



Antibacterial activity and toxicity of *Halymenia durvillei* red seaweed from Kayangan island, South Sulawesi, Indonesia

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Abstract

This study aimed to determine the antibacterial activity and toxicity of methanol and hexane extracts of *Halymenia durvillei* red seaweed which were found abundantly in Kayangan island, South Sulawesi. The antibacterial activity of the crude extract was tested against five gram-negative bacteria, namely *Escherichia coli*, *Salmonella typhi*, *Pseudomonas aeruginosa*, *Aeromonas hydrophila*, and *Vibrio harveyi* at a dose of 200 g/disk. Extract toxicity was tested on *Artemia salina* larvae at concentrations of 1,000, 500, 250, 125, 62.5, and 31.25 µg/mL. The results showed that the methanol and hexane extracts of *H. durvillei* had the highest activity against *S. typhi* and *A. hydrophila*, respectively, with inhibition zones of 26.2 mm and 21.0 mm. On the other hand, the two extracts did not show activity against *E. coli* and *P. aeruginosa*, respectively. The toxicity of the methanol extract of *H. durvillei* was twice as high as that of the hexane extract with half-maximal inhibitory concentration of 98.24 and 184.21 µg/mL, respectively. Thus, the methanol and hexane extracts of red seaweed *H. durvillei* have the potential as new antibacterial agents respectively against the pathogenic bacteria *S. typhi* and *A. hydrophila*, but also have the opportunity to be developed into antitumor herbal compounds.

Keywords: Seaweed, *Halymenia durvillei*, Antibacterial activity, Toxicity activity

Introduction

In recent years marine organisms have attracted a lot of attention and have been commercially produced for industrial use. Its various potentials can be utilized in various industrial applications for human benefit (Baptista et al., 2020; Puspita et

al., 2020). Marine organisms such as algae, sponges, and corals produce various compounds with diverse bioactivity and are potential sources of new therapeutic agents. For example, seaweed has been used as a therapeutic and traditional medicine in several countries such as China and Korea (Kang et al., 2016; Sanjeewa & Jeon, 2018).

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Marine organisms can survive in complex communities and competitive environments by producing various secondary metabolites in response to environmental stresses such as competition, predators, and tidal variations (Abu-Ghannam & Rajauria, 2013). Some of these compounds function as antimicrobials to interfere with the growth of competitive microorganisms (Kasmia et al., 2018; Pérez et al., 2016; Silva et al., 2020). Compared to other marine organisms, seaweed can play an important role in many ways. Macroalgae or seaweeds have developed physiological adaptations including synthesizing active compounds as a defense against epiphytes and other organisms fouling (Águila-Ramírez et al., 2012; Bhadury & Wright, 2004).

Several studies suggest that in crude extracts, fractionated parts and/or pure compounds of green, brown, and red seaweeds exhibit antioxidant activity (Corsetto et al., 2020), anticoagulant (Wijesinghe et al., 2011), antiviral (Cheung et al., 2014), antifungal (De Corato et al., 2017), anticancer (Gutiérrez-Rodríguez et al., 2018), cytotoxic (Gunasekaran et al., 2017), anti-inflammatory (Kazłowska et al., 2010), antibacterial (Agba-je-Daniels et al., 2017), and toxicity (Kim & Choi, 2017).

Recently, seaweed has been increasingly studied because of its potential to produce new compounds with various biological activities that are important for solving food and health problems (Brown et al., 2014). One of the challenges in recent years, microorganisms have developed a new strategy so that they are resistant to the action of antibiotics known as multiple drug-resistant bacterial strains (Hampton et al., 2020). Common antibiotics have lost most of their efficacy. Thus, it is urgently needed to continue to find new antimicrobial drugs that can address the never-ending problem of antimicrobial resistance. Antimicrobial resistance has become an important threat to human and animal health and the environment (Reed et al., 2019). The increasing resistance of pathogens to antibiotics has inspired experts to intensively explore and develop cheaper and more effective natural antimicrobial agents with better potency. It is a priority in the field of public health to find new antimicrobials with fewer side effects than antibiotics, good bio-resource availability, and minimal toxicity (Thanigaivel et al., 2015).

Diverse studies have proven that seaweed is a source of strong antibacterial agents such as *Ulva rigida* (Ismail et al., 2018; Sirakov et al., 2019); *Caulerpa racemosa* and *Caulerpa lentilifera* (Nagappan & Vairappan, 2014). Thus, it is also possible to examine the possibility of synergizing antimicrobial compounds from marine organisms with currently available drugs (Cheung et al., 2014). In addition to the promising anti-

bacterial activity of seaweed, it is also necessary to explore the activity toxicity, because it is very important for a compound to be non-toxic and have a promising effect at the same time.

In Indonesia, one of the marine algae species that is currently considered cheap and underutilized is red seaweed of the genus *Halymenia*. Even though this type of algae has wide potential for use, especially its bioactive compounds. *Halymenia* is a supply of natural pigments and also contains bioactive components which have been broadly reported, particularly within the last 5 years, specifically as antioxidants, antibacterial, anti-malarial, and antiviral (Deepak et al., 2019; Malik et al., 2020; Vinosha et al., 2019; Vinosha et al., 2020). *Halymenia* sp. grows wild in Indonesian waters which so far have not been widely used. The results of the latest research presented by Sanger et al. (2019) and Khatulistiani et al. (2020) showed that *Halymenia* sp. from the waters of North Sulawesi and Banten have potential as antidiabetic and antioxidant agents.

Halymenia durvillei is found in abundance in the waters around Makassar city, especially on Kayangan island. So far, studies on its potential bioactivity are still limited. Therefore, this study needs to be carried out to determine the antibacterial activity and toxicity of *H. durvillei* found on Kayangan island, South Sulawesi.

Materials and Methods

Materials

The main ingredients used were *H. durvillei* seaweed, methanol and hexane PA grade (Merck, Darmstadt, Germany), and dimethyl sulfoxide (DMSO; Merck). The test bacteria *Escherichia coli*, *Salmonella typhi*, and *Pseudomonas aeruginosa* had been obtained from the Faculty of Medicine, Hasanuddin University, while *Aeromonas hydrophila*, and *Vibrio harveyi* had been received from the Center for Brackish Water Aquaculture Fisheries (BPBAP) Takalar. *Artemia salina* egg test animals had been acquired at the Marine Biology Laboratory, Hasanuddin University. The growth media for bacteria are nutrient broth (NB; Oxoid, Basingstoke, UK), nutrient agar (NA; Oxoid), and tryptone soya agar (TSA; Oxoid). 6 mm Diameter paper disk, Whatman No. 1 filter paper (Merck), and the commercial antibiotic ciprofloxacin. Seawater was taken from the waters around Kayangan island. The equipment used was a blender (Philips HR 2221, Philips, Amsterdam, Netherlands), autoclave, and oven.

Seaweed collection and preparation

H. durvillei red seaweed (Fig. 1) was collected from Kayangan island waters in June 2020. The sampling location was about 3 km west of Makassar city. Samples were taken at a depth of 1 to 1.5 meters using snorkel equipment and then washed with seawater to remove adhering dirt such as sand and shellfish. The samples had been placed into sample plastics and stored in a coolbox containing ice as a cooling medium to keep the freshness of the seaweed for the duration of transportation to the laboratory. Moreover, the samples were washed with running water to clean seawater and remaining dirt, sooner rinsed with distilled water, after which drained and weighed to determine the wet weight of the seaweed. The samples were dried for 10 days then mashed using a blender after which sieved to attain seaweed flour. Seaweed flour was weighed and packed in an airtight container and stored at room temperature before use. The yield was expressed in the weight percentage of seaweed flour produced to the weight of wet seaweed.

Seaweed extraction

Extraction of samples by using maceration method the use of organic solvents methanol and hexane according to El Shafay et al. (2016) which had been modified. A total of 150 g samples had been immersed in 600 mL of every solvent (1:4 w/v) and left at room temperature for three days while stirring occasionally with the use of a magnetic stirrer. The walls and glass surface of the maceration container were protected with aluminum

foil to keep away from touch with open-air and light exposure. The maceration results were vacuum filtered with Whatman paper No. 1 to obtain the filtrate which was then concentrated using a rotary vacuum evaporator at a temperature of 40 °C – 45 °C. The resulting extract was weighed and then stored in the freezer before use. The yield of the extract was expressed as a percentage of the weight of the resulting extract to the weight of the seaweed flour used in extraction.

Antibacterial activity

Preparation of bacteria

NB and NA media had been prepared one after the other dissolving 13 g NB and 23 g NA in 1 L of distilled water and after which heated till dissolved. Both media had been sterilized in an autoclave at 121 °C for 15 minutes. Similarly, TSA media become prepared in an equal way as NA media. The agar medium was cooled to about 50 °C before being poured into the Petri dish. The test bacteria had been rejuvenated on sterile sloping media and incubated at 37 °C for 24 hours. A complete of 1 ose of bacteria on slanted agar turned into inoculated on NB media, homogenized the use of a vortex, after which incubated at 37 °C for 24 hours to obtain viability of 10⁸ colonies/mL.

Antibacterial activity test

The antibacterial activity test of crude extracts of methanol and hexane refers to Bauer et al. (1966) and Christobel et al. (2011) with the modified agar diffusion method. A complete of 1 mL of each bacterial culture turned into inoculated on NA media (besides *V. harveyi* on TSA media) in a Petri dish. Every extract of methanol and hexane turned into taken as plenty as 1,000 µg and dissolved in 50 µL of the solvent till dissolved. 10 µL turned into taken and carried out to a 6 mm diameter paper disk at a dose of 200 µg/disk. After the solvent evaporated, every disc changed into dripped with 5 µL of DMSO after which placed on agar media that already contained the test bacteria. As a positive control, the commercial antibiotic ciprofloxacin 5 µg was used (El Shafay et al., 2016) at the same time as the negative control used DMSO 5 µL. Petri dishes that already comprise the test sample are wrapped with plastic wrap and incubated at 37 °C for 24 hours. The clear zone formed around the disc indicated the inhibition of bacterial activity by the extract which turned into expressed in mm as the average value of three replications.



Fig. 1. Map showing the geographic location of seaweed sampling in the waters of Kayangan island, Makassar city, South Sulawesi province, Indonesia.

Toxicity

Hatching of *Artemia salina* shrimp larvae

An egg hatching container was prepared, namely a two insulated vessel with a 2 mm diameter hole as a connector, one room was left dark and the other room was lit with lamps. A total of 250 mL of seawater was filled into the dark and aerated. Inserted 2.5 mg of artemia eggs then covered with aluminum foil, the other side was left to stay bright. The eggs hatch after 24 hours into larvae and move to a bright room through a connecting hole. After 48 hours larvae or nauplii are ready to use for toxicity tests (Nurhayati et al., 2006; Puspitasari et al., 2018).

Toxicity test

Stock solutions of each methanol and hexane extract were prepared by weighing 100 mg of extract and then dissolved in 10 mL of solvent. A series of dilutions were made by taking 1,000, 500, 250, 125, 62.5, and 31.25 µL of the stock solution, respectively, into the vial, and the solvent was evaporated. Each was added 100 µL of DMSO and 2 mL of seawater and stirred until the extract was completely dissolved. Furthermore, the extract solution was transferred to a 10 mL volumetric flask and then added with seawater to a volume of 10 mL, homogenized to obtain an extract solution with concentrations of 1,000, 500, 250, 125, 62.5, and 31.5 µg/mL. The solution was transferred into a glass vial and then 10 larvae of *A. salina* were added to each. Each dilution was made 3 times with a solution without the addition of extract as a negative control. Observation of larval mortality was carried out after 24 hours at room temperature. Data analysis to calculate half-maximal inhibitory concentration (IC₅₀) µg/mL with sigmoid type inhibition curve (Kasmiati et al., 2018).

Results

Yield

The yield of *H. durvillei* flour is the percentage ratio of the weight of seaweed flour produced (337 g) to the weight of wet seaweed (6,600 g) which is 5.11%. Furthermore, the yield of methanol and hexane extracts were 5.32% and 4.11%, respectively, which was expressed in terms of the percentage by weight of the crude extract produced, namely 7.98 and 6.16 g to the weight of 150 g seaweed flour, respectively. This indicates that the biomass of *H. durvillei* contains more polar compounds.

Antibacterial activity

In this study, we tested the antibacterial activity of methanol and

hexane extracts of *H. durvillei* against gram-negative bacteria. Three of the five gram-negative bacteria used are bacteria that cause disease in humans, namely *E. coli*, *S. typhi*, and *P. aeruginosa*, while the other two bacteria *A. hydrophila* and *V. harveyi* are bacteria that commonly attack well-cultivated organisms shrimp and fish. Methanol extract from *H. durvillei* confirmed antibacterial activity against all test bacteria besides *E. coli*.

An important finding resulting from this study is the methanol extract of *H. durvillei* showed the highest activity against *S. typhi* with an inhibition zone of 26.2 mm. The inhibitory ability of the *H. durvillei* methanol extract was two times higher than the commercial antibiotic activity of ciprofloxacin as a positive comparison (Table 1). This indicates that the methanol extract of *H. durvillei* has excellent potential as a supply of new antibiotics, especially against *S. typhi*. Another thing that was found in this study was that the methanolic extract of *H. durvillei* had moderate inhibitory activity against *P. aeruginosa*, *A. hydrophila*, and *V. harveyi* with inhibition zones of 9.6, 9.3, and 8.9 mm, respectively (Table 1). The zone of inhibition in question is the diameter of the clear zone formed including a 6 mm diameter paper disk (Fig. 2).

Hexane extract *H. durvillei* showed inhibitory activity against all test bacteria except against bacteria *P. aeruginosa* (Fig. 3). Extract activity against *A. hydrophila* showed the highest response with a zone of inhibition 21.0 mm higher than the positive control of ciprofloxacin antibiotics (Table 1). The potency of the inhibition of hexane extract against the other three bacteria was moderate with inhibition of 8.5, 11.3, and 8.0 mm for each of the tested bacteria *E. coli*, *S. typhi*, and *V. harveyi*.

Table 1. Inhibitory activity of methanol and hexane extracts of *Halymenia durvillei* red seaweed against five types of bacterial pathogens

Pathogenic bacteria	Zone of inhibition (mm)	
	Methanol extract ¹⁾	Hexane extract ¹⁾
<i>Escherichia coli</i>	0	8.5
<i>Salmonella typhi</i>	26.2	11.3
<i>Pseudomonas aeruginosa</i>	9.6	0
<i>Aeromonas hydrophila</i>	9.3	21.0
<i>Vibrio harveyi</i>	8.9	8.0
DMSO (negative control)		0
Ciprofloxacin (positive control)		13.3

¹⁾ Data is the average of three repetitions. DMSO, dimethyl sulfoxide.

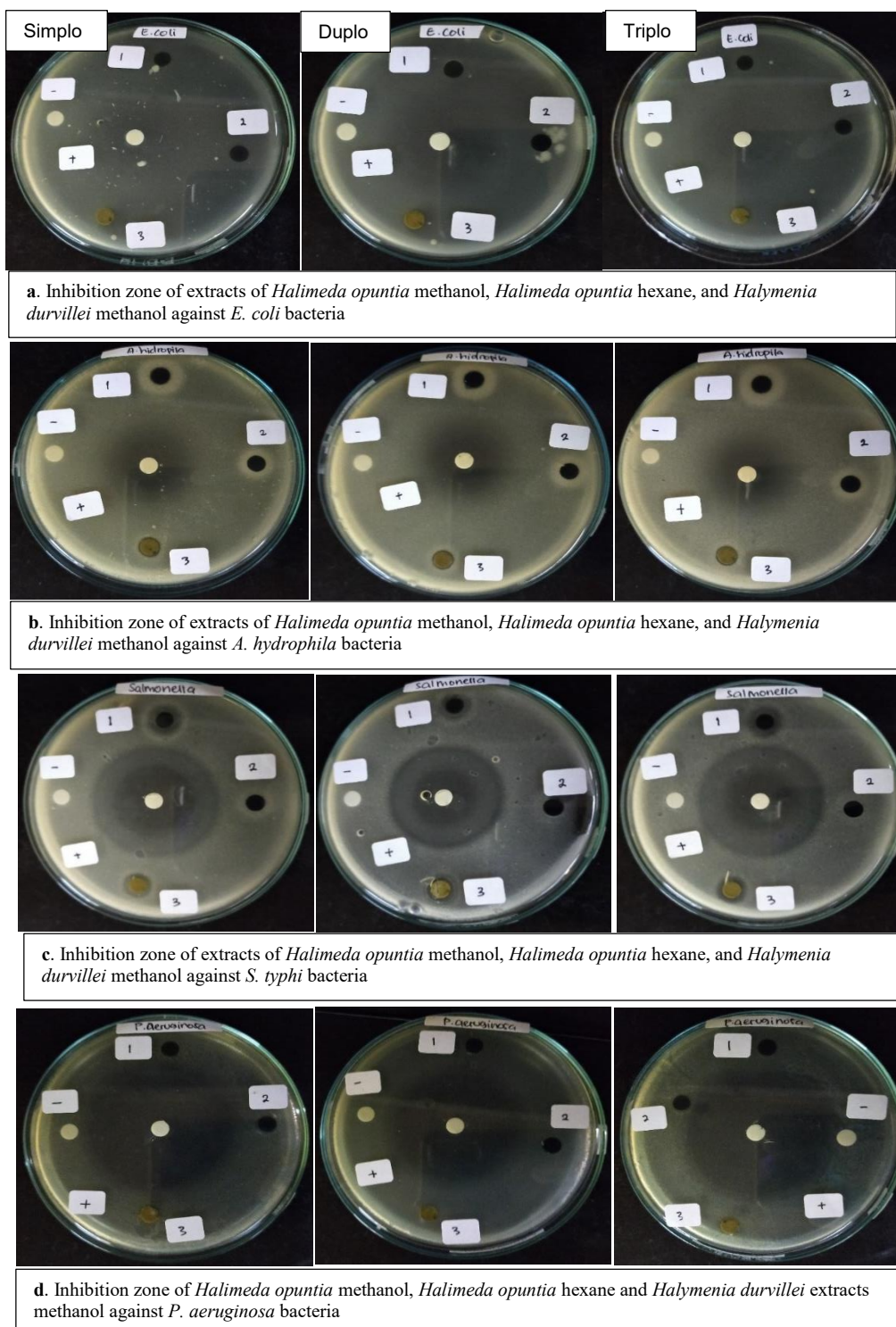


Fig. 2. Antibacterial inhibition zone of *Halymenia durvillei* methanol extract against *Escherichia coli* (a), *Aeromonas hydrophila* (b), *Salmonella typhi* (c), *Pseudomonas aeruginosa* (d). No image antibacterial inhibition zone of *H. durvillei* against *Vibrio harveyi*.

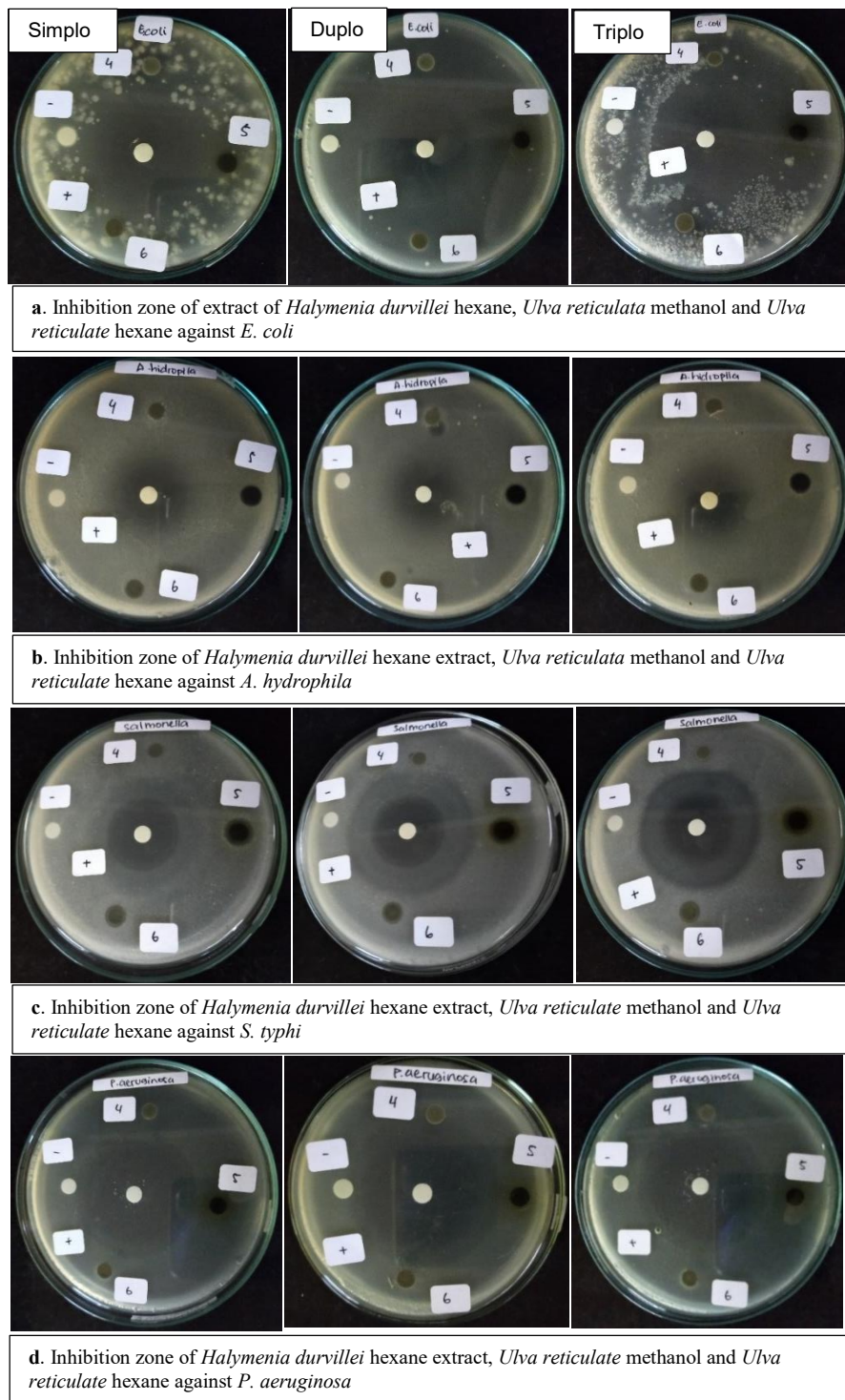


Fig. 3. Antibacterial inhibition zone of hexane extract of *Halymenia durvillei* against *Escherichia coli* (a), *Aeromonas hydrophila* (b), *Salmonella typhi* (c), *Pseudomonas aeruginosa* (d). No image antibacterial inhibition zone of *H. durvillei* against *Vibrio harveyi*.

Toxicity

In this study, the potential toxicity of red seaweed extract *H. durvillei* was also tested against larvae *A. salina* as a preliminary study to determine the potential of methanol and hexane extracts as anticancer agents as listed in Table 2.

Toxicity testing was based on the ability of the extract to inhibit the growth of the larvae of the test animals, the lower the concentration of the extract to inhibit 50% of larval growth, the more potent the extract was. Toxicity of seaweed methanol and hexane extracts *H. durvillei* evaluated after exposure for 24 hours at room temperature. After 24 hours, the percentage of dead larvae was calculated for each concentration. Methanol extract causes mortality rate *A. salina* respectively 10%, 42%, 58%, 75%, 80%, and 100% at a concentration of 31.25; 62.5; 125; 250; 500; and 1,000 µg/mL. Meanwhile, the hexane extract at the same concentration caused average mortality of larvae of test animals of 30%, 36%, 41%, 48%, 61%, and 82% after exposure for 24 hours.

The results of this study indicate the potential toxicity of methanol extract *H. durvillei* higher than the hexane extract activity with IC₅₀ of 96.24 and 184.21 µg/mL, respectively.

Discussion

Antibacterial activity

Seaweed is a unique resource from the ocean because of its

ability to thrive in the dynamic marine environment. A study showed that the antibacterial activity of seaweed is due to the symbiotic relationship between bacteria and seaweed which plays an important role in the development and growth of seaweed (Singh & Reddy, 2014). Seaweed contains epiphytic bacteria which are an excellent source of natural antimicrobial and antioxidant compounds (Horta et al., 2014). The potential broad-spectrum antibacterial effect of epiphytic bacteria (Phylum: *Firmicutes*; closely related to *Bacillus pumilus*) suggests that seaweed-associated bacteria may be producers of antibacterial compounds (Ismail et al., 2016). Another study by Sujuliyani et al. (2019) showed that *C. racemosa* had symbiotic bacteria belonging to the genus *Neisseria* that we're able to synthesize potential antibacterial compounds against *Staphylococcus aureus* and *S. typhi*. Our results show that *H. durvillei* seaweed is a potential source of antibacterial compounds.

The results of the study showed that the methanol extract of *H. durvillei* showed antibacterial activity against all test bacteria except *E. coli*. The results obtained may be greater or much less similar to results reported through previous experiments. For instance, about the effect of different extraction solvents on the antibacterial activity of the different seaweeds, ethanol extract of *Sargassum vulgare* was reported to exhibit inhibitory activity against *S. aureus* and *Klebsiella pneumoniae* (Bolaños et al., 2017). Another previous study reported alkaloid extracts from brown (*S. vulgare*, *Dictyopteris membranacea*, and *Cystoseira barbata*), green (*Ulva lactuca* and *Codium tomentosum*), and red seaweeds (*Gelidium latifolium*), respectively, exhibited remarkable antibacterial activity against *S. typhi*, *S. aureus*, *Staphylococcus epidermidis*, *Bacillus* spp., *Bacillus subtilis*, *E. coli*, *Klebsiella* spp. and *P. aeruginosa* (Alghazeer et al., 2013). Our results might be different from the previous findings which demonstrated greater activity, but our results showed antibacterial activity against all test bacteria except *E. coli*. According to Nazarudin et al. (2020), the absence of the inhibition zone is probably due to the low concentration of antibacterial active metabolites, which causes resistance of microorganisms to seaweed extracts. The research found by Yap et al. (2019) stated that the methanol extract of *C. racemosa* had moderate antibacterial activity against *E. coli* K1 (gram-negative bacteria). Another research also reported by Al-Saif et al. (2014) using the disc diffusion method showed that the chloroform extract from *Caulerpa occidentalis* was very effective against *E. coli* but has moderate antibacterial activity against *S. aureus* and *Enterococcus faecalis*. The difference in results may be due to the effect of

Table 2. Toxicity of methanol and hexane extracts of *Halymenia durvillei* to *Artemia salina* larvae

Extract	Concentration (µg/mL)	Concentration log	Mortality (%)	IC ₅₀ (µg/mL) ¹⁾
Methanol	1,000	3	100	98.24
	500	2.7	80	
	250	2.4	75	
	125	2.1	58	
	62.5	1.8	42	
	31.25	1.45	10	
Hexane	1,000	3	82	184.21
	500	2.7	61	
	250	2.4	48	
	125	2.1	41	
	62.5	1.8	36	
	31.25	1.45	30	

¹⁾ Data is the average of three repetitions.
IC₅₀: half-maximal inhibitory concentration.

seaweed's antibacterial activity, such as the different species of seaweed tested, the solvent used for extraction, the geographical location of the seaweed, and so on (Anjali et al., 2019; Nazardin et al., 2020).

As mentioned before methanol extract exhibit a higher antibacterial activity against *S. typhi*. This indicates that the methanol extract of *H. durvillei* has great potential as a source of new antibiotics, especially against *S. typhi*. *S. typhi* is the causative agent of systemic infection and typhoid fever in humans. *Salmonellosis* is common in people in developing countries and is a high-risk group (Hussain et al., 2019; Sah et al., 2020). To prevent complications of death and disease, antibiotics are the first treatment modality in an enteric fever until the prevalence of *S. typhi* (Wong et al., 2019). *Salmonella* is known to be resistant to various types of antibiotics such as ampicillin, chloramphenicol, and trimethoprim-sulfamethoxazole (Akram et al., 2020). Thus, the results of this study are important findings as an initial step in efforts to discover natural compounds from seaweed as an alternative solution to overcome resistance *Salmonella* to commercial antibiotics. Substitution of antibiotics with natural functional molecules will help overcome the currently developing resistance to antibiotics by pathogenic microorganisms (Fraga-Corral et al., 2020; Kini et al., 2020). In addition to having activity against the test bacteria in this study, the methanol extract *H. durvillei* also shows activity against *K. pneumoniae* (Hamza et al., 2014). In a study of the antibacterial activity of seaweed, *S. typhi* is a species of *Salmonella* the most widely used as test bacteria. This is because testing with *S. typhi* allows the classification of moderate or moderate to susceptible antibacterial activity (inhibition zone diameter more than 15 mm) (Akremi et al., 2017; Silva et al., 2020). Thus the antibacterial activity of red seaweed methanol extract *H. durvillei* found in this study is categorized as high in other words *S. typhi* is susceptible to methanol extract *H. durvillei*. Therefore, the abundance of seaweed as a source of active compounds can be used to obtain extracts rich in antimicrobial components to be applied in the food, cosmetic, and pharmaceutical fields (Silva et al., 2020).

The results of the study showed that the hexane *H. durvillei* extract showed inhibitory activity against all the tested bacteria except for *P. aeruginosa*. The activity of the extract against *A. hydrophila* showed the highest response. This indicates that the red seaweed hexane extract *H. durvillei* has promising potential as an antibacterial agent, especially against pathogenic bacteria *A. hydrophila* causes disease and crop failure in cultured fish (Rozi et al., 2018). The results of this study are supported by the

report of Vinosha et al. (2020) that the sulfate galactan component *Halymenia* sp., besides having antioxidant activity, is also able to increase the resistance of fish *Oreochromis* against bacteria *Aeromonas hydrophilic*. Thus the antibacterial activity of red seaweed hexane extract *H. durvillei* was found in research. This could be a promising finding to reduce the use of antibiotics in the aquaculture sector where antibiotic-resistant bacteria commonly thrive.

To the best of our knowledge, the difference in results on antimicrobial activity may be due to differences in bacterial cell wall configuration and permeability, bacterial composition, and resistance charge between the cell wall of pathogenic and secondary bacteria metabolism (Amorim et al., 2012; Lin et al., 2014; Peng et al., 2017). Regarding secondary metabolites, phenolic compounds are a group of secondary metabolites which are components of seaweed cell walls or as a defense mechanism against abiotic and biotic stressors (Generalić Mekinić et al., 2019). Several mechanisms of antimicrobial activity of phenolic compounds are inhibition of enzymes and metabolic pathways, changing membrane permeability that causes cell lysis, and binding to surface molecules (Silva et al., 2020). The mechanism of action of antibacterial activity disrupts membrane stability and cellular function in microorganisms by interacting with bacterial cell wall glycoreceptors, membrane compounds, and nucleic acids (Silva et al., 2020). Another report states that the mechanism of antimicrobial activity is the accumulation of lysozyme (immune enzyme) which digests bacterial cell walls (Perez-Santaescolástica et al., 2022). In addition to the phenolic components, previous reports stated flavonoids (Górniak et al., 2018) and alkaloids (Casciaro et al., 2020) are also important antimicrobial agents against a variety of microorganisms.

Toxicity

Method Brine Shrimp Lethality Test (BSLT) uses shrimp larvae *A. salina* as test animals to find new anticancer compounds derived from plants (Kim & Choi, 2017). This was also stated by Zamani et al. (2017) that the determination of toxicity as an initial test is a very important part of research in the search for drug substances or drug discovery. Several methods have been developed to study the viability and proliferation of cultured cells. The most widely applied method is to use a 96-well-format which is based on the principle of colorimetry. Through this method, a large number of samples can be tested quickly and simultaneously. The cytotoxic assay with this platform can directly measure the enzymatic activity in the microplate using

a microplate reader (Weyermann et al., 2005).

The results of this study showed that the potential toxicity of the methanol extract of *H. durvillei* was higher than that of the hexane extract. The toxicity of the methanol extract is slightly lower than the toxicity of the red algae ethanol extract *Eucheuma cottonii* namely 58.01 µg/mL (Sharo et al., 2013). On the other hand, the extract potency of *H. durvillei* in this study was higher than the toxicity *Sargassum* sp. proposed by Riyanto et al. (2014) and Jannah et al. (2014) which are 170 and 139 µg/mL, respectively. The potential toxicity of the methanol extract found in this study is similar to the toxicity of the red seaweed ethanol extract *Eucheuma spinosum* to *A. salina* with an IC₅₀ of less than 100 µg/mL (Hamrun et al., 2020). Red seaweed toxicity was also reported by De Alencar et al. (2014) that the ethanol extract *Amansia multifida* and *Meristiella echinocarpa* besides having antioxidant activity, the total phenolic content of 45.40 and 28.46 mg gallic acid equivalent g⁻¹ also showed toxicity to *A. salina*.

Thus, red seaweed *H. durvillei* which is found abundantly in Kayangan island, South Sulawesi waters, has the opportunity to be developed as a sourcing agent for new compounds to overcome the development of abnormal cells in human tissues.

Conclusion

Crude extract of methanol and red seaweed hexane *H. durvillei* from Kayangan island, South Sulawesi, each has high antibacterial activity, especially against gram-negative bacteria *S. typhi* and *A. hydrophila*. Methanol extract has promising toxicity with IC₅₀ value of less than 100 µg/mL which indicates that the extract has potential as a source of new anticancer compounds.

Competing interests

No potential conflict of interest relevant to this article was reported.

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Availability of data and materials

The data used to support the findings of this study are available from the corresponding author upon request.

Ethics approval and consent to participate

This article does not require IRB/IACUC approval because there are no human and animal participants.

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