



Periprosthetic Hip Joint Infection with *Flavonifractor plautii*: A Literature Review and Case Report

Alexander Wilton, MBBS*, Constantine Michael Glezos, FRACS (Orth)*+,
Hasitha Pananwala, MBBS*, Han Kiong Lim, MBBS*

Department of Orthopaedics, Ryde Hospital, Eastwood, NSW, Australia*
Department of Orthopaedics, Sydney Adventist Hospital, Wahroonga, NSW, Australia†

The purpose of this case report and review of the literature is to provide documentation on periprosthetic hip joint infection with *Flavonifractor plautii* (formerly known as *Eubacterium plautii*), a strictly anaerobic bacterium, and to report on a successful pathway for management including staged surgical revisions and extended antibiotic therapy. A systematic review of the literature was conducted, which identified this case as only the fifth documented case of human infection with this organism; as a result, conduct of further research is warranted, based on the paucity of reports in the literature addressing anaerobic periprosthetic joint infection.

Key Words: Hip joint, Prosthesis-related infections, Anaerobic bacteria, Clostridium, Total hip arthroplasty

A low incidence of periprosthetic joint infection (PJI) after elective primary total hip replacement, 2%, has been reported^{1,2}. PJI with the onset of clinical features >12 months post-implantation most commonly occurs as a result of haematogenous seeding to the joint. The rates of PJI may be higher for metal on metal bearing implants; however, the responsible organisms are similar^{3,4}. The majority of infective organisms are gram-positive organisms, such as

Staphylococcus aureus and *Staphylococcus epidermidis*⁵. The reported incidence of atypical anaerobic PJI is higher in hip arthroplasty compared with knee arthroplasty⁶ and is increasing⁷.

Anaerobic bacteria have been implicated in only 3-6% of PJI cases⁸. Of these, isolation of *Cutibacterium acnes*, which represents the majority, commonly occurs 3-12 months post-implantation from direct inoculation at the time of surgery. The literature includes only a few reports addressing management of PJI with *Clostridia* species, which is closely related to *Flavonifractor plautii*, and publications are limited to only case reports⁹⁻¹⁸. *F. plautii* (formerly known as *Eubacterium plautii* or *Fusobacterium plautii*) is a strictly anaerobic, non-spore forming, non-motile, rod-shaped bacterium; understanding of its biological and pathologic properties is limited¹⁹. Regarding genomic sequencing, the organism shares a 99% similarity to *Clostridium orbiscindens*²⁰. To the best of our knowledge, we report on the first documented case of PJI with this organism and the only reported study of human infection in an immunocompetent individual. The purpose of this case report and review of the lit-

Submitted: August 13, 2022 1st revision: September 16, 2022

Final acceptance: October 6, 2022

Address reprint request to

Alexander Wilton, MBBS

(<https://orcid.org/0000-0002-4796-9375>)

Department of Orthopaedics, Ryde Hospital, Denistone Road,
Eastwood, NSW 2122, Australia

TEL: +61-2-9858-7888 FAX: +61-2-9858-7718

E-mail: Awil2853@uni.sydney.edu.au

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/4.0>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

erature is to provide documentation of PJI with this rare pathogen and to report on a successful pathway for management including staged surgical revisions and extended antibiotic therapy.

CASE REPORT

Our patient was a 62-year-old male non-smoker who presented to the emergency department with a complaint of worsening pain in the left groin and subjective fevers for approximately four weeks. His second dose of the ChAdOx1 nCoV-19 (Oxford–AstraZeneca) vaccine was administered two weeks prior to presentation and he experienced one episode of diarrhoea 10 days prior to presentation. He had no recent history of taking antibiotics.

The patient had undergone a total replacement of the left hip 15 years prior for treatment of arthrosis secondary to hip dysplasia. Until this current episode, no complication had been reported in association with the implant except for minor asymptomatic osteolysis secondary to metal-on-metal wear. His medical history included hypertension and paroxysmal atrial fibrillation for which he took rivaroxaban. He was admitted to a hospital six years prior with diarrhoea, haematochezia, and *Bacteroides vulgatus* bacteremia, which was treated successfully with antibiotics. No significant gastrointestinal abnormality was observed by inpatient gastroscopy and colonoscopy performed at that time and the patient made a full recovery. The patient had no additional medical history and no risk factors for immunocompro-

mise.

Upon arrival in the emergency department, the patient was afebrile and mild tachycardia was detected with a heart rate of 100-110 bpm in atrial fibrillation and he was hypotensive with a blood pressure of 80/40 mmHg. His respiratory rate was 19 breaths per minute and his S_pO_2 was 99% on room air. The left hip joint showed irritability to passive movements. Findings from cardiovascular, gastrointestinal, and respiratory examinations were otherwise unremarkable.

The results of blood analysis indicated elevated levels of inflammatory markers (Fig. 1). The levels of blood metal ion were normal. The pelvic x-ray showed no new changes to the prosthesis, which showed longstanding, unchanged features of femoral osteolysis secondary to metal wear (Fig. 2). Aspiration of the hip performed in the emergency department yielded 20 mL of haemoserous fluid, returning 250,200 white blood cell count (WBC)/ μ L; gram-negative and positive bacilli were detected on microscopy (Fig. 3).

The patient was started on empirical intravenous antibiotic therapy (cefazolin) and transferred to the intensive care unit for further resuscitation. On day 2, light growth of tiny, grey, translucent colonies was isolated on an anaerobic agar plate inoculated with the joint aspirate (Fig. 4).

On day 4, these colonies were identified as *F. plautii* by matrix-assisted laser desorption/ionisation-time-of-flight mass spectroscopy (MALDI-TOF MS, MALDI Biotyper[®]; Bruker, Billerica, MA, USA) with a confidence score of 1.96. Growth of the same organism was observed in a peripheral blood culture anaerobic bottle. According to recommenda-

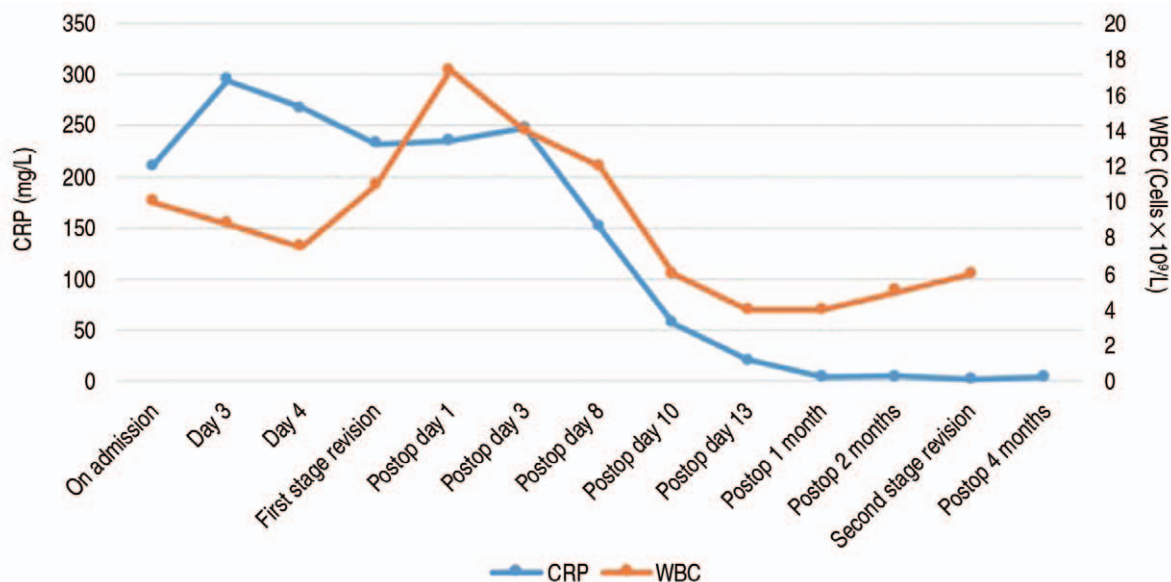


Fig. 1. Serum inflammatory markers during admission. C-reactive protein (CRP) (mg/L) on left y-axis (blue line); white blood cell count (WBC) (cells × 10⁹/L) on right y-axis (orange line). Postop: postoperative.

tion by the infectious disease team, the antibiotic regimen was changed to include amoxicillin-clavulanate, gentamicin, metronidazole, and vancomycin. Antibiotic susceptibility was determined on day 4 and therapy was adjusted accordingly (Table 1).

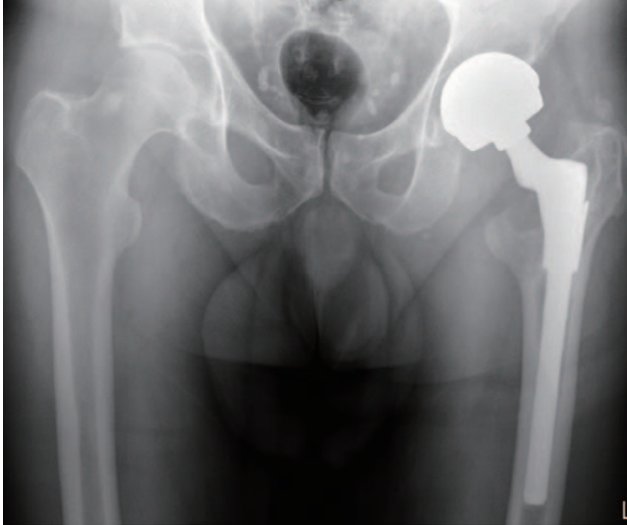


Fig. 2. Anteroposterior pelvis x-ray at time of acute presentation with hip pain and sepsis. S-ROM femoral components (DePuy Orthopaedics, Warsaw, IN, USA) with metal on metal bearing. The proximal femoral erosion around the lesser trochanter was longstanding and unchanged in size over many years.

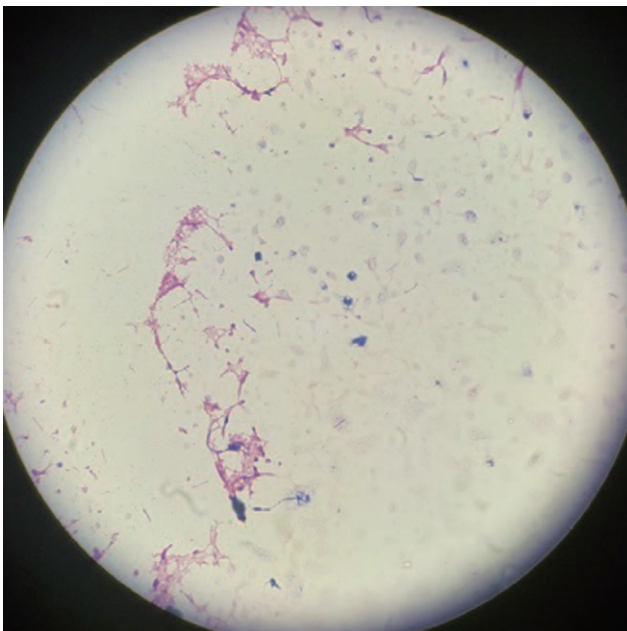


Fig. 3. Gram-positive and negative bacilli on gram stain microscopy (20× magnification) of left hip joint synovial fluid aspirate.

Surgical intervention was delayed due to limited access to operating theatres during the coronavirus disease 2019 (COVID-19) pandemic, thus, the patient underwent medical stabilisation in the intensive care unit. On day 7, the patient underwent a first-stage revision hip arthroplasty. Exploration of the hip was performed using the posterior approach, resulting in detection of extensive abscess formation and destruction of soft tissue deep to the fascia. No gross evidence of metallosis was observed. No loosening of the stem was observed and the use of a fine oscillating saw around the implant/bone interface was required in order to facilitate its removal. Good fixation of the acetabular component was observed and use of the Explant[®] acetabular cup removal system (Zimmer Biomet, Warsaw, IN, USA) was required. Debridement of extensive erosion of pericapsular soft tissue was performed. Pulse lavage and intermittent irrigation with hydrogen peroxide were performed for irrigation of the field with 6 L of normal saline. Reaming of the canal was performed using a 14 mm flexible reamer, followed by thorough washing with a canal brush. Extraction of all hardware was followed by implantation of an antibiotic cement articulating spacer containing a mixture of gentamycin and vancomycin (Fig. 5). No organisms were isolated on culture from intraoperative specimens or subsequent postoperative blood cultures.

A thickened ascending colon was observed on a postoperative computed tomography of the abdomen, and a sub-

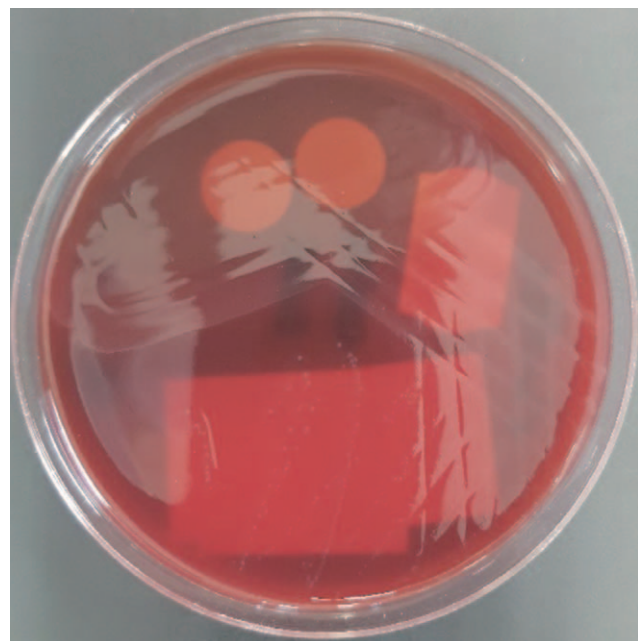


Fig. 4. *Flavonifractor plautii* colonies isolated on anaerobic blood agar.

sequent colonoscopic biopsy showed a single sigmoid tubular polyp and diverticular disease. Conservative management was administered. Findings from a chest x-ray, uranalysis, and cardiac echocardiogram performed during admission were normal.

The patient received 24-hour intravenous administration of ceftriaxone 2 g for six weeks via a central line followed by oral amoxicillin/clavulanic acid and metronidazole for six weeks. Normalisation of the C-reactive protein occurred during this period (Fig. 1). After completion of oral antibiotics, a further aspiration of the hip was performed, which yielded 832 WBC/ μ L, 20% polymorphonuclear cells and negative cultures after incubation for two weeks.

Second stage revision arthroplasty was performed three months after the first stage. A posterior approach was used for exposure of the hip. Extraction of the antibiotic cement

and all spacer implants was performed. No macroscopic evidence of infection or bone necrosis was observed. Multiple swabs and tissue cultures sent from both acetabular and femoral sites showed no growth on extended culture. Administration of routine intraoperative surgical antibiotic prophylaxis was subsequently performed (cefazolin and vancomycin). Preparation of acetabular and femoral sides was performed with implantation of uncemented components (Fig. 6). The patient was discharged on day 7 and his recovery was uncomplicated. At 12-month follow-up the patient remains in good health without ongoing infective issues. He remains off antibiotics. Normal, pain free gait with near normal hip range of movement has been observed by physical examination.

DISCUSSION

A PubMed search for the terms: ‘*Flavonifractor plautii*’ OR ‘*Eubacterium plautii*’ was conducted, resulting in identification of 51 articles; all abstracts were read. Abstracts that failed to provide a discussion of either organism in the context of pathological human infection were excluded (Fig. 7). Discussion of human infection with either organism was included in four abstracts²¹⁻²⁴ (Table 2). After these papers were read, data extraction was performed, including patient age, sex, concurrent pathology, immunosuppression, time to culture, and culture technique.

The results of our systematic search of the literature indi-

Table 1. Antibiotic Susceptibility Testing of the *Flavonifractor plautii* Blood Culture Isolate Performed via Gradient Diffusion*

| Antibiotics | Susceptibility | MIC (mg/L) |
|-------------------------|----------------|------------|
| Amoxicillin-clavulanate | Sensitive | 0.125 |
| Ceftriaxone | Sensitive | 0.38 |
| Clindamycin | Sensitive | 0.38 |
| Metronidazole | Sensitive | 0.016 |
| Moxifloxacin | Sensitive | 0.38 |
| Penicillin | Intermediate | 0.38 |
| Vancomycin | Resistant | 3 |

MIC: minimum inhibitory concentration.

* Etest[®]; bioMérieux, Marcy-l’Étoile, France.



Fig. 5. X-ray pelvis after first stage revision with antibiotic cemented articulating spacer.

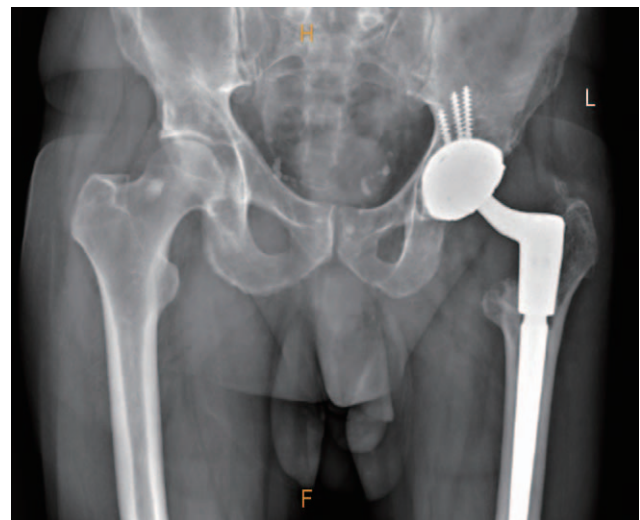


Fig. 6. Anteroposterior pelvis x-ray at 1-year follow-up. The definitive implants include Stryker Trident[®] Tritanium[®] shell with 3 screws, Restoration[®] MDM[®] X3[®] polyethylene insert, Restoration[®] Modular Hip System femoral components, Biolox[®] delta 28 mm ceramic head.

cated that most studies of *F. plautii* have been in relation to its role in the gut microbiome^{25,26} with potential associations with colorectal cancer²⁷, altered immune function²⁸, and inflammatory bowel disease²⁹. In all four documented cases, human infection occurred in the setting of immunosuppression and diagnosis was delayed. The slow growth rate of the bacterium, the requirement for use of enriched media, and the need for strictly anaerobic incubation complicate identification of *F. plautii*. Identification is further complicated by its inconsistent appearance with use of conventional gram staining processes³⁰. Accurate identification of the organism as gram-positive on initial staining was reported in only one previous study²¹.

Identification of anaerobic bacteria, including *C. acnes*, occurs more frequently in hip PJI than in knee PJI⁸. In general, there is a paucity of evidence for use in guiding diagnosis and management of anaerobic PJI, and the revised criteria established by the Musculoskeletal Infection Society (MSIS) limit discussion to *C. acnes* only³¹. Clostridium PJI is most frequently observed in the setting of polymicrobial infection, usually in patients who have a history of gastroin-

testinal disease³². Isolation of *C. difficile* has been reported in prostheses of the hip^{12,14}, knee¹⁵, and shoulder⁹. PJI with this organism can be catastrophic; amputation was reported in two of the five recorded cases³³.

We believe that gut translocation and haematogenous seeding to the prosthesis following a recent diarrhoeal illness in our patient was the most plausible mechanism of infection. Although the patient's history of previous diarrhoeal illness and *B. vulgatus* bacteremia was suspicious, extensive investigation for assessment of intestinal pathology showed no troublesome findings. Recent administration of the ChAdOx1 nCoV-19 (Oxford–AstraZeneca) vaccine may have been a factor in development of his newly diagnosed diarrheal illness; one large study reported that the incidence of this side effect was 2.2%³⁴.

Selection of our strategy of administering high-dose intravenous antibiotics and performing 2-stage revision arthroplasty, which was based on the apparent infection chronicity (~4 weeks), adhered to accepted practice for management of established PJI³⁵. Ideally, the first operative intervention should have been performed prior to day 7 and sam-

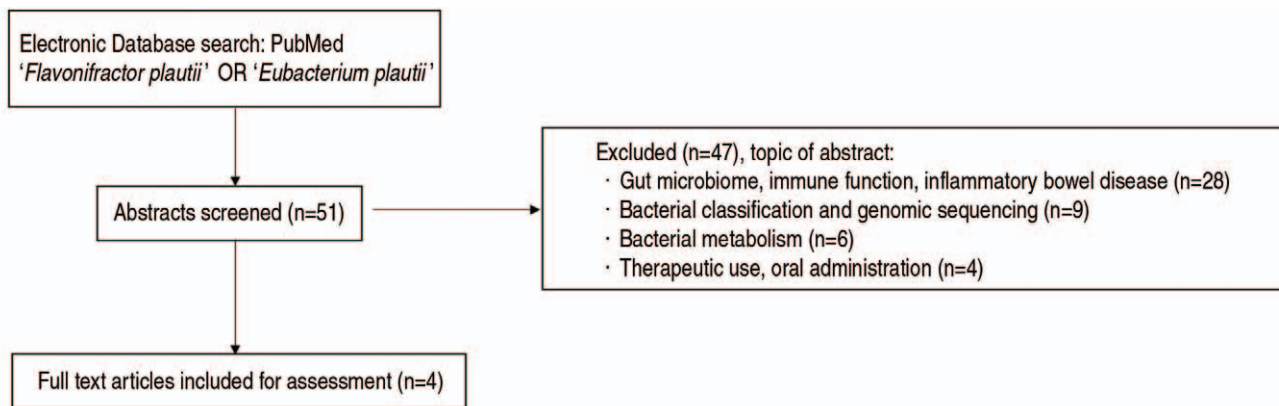


Fig. 7. Flow chart for the literature review.

Table 2. Summary of Previously Documented Human Infections with *Flavonifractor plautii*

| Study | Age (yr)/sex | Pathology | Immuno-suppression | Site (s) cultured | Time to culture | Gram stain |
|--------------------------------------|--------------|---------------------------------|----------------------------------|------------------------|--------------------------|------------|
| Garre et al. ²³ [1991] | 45/M | Dog bite | Asplenic | Blood CSF | After patient discharged | Negative |
| Orlando et al. ²⁴ [2008] | 33/M | Laparotomy for perforated ileum | Renal transplant graft rejection | Blood Pleural fluid | Day 4 | Irregular |
| Berger et al. ²¹ [2018] | 69/M | Necrotising cholecystitis | Chemotherapy | Blood | Day 4 | Positive |
| Costescu et al. ²² [2022] | 45/M | Severe burns | Burns | Blood | Day 3 | Negative |

M: male.

pling of intraoperative tissue should have been performed prior to the start of antibiotic therapy; however, access to operating theatres was limited by the impact of the COVID-19 pandemic and urgent antibiotic therapy was required for treatment of the patient's clinically septic condition. This was likely the reason that subsequent intraoperative specimens remained culture negative. Accurate diagnosis may be compromised by administration of preoperative antibiotics, which has been implicated in cases of culture negative PJI³⁶⁻³⁸). Nonetheless, the patient fulfilled the definition for PJI that was established by the 2018 validated International Consensus Meeting (ICM), and elevated serum and synovial markers met the minor criteria for diagnosis³¹).

Sampling of multiple hip aspirate cultures was also performed prior to the second stage for augmentation of the standard PJI protocol. Catastrophic outcomes related to antibiotic resistance and high virulence of organisms have been reported in multiple case reports of clostridium PJI; therefore, these negative cultures were obtained in order to ensure confidence in the eradication of the atypical organism^{10,15}).

This study describes only a single patient infected with a rare organism, thus, it is limited in its utility for use in guiding the management of anaerobic PJI. With increasing rates of arthroplasty and PJI, conduct of further research for stratification of patient risk and for better understanding of the implications of atypical anaerobic infections will be required.

The case described in this report highlights a successful pathway for management of treatment for atypical anaerobic PJI and the results of a review of the literature confirmed the organism as novel. The pathway included staged surgical revisions with explantation of the prosthesis and use of a temporary antimicrobial-impregnated cement spacer. Effective eradication of the organism was achieved by administration of extended therapy with ceftriaxone, amoxicillin/clavulanic acid, and metronidazole. Secondary definitive arthroplasty was performed safely following fine needle aspiration in order to confirm eradication of the organism.

CONFLICT OF INTEREST

The authors declare that there is no potential conflict of interest relevant to this article.

REFERENCES

1. Bozic KJ, Ries MD. *The impact of infection after total hip arthroplasty on hospital and surgeon resource utilization. J Bone Joint Surg Am.* 2005;87:1746-51.

2. Ong KL, Kurtz SM, Lau E, Bozic KJ, Berry DJ, Parvizi J. *Prosthetic joint infection risk after total hip arthroplasty in the Medicare population. J Arthroplasty.* 2009;24(6 Suppl):105-9. <https://doi.org/10.1016/j.arth.2009.04.027>
3. Grammatopoulos G, Munemoto M, Inagaki Y, Tanaka Y, Athanasou NA. *The diagnosis of infection in metal-on-metal hip arthroplasties. J Arthroplasty.* 2016;31:2569-73. <https://doi.org/10.1016/j.arth.2016.03.064>
4. Pace F, Randelli F, Serrao L, Banci L. *Infection rate in ASR metal on metal hip replacement: possible correlation between high serum levels of metal ions and infection. Orthop Proc.* 2013;95-B(Suppl 34):495.
5. Pulido L, Ghanem E, Joshi A, Purtill JJ, Parvizi J. *Periprosthetic joint infection: the incidence, timing, and predisposing factors. Clin Orthop Relat Res.* 2008;466:1710-5. <https://doi.org/10.1007/s11999-008-0209-4>
6. Tsai Y, Chang CH, Lin YC, Lee SH, Hsieh PH, Chang Y. *Different microbiological profiles between hip and knee prosthetic joint infections. J Orthop Surg (Hong Kong).* 2019;27:2309499019847768. <https://doi.org/10.1177/2309499019847768>
7. Bjerke-Kroll BT, Christ AB, McLawhorn AS, Sculco PK, Jules-Elysée KM, Sculco TP. *Periprosthetic joint infections treated with two-stage revision over 14 years: an evolving microbiology profile. J Arthroplasty.* 2014;29:877-82. <https://doi.org/10.1016/j.arth.2013.09.053>
8. Shah NB, Tande AJ, Patel R, Barbari EF. *Anaerobic prosthetic joint infection. Anaerobe.* 2015;36:1-8. <https://doi.org/10.1016/j.anaerobe.2015.08.003>
9. Ranganath S, Midturi JK. *Unusual case of prosthetic shoulder joint infection due to Clostridium difficile. Am J Med Sci.* 2013;346:422-3. <https://doi.org/10.1097/MAJ.0b013e3182987d05>
10. Brassinne L, Rodriguez-Villalobos H, Jonckheere S, Dubuc JE, Yombi JC. *Early infection of hip joint prosthesis by Clostridium difficile in an HIV-1 infected patient. Anaerobe.* 2014;27:96-9. <https://doi.org/10.1016/j.anaerobe.2014.03.007>
11. Curtis L, Lipp MJ. *Clostridium difficile infection of a prosthetic knee joint requiring amputation. Surg Infect (Larchmt).* 2013;14:163-4. <https://doi.org/10.1089/sur.2012.098>
12. Morshed S, Malek F, Silverstein RM, O'Donnell RJ. *Clostridium cadaveris septic arthritis after total hip arthroplasty in a metastatic breast cancer patient. J Arthroplasty.* 2007;22:289-92. <https://doi.org/10.1016/j.arth.2006.02.158>
13. McCarthy J, Stingemore N. *Clostridium difficile infection of a prosthetic joint presenting 12 months after antibiotic-associated diarrhoea. J Infect.* 1999;39:94-6. [https://doi.org/10.1016/s0163-4453\(99\)90110-x](https://doi.org/10.1016/s0163-4453(99)90110-x)
14. Achong DM, Oates E. *Periprosthetic Clostridium difficile hip abscess imaged with In-111 WBCs. Clin Nucl Med.* 1994;19:860-2. <https://doi.org/10.1097/00003072-199410000-00002>
15. Pron B, Merckx J, Touzet P, et al. *Chronic septic arthritis and osteomyelitis in a prosthetic knee joint due to Clostridium difficile. Eur J Clin Microbiol Infect Dis.* 1995;14:599-601. <https://doi.org/10.1007/BF01690732>
16. Stern SH, Sculco TP. *Clostridium perfringens infection in a total knee arthroplasty. A case report. J Arthroplasty.* 1988;3 Suppl:S37-40. [https://doi.org/10.1016/s0883-5403\(88\)80006-8](https://doi.org/10.1016/s0883-5403(88)80006-8)

17. Wilde AH, Sweeney RS, Borden LS. Hematogenously acquired infection of a total knee arthroplasty by *Clostridium perfringens*. *Clin Orthop Relat Res*. 1988;(229):228-31. <https://doi.org/10.1097/00003086-198804000-00031>
18. Kibbler CC, Jackson AM, Grüneberg RN. Successful antibiotic therapy of clostridial septic arthritis in a patient with bilateral total hip prostheses. *J Infect*. 1991;23:293-5. [https://doi.org/10.1016/0163-4453\(91\)93008-z](https://doi.org/10.1016/0163-4453(91)93008-z)
19. Hill GB, Ayers OM, Kohan AP. Characteristics and sites of infection of *Eubacterium nodatum*, *Eubacterium timidum*, *Eubacterium brachy*, and other asaccharolytic eubacteria. *J Clin Microbiol*. 1987;25:1540-5. <https://doi.org/10.1128/jcm.25.8.1540-1545.1987>
20. Carlier JP, Bedora-Faure M, K'ouas G, Alauzet C, Mory F. Proposal to unify *Clostridium orbiscindens* Winter et al. 1991 and *Eubacterium plautii* (Séguin 1928) Hofstad and Aasjord 1982, with description of *Flavonifractor plautii* gen. nov., comb. nov., and reassignment of *Bacteroides capillosus* to *Pseudoflavonifractor capillosus* gen. nov., comb. nov. *Int J Syst Evol Microbiol*. 2010;60(Pt 3):585-90. <https://doi.org/10.1099/ijs.0.016725-0>
21. Berger FK, Schwab N, Glanemann M, Bohle RM, Gärtner B, Groesdonk HV. *Flavonifractor (Eubacterium) plautii* bloodstream infection following acute cholecystitis. *IDCases*. 2018;14:e00461. <https://doi.org/10.1016/j.idcr.2018.e00461>
22. Costescu Strachinaru DI, Gallez JL, Daras S, et al. A case of *Flavonifractor plautii* blood stream infection in a severe burn patient and a review of the literature. *Acta Clin Belg*. 2022;77:693-97. <https://doi.org/10.1080/17843286.2021.1944584>
23. Garre M, le Henaff C, Tande, et al. Fulminant *Eubacterium plautii* infection following dog bite in asplenic man. *Lancet*. 1991;338:384-5. [https://doi.org/10.1016/0140-6736\(91\)90518-t](https://doi.org/10.1016/0140-6736(91)90518-t)
24. Orlando G, Pisani F, Mastrantonio P, et al. *Eubacterium plautii* infection in a kidney transplant recipient: a noteworthy case of pleural effusion and fever. *Clin Transplant*. 2008;22:520-4. <https://doi.org/10.1111/j.1399-0012.2008.00805.x>
25. Gupta A, Dhakan DB, Maji A, et al. Association of *Flavonifractor plautii*, a flavonoid-degrading bacterium, with the gut microbiome of colorectal cancer patients in India. *mSystems*. 2019;4:e00438-19. <https://doi.org/10.1128/mSystems.00438-19>
26. Huang R, Li F, Zhou Y, et al. Metagenome-wide association study of the alterations in the intestinal microbiome composition of ankylosing spondylitis patients and the effect of traditional and herbal treatment. *J Med Microbiol*. 2020;69:797-805. <https://doi.org/10.1099/jmm.0.001107>
27. Yang Y, Du L, Shi D, et al. Dysbiosis of human gut microbiome in young-onset colorectal cancer. *Nat Commun*. 2021;12:6757. <https://doi.org/10.1038/s41467-021-27112-y>
28. Ogita T, Yamamoto Y, Mikami A, Shigemori S, Sato T, Shimosato T. Oral administration of *Flavonifractor plautii* strongly suppresses Th2 immune responses in mice. *Front Immunol*. 2020;11:379. <https://doi.org/10.3389/fimmu.2020.00379>
29. Li W, Sun Y, Dai L, et al. Ecological and network analyses identify four microbial species with potential significance for the diagnosis/treatment of ulcerative colitis (UC). *BMC Microbiol*. 2021;21:138. <https://doi.org/10.1186/s12866-021-02201-6>
30. Johnson MJ, Thatcher E, Cox ME. Techniques for controlling variability in gram staining of obligate anaerobes. *J Clin Microbiol*. 1995;33:755-8. <https://doi.org/10.1128/jcm.33.3.755-758.1995>
31. Parvizi J, Tan TL, Goswami K, et al. The 2018 definition of periprosthetic hip and knee infection: an evidence-based and validated criteria. *J Arthroplasty*. 2018;33:1309-14.e2. <https://doi.org/10.1016/j.arth.2018.02.078>
32. Marculescu CE, Cantey JR. Polymicrobial prosthetic joint infections: risk factors and outcome. *Clin Orthop Relat Res*. 2008;466:1397-404. <https://doi.org/10.1007/s11999-008-0230-7>
33. Song Y, Shao HY, Cheng X, Guo Y. First case of periprosthetic joint infection due to *Clostridioides difficile* in China. *BMC Infect Dis*. 2021;21:462. <https://doi.org/10.1186/s12879-021-06171-y>
34. Menni C, Klaser K, May A, et al. Vaccine side-effects and SARS-CoV-2 infection after vaccination in users of the COVID Symptom Study app in the UK: a prospective observational study. *Lancet Infect Dis*. 2021;21:939-49. [https://doi.org/10.1016/S1473-3099\(21\)00224-3](https://doi.org/10.1016/S1473-3099(21)00224-3)
35. Cooper HJ, Della Valle CJ. The two-stage standard in revision total hip replacement. *Bone Joint J*. 2013;95-B(11 Suppl A):84-7. <https://doi.org/10.1302/0301-620X.95B11.32906>
36. Shahi A, Deirmengian C, Higuera C, et al. Premature therapeutic antimicrobial treatments can compromise the diagnosis of late periprosthetic joint infection. *Clin Orthop Relat Res*. 2015;473:2244-9. <https://doi.org/10.1007/s11999-015-4142-z>
37. Malekzadeh D, Osmon DR, Lahr BD, Hanssen AD, Barbari EF. Prior use of antimicrobial therapy is a risk factor for culture-negative prosthetic joint infection. *Clin Orthop Relat Res*. 2010;468:2039-45. <https://doi.org/10.1007/s11999-010-1338-0>
38. Cascone VJ, Cohen RS, Dodson NP, Cannon CM. Implications of culture collection after the first antimicrobial dose in septic emergency department patients. *Am J Emerg Med*. 2019;37:947-51. <https://doi.org/10.1016/j.ajem.2019.02.016>