

# FRMD7-associated Infantile Nystagmus Syndrome

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Infantile nystagmus syndrome (INS) is a genetically heterogeneous disorder. To date, more than 100 genes have been reported to cause INS and there is significant overlap in phenotypic characteristics. The most common form of X-linked INS is attributed to *FRMD7* at Xq26. Recent advances in molecular genetics have facilitated the identification of pathogenic variants of *FRMD7* and the investigation for underlying mechanisms of *FRMD7*-associated INS. This review summarizes genetic and clinical features of *FRMD7*-associated INS, and introduces updates on the pathogenesis of *FRMD7* mutation.

**Key words:** Infantile nystagmus syndrome, *FRMD7*

## REVIEW

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## INTRODUCTION

Infantile nystagmus syndrome (INS), formerly called congenital nystagmus is characterized by rhythmic involuntary oscillations of the eyes that are present at birth or during infancy [1]. It can be associated with afferent visual system disorders such as ocular albinism, anterior segment dysgenesis, and foveal hypoplasia (Fig. 1). On the other hand, idiopathic INS arises independently of any other visual or neurological disorders. This has led to speculation that idiopathic INS may be caused by abnormal development of the ocular motor system itself rather than disorders of the afferent visual pathway [2,3].

The inheritance patterns of idiopathic INS are heterogeneous and have been reported as autosomal dominant, autosomal recessive, or X-linked trait [4-6]. However, the most common form of inheritance is X-linked, which can be dominant or recessive. Three loci have been identified at Xp11.4-p11.3, Xp22, and Xq26-q27, but approximately 50% of idiopathic INS families have been linked to Xq26-q27. After Tarpey et al. first identified pathogenic mutations in *FRMD7* (MIM#300628) at Xq26, over 90 different mutations have been reported in patients with idiopathic INS [7].

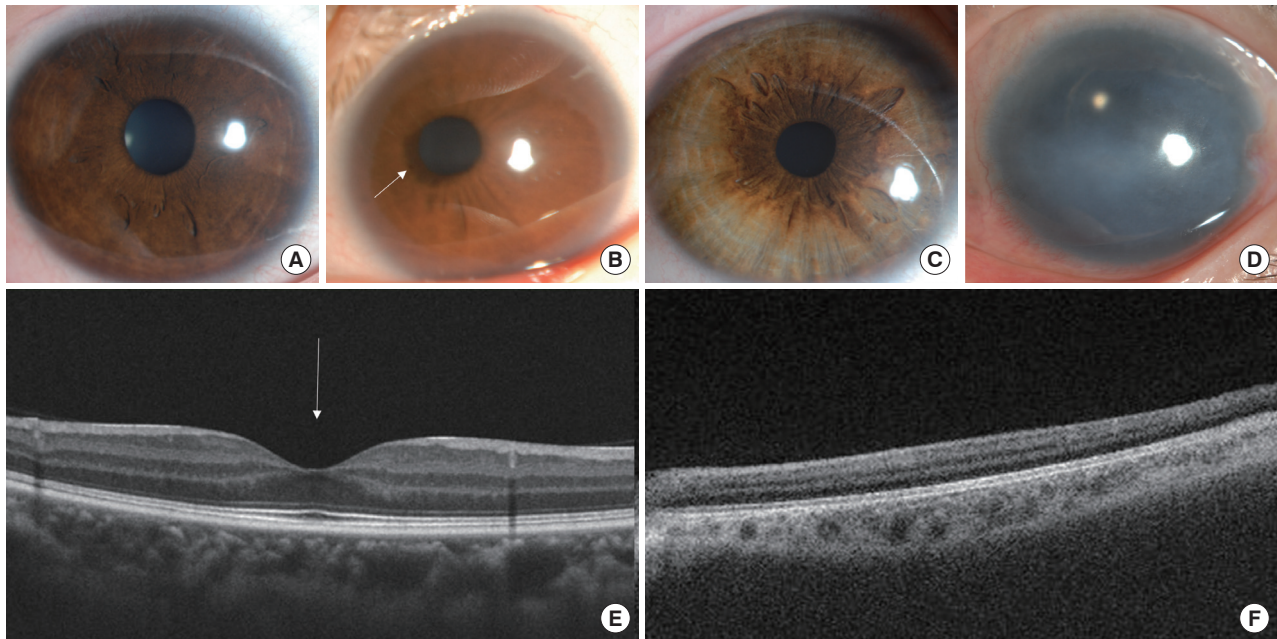
This review summarizes genetic and clinical features of *FRMD7*-associated INS, and introduces updates on the pathogenesis of *FRMD7* mutation.

### FRMD7 (FERM Domain-Containing 7) Structure and Function

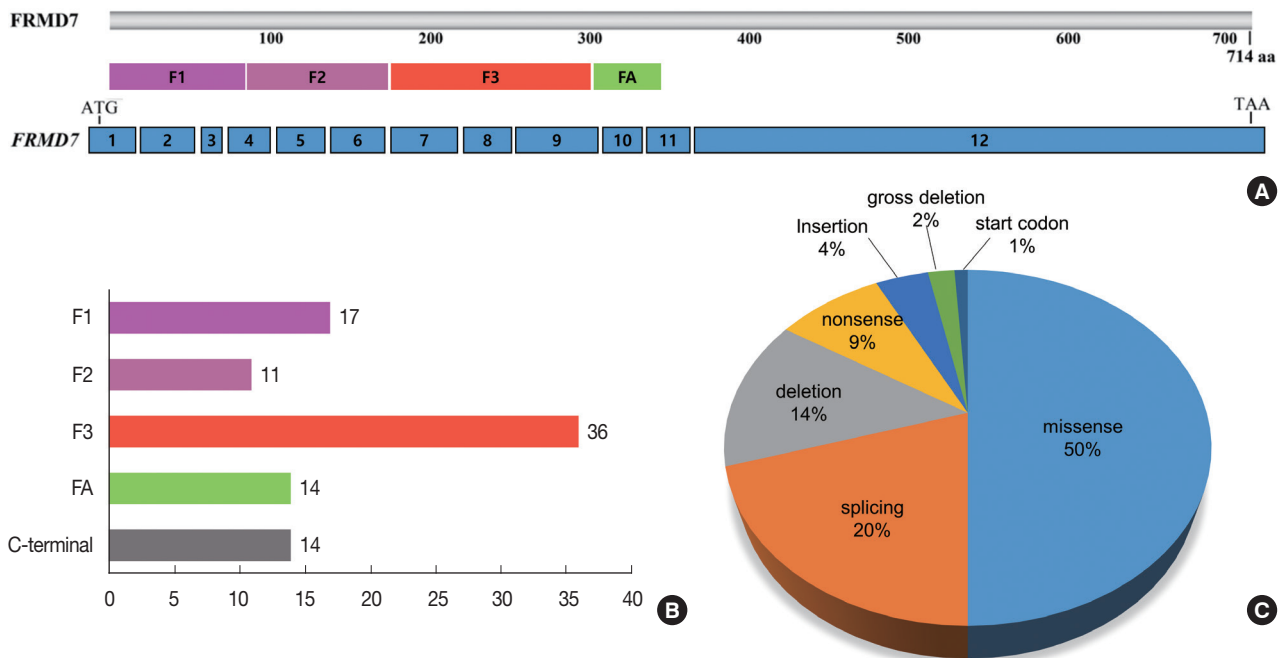
The *FRMD7* gene consists of 12 exons and encodes a 714-residue polypeptide [6]. It contains a conserved N-terminal FERM domain (amino acids 2-282) and FERM-adjacent (FA) domain (amino acids 288-336), whereas the C-terminal region has no significant homology to other proteins (Fig. 2A). The FERM domain has 3 lobed “cloverleaf” structures: F1 (lobe A), F2 (lobe B), and F3 (lobe C). These are plasma membrane-cytoskeleton coupling proteins which bind to actin

or other cytoskeleton components. The FA domain contains conserved motifs that are potential substrates for kinases, suggesting its regulatory effect in FRMD7 protein. In situ hybrid-

ization experiments in human embryonic brain showed FRMD7 expression in neuronal tissues involved in the vestibulo-ocular reflex and optokinetic reflex such as the developing



**Fig. 1.** Afferent visual system disorders associated with infantile nystagmus syndrome. (A) Anterior segment photography of normal eye. (B) Nasally displaced pupil with iris ectropion uvea (white arrow). (C) Iris hypopigmentation seen in ocular albinism. (D) Complete absence of the iris. (E) Optical coherence tomogram showing a normal foveal pit (white arrow). (F) Absence of foveal pit.



**Fig. 2.** (A) Schematic representation of FRMD7 protein. The FRMD7 protein contains an N-terminal FERM domain (F1, F2, and F3 lobes) and a FERM-adjacent (FA) domain. (B) Localization of *FRMD7* mutations. F3 lobe is the most mutation-rich domain. (C) Spectrum of *FRMD7* mutations. The missense mutation accounts for a half of all *FRMD7* mutations.

neural retina, optic stalk, otic vesicle, vestibulocochlear nerve, vestibular nucleus, and cerebellum [7-9].

The *FRMD7* protein is highly co-localized with the actin of primary neurites in differentiating Neuro2A cells, which promotes elongation of axons and dendrites [10]. Knockdown of *FRMD7* protein causes a reduction in average neurite length [11]. Several studies have proposed a mechanism of *FRMD7* regulation for neuronal cytoskeletal dynamics. The *FRMD7* protein shares close amino acid sequence homology with two other FERM domain containing proteins: FARP1 and FARP2 [6]. They are involved in neurite outgrowth and branching through activating Rho GTPase signaling. Rho GTPases are key regulators of the actin cytoskeleton in eukaryotic cells and mediate morphological changes during neuronal development and plasticity. It was found that wild-type human *FRMD7* activated Rac1 signaling by interacting with RhoGDI $\alpha$ , the main regulator of Rho GTPase, while mutant *FRMD7* failed to interact with RhoGDI $\alpha$  and to activate Rac1 signaling [12]. Recent studies also demonstrated that *FRMD7* regulates the expression of Rac1 in stable SHSY-5Y cells [13], and mutant *FRMD7* significantly influences the expression of Rac1, Cdc42, and RhoA during the induction period of human fibroblasts-reprogrammed neurons [14]. Thus, it is possible that *FRMD7* is involved in the regulation of neuronal cytoskeletal dynamics through Rho GTPase signaling at the growth cone. Alternatively, the interaction between *FRMD7* and calcium/calmodulin-dependent serine protein kinase (CASK) may promote the membrane extension during neurite outgrowth since the function of CASK is to link the plasma membrane to the actin cytoskeleton [15]. Furthermore, *FRMD7* is specially localized in starburst cells of the mouse retina and the directional selective (DS) inhibitory input from starburst cells to DS ganglion cells is lost in *FRMD7* mutant mice [16]. A recent study found *FRMD7* to directly interact with the loop between transmembrane domains 3 and 4 of GABRA2, a type A gamma-aminobutyric acid (GABA) receptor subunit, and colocalization of *FRMD7* and GABRA2 was found in the mouse retina [17]. Thus, *FRMD7* mutations perturb the interaction between *FRMD7* and GABRA2, which may impair GABA inhibitory inputs from starburst cells to DS ganglion cells, eventually leading to the loss of optokinetic reflex that can be seen in INS patients. All of these findings support that nystagmus by *FRMD7* mutations may result from defective axogenesis, dendritogenesis, and neuronal guidance in the areas of the brain which control eye movements.

### *FRMD7* Mutations

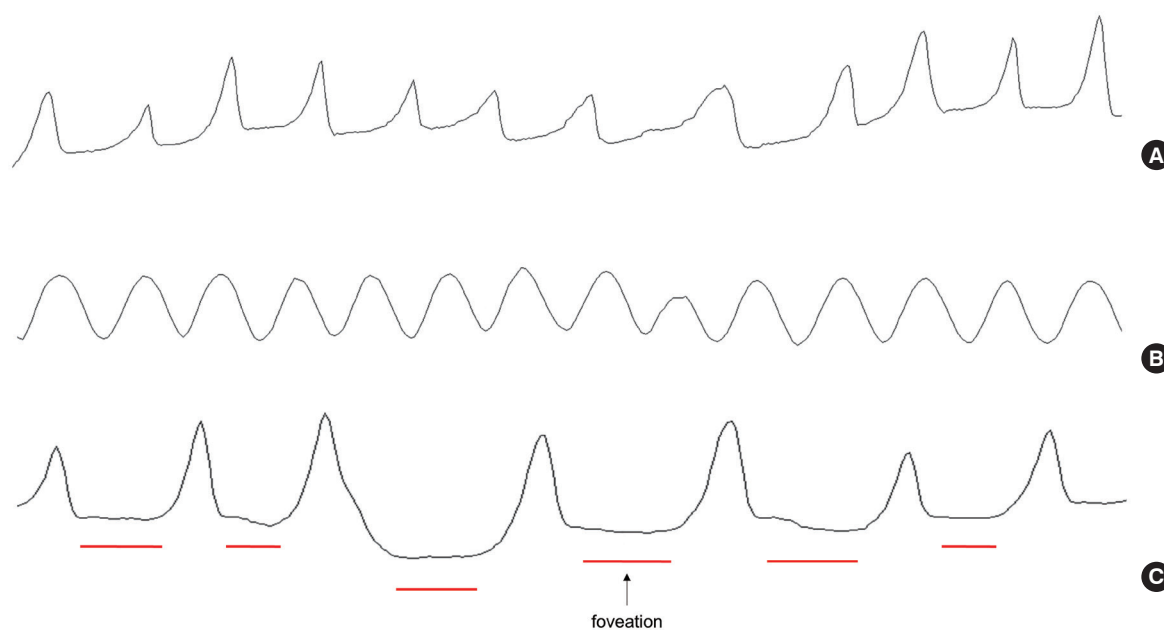
To date, over 90 different mutations within *FRMD7* have been reported (Supplementary Table S1) [18,19]. Approximately 84% of mutations concentrates heavily within the N-terminal FERM and FA domain without any consistent hot spots (Fig. 2B). Most have been identified in single case with idiopathic INS, but some mutations including c.41delAGA (p.K14del), c.70G>A (p.G24R), c.875T>C (p.L292P), c.910C>T (p.R303X), and c.1003C>T (p.R335X) have been detected in different racial groups. Especially, c.875T>C (p.L292P) accounted for more than 50% of Korean patients carrying *FRMD7* mutations [19]. All patients with c.875T>C came from the same restricted region (Gyeongsangnam-do) of Korea, and shared two single-nucleotide polymorphisms (rs6637934, rs5977623) of exon 12 within *FRMD7*, suggesting that c.875T>C might have arisen from the founder effect in the Korean population with idiopathic INS.

A half of the mutations are missense which may destabilize the overall structure of *FRMD7* protein, while the other half are predicted to cause gross defects at the protein level due to nonsense mutations, frameshift by small deletion or insertion, aberrant splicing, and large intragenic deletion (Fig. 2C). Among the 12 exons, exon 9 represents the most common mutation-rich exon (23%), followed by exon 12 (12%) and exon 8 (11%).

Incomplete penetrance was observed in female carriers, ranged from 30 to 100% [5,20-23]. This phenomenon has been explained by skewed X-inactivation and interactions with disease-modifying genes or environmental factors. Although skewed X-inactivation has consistently been suggested as a mechanism that may influence the penetrance of X-linked disorders in females, some studies have revealed that there was no clear causal link between X-inactivation pattern and phenotype in INS families with *FRMD7* mutation [20,21]. Furthermore, affected females showed random X-inactivation, reflecting a tissue mosaicism [22]. Different methylation patterns for the X chromosome were also found between female carriers, implying that a molecular basis for variable methylation might not be involved in the dissimilar penetrance [21]. Further investigations of X-inactivation status of *FRMD7* may help understand the incomplete pattern of inheritance.

### Clinical Features of *FRMD7*-associated INS

The clinical features of *FRMD7*-associated INS are not much different from those of non-*FRMD7* INS. The nystagmus is present at birth or during infancy, and usually manifests as



**Fig. 3.** Nystagmus waveforms recorded by video-nystagmography. (A) Jerk nystagmus with slow phases that drift away from the fixation position with increasing velocity waveforms. (B) Pendular nystagmus showing sinusoidal oscillations. (C) Foveation period (red bars) which the eye velocity is at or near zero.

horizontal conjugate oscillations, while vertical nystagmus is not typical for *FRMD7*-associated INS [1,4]. The direction of the nystagmus changes with eccentric gaze (right-beating on right gaze and left-beating on left gaze) or alternates periodically with time (periodic alternating nystagmus) [8,19]. The nystagmus is often accentuated by anxiety, attention, and attempts to fixate an object, while attenuated with eyelid closure or on convergence. The nystagmus waveform can be pendular or jerk with increasing exponential slow phases (Fig. 3A and 3B), but a pendular waveform is more common in adults with *FRMD7* mutations.

The nystagmus decreases when the eyes are moved into a particular position within the orbit, called the null point or zone [1]. Some individuals with INS tend to turn their head close to the null point or zone, resulting in abnormal head posture (AHP). The presence of AHP often leads parents to bring a child for medical evaluation and treatment.

Despite continuous eye oscillations, individuals with *FRMD7*-associated INS show relatively good visual acuity and no oscillopsia due to the presence of foveation period which the eye velocity is at or near zero (Fig. 3C). During this brief period, the image of the target is relatively stationary in the foveal area, leading to good visual acuity without oscillopsia [1]. Furthermore, *FRMD7*-associated INS is not accompanied with afferent visual system disorders causing reduced visual acuity

such as foveal anomaly or retina dystrophy. Although previous studies have revealed morphological changes of retina and optic nerve such as decreased peripapillary retinal nerve fiber layer and shallow foveal pit and optic nerve head, these changes may be subclinical [9,18,19]. However, some individuals may complain of oscillopsia when the nystagmus is pronounced or the individual is tired.

## CONCLUSION

*FRMD7* is a major disease-causing gene of idiopathic INS. Although the molecular pathogenesis of *FRMD7* is still unclear, it is thought that *FRMD7* may participate in neuronal development in the areas of the brain controlling ocular motor and gaze stability. Further functional investigation and mutant analysis are needed to reveal the pathogenic mechanisms of *FRMD7*-associated INS.

## ACKNOWLEDGMENTS

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## CONFLICTS OF INTEREST

The authors have no financial conflicts of interest.

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