster P, Badamchi-Zadeh A, Borducchi EN, Yates KB, Abbink P, Kirilova M, Ng'ang'a D, et al. Immediate dysfunction of vaccine-elicited CD8⁺ T cells primed in the absence of CD4⁺ T cells. *J Immunol* 2016;197:1809-1822.
[PUBMED](#) | [CROSSREF](#)
28. Wherry EJ, Ha SJ, Kaech SM, Haining WN, Sarkar S, Kalia V, Subramaniam S, Blattman JN, Barber DL, Ahmed R. Molecular signature of CD8⁺ T cell exhaustion during chronic viral infection. *Immunity* 2007;27:670-684.
[PUBMED](#) | [CROSSREF](#)
29. West EE, Youngblood B, Tan WG, Jin HT, Araki K, Alexe G, Konieczny BT, Calpe S, Freeman GJ, Terhorst C, et al. Tight regulation of memory CD8⁺ T cells limits their effectiveness during sustained high viral load. *Immunity* 2011;35:285-298.
[PUBMED](#) | [CROSSREF](#)
30. Zajac AJ, Blattman JN, Murali-Krishna K, Sourdive DJ, Suresh M, Altman JD, Ahmed R. Viral immune evasion due to persistence of activated T cells without effector function. *J Exp Med* 1998;188:2205-2213.
[PUBMED](#) | [CROSSREF](#)
31. Snyder CM, Loewendorf A, Bonnett EL, Croft M, Benedict CA, Hill AB. CD4⁺ T cell help has an epitope-dependent impact on CD8⁺ T cell memory inflation during murine cytomegalovirus infection. *J Immunol* 2009;183:3932-3941.
[PUBMED](#) | [CROSSREF](#)
32. Kemball CC, Pack CD, Guay HM, Li ZN, Steinhauer DA, Szomolanyi-Tsuda E, Lukacher AE. The antiviral CD8⁺ T cell response is differentially dependent on CD4⁺ T cell help over the course of persistent infection. *J Immunol* 2007;179:1113-1121.
[PUBMED](#) | [CROSSREF](#)
33. Cardin RD, Brooks JW, Sarawar SR, Doherty PC. Progressive loss of CD8⁺ T cell-mediated control of a gamma-herpesvirus in the absence of CD4⁺ T cells. *J Exp Med* 1996;184:863-871.
[PUBMED](#) | [CROSSREF](#)
34. Aubert RD, Kamphorst AO, Sarkar S, Vezys V, Ha SJ, Barber DL, Ye L, Sharpe AH, Freeman GJ, Ahmed R. Antigen-specific CD4 T-cell help rescues exhausted CD8 T cells during chronic viral infection. *Proc Natl Acad Sci U S A* 2011;108:21182-21187.
[PUBMED](#) | [CROSSREF](#)
35. Battagay M, Moskophidis D, Rahemtulla A, Hengartner H, Mak TW, Zinkernagel RM. Enhanced establishment of a virus carrier state in adult CD4⁺ T-cell-deficient mice. *J Virol* 1994;68:4700-4704.
[PUBMED](#) | [CROSSREF](#)
36. Matloubian M, Concepcion RJ, Ahmed R. CD4⁺ T cells are required to sustain CD8⁺ cytotoxic T-cell responses during chronic viral infection. *J Virol* 1994;68:8056-8063.
[PUBMED](#) | [CROSSREF](#)
37. Fröhlich A, Kisielow J, Schmitz I, Freigang S, Shamshiev AT, Weber J, Marsland BJ, Oxenius A, Kopf M. IL-21R on T cells is critical for sustained functionality and control of chronic viral infection. *Science* 2009;324:1576-1580.
[PUBMED](#) | [CROSSREF](#)
38. Elsaesser H, Sauer K, Brooks DG. IL-21 is required to control chronic viral infection. *Science* 2009;324:1569-1572.
[PUBMED](#) | [CROSSREF](#)
39. Yi JS, Du M, Zajac AJ. A vital role for interleukin-21 in the control of a chronic viral infection. *Science* 2009;324:1572-1576.
[PUBMED](#) | [CROSSREF](#)
40. Chevalier ME, Jülg B, Pyo A, Flanders M, Ranasinghe S, Soghoian DZ, Kwon DS, Rychert J, Lian J, Muller MI, et al. HIV-1-specific interleukin-21⁺ CD4⁺ T cell responses contribute to durable viral control through the modulation of HIV-specific CD8⁺ T cell function. *J Virol* 2011;85:733-741.
[PUBMED](#) | [CROSSREF](#)

41. Iannello A, Boulassel MR, Samarani S, Debbeche O, Tremblay C, Toma E, Routy JP, Ahmad A. Dynamics and consequences of IL-21 production in HIV-infected individuals: a longitudinal and cross-sectional study. *J Immunol* 2010;184:114-126.
[PUBMED](#) | [CROSSREF](#)
42. Williams LD, Bansal A, Sabbaj S, Heath SL, Song W, Tang J, Zajac AJ, Goepfert PA. Interleukin-21-producing HIV-1-specific CD8 T cells are preferentially seen in elite controllers. *J Virol* 2011;85:2316-2324.
[PUBMED](#) | [CROSSREF](#)
43. Feng G, Zhang JY, Zeng QL, Jin L, Fu J, Yang B, Sun Y, Jiang T, Xu X, Zhang Z, et al. HCV-specific interleukin-21⁺CD4⁺ T cells responses associated with viral control through the modulation of HCV-specific CD8⁺ T cells function in chronic hepatitis C patients. *Mol Cells* 2013;36:362-367.
[PUBMED](#) | [CROSSREF](#)
44. Hale JS, Youngblood B, Latner DR, Mohammed AU, Ye L, Akondy RS, Wu T, Iyer SS, Ahmed R. Distinct memory CD4⁺ T cells with commitment to T follicular helper- and T helper 1-cell lineages are generated after acute viral infection. *Immunity* 2013;38:805-817.
[PUBMED](#) | [CROSSREF](#)
45. Zander R, Kasmani MY, Chen Y, Topchyan P, Shen J, Zheng S, Burns R, Ingram J, Cui C, Joshi N, et al. Tfh-cell-derived interleukin 21 sustains effector CD8⁺ T cell responses during chronic viral infection. *Immunity* 2022;55:475-493.e5.
[PUBMED](#) | [CROSSREF](#)
46. Fahey LM, Wilson EB, Elsaesser H, Fistonich CD, McGavern DB, Brooks DG. Viral persistence redirects CD4 T cell differentiation toward T follicular helper cells. *J Exp Med* 2011;208:987-999.
[PUBMED](#) | [CROSSREF](#)
47. Harker JA, Lewis GM, Mack L, Zuniga EI. Late interleukin-6 escalates T follicular helper cell responses and controls a chronic viral infection. *Science* 2011;334:825-829.
[PUBMED](#) | [CROSSREF](#)
48. Crotty S. T follicular helper cell differentiation, function, and roles in disease. *Immunity* 2014;41:529-542.
[PUBMED](#) | [CROSSREF](#)
49. Crotty S. Follicular helper CD4 T cells (Tfh). *Annu Rev Immunol* 2011;29:621-663.
[PUBMED](#) | [CROSSREF](#)
50. Greczmiel U, Kräutler NJ, Pedrioli A, Bartsch I, Agnellini P, Bedenikovic G, Harker J, Richter K, Oxenius A. Sustained T follicular helper cell response is essential for control of chronic viral infection. *Sci Immunol* 2017;2:eaam8686.
[PUBMED](#) | [CROSSREF](#)
51. Cook KD, Shpargel KB, Starmer J, Whitfield-Larry F, Conley B, Allard DE, Rager JE, Fry RC, Davenport ML, Magnuson T, et al. T follicular helper cell-dependent clearance of a persistent virus infection requires T cell expression of the histone demethylase UTX. *Immunity* 2015;43:703-714.
[PUBMED](#) | [CROSSREF](#)
52. Linterman MA, Beaton L, Yu D, Ramiscal RR, Srivastava M, Hogan JJ, Verma NK, Smyth MJ, Rigby RJ, Vinuesa CG. IL-21 acts directly on B cells to regulate Bcl-6 expression and germinal center responses. *J Exp Med* 2010;207:353-363.
[PUBMED](#) | [CROSSREF](#)
53. Zotos D, Coquet JM, Zhang Y, Light A, D'Costa K, Kallies A, Corcoran LM, Godfrey DI, Toellner KM, Smyth MJ, et al. IL-21 regulates germinal center B cell differentiation and proliferation through a B cell-intrinsic mechanism. *J Exp Med* 2010;207:365-378.
[PUBMED](#) | [CROSSREF](#)
54. Rasheed MA, Latner DR, Aubert RD, Gourley T, Spolski R, Davis CW, Langley WA, Ha SJ, Ye L, Sarkar S, et al. Interleukin-21 is a critical cytokine for the generation of virus-specific long-lived plasma cells. *J Virol* 2013;87:7737-7746.
[PUBMED](#) | [CROSSREF](#)
55. Wherry EJ, Kurachi M. Molecular and cellular insights into T cell exhaustion. *Nat Rev Immunol* 2015;15:486-499.
[PUBMED](#) | [CROSSREF](#)
56. Utzschneider DT, Charmoy M, Chennupati V, Pousse L, Ferreira DP, Calderon-Copete S, Danilo M, Alfei F, Hofmann M, Wieland D, et al. T cell factor 1-expressing memory-like CD8⁺ T cells sustain the immune response to chronic viral infections. *Immunity* 2016;45:415-427.
[PUBMED](#) | [CROSSREF](#)
57. Snell LM, MacLeod BL, Law JC, Osokine I, Elsaesser HJ, Hezaveh K, Dickson RJ, Gavin MA, Guidos CJ, McGaha TL, et al. CD8⁺ T cell priming in established chronic viral infection preferentially directs differentiation of memory-like cells for sustained immunity. *Immunity* 2018;49:678-694.e5.
[PUBMED](#) | [CROSSREF](#)

58. Lin WW, Nish SA, Yen B, Chen YH, Adams WC, Kratchmarov R, Rothman NJ, Bhandoola A, Xue HH, Reiner SL. CD8⁺ T lymphocyte self-renewal during effector cell determination. *Cell Reports* 2016;17:1773-1782.
[PUBMED](#) | [CROSSREF](#)
59. He R, Hou S, Liu C, Zhang A, Bai Q, Han M, Yang Y, Wei G, Shen T, Yang X, et al. Follicular CXCR5-expressing CD8⁺ T cells curtail chronic viral infection. *Nature* 2016;537:412-428.
[PUBMED](#) | [CROSSREF](#)
60. Im SJ, Hashimoto M, Gerner MY, Lee J, Kissick HT, Burger MC, Shan Q, Hale JS, Lee J, Nasti TH, et al. Defining CD8⁺ T cells that provide the proliferative burst after PD-1 therapy. *Nature* 2016;537:417-421.
[PUBMED](#) | [CROSSREF](#)
61. Leong YA, Chen Y, Ong HS, Wu D, Man K, Deleage C, Minnich M, Meckiff BJ, Wei Y, Hou Z, et al. CXCR5⁺ follicular cytotoxic T cells control viral infection in B cell follicles. *Nat Immunol* 2016;17:1187-1196.
[PUBMED](#) | [CROSSREF](#)
62. Zander R, Schauder D, Xin G, Nguyen C, Wu X, Zajac A, Cui W. CD4⁺ T cell help is required for the formation of a cytolytic CD8⁺ T cell subset that protects against chronic infection and cancer. *Immunity* 2019;51:1028-1042.e4.
[PUBMED](#) | [CROSSREF](#)
63. Hudson WH, Gensheimer J, Hashimoto M, Wieland A, Valanparambil RM, Li P, Lin JX, Konieczny BT, Im SJ, Freeman GJ, et al. Proliferating transitory T cells with an effector-like transcriptional signature emerge from PD-1⁺ stem-like CD8⁺ T cells during chronic infection. *Immunity* 2019;51:1043-1058.e4.
[PUBMED](#) | [CROSSREF](#)
64. Chen Z, Ji Z, Ngiow SF, Manne S, Cai Z, Huang AC, Johnson J, Staup RP, Bengsch B, Xu C, et al. TCF-1-centered transcriptional network drives an effector versus exhausted CD8 T cell-fate decision. *Immunity* 2019;51:840-855.e5.
[PUBMED](#) | [CROSSREF](#)
65. Kanev K, Wu M, Drews A, Roelli P, Wurmser C, von Hösslin M, Zehn D. Proliferation-competent Tcf1⁺ CD8 T cells in dysfunctional populations are CD4 T cell help independent. *Proc Natl Acad Sci U S A* 2019;116:20070-20076.
[PUBMED](#) | [CROSSREF](#)
66. Miller BC, Sen DR, Al Abosy R, Bi K, Virkud YV, LaFleur MW, Yates KB, Lako A, Felt K, Naik GS, et al. Subsets of exhausted CD8⁺ T cells differentially mediate tumor control and respond to checkpoint blockade. *Nat Immunol* 2019;20:326-336.
[PUBMED](#) | [CROSSREF](#)
67. Beltra JC, Manne S, Abdel-Hakeem MS, Kurachi M, Giles JR, Chen Z, Casella V, Ngiow SF, Khan O, Huang YJ, et al. Developmental relationships of four exhausted CD8⁺ T cell subsets reveals underlying transcriptional and epigenetic landscape control mechanisms. *Immunity* 2020;52:825-841.e8.
[PUBMED](#) | [CROSSREF](#)
68. Zander R, Cui W. Exhausted CD8⁺ T cells face a developmental fork in the road. *Trends Immunol* 2023;44:276-286.
[PUBMED](#) | [CROSSREF](#)
69. Guo X, Zhang Y, Zheng L, Zheng C, Song J, Zhang Q, Kang B, Liu Z, Jin L, Xing R, et al. Global characterization of T cells in non-small-cell lung cancer by single-cell sequencing. *Nat Med* 2018;24:978-985.
[PUBMED](#) | [CROSSREF](#)
70. Seo N, Hayakawa S, Takigawa M, Tokura Y. Interleukin-10 expressed at early tumour sites induces subsequent generation of CD4⁺ T-regulatory cells and systemic collapse of antitumour immunity. *Immunology* 2001;103:449-457.
[PUBMED](#) | [CROSSREF](#)
71. Antony PA, Piccirillo CA, Akpınarlı A, Finkelstein SE, Speiss PJ, Surman DR, Palmer DC, Chan CC, Klebanoff CA, Overwijk WW, et al. CD8⁺ T cell immunity against a tumor/self-antigen is augmented by CD4⁺ T helper cells and hindered by naturally occurring T regulatory cells. *J Immunol* 2005;174:2591-2601.
[PUBMED](#) | [CROSSREF](#)
72. Tada Y, Togashi Y, Kotani D, Kuwata T, Sato E, Kawazoe A, Doi T, Wada H, Nishikawa H, Shitara K. Targeting VEGFR2 with ramucirumab strongly impacts effector/activated regulatory T cells and CD8⁺ T cells in the tumor microenvironment. *J Immunother Cancer* 2018;6:106.
[PUBMED](#) | [CROSSREF](#)
73. Kim JH, Kim BS, Lee SK. Regulatory T cells in tumor microenvironment and approach for anticancer immunotherapy. *Immune Netw* 2020;20:e4.
[PUBMED](#) | [CROSSREF](#)
74. Eberhardt CS, Kissick HT, Patel MR, Cardenas MA, Prokhnevskaya N, Obeng RC, Nasti TH, Griffith CC, Im SJ, Wang X, et al. Functional HPV-specific PD-1⁺ stem-like CD8 T cells in head and neck cancer. *Nature* 2021;597:279-284.
[PUBMED](#) | [CROSSREF](#)

75. Bauer CA, Kim EY, Marangoni F, Carrizosa E, Claudio NM, Mempel TR. Dynamic Treg interactions with intratumoral APCs promote local CTL dysfunction. *J Clin Invest* 2014;124:2425-2440.
[PUBMED](#) | [CROSSREF](#)
76. Joshi NS, Akama-Garren EH, Lu Y, Lee DY, Chang GP, Li A, DuPage M, Tammela T, Kerper NR, Farago AF, et al. Regulatory T cells in tumor-associated tertiary lymphoid structures suppress anti-tumor T cell responses. *Immunity* 2015;43:579-590.
[PUBMED](#) | [CROSSREF](#)
77. Johnson DB, Bordeaux J, Kim JY, Vaupel C, Rimm DL, Ho TH, Joseph RW, Daud AI, Conry RM, Gaughan EM, et al. Quantitative spatial profiling of PD-1/PD-L1 interaction and HLA-DR/IDO-1 predicts improved outcomes of anti-PD-1 therapies in metastatic melanoma. *Clin Cancer Res* 2018;24:5250-5260.
[PUBMED](#) | [CROSSREF](#)
78. Rodig SJ, Gusenleitner D, Jackson DG, Gjini E, Giobbie-Hurder A, Jin C, Chang H, Lovitch SB, Horak C, Weber JS, et al. MHC proteins confer differential sensitivity to CTLA-4 and PD-1 blockade in untreated metastatic melanoma. *Sci Transl Med* 2018;10:eaar3342.
[PUBMED](#) | [CROSSREF](#)
79. Roemer MG, Redd RA, Cader FZ, Pak CJ, Abdelrahman S, Ouyang J, Sasse S, Younes A, Fanale M, Santoro A, et al. Major histocompatibility complex class II and programmed death ligand 1 expression predict outcome after programmed death 1 blockade in classic hodgkin lymphoma. *J Clin Oncol* 2018;36:942-950.
[PUBMED](#) | [CROSSREF](#)
80. Park IA, Hwang SH, Song IH, Heo SH, Kim YA, Bang WS, Park HS, Lee M, Gong G, Lee HJ. Expression of the MHC class II in triple-negative breast cancer is associated with tumor-infiltrating lymphocytes and interferon signaling. *PLoS One* 2017;12:e0182786.
[PUBMED](#) | [CROSSREF](#)
81. Muranski P, Borman ZA, Kerkar SP, Klebanoff CA, Ji Y, Sanchez-Perez L, Sukumar M, Reger RN, Yu Z, Kern SJ, et al. Th17 cells are long lived and retain a stem cell-like molecular signature. *Immunity* 2011;35:972-985.
[PUBMED](#) | [CROSSREF](#)
82. Végran F, Apetoh L, Ghiringhelli F. Th9 cells: a novel CD4 T-cell subset in the immune war against cancer. *Cancer Res* 2015;75:475-479.
[PUBMED](#) | [CROSSREF](#)
83. Cachot A, Bilous M, Liu YC, Li X, Saillard M, Cenerenti M, Rockinger GA, Wyss T, Guillaume P, Schmidt J, et al. Tumor-specific cytolytic CD4 T cells mediate immunity against human cancer. *Sci Adv* 2021;7:eabe3348.
[PUBMED](#) | [CROSSREF](#)
84. Quezada SA, Simpson TR, Peggs KS, Merghoub T, Vider J, Fan X, Blasberg R, Yagita H, Muranski P, Antony PA, et al. Tumor-reactive CD4⁺ T cells develop cytotoxic activity and eradicate large established melanoma after transfer into lymphopenic hosts. *J Exp Med* 2010;207:637-650.
[PUBMED](#) | [CROSSREF](#)
85. Muranski P, Boni A, Antony PA, Cassard L, Irvine KR, Kaiser A, Paulos CM, Palmer DC, Touloukian CE, Ptak K, et al. Tumor-specific Th17-polarized cells eradicate large established melanoma. *Blood* 2008;112:362-373.
[PUBMED](#) | [CROSSREF](#)
86. Liotta F, Gacci M, Frosali F, Querci V, Vittori G, Lapini A, Santarlasci V, Serni S, Cosmi L, Maggi L, et al. Frequency of regulatory T cells in peripheral blood and in tumour-infiltrating lymphocytes correlates with poor prognosis in renal cell carcinoma. *BJU Int* 2011;107:1500-1506.
[PUBMED](#) | [CROSSREF](#)
87. Hanagiri T, Shigematsu Y, Shinohara S, Takenaka M, Oka S, Chikaishi Y, Nagata Y, Iwata T, Uramoto H, So T, et al. Clinical significance of the frequency of regulatory T cells in regional lymph node lymphocytes as a prognostic factor for non-small-cell lung cancer. *Lung Cancer* 2013;81:475-479.
[PUBMED](#) | [CROSSREF](#)
88. Ben Khelil M, Godet Y, Abdeljaoued S, Borg C, Adotévi O, Loyon R. Harnessing antitumor CD4⁺ T cells for cancer immunotherapy. *Cancers (Basel)* 2022;14:260.
[PUBMED](#) | [CROSSREF](#)
89. Schmidt A, Oberle N, Krammer PH. Molecular mechanisms of Treg-mediated T cell suppression. *Front Immunol* 2012;3:51.
[PUBMED](#) | [CROSSREF](#)
90. Lucca LE, Dominguez-Villar M. Modulation of regulatory T cell function and stability by co-inhibitory receptors. *Nat Rev Immunol* 2020;20:680-693.
[PUBMED](#) | [CROSSREF](#)

91. Kumar P, Saini S, Prabhakar BS. Cancer immunotherapy with check point inhibitor can cause autoimmune adverse events due to loss of Treg homeostasis. *Semin Cancer Biol* 2020;64:29-35.
[PUBMED](#) | [CROSSREF](#)
92. Kim MJ, Kim K, Park HJ, Kim GR, Hong KH, Oh JH, Son J, Park DJ, Kim D, Choi JM, et al. Deletion of PD-1 destabilizes the lineage identity and metabolic fitness of tumor-infiltrating regulatory T cells. *Nat Immunol* 2023;24:148-161.
[PUBMED](#) | [CROSSREF](#)
93. Knocke S, Fleischmann-Mundt B, Saborowski M, Manns MP, Kühnel F, Wirth TC, Woller N. Tailored tumor immunogenicity reveals regulation of CD4 and CD8 T cell responses against cancer. *Cell Reports* 2016;17:2234-2246.
[PUBMED](#) | [CROSSREF](#)
94. Marzo AL, Kinnear BF, Lake RA, Frelinger JJ, Collins EJ, Robinson BW, Scott B. Tumor-specific CD4⁺ T cells have a major “post-licensing” role in CTL mediated anti-tumor immunity. *J Immunol* 2000;165:6047-6055.
[PUBMED](#) | [CROSSREF](#)
95. Wong SB, Bos R, Sherman LA. Tumor-specific CD4⁺ T cells render the tumor environment permissive for infiltration by low-avidity CD8⁺ T cells. *J Immunol* 2008;180:3122-3131.
[PUBMED](#) | [CROSSREF](#)
96. Bos R, Sherman LA. CD4⁺ T-cell help in the tumor milieu is required for recruitment and cytolytic function of CD8⁺ T lymphocytes. *Cancer Res* 2010;70:8368-8377.
[PUBMED](#) | [CROSSREF](#)
97. Hashimoto M, Kamphorst AO, Im SJ, Kissick HT, Pillai RN, Ramalingam SS, Araki K, Ahmed R. CD8 T cell exhaustion in chronic infection and cancer: opportunities for interventions. *Annu Rev Med* 2018;69:301-318.
[PUBMED](#) | [CROSSREF](#)
98. Baitsch L, Baumgaertner P, Devèvre E, Raghav SK, Legat A, Barba L, Wieckowski S, Bouzourene H, Deplancke B, Romero P, et al. Exhaustion of tumor-specific CD8⁺ T cells in metastases from melanoma patients. *J Clin Invest* 2011;121:2350-2360.
[PUBMED](#) | [CROSSREF](#)
99. Pauken KE, Wherry EJ. Overcoming T cell exhaustion in infection and cancer. *Trends Immunol* 2015;36:265-276.
[PUBMED](#) | [CROSSREF](#)
100. Schietinger A, Delrow JJ, Basom RS, Blattman JN, Greenberg PD. Rescued tolerant CD8 T cells are preprogrammed to reestablish the tolerant state. *Science* 2012;335:723-727.
[PUBMED](#) | [CROSSREF](#)
101. Schietinger A, Greenberg PD. Tolerance and exhaustion: defining mechanisms of T cell dysfunction. *Trends Immunol* 2014;35:51-60.
[PUBMED](#) | [CROSSREF](#)
102. Pauken KE, Sammons MA, Odorizzi PM, Manne S, Godec J, Khan O, Drake AM, Chen Z, Sen DR, Kurachi M, et al. Epigenetic stability of exhausted T cells limits durability of reinvigoration by PD-1 blockade. *Science* 2016;354:1160-1165.
[PUBMED](#) | [CROSSREF](#)
103. Ribas A, Wolchok JD. Cancer immunotherapy using checkpoint blockade. *Science* 2018;359:1350-1355.
[PUBMED](#) | [CROSSREF](#)
104. Wei SC, Levine JH, Cogdill AP, Zhao Y, Anang NA, Andrews MC, Sharma P, Wang J, Wargo JA, Pe'er D, et al. Distinct cellular mechanisms underlie anti-CTLA-4 and anti-PD-1 checkpoint blockade. *Cell* 2017;170:1120-1133.e17.
[PUBMED](#) | [CROSSREF](#)
105. Ahmadzadeh M, Johnson LA, Heemskerk B, Wunderlich JR, Dudley ME, White DE, Rosenberg SA. Tumor antigen-specific CD8 T cells infiltrating the tumor express high levels of PD-1 and are functionally impaired. *Blood* 2009;114:1537-1544.
[PUBMED](#) | [CROSSREF](#)
106. Shin DS, Zaretsky JM, Escuin-Ordinas H, Garcia-Diaz A, Hu-Lieskovan S, Kalbasi A, Grasso CS, Hugo W, Sandoval S, Torrejon DY, et al. Primary resistance to PD-1 blockade mediated by JAK1/2 mutations. *Cancer Discov* 2017;7:188-201.
[PUBMED](#) | [CROSSREF](#)
107. Sen DR, Kaminski J, Barnitz RA, Kurachi M, Gerdemann U, Yates KB, Tsao HW, Godec J, LaFleur MW, Brown FD, et al. The epigenetic landscape of T cell exhaustion. *Science* 2016;354:1165-1169.
[PUBMED](#) | [CROSSREF](#)
108. Siddiqui I, Schaeuble K, Chennupati V, Fuertes Marraco SA, Calderon-Copete S, Pais Ferreira D, Carmona SJ, Scarpellino L, Gfeller D, Pradervand S, et al. Intratumoral Tcf1⁺PD-1⁺CD8⁺ T cells with stem-like

- properties promote tumor control in response to vaccination and checkpoint blockade immunotherapy. *Immunity* 2019;50:195-211.e10.
[PUBMED](#) | [CROSSREF](#)
109. Kurtulus S, Madi A, Escobar G, Klapholz M, Nyman J, Christian E, Pawlak M, Dionne D, Xia J, Rozenblatt-Rosen O, et al. Checkpoint blockade immunotherapy induces dynamic changes in PD-1⁺ CD8⁺ tumor-infiltrating T cells. *Immunity* 2019;50:181-194.e6.
[PUBMED](#) | [CROSSREF](#)
110. Alspach E, Lussier DM, Miceli AP, Kizhvatov I, DuPage M, Luoma AM, Meng W, Lichti CF, Esaulova E, Vomund AN, et al. MHC-II neoantigens shape tumour immunity and response to immunotherapy. *Nature* 2019;574:696-701.
[PUBMED](#) | [CROSSREF](#)
111. Kagamu H, Kitano S, Yamaguchi O, Yoshimura K, Horimoto K, Kitazawa M, Fukui K, Shiono A, Mouri A, Nishihara F, et al. CD4⁺ T-cell immunity in the peripheral blood correlates with response to anti-PD-1 therapy. *Cancer Immunol Res* 2020;8:334-344.
[PUBMED](#) | [CROSSREF](#)
112. Martens A, Wistuba-Hamprecht K, Yuan J, Postow MA, Wong P, Capone M, Madonna G, Khammari A, Schilling B, Sucker A, et al. Increases in absolute lymphocytes and circulating CD4⁺ and CD8⁺ T cells are associated with positive clinical outcome of melanoma patients treated with ipilimumab. *Clin Cancer Res* 2016;22:4848-4858.
[PUBMED](#) | [CROSSREF](#)
113. Zuazo M, Arasanz H, Fernández-Hinojal G, García-Granda MJ, Gato M, Bocanegra A, Martínez M, Hernández B, Teijeira L, Morilla I, et al. Functional systemic CD4 immunity is required for clinical responses to PD-L1/PD-1 blockade therapy. *EMBO Mol Med* 2019;11:e10293.
[PUBMED](#) | [CROSSREF](#)
114. Restifo NP, Dudley ME, Rosenberg SA. Adoptive immunotherapy for cancer: harnessing the T cell response. *Nat Rev Immunol* 2012;12:269-281.
[PUBMED](#) | [CROSSREF](#)
115. Kochenderfer JN, Wilson WH, Janik JE, Dudley ME, Stetler-Stevenson M, Feldman SA, Maric I, Raffeld M, Nathan DA, Lanier BJ, et al. Eradication of B-lineage cells and regression of lymphoma in a patient treated with autologous T cells genetically engineered to recognize CD19. *Blood* 2010;116:4099-4102.
[PUBMED](#) | [CROSSREF](#)
116. Brentjens RJ, Rivière I, Park JH, Davila ML, Wang X, Stefanski J, Taylor C, Yeh R, Bartido S, Borquez-Ojeda O, et al. Safety and persistence of adoptively transferred autologous CD19-targeted T cells in patients with relapsed or chemotherapy refractory B-cell leukemias. *Blood* 2011;118:4817-4828.
[PUBMED](#) | [CROSSREF](#)
117. Rosenberg SA, Yang JC, Sherry RM, Kammula US, Hughes MS, Phan GQ, Citrin DE, Restifo NP, Robbins PF, Wunderlich JR, et al. Durable complete responses in heavily pretreated patients with metastatic melanoma using T-cell transfer immunotherapy. *Clin Cancer Res* 2011;17:4550-4557.
[PUBMED](#) | [CROSSREF](#)
118. Porter DL, Levine BL, Kalos M, Bagg A, June CH. Chimeric antigen receptor-modified T cells in chronic lymphoid leukemia. *N Engl J Med* 2011;365:725-733.
[PUBMED](#) | [CROSSREF](#)
119. Robbins PF, Morgan RA, Feldman SA, Yang JC, Sherry RM, Dudley ME, Wunderlich JR, Nahvi AV, Helman LJ, Mackall CL, et al. Tumor regression in patients with metastatic synovial cell sarcoma and melanoma using genetically engineered lymphocytes reactive with NY-ESO-1. *J Clin Oncol* 2011;29:917-924.
[PUBMED](#) | [CROSSREF](#)
120. Rosenberg SA, Restifo NP. Adoptive cell transfer as personalized immunotherapy for human cancer. *Science* 2015;348:62-68.
[PUBMED](#) | [CROSSREF](#)
121. Tran E, Turcotte S, Gros A, Robbins PF, Lu YC, Dudley ME, Wunderlich JR, Somerville RP, Hogan K, Hinrichs CS, et al. Cancer immunotherapy based on mutation-specific CD4⁺ T cells in a patient with epithelial cancer. *Science* 2014;344:641-645.
[PUBMED](#) | [CROSSREF](#)
122. Atrash S, Bano K, Harrison B, Abdallah AO. CAR-T treatment for hematological malignancies. *J Investig Med* 2020;68:956-964.
[PUBMED](#) | [CROSSREF](#)
123. Braendstrup P, Levine BL, Ruella M. The long road to the first FDA-approved gene therapy: chimeric antigen receptor T cells targeting CD19. *Cytotherapy* 2020;22:57-69.
[PUBMED](#) | [CROSSREF](#)

124. Xu X, Sun Q, Liang X, Chen Z, Zhang X, Zhou X, Li M, Tu H, Liu Y, Tu S, et al. Mechanisms of relapse after CD19 CAR T-cell therapy for acute lymphoblastic leukemia and its prevention and treatment strategies. *Front Immunol* 2019;10:2664.
[PUBMED](#) | [CROSSREF](#)
125. Stock S, Schmitt M, Sellner L. Optimizing manufacturing protocols of chimeric antigen receptor T cells for improved anticancer immunotherapy. *Int J Mol Sci* 2019;20:6223.
[PUBMED](#) | [CROSSREF](#)
126. Turtle CJ, Hanafi LA, Berger C, Gooley TA, Cherian S, Hudecek M, Sommermeyer D, Melville K, Pender B, Budiarto TM, et al. CD19 CAR-T cells of defined CD4⁺:CD8⁺ composition in adult B cell ALL patients. *J Clin Invest* 2016;126:2123-2138.
[PUBMED](#) | [CROSSREF](#)
127. Moeller M, Haynes NM, Kershaw MH, Jackson JT, Teng MW, Street SE, Cerutti L, Jane SM, Trapani JA, Smyth MJ, et al. Adoptive transfer of gene-engineered CD4⁺ helper T cells induces potent primary and secondary tumor rejection. *Blood* 2005;106:2995-3003.
[PUBMED](#) | [CROSSREF](#)
128. Guedan S, Chen X, Madar A, Carpenito C, McGettigan SE, Frigault MJ, Lee J, Posey AD Jr, Scholler J, Scholler N, et al. ICOS-based chimeric antigen receptors program bipolar T_H17/T_H1 cells. *Blood* 2014;124:1070-1080.
[PUBMED](#) | [CROSSREF](#)
129. Melenhorst JJ, Chen GM, Wang M, Porter DL, Chen C, Collins MA, Gao P, Bandyopadhyay S, Sun H, Zhao Z, et al. Decade-long leukaemia remissions with persistence of CD4⁺ CAR T cells. *Nature* 2022;602:503-509.
[PUBMED](#) | [CROSSREF](#)
130. Ostroumov D, Fekete-Drimusz N, Saborowski M, Kühnel F, Woller N. CD4 and CD8 T lymphocyte interplay in controlling tumor growth. *Cell Mol Life Sci* 2018;75:689-713.
[PUBMED](#) | [CROSSREF](#)
131. Ahrends T, Spanjaard A, Pilzecker B, Bąbała N, Bovens A, Xiao Y, Jacobs H, Borst J. CD4⁺ T cell help confers a cytotoxic T cell effector program including coinhibitory receptor downregulation and increased tissue invasiveness. *Immunity* 2017;47:848-861.e5.
[PUBMED](#) | [CROSSREF](#)
132. Muranski P, Restifo NP. Adoptive immunotherapy of cancer using CD4⁺ T cells. *Curr Opin Immunol* 2009;21:200-208.
[PUBMED](#) | [CROSSREF](#)
133. Hunder NN, Wallen H, Cao J, Hendricks DW, Reilly JZ, Rodmyre R, Jungbluth A, Gnjjatic S, Thompson JA, Yee C. Treatment of metastatic melanoma with autologous CD4⁺ T cells against NY-ESO-1. *N Engl J Med* 2008;358:2698-2703.
[PUBMED](#) | [CROSSREF](#)
134. Xie Y, Akpınarli A, Maris C, Hipkiss EL, Lane M, Kwon EK, Muranski P, Restifo NP, Antony PA. Naive tumor-specific CD4⁺ T cells differentiated *in vivo* eradicate established melanoma. *J Exp Med* 2010;207:651-667.
[PUBMED](#) | [CROSSREF](#)
135. Lu Y, Hong S, Li H, Park J, Hong B, Wang L, Zheng Y, Liu Z, Xu J, He J, et al. Th9 cells promote antitumor immune responses *in vivo*. *J Clin Invest* 2012;122:4160-4171.
[PUBMED](#) | [CROSSREF](#)
136. Végran F, Berger H, Boidot R, Mignot G, Bruchard M, Dosset M, Chalmin F, Rébé C, Dérangère V, Ryffel B, et al. The transcription factor IRF1 dictates the IL-21-dependent anticancer functions of T_H9 cells. *Nat Immunol* 2014;15:758-766.
[PUBMED](#) | [CROSSREF](#)
137. Lee J, Lozano-Ruiz B, Yang FM, Fan DD, Shen L, González-Navajas JM. The multifaceted role of Th1, Th9, and Th17 cells in immune checkpoint inhibition therapy. *Front Immunol* 2021;12:625667.
[PUBMED](#) | [CROSSREF](#)
138. Leonard WJ, Wan CK. IL-21 signaling in immunity. *F1000 Res* 2016;5:224.
[PUBMED](#) | [CROSSREF](#)
139. Hinrichs CS, Spolski R, Paulos CM, Gattinoni L, Kerstann KW, Palmer DC, Klebanoff CA, Rosenberg SA, Leonard WJ, Restifo NP. IL-2 and IL-21 confer opposing differentiation programs to CD8⁺ T cells for adoptive immunotherapy. *Blood* 2008;111:5326-5333.
[PUBMED](#) | [CROSSREF](#)
140. Spolski R, Leonard WJ. Interleukin-21: a double-edged sword with therapeutic potential. *Nat Rev Drug Discov* 2014;13:379-395.
[PUBMED](#) | [CROSSREF](#)

141. Santegoets SJ, Turksma AW, Suhoski MM, Stam AG, Albelda SM, Hooijberg E, Scheper RJ, van den Eertwegh AJ, Gerritsen WR, Powell DJ Jr, et al. IL-21 promotes the expansion of CD27⁺ CD28⁺ tumor infiltrating lymphocytes with high cytotoxic potential and low collateral expansion of regulatory T cells. *J Transl Med* 2013;11:37.
[PUBMED](#) | [CROSSREF](#)
142. Xin G, Schauder DM, Lainez B, Weinstein JS, Dai Z, Chen Y, Esplugues E, Wen R, Wang D, Parish IA, et al. A critical role of IL-21-induced BATF in sustaining CD8-T-cell-mediated chronic viral control. *Cell Reports* 2015;13:1118-1124.
[PUBMED](#) | [CROSSREF](#)
143. Topchyan P, Xin G, Chen Y, Zheng S, Burns R, Shen J, Kasmani MY, Kudek M, Yang N, Cui W. Harnessing the IL-21-BATF pathway in the CD8⁺ T cell anti-tumor response. *Cancers (Basel)* 2021;13:1263.
[PUBMED](#) | [CROSSREF](#)
144. Gaffen SL, Jain R, Garg AV, Cua DJ. The IL-23-IL-17 immune axis: from mechanisms to therapeutic testing. *Nat Rev Immunol* 2014;14:585-600.
[PUBMED](#) | [CROSSREF](#)
145. Kurachi M, Barnitz RA, Yosef N, Odorizzi PM, DiIorio MA, Lemieux ME, Yates K, Godec J, Klatt MG, Regev A, et al. The transcription factor BATF operates as an essential differentiation checkpoint in early effector CD8⁺ T cells. *Nat Immunol* 2014;15:373-383.
[PUBMED](#) | [CROSSREF](#)
146. Ciofani M, Madar A, Galan C, Sellars M, Mace K, Pauli F, Agarwal A, Huang W, Parkhurst CN, Muratet M, et al. A validated regulatory network for Th17 cell specification. *Cell* 2012;151:289-303.
[PUBMED](#) | [CROSSREF](#)
147. Betz BC, Jordan-Williams KL, Wang C, Kang SG, Liao J, Logan MR, Kim CH, Taparowsky EJ. Batf coordinates multiple aspects of B and T cell function required for normal antibody responses. *J Exp Med* 2010;207:933-942.
[PUBMED](#) | [CROSSREF](#)
148. Ise W, Kohyama M, Schraml BU, Zhang T, Schwer B, Basu U, Alt FW, Tang J, Oltz EM, Murphy TL, et al. The transcription factor BATF controls the global regulators of class-switch recombination in both B cells and T cells. *Nat Immunol* 2011;12:536-543.
[PUBMED](#) | [CROSSREF](#)
149. Tussiwand R, Lee WL, Murphy TL, Mashayekhi M, Kc W, Albring JC, Satpathy AT, Rotondo JA, Edelson BT, Kretzer NM, et al. Compensatory dendritic cell development mediated by BATF-IRF interactions. *Nature* 2012;490:502-507.
[PUBMED](#) | [CROSSREF](#)
150. Jabeen R, Goswami R, Awe O, Kulkarni A, Nguyen ET, Attenasio A, Walsh D, Olson MR, Kim MH, Tepper RS, et al. Th9 cell development requires a BATF-regulated transcriptional network. *J Clin Invest* 2013;123:4641-4653.
[PUBMED](#) | [CROSSREF](#)
151. Sahoo A, Alekseev A, Tanaka K, Obertas L, Lerman B, Haymaker C, Clise-Dwyer K, McMurray JS, Nurieva R. Batf is important for IL-4 expression in T follicular helper cells. *Nat Commun* 2015;6:7997.
[PUBMED](#) | [CROSSREF](#)
152. Schraml BU, Hildner K, Ise W, Lee WL, Smith WA, Solomon B, Sahota G, Sim J, Mukasa R, Cemerski S, et al. The AP-1 transcription factor BATF controls T_H17 differentiation. *Nature* 2009;460:405-409.
[PUBMED](#) | [CROSSREF](#)
153. Chen Y, Zander RA, Wu X, Schauder DM, Kasmani MY, Shen J, Zheng S, Burns R, Taparowsky EJ, Cui W. BATF regulates progenitor to cytolytic effector CD8⁺ T cell transition during chronic viral infection. *Nat Immunol* 2021;22:996-1007.
[PUBMED](#) | [CROSSREF](#)
154. Seo H, González-Avalos E, Zhang W, Ramchandani P, Yang C, Lio CJ, Rao A, Hogan PG. BATF and IRF4 cooperate to counter exhaustion in tumor-infiltrating CAR T cells. *Nat Immunol* 2021;22:983-995.
[PUBMED](#) | [CROSSREF](#)
155. Topchyan P, Zander R, Kasmani MY, Nguyen C, Brown A, Lin S, Burns R, Cui W. Spatial transcriptomics demonstrates the role of CD4 T cells in effector CD8 T cell differentiation during chronic viral infection. *Cell Reports* 2022;41:111736.
[PUBMED](#) | [CROSSREF](#)
156. Cui C, Wang J, Fagerberg E, Chen PM, Connolly KA, Damo M, Cheung JF, Mao T, Askari AS, Chen S, et al. Neoantigen-driven B cell and CD4 T follicular helper cell collaboration promotes anti-tumor CD8 T cell responses. *Cell* 2021;184:6101-6118.e13.
[PUBMED](#) | [CROSSREF](#)

157. Jansen CS, Prokhnevskaya N, Master VA, Sanda MG, Carlisle JW, Bilen MA, Cardenas M, Wilkinson S, Lake R, Sowalsky AG, et al. An intra-tumoral niche maintains and differentiates stem-like CD8 T cells. *Nature* 2019;576:465-470.
[PUBMED](#) | [CROSSREF](#)
158. Schumacher TN, Thommen DS. Tertiary lymphoid structures in cancer. *Science* 2022;375:eabf9419.
[PUBMED](#) | [CROSSREF](#)
159. Bindea G, Mlecnik B, Tosolini M, Kirilovsky A, Waldner M, Obenauf AC, Angell H, Fredriksen T, Lafontaine L, Berger A, et al. Spatiotemporal dynamics of intratumoral immune cells reveal the immune landscape in human cancer. *Immunity* 2013;39:782-795.
[PUBMED](#) | [CROSSREF](#)
160. Cillo AR, Kürten CH, Tabib T, Qi Z, Onkar S, Wang T, Liu A, Duvvuri U, Kim S, Soose RJ, et al. Immune landscape of viral- and carcinogen-driven head and neck cancer. *Immunity* 2020;52:183-199.e9.
[PUBMED](#) | [CROSSREF](#)
161. Gu-Trantien C, Loi S, Garaud S, Equeter C, Libin M, de Wind A, Ravoet M, Le Buanec H, Sibille C, Manfouo-Foutsop G, et al. CD4⁺ follicular helper T cell infiltration predicts breast cancer survival. *J Clin Invest* 2013;123:2873-2892.
[PUBMED](#) | [CROSSREF](#)
162. Garaud S, Buisseret L, Solinas C, Gu-Trantien C, de Wind A, Van den Eynden G, Naveaux C, Lodewyckx JN, Boisson A, Duvillier H, et al. Tumor infiltrating B-cells signal functional humoral immune responses in breast cancer. *JCI Insight* 2019;5:e129641.
[PUBMED](#) | [CROSSREF](#)
163. Germain C, Gnjatic S, Tamzalit F, Knockaert S, Remark R, Goc J, Lepelley A, Becht E, Katsahian S, Bizouard G, et al. Presence of B cells in tertiary lymphoid structures is associated with a protective immunity in patients with lung cancer. *Am J Respir Crit Care Med* 2014;189:832-844.
[PUBMED](#) | [CROSSREF](#)
164. Griss J, Bauer W, Wagner C, Simon M, Chen M, Grabmeier-Pfistershammer K, Maurer-Granofszky M, Roka F, Penz T, Bock C, et al. B cells sustain inflammation and predict response to immune checkpoint blockade in human melanoma. *Nat Commun* 2019;10:4186.
[PUBMED](#) | [CROSSREF](#)
165. Helmink BA, Reddy SM, Gao J, Zhang S, Basar R, Thakur R, Yizhak K, Sade-Feldman M, Blando J, Han G, et al. B cells and tertiary lymphoid structures promote immunotherapy response. *Nature* 2020;577:549-555.
[PUBMED](#) | [CROSSREF](#)
166. Hennequin A, Derangère V, Boidot R, Apetoh L, Vincent J, Orry D, Fraisse J, Causeret S, Martin F, Arnould L, et al. Tumor infiltration by Tbet⁺ effector T cells and CD20⁺ B cells is associated with survival in gastric cancer patients. *Oncotmunology* 2015;5:e1054598.
[PUBMED](#) | [CROSSREF](#)
167. Kroeger DR, Milne K, Nelson BH. Tumor-infiltrating plasma cells are associated with tertiary lymphoid structures, cytolytic T-cell responses, and superior prognosis in ovarian cancer. *Clin Cancer Res* 2016;22:3005-3015.
[PUBMED](#) | [CROSSREF](#)
168. Petitprez F, de Reyniès A, Keung EZ, Chen TW, Sun CM, Calderaro J, Jeng YM, Hsiao LP, Lacroix L, Bougouin A, et al. B cells are associated with survival and immunotherapy response in sarcoma. *Nature* 2020;577:556-560.
[PUBMED](#) | [CROSSREF](#)
169. Truxova I, Kasikova L, Hensler M, Skapa P, Laco J, Pecen L, Belicova L, Praznovec I, Halaska MJ, Brtnicky T, et al. Mature dendritic cells correlate with favorable immune infiltrate and improved prognosis in ovarian carcinoma patients. *J Immunother Cancer* 2018;6:139.
[PUBMED](#) | [CROSSREF](#)
170. Wieland A, Patel MR, Cardenas MA, Eberhardt CS, Hudson WH, Obeng RC, Griffith CC, Wang X, Chen ZG, Kissick HT, et al. Defining HPV-specific B cell responses in patients with head and neck cancer. *Nature* 2021;597:274-278.
[PUBMED](#) | [CROSSREF](#)
171. Ruffin AT, Cillo AR, Tabib T, Liu A, Onkar S, Kunning SR, Lampenfeld C, Atiya HI, Abecassis I, Kürten CH, et al. B cell signatures and tertiary lymphoid structures contribute to outcome in head and neck squamous cell carcinoma. *Nat Commun* 2021;12:3349.
[PUBMED](#) | [CROSSREF](#)
172. Sharonov GV, Serebrovskaya EO, Yuzhakova DV, Britanova OV, Chudakov DM. B cells, plasma cells and antibody repertoires in the tumour microenvironment. *Nat Rev Immunol* 2020;20:294-307.
[PUBMED](#) | [CROSSREF](#)

173. Sautès-Fridman C, Lawand M, Giraldo NA, Kaplon H, Germain C, Fridman WH, Dieu-Nosjean MC. Tertiary lymphoid structures in cancers: prognostic value, regulation, and manipulation for therapeutic intervention. *Front Immunol* 2016;7:407.
[PUBMED](#) | [CROSSREF](#)
174. Cho JW, Park S, Kim G, Han H, Shim HS, Shin S, Bae YS, Park SY, Ha SJ, Lee I, et al. Dysregulation of T_{HH}-B-T_{RM} lymphocyte cooperation is associated with unfavorable anti-PD-1 responses in EGFR-mutant lung cancer. *Nat Commun* 2021;12:6068.
[PUBMED](#) | [CROSSREF](#)
175. Luo S, Zhu R, Yu T, Fan H, Hu Y, Mohanta SK, Hu D. Chronic inflammation: a common promoter in tertiary lymphoid organ neogenesis. *Front Immunol* 2019;10:2938.
[PUBMED](#) | [CROSSREF](#)
176. Pitzalis C, Jones GW, Bombardieri M, Jones SA. Ectopic lymphoid-like structures in infection, cancer and autoimmunity. *Nat Rev Immunol* 2014;14:447-462.
[PUBMED](#) | [CROSSREF](#)
177. Manzo A, Bombardieri M, Humby F, Pitzalis C. Secondary and ectopic lymphoid tissue responses in rheumatoid arthritis: from inflammation to autoimmunity and tissue damage/remodeling. *Immunol Rev* 2010;233:267-285.
[PUBMED](#) | [CROSSREF](#)
178. Pipi E, Nayar S, Gardner DH, Colafrancesco S, Smith C, Barone F. Tertiary lymphoid structures: autoimmunity goes local. *Front Immunol* 2018;9:1952.
[PUBMED](#) | [CROSSREF](#)
179. Korpos É, Kadri N, Loismann S, Findeisen CR, Arfuso F, Burke GW 3rd, Richardson SJ, Morgan NG, Bogdani M, Pugliese A, et al. Identification and characterisation of tertiary lymphoid organs in human type 1 diabetes. *Diabetologia* 2021;64:1626-1641.
[PUBMED](#) | [CROSSREF](#)
180. de Chaisemartin L, Goc J, Damotte D, Validire P, Magdeleinat P, Alifano M, Cremer I, Fridman WH, Sautès-Fridman C, Dieu-Nosjean MC. Characterization of chemokines and adhesion molecules associated with T cell presence in tertiary lymphoid structures in human lung cancer. *Cancer Res* 2011;71:6391-6399.
[PUBMED](#) | [CROSSREF](#)
181. Dieu-Nosjean MC, Antoine M, Danel C, Heudes D, Wislez M, Poulot V, Rabbe N, Laurans L, Tartour E, de Chaisemartin L, et al. Long-term survival for patients with non-small-cell lung cancer with intratumoral lymphoid structures. *J Clin Oncol* 2008;26:4410-4417.
[PUBMED](#) | [CROSSREF](#)
182. Bergomas F, Grizzi F, Doni A, Pesce S, Laghi L, Allavena P, Mantovani A, Marchesi F. Tertiary intratumor lymphoid tissue in colo-rectal cancer. *Cancers (Basel)* 2011;4:1-10.
[PUBMED](#) | [CROSSREF](#)
183. Coppola D, Nebozhyn M, Khalil F, Dai H, Yeatman T, Loboda A, Mulé JJ. Unique ectopic lymph node-like structures present in human primary colorectal carcinoma are identified by immune gene array profiling. *Am J Pathol* 2011;179:37-45.
[PUBMED](#) | [CROSSREF](#)
184. Cabrita R, Lauss M, Sanna A, Donia M, Skaarup Larsen M, Mitra S, Johansson I, Phung B, Harbst K, Vallon-Christersson J, et al. Tertiary lymphoid structures improve immunotherapy and survival in melanoma. *Nature* 2020;577:561-565.
[PUBMED](#) | [CROSSREF](#)
185. van Dijk N, Gil-Jimenez A, Silina K, Hendricksen K, Smit LA, de Feijter JM, van Montfoort ML, van Rooijen C, Peters D, Broeks A, et al. Preoperative ipilimumab plus nivolumab in locoregionally advanced urothelial cancer: the NABUCCO trial. *Nat Med* 2020;26:1839-1844.
[PUBMED](#) | [CROSSREF](#)
186. Siliņa K, Soltermann A, Attar FM, Casanova R, Uckelely ZM, Thut H, Wandres M, Isajevs S, Cheng P, Curioni-Fontecedro A, et al. Germinal centers determine the prognostic relevance of tertiary lymphoid structures and are impaired by corticosteroids in lung squamous cell carcinoma. *Cancer Res* 2018;78:1308-1320.
[PUBMED](#) | [CROSSREF](#)
187. Cottrell TR, Thompson ED, Forde PM, Stein JE, Duffield AS, Anagnostou V, Rekhman N, Anders RA, Cuda JD, Illei PB, et al. Pathologic features of response to neoadjuvant anti-PD-1 in resected non-small-cell lung carcinoma: a proposal for quantitative immune-related pathologic response criteria (irPRC). *Ann Oncol* 2018;29:1853-1860.
[PUBMED](#) | [CROSSREF](#)
188. Thommen DS, Koelzer VH, Herzig P, Roller A, Trefny M, Dimeloe S, Kiiäläinen A, Hanhart J, Schill C, Hess C, et al. A transcriptionally and functionally distinct PD-1⁺ CD8⁺ T cell pool with predictive potential in non-small-cell lung cancer treated with PD-1 blockade. *Nat Med* 2018;24:994-1004.
[PUBMED](#) | [CROSSREF](#)

189. Johansson-Percival A, He B, Li ZJ, Kjellén A, Russell K, Li J, Larma I, Ganss R. De novo induction of intratumoral lymphoid structures and vessel normalization enhances immunotherapy in resistant tumors. *Nat Immunol* 2017;18:1207-1217.
[PUBMED](#) | [CROSSREF](#)
190. Allen E, Jabouille A, Rivera LB, Lodewijckx I, Missiaen R, Steri V, Feyen K, Tawney J, Hanahan D, Michael IP, et al. Combined antiangiogenic and anti-PD-L1 therapy stimulates tumor immunity through HEV formation. *Sci Transl Med* 2017;9:eaak9679.
[PUBMED](#) | [CROSSREF](#)
191. Overacre-Delgoffe AE, Bumgarner HJ, Cillo AR, Burr AH, Tometch JT, Bhattacharjee A, Bruno TC, Vignali DA, Hand TW. Microbiota-specific T follicular helper cells drive tertiary lymphoid structures and anti-tumor immunity against colorectal cancer. *Immunity* 2021;54:2812-2824.e4.
[PUBMED](#) | [CROSSREF](#)
192. Lutz ER, Wu AA, Bigelow E, Sharma R, Mo G, Soares K, Solt S, Dorman A, Wamwea A, Yager A, et al. Immunotherapy converts nonimmunogenic pancreatic tumors into immunogenic foci of immune regulation. *Cancer Immunol Res* 2014;2:616-631.
[PUBMED](#) | [CROSSREF](#)