Review Article

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Extracellular Mechanisms of Neutrophils in Immune Cell Crosstalk

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OPEN ACCESS

Received: Jul 19, 2023 Revised: Aug 24, 2023 Accepted: Sep 5, 2023 Published online: Sep 22, 2023

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Conflict of Interest

The authors declare no potential conflicts of interest.

Abbreviations

APRIL, a proliferation-inducing ligand; ARDS, acute respiratory distress syndrome; BAFF, B-cell activating factor; BCG, Bacillus Calmette-Guérin; DC, dendritic cell; EV, extracellular vesicle; G-MDSC, Department of Physiology, CMRI, School of Medicine, Kyungpook National University, Daegu 41944, Korea

ABSTRACT

Neutrophils are professional phagocytes that provide defense against invading pathogens through phagocytosis, degranulation, generation of ROS, and the formation of neutrophil extracellular traps (NETs). Although long been considered as short-lived effector cells with limited biosynthetic activity, recent studies have revealed that neutrophils actively communicate with other immune cells. Neutrophils employ various types of soluble mediators, including granules, cytokines, and chemokines, for crosstalk with immune cells. Additionally, ROS and NETs, major arsenals of neutrophils, are utilized for intercellular communication. Furthermore, extracellular vesicles play a crucial role as mediators of neutrophil crosstalk. In this review, we highlight the extracellular mechanisms of neutrophils and their roles in crosstalk with other cells.

Keywords: Neutrophils; Neutrophil extracellular trap; Extracellular vesicles; Immune cell crosstalk

INTRODUCTION

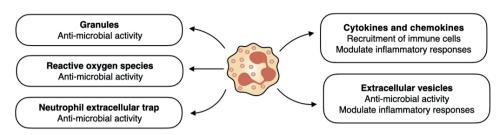
Neutrophils are one of the first responders of immune system and have long been considered as short-lived effector cells with limited biosynthetic activity (1,2). They provide defense against invading pathogens through phagocytosis, degranulation, generation of ROS, and the formation of neutrophil extracellular trap (NET) (3). In addition to these defense mechanisms, neutrophils have been shown to have additional biological functions, such as the release of soluble factors, cytokines, chemokines, growth factors, alarmins, and specialized pro-resolving mediators (4-7). They also generate extracellular vesicles (EVs) that exert either proinflammatory or anti-inflammatory responses (8,9). Recent studies have also shown that neutrophils play a role in antigen (Ag) presentation by releasing immunomodulatory molecules (4,6,7). This suggests that neutrophils actively communicate with other innate and adaptive immune cells rather just exerting their phagocytic role (7,10).

Neutrophils are known to indirectly regulate adaptive immune cells by regulating innate immune cells, such as macrophages or dendritic cells (DCs). However, recent studies

granulocytic myeloid derived suppressor cell; IRFI, interferon regulatory factor 1; MPO, myeloperoxidase; Mtb, *Mycobacterium tuberculosis*; NDEV, neutrophil-derived extracellular vesicle; NE, neutrophil elastase; NET, neutrophil extracellular trap; TAN, tumorassociated neutrophil; pDC, plasmacytoid dendritic cell.

Author Contributions

Conceptualization: Shrestha S, Hong CW; Funding acquisition: Hong CW; Supervision: Hong CW; Visualization: Shrestha S, Hong CW; Writing - original draft: Shrestha S, Hong CW; Writing - review & editing: Hong CW.



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Figure 1. Overview of neutrophil interactions and immune responses. As the first responders of immune system, neutrophils employ a variety of defense mechanisms against pathogens. These mechanisms include granule release, production of ROS, formation of NETs, and secretion of cytokines, chemokines, and EVs. These arsenals not only facilitate the elimination of pathogens, but also serve as means for neutrophils to communicate with neighboring immune cells.

have shown that they can also directly interact with adaptive immune cells through Ag presentation (6). Neutrophils can populate in secondary lymphoid tissues, such as spleen and lymph nodes, and present Ags to adaptive immune cells through expression of MHC class II (6,7). They also interact with other immune cells through contact-dependent mechanisms and soluble mediators, leading to recruitment, activation, and maturation of macrophages, DCs, NK cells, B cells, and T cells (2, 9,11-16).

Neutrophils also generate EVs that mediate immune modulation and cell-to-cell communications with both immune and non-immune cells (8,9). Neutrophils thus have emerged as key components of cell crosstalk rather than merely bystander of complex immune responses (**Fig. 1**). In this review, we highlight the extracellular arsenals of neutrophils, such as soluble mediators, NETs, and EVs, and their role in crosstalk with other cells.

NEUTROPHIL-DERIVED SOLUBLE MEDIATORS

Neutrophils are equipped with various types of soluble mediators, such as granules, cytokines, and chemokines. During differentiation, neutrophils produce different types of granule proteins which are stored within intracellular pools (17). The production of these granules is stringently controlled across different stages of differentiation (1,18). Notably, neutrophils rapidly release these pre-formed granules in response to external stimuli (18). Neutrophils serve as a significant source for over 70 distinct cytokines, chemokines, and growth factors (17). Neutrophils not only store pre-formed cytokines, but also synthesize cytokines *de novo*. Mature neutrophils have mRNAs for cytokine (17), hence they *de novo* synthesize cytokines in response to external stimulation such as TLR 8 agonists (19). These soluble mediators are used by neutrophils for intercellular communication (**Fig. 2**).

Neutrophils are equipped with various types of granules, which are categorized into four groups: i) primary(azurophil) granules, containing myeloperoxidase (MPO), neutrophil elastase (NE), cathepsin G, proteinase 3, defensin, lysozyme, azurocidin, arginase, and serine proteases; ii) secondary(specific) granules, containing lysozyme, lactoferrin, nicotinamide adenine dinucleotide phosphate oxidase components, cathelicidin, collagenase, and plasminogen activator inhibitor-1; iii) tertiary granules, containing gelatinase, leukolysin, collagenase, cathepsin D, and neutrophil gelatinase-associated lipocalin; and iv) secretory vesicles, containing complement receptors, Fc receptors, chemokine receptors, adhesion molecules, cytokines, chemokines, and growth factors (1). Neutrophils release these granule proteins to extracellular milieu through exocytosis (1,17). Neutrophils eliminate pathogens



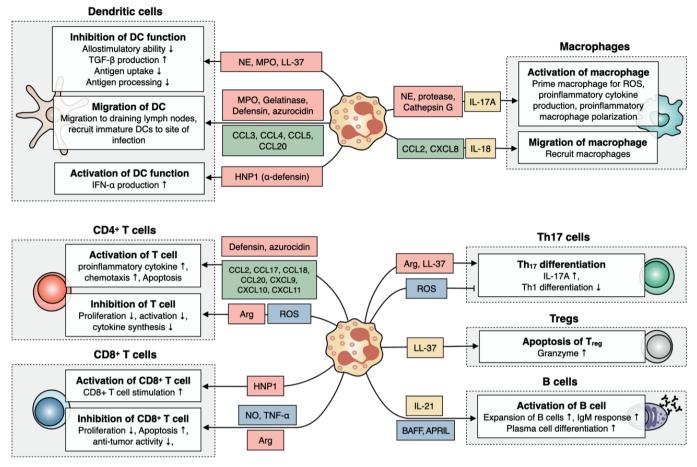


Figure 2. Intercellular crosstalk by neutrophils through soluble mediators. Neutrophils employ various soluble mediators, including granules, cytokines, chemokines, and ROS, for intercellular communication with neighboring immune cells. Neutrophils activate macrophages through IL-17A, protease, NE, and cathepsin G, and induce macrophage migration via CCL2, CXCL8, and IL-18. While neutrophils suppress DC functions through NEs, MPO, and LL-37, they enhance DC activities via HNP1. Granules and chemokines released by neutrophils stimulate DC migration. Neutrophils also communicate with adaptive immune cells through soluble mediators. Neutrophils inhibit T cell function through Arg, and ROS, and promote differentiation into Th17 cells via Arg, LL-37, and ROS. Certain granule proteins, such as defensin and azurocidin, induce proinflammatory cytokine production and apoptosis in CD4⁺ T cells, while chemokines induce chemotaxis of CD4⁺ T cells. LL-37 triggers apoptosis in Tregs. Arg, NO, and TNF-a inhibit functions of CD8⁺ T cells through suppression of proliferation and induction of apoptosis, resulting in the inhibition of anti-tumor activities of CD8⁺ T cells. HNP1 rather enhances the function of CD8⁺ T cell. Neutrophils also communicate with B cells via BAFF, APRIL, and IL-21.

HNP1, human neutrophil peptide 1; Arg, arginase; NO, nitric oxide.

through antimicrobial activity of these granules and also utilize them for transmigration through tissue destruction. However, these granules also allow intercellular communications by neutrophils.

NE induces differentiation of macrophages into a pro-inflammatory phenotype through activation of the Srk kinase family (20) and attenuates allostimulatory ability of DCs through skewing maturation of DCs into TGF- β secreting cells (21). MPO secreted by neutrophils inhibits DC activation through its catalytic activity, suppressing adaptive functions of DCs such as migration, Ag uptake, and Ag processing (22). In contrast, MPO and gelatinase are essential for DC migration into draining lymph nodes and subsequent T cell priming in contact hypersensitivity (23). Cathepsin G derived from neutrophils induce chemotaxis of macrophages and activates them to produce pro-inflammatory cytokines, resulting in increased susceptibility to acute HIV-1 infection (24). Human neutrophil peptide 1 (α -defensin) stimulates DCs to produce IFN- α by activating and promoting the nuclear

translocation of IFN regulatory factor 1 (IRF1) (25). Indeed, defensin triggers the secretion of proinflammatory cytokines such as IFN- γ , IL-2, and IL-8 from T cells through NF- κ B activation, and also induce T cell apoptosis (26). Defensin and azurocidin induce chemotaxis and regulate functions of naïve T cells and DCs (27,28). Proteases released from neutrophils prime macrophages for more effective oxidative response against microorganism and tumor cells (29). Arginase released from neutrophils suppresses the proliferation, activation, and cytokine synthesis in T cells (30,31) and promotes Th17 cell differentiation (32). Arginase generated from granulocytic myeloid derived suppressor cells (G-MDSCs) suppress proliferation of both CD4⁺ and CD8⁺ T cells (33). LL-37, a member of the cathelicidin family of antimicrobial peptides, inhibits DC maturation and activation triggered by TLR ligands, resulting in a reduced ability to activate T cells (34). LL-37 has a broader role beyond its effect on DCs. LL-37 suppresses the differentiation of Th1 cells while promoting IL-17A production from T cells by enhancing aryl hydrocarbon receptor and ROR γ t expression in a TGF- β dependent manner (35). Moreover, LL-37 induces granzyme-mediated apoptosis in Tregs and stimulated CD8⁺ T cells (36,37).

Neutrophils are also major sources of chemokines. Activated neutrophils release CXCL1, CXCL8 (also known as IL-8), CXCL9 (monokine induced by gamma IFN), CXCL10 (IFN gamma-induced protein 10), CXCL11 (IFN-inducible T-cell alpha chemoattractant), CXCL12 (stromal cell-derived factor 1), CXCL13 (B cell-attracting chemokine 1), CCL2 (MCP-1), CCL3 (MIP-1 α), CCL4 (MIP-1 β), CCL17 (MIP-3 β), and CCL20 (MIP-3 α). These neutrophil-derived chemokines orchestrate the recruitment of various immune cells, including monocytes, macrophages, DCs, NK cells, various T cell subsets, resulting in an amplification of immune responses (27). Neutrophils release CCL2, recruiting macrophages and enhancing the replication of HIV (38). Neutrophils synthesize chemokines *de novo* and recruit immature DCs to the site of infection via CCL3, CCL4, CCL5, and CCL20 during *Toxoplasma gondii* infection (39). Bacillus Calmette-Guérin (BCG)-activated neutrophils produce chemokines that promote T cell chemotaxis, thereby enhancing the effectiveness of BCG immunotherapy (40). Furthermore, activated neutrophils attract various T cell subsets, including Th1, Th17, naïve T cells and CD8⁺ T cells, to inflammation sites through the release of chemokines such as CCL2, CXCL9, CXCL10, CXCL11, CCL17, CCL18, and CCL20 (27).

In addition to chemokines, neutrophils produce various cytokines and growth factors including IL-1 β , IL-1 α (IL-1 receptor antagonist), IL-6, IL-12, IL-17A, IL-18, IL-21, IFN- α , TGF- β 1, TNF- α , G-CSF, M-CSF, and GM-CSF (28,41,42). While many of these neutrophil-derived cytokines foster proinflammatory responses, they also mediate intercellular communication between neutrophils and other immune cells. Neutrophil derived IL-17A coordinates IFN- γ -mediated programming of M1 proinflammatory macrophages during acute pneumonic plague (43). IL-18, another cytokine produced by neutrophils, is involved in recruitment of macrophages and replication of R5HIV (38). TNF- α released from tumor-associated neutrophils (TANs) triggers apoptosis of non-activated CD8⁺ T cells and inhibits their anti-tumor activity (44). Inflammatory cytokines derived from BCG-activated neutrophils also indirectly stimulate T cell chemotaxis and improve effectiveness of BCG immunotherapy (40).

Neutrophils also produce large amounts of ROS in response to external stimulation, causing oxidative damage against pathogens. However, neutrophils utilize ROS production as a localized signaling mechanism to communicate with neighboring immune cells (45,46). Both mature neutrophils and G-MDSCs utilize localized ROS production to suppress T cell

immune responses (46,47). Additionally, TANs inhibit the proliferation of IL-17 producing T cells through ROS production (48). Furthermore, nitric oxide released from TANs impairs the anti-tumor activity of CD8⁺ T cells (44).

Neutrophils also communicate with nearby immune cells through expression of certain ligands. They express B-cell activating factor (BAFF) and a proliferation-inducing ligand (APRIL), both of which are crucial for the survival, maturation, and differentiation of B cells (42). Neutrophil-derived BAFF drives protective B cell immunity against lethal *Salmonella typhimurium* infection by fostering expansion of B cells and promoting IgM responses (49). B helper neutrophils, found in the marginal zone of secondary lymphoid tissue, secrete higher amounts of BAFF, APRIL, and IL-21, thereby activating marginal B cells (42). Moreover, APRIL produced by inflammation-recruited neutrophils contributes to establishment of plasma cell niches in mucosa-associated lymphoid tissue, ensuring sustained local Ab production (50).

NETS

NETs are complex structures composed of extracellular DNA, granule proteins, and histones. They were first identified as an arsenal of neutrophils to trap and eliminate pathogens extracellularly (51,52). Initial studies on NETs focused on their role in inflammation, but recent findings have uncovered their wider biological implications that extend beyond their antimicrobial activities (**Fig. 3**). NETs have been associated with a range of immunological diseases, including autoimmune diseases, cardiovascular diseases, pulmonary diseases, thrombosis, sepsis, and cancer (reviewed in (52,53)). Furthermore, the persistence of NETs in circulation provides a crucial source of autoantigen that incites autoantibody production,

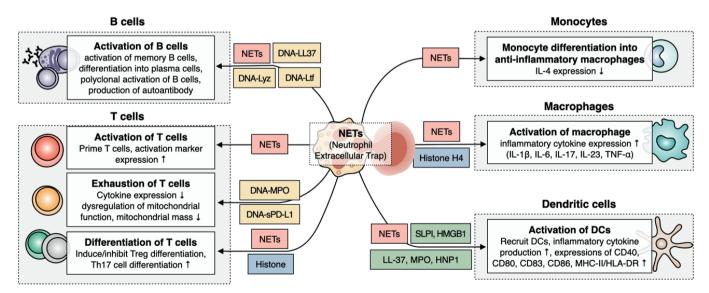


Figure 3. Intercellular crosstalk by neutrophils through NET. NETs, composed of DNA, granule proteins, and histones, regulate neighboring immune cells either directly or via tangled granule proteins and histones. They stimulate macrophages to produce inflammatory cytokines and induce monocyte differentiation into anti-inflammatory macrophages through reduced IL-4 expression. NETs and their granule proteins recruit DCs, trigger inflammatory cytokine production, and induce upregulation of activation marker expressions. Prolonged exposure to NETs cause apoptosis of macrophages and DCs through mitochondrial dysfunction. NETs and granule proteins also activate memory B cells, stimulate the differentiation of naïve B cells into plasma cells, and drive auto-antibody production in B cells. NETs exhibit ambivalent effects on T cells, leading to either activation or exhaustion. NETs either stimulate or inhibit the differentiation of Tregs, while consistently inducing Th17 cell differentiation.

SLPI, secretory leukoprotease inhibitor; HMGB1, high mobility group box 1; HNP1, human neutrophil peptide 1; HLA-DR, human leukocyte antigen-DR isotype.

thereby exacerbating autoimmune reactions (reviewed in (5,54)). Moreover, NETs can directly stimulate various immune cells, such as B cells, T cells, and Ag-presenting cells, leading to autoimmune reactions (reviewed in (5,54)). Additionally, NETs have been shown to entrap CD4⁺ T cells, CD8⁺ T cells, B cells, and monocytes, contributing to either depletion or loss of immune cells during HIV/simian immunodeficiency virus infections (55).

Macrophages are one of the first responders to NETs and play a key role in their clearance. NETs trigger the activation of macrophages, facilitating transfer of antimicrobial peptides to enhance their antimicrobial functions (11). Additionally, NETs stimulate macrophages to produce inflammatory cytokines such as IL-16, IL-6, IL-17, IL-23, and TNF- α , which exacerbate autoimmune and inflammatory diseases (56,57). In patients with Behcet's disease, elevated levels of histone H4 found in NETs have been associated with increased macrophage activation and the production of IL-8 and TNF- α (58). NETs produced by neutrophils stimulated with IL-8 via CXCR2 exacerbate the progression of atherosclerosis by activating TLR9-dependent NF-κB pathways in macrophages (59). Interestingly, NETs can reprogram the differentiation of monocytes into anti-inflammatory macrophages by downregulating the expression of IL-4 receptors (60). However, prolonged exposure of NETs can induce macrophage death through caspase-induced mitochondrial damage and activation of apoptosis induced factor-dependent pathways (61). Macrophages play a crucial role in clearing NETs from both circulation and tissues. They degrade NETs through extracellular digestion and then uptake them through micropinocytosis (62,63). Impaired NET clearance by macrophages has been closely associated with aggravation of inflammatory diseases. as observed in patients with acute respiratory distress syndrome (ARDS) (64). A failure of macrophages to effectively phagocytose NETs triggers pyroptosis of alveolar macrophages, leading to exacerbation of inflammation during ARDS (65). Either a complete failure or a frustrated phagocytosis of NETs activates innate immune sensors, such as cyclic GMP-AMP synthase-STING, leading to type I IFN production in macrophages (66).

DCs are also an important responder of NETs. NETs recruit DCs and activate them via Fc fragment of IgG through low affinity IIa receptor (FC γ II, CD32), leading to production of IFN- α through TLR9 (61,67,68). Certain granule proteins on NETs, such as LL-37, α -defensin, β -defensin, high mobility group box 1, MPO, and secretory leukocyte proteinase inhibitor, activate plasmacytoid DCs (pDCs) to produce inflammatory cytokines including TNF- α , IL-6, and IFN- α during autoimmune diseases (22,67,68). Upon exposure to NETs, pDCs exhibit increased surface expressions of CD40, CD80, CD83, CD86, and MHC-II/human leukocyte antigen-DR isotype, and elevated secretion of proinflammatory cytokines, such as IL-1 β , IL-6, IL-8, IL-10, IL-12, and TNF- α (69). However, NETs can hinder differentiation of DCs (60) and maturation of LPS-stimulated monocytic DCs (70). Furthermore, prolonged exposure to NETs can induce the death of DCs through mitochondrial damage (61). These studies suggest the dual roles of NETs on influencing the function of DCs.

Recent studies suggest that NETs also interact with adaptive immune cells. NETs modulate T cells through direct contact with TCR signaling (71). NETs prime CD4⁺ T cells through direct contact with TCR, leading to a reduced activation threshold and enhanced Agspecific responses (71). Interestingly, NET-exposed CD4⁺ T cells also exhibit increased expressions of activation markers, CD25 and CD69 (71). A similar pattern of activation marker expression is found in NET-exposed CD8⁺ T cells (71). MPO-DNA components in NETs, in conjunction with sPD-L1, contribute to the exhaustion and dysfunction of T cells within the tumor microenvironment (72). Moreover, NETs bearing PD-L1 have been found

to exhaust T cells during ischemia and reperfusion injury in the liver, leading to decreased cytokine expressions, dysregulation of mitochondrial function, diminished mitochondrial mass, and reduced glucose and lipid uptake (72). NET-associated histones can also drive the differentiation and cytokine production in Th17 cells via TLR2/MyD88/STAT3/RORydependent pathway (14). Additionally, NETs can affect T cell differentiation. NETs induce selective differentiation of naïve T cells into Tregs through metabolic reprogramming toward mitochondrial oxidative phosphorylation through TLR4 (13). In contrast, NETs inhibit Treg differentiation in in vitro and decrease Treg to Th17 cell ratio during neutrophilic asthma (73). NETs directly activate memory B cells (10) and promote the progression of diffuse B cell lymphoma through TLR9-dependent pathway (74). LL-37 and DNA complex in NETs stimulate a polyclonal activation of memory B cells and BCR, leading to production of Ab against LL-37 during Systemic Lupus Ervthematosus (10), Lysozymes and lactoferrin complexes in NETs also activate B cells via TLR9, prompting the production of autoantibodies (10). In patients with Bullous Pemphigoid, NETs have been found to induce the differentiation of B cells into CD19+ CD38+ plasma cells via p38 MAPK pathway, thereby augmenting autoantibody release (9).

NEUTROPHIL-DERIVED EXTRACELLULAR VESICLES (NDEVs)

EVs are membrane-derived vesicles that are released by almost all types of cells, including immune cells. They are categorized into three major groups based on their size and generation mechanism: exosomes, microvesicles, and apoptotic bodies. Neutrophils can spontaneously produce EVs, or generate them in response to certain stimuli (15,16,75). The production and function of these NDEVs varies depending on the prevailing conditions of neutrophils (75). NDEVs are composed of proteins, lipids, amino-acids, glycoconjugates, and RNAs (5,15). NDEVs exert antimicrobial activity, stimulate chemotaxis of immune cells such as macrophages and CD8⁺ T cells (16,76), and facilitate intercellular communication through their enclosed proteins and RNAs (5) (**Fig. 4**).

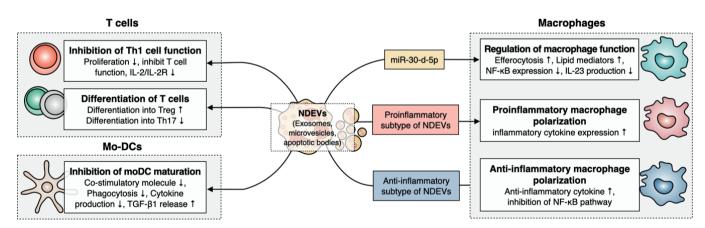


Figure 4. Intercellular crosstalk by neutrophils through NDEVs. Neutrophils produce EVs either spontaneously or in response to stimuli. The contents in NDEVs, such as proteins and RNAs, vary depending on the prevailing condition of neutrophils, resulting in different effects on neighboring immune cells. The proinflammatory subtype of NDEVs drives proinflammatory phenotype polarization of macrophages, whereas the anti-inflammatory subtype of NDEVs induces the anti-inflammatory phenotype polarization of macrophages. NDEVs also regulate macrophage functions through enclosed miRNAs. NDEVs induce the differentiation of Tregs and inhibit Th17 differentiation. They also inhibit the function of Th1 cells through downregulation of IL-2 and IL-2R, and they inhibit maturation of moDCs.

moDC, monocytic dendritic cell.

NDEVs trigger the release of TGF- β from macrophages, thereby promoting their antiinflammatory functions (77-79). They activate various signaling pathways, including Mer Tyrosine Kinase activation, NF-κB translocation inhibition, Ca2+ signaling induction, and TGF- β release induction, thereby regulating macrophage responses at the posttranscriptional level (77-79). Anti-inflammatory subtypes of NDEV drive macrophages toward an anti-inflammatory phenotype, resulting in the inhibition of inflammatory functions (16,80). However, NDEVs from Mucobacterium tuberculosis (Mtb)-infected neutrophils induce the differentiation of macrophages into proinflammatory phenotype (81,82). Similarly, NDEVs generated by activated neutrophils during sepsis can skew the polarization of macrophages toward proinflammatory phenotype and prime them for pyroptosis through the exosomal miR-30-d-5p-mediated activation of NLRP3 (83). This proinflammatory subtype of NDEVs also induces polarization of macrophages into proinflammatory phenotype through miRs (16). Macrophages phagocytose NDEVs via Tim-4 and MFG-E8, and these macrophages exhibit an increase in IL-10 secretion with reduced numbers during sepsis (84). NDEVs also enhance the efferocytosis of apoptotic neutrophils by macrophages and increase production of specific lipid mediators (85). They also inhibit the maturation of Mo-DCs, impairing the expression of costimulatory molecules, cytokine secretion, and phagocytic activity while increasing the release of TGF-β1 (70). EVs generated by PMN-MDSCs inhibit the differentiation of naïve T cells into Th1 and Th17, rather promoting differentiation into Tregs (86,87). Furthermore, NDEVs suppress the activation and proliferation of T-helper cells by down-regulating IL-2 and IL-2 receptor (88). Marcrophages and DCs phagocytose apoptotic neutrophils and clear them through efferocytosis, which provides antimicrobial molecules and stimulates the production of specialized pro-resolving mediators (85).

Apoptotic bodies from neutrophils elicit an anti-inflammatory response in macrophages through the suppression of NF- κ B activation (89,90). They also enhance the production of immunosuppressive cytokine from LPS-stimulated monocytes (91) and suppress the production of anti-inflammatory cytokines in macrophages through expression of proteinase 3 (92). In contrast, apoptotic bodies of Mtb-stimulated neutrophils stimulate proinflammatory responses from macrophages (93,94). Both macrophages and DCs clear apoptotic bodies through efferocytosis (95). The efferocytosis of neutrophil apoptotic bodies dampen the production of IL-23 in macrophages (95) and inhibit DC activation by suppressing the expression of costimulatory molecule (8,96). Notably, LPS accelerate the clearance of neutrophil apoptotic bodies by monocyte-derived macrophages (97). Furthermore, apoptotic bodies from neutrophils stimulated with progesterone induce differentiation of naïve T cells into Tregs through transfer neutrophil proteins, such as forehead box protein, while inhibiting differentiation into Th17 cells (12).

CONCLUSION

Neutrophils, as pivotal components of the innate immune system, interact with a variety of immune cells through various mechanisms. They communicate with both innate and adaptive immune cells, either directly or indirectly, via soluble mediators, NETs, and EVs. The complex and dynamic nature of neutrophil communication highlights their multifaceted role within the immune system and provides insights into their contribution to both physiological function of the immune system and the pathogenesis of various diseases.



ACKNOWLEDGEMENTS

This work was supported by National Research Foundation of Korea Grants 2022R1A2C2006898(CWH), 2021H1D3A2A01099688 and RS-2023-00222188(SS).

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