

RESEARCH PAPER

Isolation of Three Unrecorded Yeasts from the Guts of Earthworms Collected from Korea

Hyejin Oh, and Myung Kyum Kim*

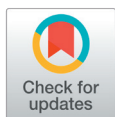
Department of Bio & Environmental Technology, College of Natural Science, Seoul Women's University, Seoul 01797, Korea

*Corresponding author: biotech@swu.ac.kr

ABSTRACT

In 2021, seven yeast strains were isolated from earthworm (*Eisenia andrei*) gut samples collected from the Nanji Water Regeneration Center in Goyang City, Gyeonggi Province, Korea. A total of seven yeasts were isolated, of which three strains have not been previously reported in Korea. To identify the yeasts, pairwise sequence comparisons of large subunit (LSU) rDNA sequences were performed using the basic local alignment search tool (BLAST). Assimilation test and cell morphology analysis were performed using the API 20C AUX kit and phase contrast microscope, respectively. Five of the seven strains were assigned to the genus *Candida* of the order Saccharomycetales of the class Saccharomycetes, and two to the genus *Apiotrichum* of the order Trichosporonales of the class Tremellomycetes. The yeast strain *Candida sojae* E2 belongs to the family Debaryomycetaceae, and *Apiotrichum laibachii* E8 and *A. laibachii* E9 belong to the family Trichosporonaceae. All strains were cultured in yeast mold agar for three days and showed different colony forms. *C. sojae* E2 was round and entire shaped, while *A. laibachii* E8 and *A. laibachii* E9 was round and convex shaped. This study focuses on the description of the three yeast strains that have not been officially reported in Korea.

Keywords: *Apiotrichum*, *Candida*, Unreported yeasts, 26S rRNA



OPEN ACCESS

pISSN : 0253-651X
eISSN : 2383-5249

Kor. J. Mycol. 2021 December, 49(4): 547-553
<https://doi.org/10.4489/KJM.20210054>

Received: October 08, 2021

Revised: November 15, 2021

Accepted: November 20, 2021

© 2021 THE KOREAN SOCIETY OF MYCOLOGY.



This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/4.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

INTRODUCTION

Microbiome research of soil invertebrates have received much attention [1,2]. Recently, a study reported that terrestrial insects, such as collembolans, earthworms, and nematodes, are associated with an abundant microbiome and putative symbionts [3]. Earthworms are excellent bioindicators of soil quality and perform an important role in increasing soil aeration, infiltration, structure, nutrient cycling, water movement, and plant growth [4].

In 2021, seven strains were isolated from earthworm (*Eisenia andrei*) gut samples collected from the Nanji Water Regeneration Center in Korea. Among the seven strains, five were Ascomycota yeast and two were Basidiomycota yeast. The five strains were assigned to the genus *Candida* of the order Saccharomycetales of class Saccharomycetes in the phylum Ascomycota, and two were assigned to the genus *Apiotrichum* (two strains) of the order Trichosporonales of the class Tremellomycetes in the phylum

Basidiomycota. Three of the isolated strains; E2, E8, and E9, have not previously been reported in Korea.

The genus *Candida* consists of 314 recognized species and has *C. vulgaria* [5] as a type species. *Candida* species have been isolated from human or animal excrement and aquatic environments containing urban sewage effluent [6-8], flowers, and insects [9-12]. The cells are globose, ellipsoidal, cylindrical, or elongate, sometimes with ogival, triangular, or lunate shape, and reproduce through holoblastic budding [5].

The genus *Apiotrichum* consists of 22 reported species [13,14] and has *Apiotrichum* (= *Trichosporon*) *porosum* [15] as the type species. The genus *Apiotrichum* was redefined to accommodate gracile/brassicae [15-18]. Yeasts of this genus are found distributed in nature, on sources such as cabbage, rotten wood, sour milk, and grassland [19] and many species have been isolated from the soil [17,18]. *Apiotrichum* presents pseudohyphae and budding cells, but basidiocarps, sexual reproduction, fermentation, and nitration [15,17].

MATERIALS AND METHODS

The host worms were collected from the Nanji Water Regeneration Center in Goyang City, Gyeonggi Province, Korea, and a total of seven strains were isolated from the intestinal tract of the worms. The intestine was separated from *E. andrei*, cut to approximately 2 cm with tweezers, pulverized, and washed continuously with distilled water [20]. The suspension containing the microbes was spread (100 μ L each) on yeast mold agar (YM agar; Difco, Franklin Lakes, USA). Single yeast colonies were purified and maintained on YM agar containing 25% (w/v) glycerol suspension at -80°C in a deep freezer [21,22]. Information on the designated strain identifications (IDs), recent associations, 26S rRNA similarities, and internal transcribed spacer (ITS) similarities for isolated strains are described in Table 1.

The incubation period is generally 23 days at 25°C, but the incubation temperature and times range between 4-42°C and 3 days or more, respectively, for each strain [23]. Growth tests were performed on different media, such as YM agar, corn meal agar (CMA; Difco, Franklin Lakes, USA), and potato dextrose agar (PDA; Difco, Franklin Lakes, USA), at 10°C (Fig. 1). Budding and cell morphology were observed using a phase-contrast microscope (Leica DM500, Wetzlar, Germany), using pure cultured cells incubated for 3-5 days in YM agar. Phase contrast microscope images and pictures of plates containing the strains are shown in Fig. 2. The API 20C AUX kit (BioMérieux, Marcy-l'Étoile, France) was used to determine the carbon source assimilation of the unrecorded yeast strains.

The yeast used for DNA extraction was subcultured on YM agar and incubated at 10°C for 3-5 days. Genomic DNA was extracted using a cDNA Synthesis Kit (NanoHelix, Daejeon, Korea) according to the manufacturer's instructions. For strain classification, the sequences were analyzed by amplifying the D1/D2 region of the large subunit (LSU) RNA gene using the universal primers NL1 (5'-GCATCAAGGAGAG-3') and NL4 (5'-GTCGTTTCAGG-3') [24]. The types of yeast closely associated with the isolated strains were collected using the MYCOBANK database (<https://www.mycobank.org/>). The LSU rDNA sequences of the strains were obtained from NCBI (<https://www.ncbi.nlm.nih.gov/>), and homology with other yeast was compared using the Basic Local Alignment Search Tool (BLAST) database of NCBI. The gene sequences were edited using the Seqman program and used for constructing phylogenetic trees using the neighbor-

joining algorithm [25] of the MEGA7 program [26]. The phylogenetic tree topology was evaluated for statistical reliability based on bootstrap values for 1,000 replications [27] and using the GenBank registration number. The evolutionary distance was calculated using the Kimura 2-parameter model [28].

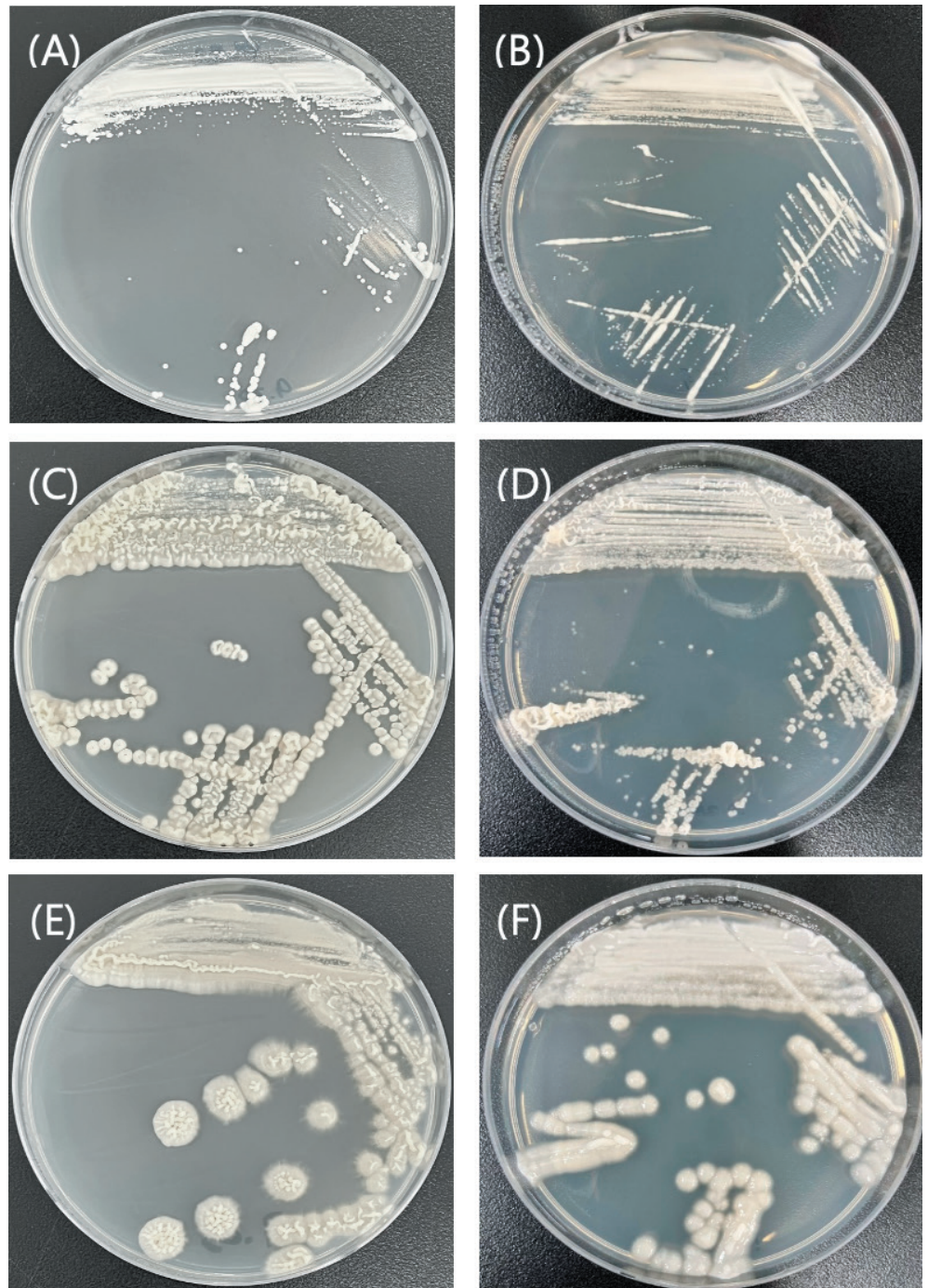


Fig. 1. Morphology of cells from the unrecorded strains incubated at 10°C. The colonies of (A) *Candida sojae* E2, (C) *Apiotrichum laibachii* E8, and (E) *A. laibachii* E9 cultured in yeast mold agar (YM). The colonies of (B) *C. sojae* E2, (D) *A. laibachii* E8, and (F) *A. laibachii* E9 cultured in potato dextrose agar (PDA).

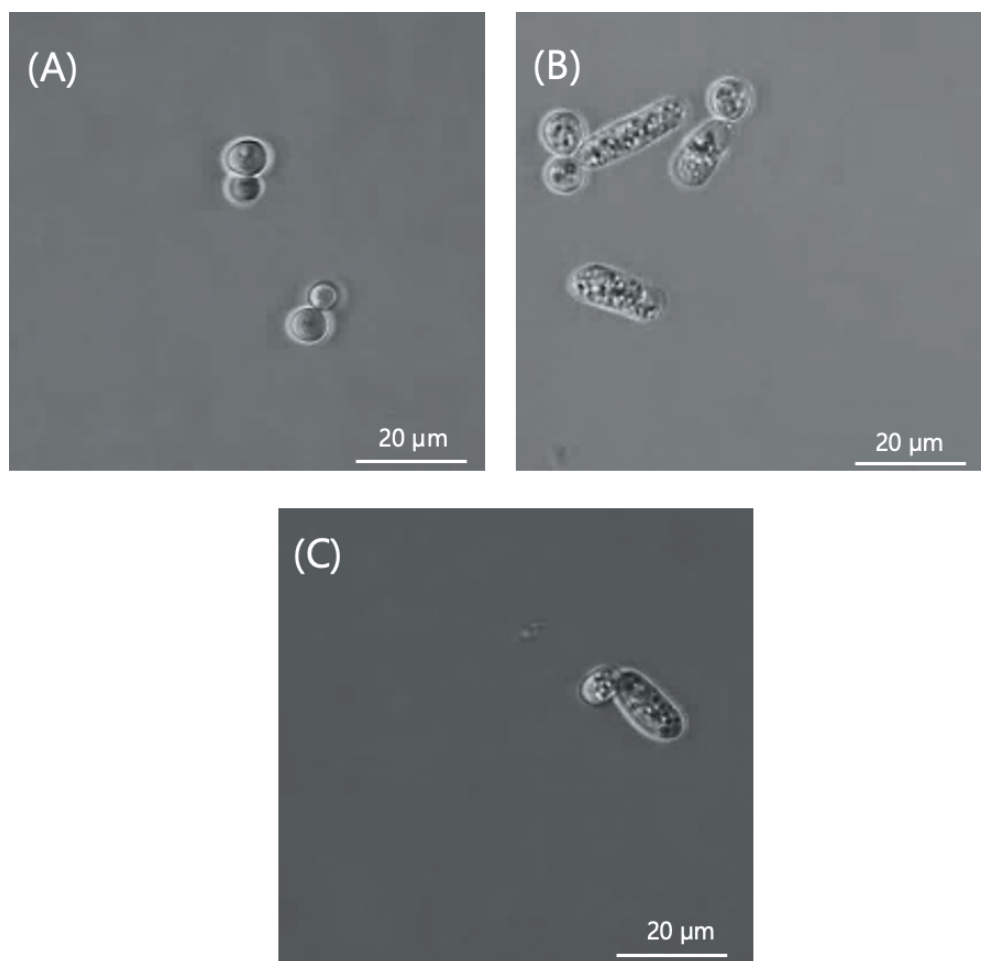


Fig. 2. Micromorphology of budding cells on yeast mold agar (YM) plates after 3-5 days at 10°C. The cells of (A) *Candida sojae* E2, (C) *Apiotrichum laibachii* E8, and (E) *A. laibachii* E9. Bars, 20 µm.

Table 1. Summary of isolated yeast strains and their taxonomic affiliations. All strains were cultured under 10°C for 3 days.

Phylum	Class	Order	Family	Strain ID	Most closely related species	D1/D2 identity	ITS identity	Record in Korea				
Ascomycota	Saccharomycetes	Saccharomycetales	Debaryomycetaceae	E2	<i>Candida sojae</i>	557/557 (100%)	501/504 (99%)	Unreported				
				E3	<i>Candida palmiophila</i>	567/567 (100%)	604/610 (99%)	Reported				
				E5	<i>Candida subhashii</i>	590/591 (100%)	685/685 (100%)	Reported				
				E6	<i>Candida palmiophila</i>	567/567 (100%)	601/607 (99%)	Reported				
				E7	<i>Candida palmiophila</i>	554/555 (100%)	595/595 (100%)	Reported				
				Basidiomycota	Tremellomycetes	Trichosporonales	Trichosporonaceae	E8	<i>Apiotrichum laibachii</i>	560/561 (100%)	527/527 (100%)	Unreported
								E9	<i>Apiotrichum laibachii</i>	561/561 (100%)	526/526 (100%)	Unreported

ITS: Internal transcribed spacer.

RESULTS AND DISCUSSION

A total of seven strains of yeast were separated from the Nanji Water Regeneration Center in Gyeonggi Province, Korea. By analyzing the similarities in the small subunit (SSU) rDNA and D1/D2 region of LSU rDNA sequence, we found that three of the separated strains were unrecorded in Korea. There was a higher proportion of Ascomycota yeast among the strains isolated from the internal contents of the *E. andrei*, compared to Basidiomycota yeast. Five of the seven strains were identified as belonging to the family Debaryomycetaceae of the phylum Ascomycota, while the remaining two strains were classified into Trichosporonaceae of the phylum Basidiomycota. The D1/D2 and ITS similarities between the strains and their closely related species and their taxonomic composition are summarized in Table 1. The characteristics of the unrecorded strains are presented in Table 2.

Three unrecorded yeasts were identified: *Candida* (1 strain) and *Apiotrichum* (2 strains). The phylogenetic consensus trees of the three strains support a close relationship by demonstrating that the isolated strains are closely linked to strains that exhibit the highest LSU rDNA sequence similarity (Fig. 3 and 4).

Based on previous phylogenetic and biochemical studies, this study identified three unrecorded yeasts in the domestic ecosystem and investigated their phenotypic characteristics.

Table 2. Characteristics of the unrecorded strains from worm gut.

Strain ID	Yeast Strains		
	E2	E8	E9
Morphological characteristics			
Shape	Oval	Oval	Oval
Vegetative reproduction	Budding	Budding	Budding
API 20C AUX			
Glucose	+	-	+
Glycerol	-	-	+
2-Keto-D-gluconate	+	-	+
L-Arabinose	-	-	+
D-Xylose	+	-	+
Adonitol	+	-	-
Xylitol	-	-	-
D-Galactose	+	-	+
Inositol	+	-	+
D-Sorbitol	+	-	-
d-Methyl-D-glucoside	-	-	+
N-Acetyl-D-glucosamine	+	+	+
D-Cellobiose	-	-	-
D-Lactose (bovine origin)	-	-	+
D-Maltose	+	-	-
D-Saccharose (Sucrose)	+	-	+
D-Trehalose	-	-	+
D-Melezitose	+	-	-
D-Raffinose	-	-	+

All data were obtained in this study. +, positive; w, weakly positive; -, negative.

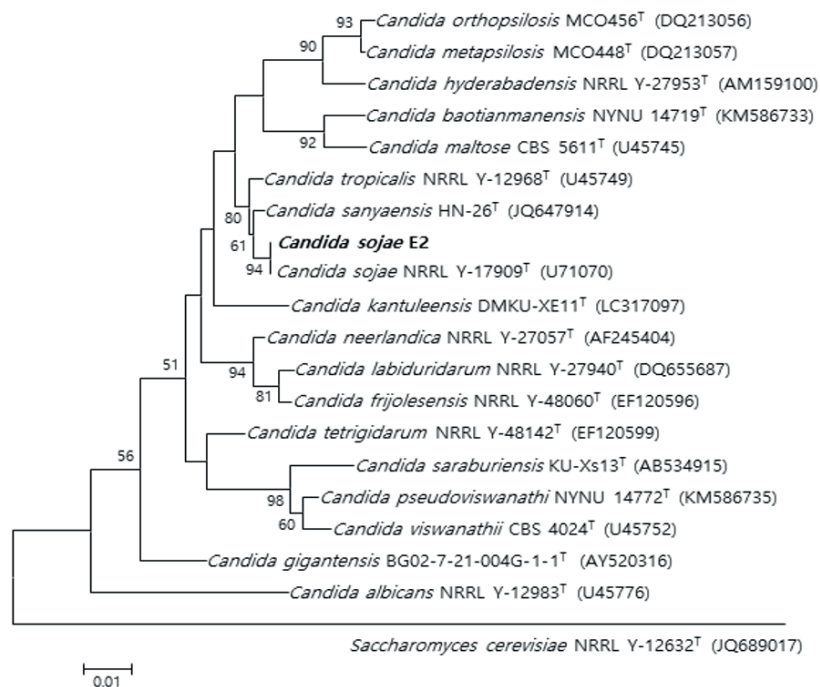


Fig. 3. Neighbor-joining phylogenetic tree based on 26S rRNA gene sequences shows the phylogenetic relationships between the strain E2 and their closest strains of the genus *Candida*. *Saccharomyces cerevisiae* NRRL Y-12632^T strain was used as the outgroup. Bootstrap values (>50 %) are shown at each branch. Bar, 0.01 substitutions per nucleotide position. ^T: Type strain. Bold type font: isolated strain.

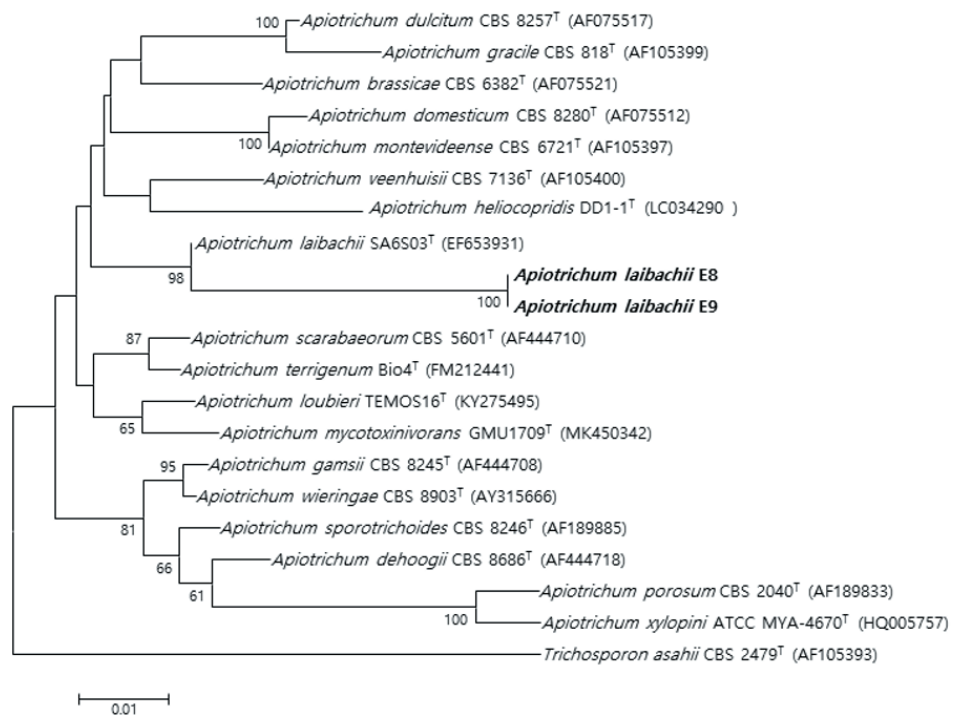


Fig. 4. Neighbor-joining phylogenetic tree based on 26S rRNA gene sequences shows the phylogenetic relationships between the strains E8, E9 and their closest strains of the genus *Apiotrichum*. *Trichosporon asahii* CBS 2479^T strain was used as the outgroup. Bootstrap values (>50 %) are shown at each branch. Bar, 0.01 substitutions per nucleotide position. ^T: Type strain. Bold type font: isolated strain.

Description of *Candida sojae* E2

Colonies are convex, entire, and cream-colored after 3 days of incubation on YM agar and PDA at 10°C. The strain grew on YM, PDA, and CMA (weakly) (Fig. 1). In the API 20C AUX test, strain E2 was positive for D-sorbitol, inositol, glucose, D-melezitose, D-saccharose (sucrose), D-maltose, D-galactose, D-xylose, adonitol, N-acetyl-D-glucosamine, and 2-keto-D-gluconate. However, it was negative for D-raffinose, D-trehalose, D-cellobiose, d-methyl-D-glucoside, xylitol, glycerol, L-arabinose, and D-lactose (bovine origin). Strain E2 (KCTC 27832) was isolated from the intestinal tract of the earthworms obtained from the Nanji Water Regeneration Center, Gyeonggi Province, Korea.

Description of *Apiotrichum laibachii* E8

Colonies were entire, umbonate, and cream-colored after 3 days of incubation on YM agar at 10°C and observed to form an avillous appearance in which the center protrudes. The strain grew on YM, PDA, and CMA (weakly) (Fig. 1). In the API 20C AUX test, strain E8 was positive for N-acetyl-D-glucosamine. However, it was negative for glucose, glycerol, 2-keto-D-gluconate, L-arabinose, D-sorbitol, d-methyl-D-glucoside, D-cellobiose, D-lactose (bovine origin), D-maltose, D-saccharose (sucrose), D-trehalose, D-melezitose, xylitol, D-galactose, adonitol, inositol, and D-raffinose. Strain E8 (KCTC 27842) was isolated from the intestinal tract of the earthworms obtained from the Nanji Water Regeneration Center, Gyeonggi Province, Korea.

Description of *Apiotrichum laibachii* E9

Colonies were entire, umbonate, and cream-colored after 3 days of incubation on YM agar and PDA at 10°C and observed to form an avillous appearance in which the center protrudes. The strain grew on YM, PDA, and CMA (weakly) (Fig. 1). In the API 20C AUX test, strain E9 was positive for D-lactose (bovine origin), N-acetyl-D-glucosamine, d-methyl-D-glucoside, D-galactose, glucose, 2-keto-D-gluconate, L-arabinose, D-xylose, D-saccharose (sucrose), D-trehalose, and D-raffinose. However, it was negative for D-melezitose, D-cellobiose, xylitol, adonitol, D-maltose, D-sorbitol, glycerol, and inositol. Strain E9 (KCTC 27843) was isolated from the intestinal tract of the earthworms from the Nanji Water Regeneration Center, Gyeonggi Province, Korea.

ACKNOWLEDGMENT

This work was supported by a grant from the National Institute of Biological Resources (NIBR), funded by the Ministry of Environment (MOE) of the Republic of Korea (NIBR202130202).

REFERENCES

1. May RM. How many species are there on earth?. *Science* 1988; 241:1441-9.
2. Bahrndorff S, Alemu T, Alemneh T, Lund Nielsen J. The microbiome of animals: implications for conservation biology. *Int J Gen* 2016; e5304028.
3. Hosokawa T, Kikuchi Y, Nikoh N, Shimada M, Fukatsu T. Strict host-symbiont cospeciation and reductive genome evolution in insect gut bacteria. *PLoS biology* 2006; 4:e337.
4. Vandecasteele B, Samyn J, Quataert P, Muys B, Tack FMG. Earthworm biomass as additional information for risk assessment of heavy metal biomagnification, a case study for dredged sediment-derived soils and polluted floodplain soils. *Environ Pollut* 2004; 129:363-75.
5. Lachance MA, Boekhout T, Scorzetti G, Fell JW, Kurtzman CP. *Candida* Berkhout (1923). In *The yeasts* 2011; Elsevier, 987-1278.
6. Cook WL, Schlitzer RL. Isolation of *Candida albicans* from freshwater and sewage. *Appl Environ Microbiol* 1981; 41:840-42.
7. Calderone R. Taxonomy and biology of *Candida*. In: R. Calderone (Ed.), *Candida and Candidiasis*. ASM Press 2002; 15-29.
8. Bougnoux ME, Aanensen DM, Morand S, Théraud M, Spratt B. G, d'Enfert C. Multilocus sequence typing of *Candida albicans*: strategies, data exchange and applications. *Infect Genet Evol* 2004; 4:243-52.
9. Lachance MA, Rosa CA, Starmer WT, Bowles JM. *Candida ipomoeae*, a new yeast species related to large-spored *Metschnikowia* species. *Can J Microbiol* 1998; 44:718-22.
10. Lachance MA, Starmer WT, Rosa CA, Bowles JM, Barker JSF, Janzen DH. Biogeography of the yeasts of ephemeral flowers and their insects. *FEMS Yeast Res.* 2001; 1:1-8.
11. Hong SG, Bae KS, Herzberg M, Titze A, Lachance MA. *Candida kunwiensis* sp. nov., a yeast associated with flowers and bumblebees. *Int J Syst Evol Microbiol* 2003; 53:367-72.
12. Nguyen NH, Suh SO, Erbil CK, Blackwell M. *Metschnikowia noctiluminum* sp. nov., *Metschnikowia corniflorae* sp. nov., and *Candida chrysolidarum* sp. nov., isolated from green lacewings and beetles. *J mycol res* 2006; 110:346-56.
13. Takashima M, Manabe RI, Nishimura Y, Endoh R, Ohkuma M, Sriswasdi S, Iwasaki W. Recognition and delineation of yeast genera based on genomic data: Lessons from Trichosporonales. *Fungal Genet Biol* 2019; 130:31-42.
14. Takashima M, Kurakado S, Cho O, Kikuchi K, Sugiyama J, Sugita T. Description of four *Apiotrichum* and two *Cutaneotrichosporon* species isolated from guano samples from bat-inhabited caves in Japan. *Int J Syst Evol Microbiol* 2020; 70:4458-69.
15. Liu XZ, Wang QM, Theelen B, Groenewald M, Bai FY, Boekhout T. Phylogeny of tremellomycetous yeasts and related dimorphic and filamentous basidiomycetes reconstructed from multiple gene sequence analyses. *Stud Mycol* 2015; 81:1-26.
16. Middelhoven WJ, Scorzetti G, Fell JW. Systematics of the anamorphic basidiomycetous yeast genus *Trichosporon* Behrend with the description of five novel species: *Trichosporon vadense*, *T. smithiae*, *T. dehoogii*, *T. scarabaeorum* and *T. gamsii*. *Int J Syst Evol Microbiol* 2004; 54: 975-86.
17. Sugita T. *Trichosporon behrend* (1890). In *The yeasts* 2011; Elsevier, 2015-2061.

18. James SA, Bond CJ, Stanley R, Ravella SR, Péter G, Dlačny D, Roberts IN. *Apiotrichum terrigenum* sp. nov., a soil-associated yeast found in both the UK and mainland Europe Int J Syst Evol Microbiol 2016; 66:5046.
19. Aliyu H, Gorte O, De Maayer P, Neumann A, Ochsenreither K. Genomic insights into the lifestyles, functional capacities and oleagenicity of members of the fungal family Trichosporonaceae. Sci Rep 2020; 10:1-12.
20. Tanahashi M, Kubota K, Matsushita N, Togashi K. Discovery of mycangia and the associated xylose-fermenting yeasts in stag beetles (Coleoptera: Lucanidae). Sci Nat 2010; 97:311-317.
21. Yarrow D. Methods for the isolation, maintenance and identification of yeasts. The yeasts 1998; Elsevier, 77–100.
22. Nguyen NH, Suh SO, Blackwell M. Five novel *Candida* species in insect-associated yeast clades isolated from *Neuroptera* and other insects. Mycologia 2007; 99:842-858.
23. Kurtzman CP, Fell JW, Boekhout T, Robert V. Methods for isolation, phenotypic characterization and maintenance of yeasts. The Yeasts 2011; Elsevier, 87–110.
24. Leaw SN, Chang HC, Sun HF, Barton R, Bouchara JP, Chang TC. Identification of medically important yeast species by sequence analysis of the internal transcribed spacer regions. J Clin Microbiol 2006; 44:693–699.
25. Saitou N, Nei M. The neighbor-joining method: a new method for reconstructing phylogenetic trees. Mol Bio Evol 1987; 4:406-425.
26. Kumar S, Stecher G, Michael Li, Knyaz C, Tamura K. MEGA X: molecular evolutionary genetics analysis across computing platforms. Mol Biol Evol 2018; 35:1547-1549
27. Felsenstein J. Confidence limits on phylogenies: an approach using the bootstrap. Evolution 1985; 39:783-791.
28. Kimura M. The neutral theory of molecular evolution. Cambridge University Press 1983.