RESEARCH ARTICLE

Two Unrecoreded Species Belonging to Penicillium Section Exilicaulis in South Korea

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ABSTRACT

Penicillium in section Exilicaulis is characterized by non-vesiculate monoverticillate and biverticillate stipes. Species in sect. Exilicaulis are commonly found in soil and plants in terrestrial environments; however, only a few species have been reported in Korea. To investigate the diversity of Penicillium sect. Exilicaulis, Penicillium species were isolated from terrestrial and marine environments. Based on sequence analyses of β -tubulin, calmodulin, and the second largest subunit of RNA polymerase II loci, 19 strains of Penicillium in sect. Exilicaulis were identified as P. citreonigrum, P. citreosulfuratum, P. corylophilum, P. menonorum, P. rubefaciens, P. velutinum, Penicillium sp. 1, and Penicillium sp. 2. Two of them, P. citreonigrum and P. citreosulfuratum, were confirmed to be new to Korea. Molecular phylogenies and detailed descriptions of the two unrecorded species are provided.

Keywords: BenA, CaM, Newly recorded species, RPB2

INTRODUCTION

The genus Penicillium is one of the most common fungi found in various environments [1-3]. They play important ecological roles as decomposers [4]. Some Penicillium species are known as producer of solubilized phosphorus, siderophore, and phytohormones, which are important for plant health [5]. To date, approximately 460 species of Penicillium in 26 sections have been reported worldwide [6-10]. Penicillium in sect. Exilicaulis is commonly isolated from air, soil, plants, and insects [8,9]. Some species can produce metabolites that cause allergies in humans [11,12].

Section Exilicaulis is characterized by non-vesiculate monoverticillate stipes [1]. Recently, based on multigene phylogenetic analysis, sect. Exilicaulis was redefined to include species with biverticillate conidiophores, and was separated into six clades [9,13]. B-tubulin (BenA), calmodulin (CaM), and the second largest subunit of RNA polymerase II (RPB2) were used for accurate identification of the species in this section [9]. So far, 55 species have been reported in this section worldwide [8,9]. In Korea, a total of nine species have been reported: P. corylophilum, P. decumbens, P. erubescens, P. melinii, P. menonorum, P. restrictum, P. rubefaciens, P. rubidurum, and P. velutinum. They were isolated from air, mushroom



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This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/bync/4.0/) which permits unrestricted non-commercial media materials, and soil. Most species were identified by morphological characteristics and/or internal transcribed spacer (ITS) sequence [14-17]. However, *Penicillium* are difficult to identify at the species level using morphology and/or ITS sequence because of their morphological plasticity and similarity and low resolution of ITS region [7]. Therefore, the accurate diversity at species level of this section in Korea is unclear.

As part of the projects organized by Basic Science Research Program through the National Research Foundation of Korea (NRF) and the Marine Biotechnology Program of the Korea Institute of Marine Science and Technology Promotion (KIMST) to excavate Korean *Penicillium* and marine fungi, we have been studying a number of *Penicillium* species from various environments in Korea. To investigate the accurate diversity of *Penicillium* sect. *Exilicaulis* in various environments, we re-identified previously isolated species in sect. *Exilicaulis* using sequence analysis of *BenA*, *CaM*, and *RPB2*. In this study, a total of eight species were detected in this section and two species–*P. citreonigrum*, and *P. citreosulfuratum* – were confirmed as unrecorded species in Korea. The detailed descriptions for the two unrecorded species have been provided in this study.

MATERIALS AND METHODS

Materials

A total of 19 *Penicillium* strains were identified in this study. Twelve strains were isolated from egg masses of *Arctoscopus japonicas* (2 strains), mudflats (4), sea sand (5), and seaweed (1) using previously described methods [18-20]. Two strains were isolated from rhizosphere soil. Five grams of soil was diluted tenfold with sterile water. Next, 100 μ L of each dilution was plated on dichloran rose bengal chloramphenicol agar (DRBC, Difco, Becton Dickinson, MD, USA). Rotten pine bark (1 strain), rotten pine sapwood (3) and sponge (1) were cut to approximately 5 mm in length and were placed on DRBC agar. All plates were incubated at 25°C for 7 days and were transferred to a PDA plate. Each strain is stored in 20% glycerol at -80°C at the Seoul National University Fungus Collection (SFC) (Table 1).

DNA extraction, amplification, and sequencing

Genomic DNA was extracted from strain grown on malt extract agar (MEA; Oxoid, Hampshire, UK) using a cetyltrimethylammonium bromide extraction protocol [21]. *BenA*, *CaM* and *RPB2* were amplified using previously described methods [22]. The PCR products were purified using the Expin[™] PCR Purification Kit (GeneAll Biotechnology, Seoul, Korea), according to the manufacturer's instructions. DNA sequencing was performed on an ABI Prism 3700 genetic analyzer (Life Technologies, Gaithersburg, MD, USA) at Macrogen (Seoul, Korea) using the PCR primers.

| | 5 | | | | | |
|------------------------------|------------------|---|-----------------------------|----------|----------|----------|
| Species | Strain No. | Site | Habitate | BenA | CaM | RPB2 |
| P. citreonigrum ^a | SFC20200821-M01 | Ocheon-dong, Yeosu-si, Jeollanam-do | Sponge | MT945077 | MT945096 | MT945115 |
| | SFCP0024 | Daebang-ri, Subuk-myeon, Damyang-gun, Jeollanam-do | Rotten Pine bark | MT945078 | MT945097 | MT945116 |
| | SFCP0026 | Sillim-dong, Gwanak-gu, Seoul | Rotten Pine sapwood | MT945079 | MT945098 | MT945117 |
| | SFCP0027 | Sodo-dong, Taebaek-si, Gangwon-do | Rotten Pine sapwood | MT945080 | MT945099 | MT945118 |
| | SFCP0028 | Sodo-dong, Taebaek-si, Gangwon-do | Rotten Pine sapwood | MT945081 | MT945100 | MT945119 |
| P. citreosulfuratum | SFC20170821-M07 | Oryu-ri, Hyeongyeong-myeon, Muan-gun, Jeollanam-do | Sea sand | MT945082 | MT945101 | MT945120 |
| | SFC20200821-M02 | Oryu-ri, Hyeongyeong-myeon, Muan-gun, Jeollanam-do | Sea sand | MT945083 | MT945102 | MT945121 |
| | SFC20200821-M03 | Oryu-ri, Hyeongyeong-myeon, Muan-gun, Jeollanam-do | Sea sand | MT945084 | MT945103 | MT945122 |
| P. corylophilum | SFC20170718-M14 | Chodo-ri, Hyeonnae-myeon, Goseong-gun, Gangwon-do | Sea sand | MT945085 | MT945104 | MT945123 |
| | SFC20141123-M44 | Gwakji-ri, Aewol-eup, Jeju-si, Jeju-do | Seaweed | MT945086 | MT945105 | MT945124 |
| P. menonorum | SFC20200821-M04 | Pyeongsan-ri, Hyeongyeong-myeon, Muan-gun, Jeollanam-do | Mud flat | MT945087 | MT945106 | MT945125 |
| | SFC20200821-M05 | Pyeongsan-ri, Hyeongyeong-myeon, Muan-gun, Jeollanam-do | Mud flat | MT945088 | MT945107 | MT945126 |
| P. rubefaciens | SFC20140101-M799 | Wando-gun, Jeollanam-do | Mud flat | MT945089 | MT945108 | MT945127 |
| P. velutinum | SFC20150915-M11 | Dongmak-ri, Hwado-myeon, Ganghwa-gun, Incheon | Sea sand | MT945090 | MT945109 | MT945128 |
| | SFC20200506-M51 | Dongdeok-ri, Yeongok-myeon, Gangneung-si, Gangwon-do | Sailfin sandfish egg masses | MT945091 | MT945110 | MT945129 |
| | SFC101366 | Jeong-am-ri, Ganghyeon-myeon, Yangyang-gun, Gangwon-do | Sailfin sandfish egg masses | MT945092 | MT945111 | MT945130 |
| Penicillium sp. 1 | SFC20200821-M06 | Jangheung-ri, Gilsang-myeon, Ganghwa-gun, Incheon | Mud flat | MT945093 | MT945112 | MT945131 |
| Penicillium sp. 2 | SFCP0509 | Ui-dong, Gangbuk-gu, Seoul | Rhizosphere soil | MT945094 | MT945113 | MT945132 |
| | SFCP0523 | Ui-dong, Gangbuk-gu, Seoul | Rhizosphere soil | MT945095 | MT945114 | MT945133 |

Table 1. Summary and GenBank accession numbers for Penicillium in section Exilicaulis.

^a The unrecorded Penicillium species in Korea are represented in bold.

Phylogenetic analysis

Each sequence was assembled and proofread using MEGA5 [23]. The resulting consensus sequences were deposited in GenBank (accession Nos. are shown in Table 1). Molecular identification was performed in two steps. First, we identified strains belonging to section *Exilicaulis* by comparison to the *BenA* sequences of type strains. Next, each strain was identified to the species level by analyzing the combined dataset of the three loci (*BenA*, *CaM*, and *RPB2*). *Penicillium trzebinskii* CBS 351.51 was used as the outgroup [9]. The sequence similarities were calculated from the three loci for each species using MEGA5 [23]. Multiple alignments were performed using MAFFT v7 [24]. Maximum likelihood phylogenetic analyses were performed with RAxML [25] implemented on CIPRES web portal [26], using the GTR+G model with 1,000 bootstrap replicates.

Morphological analysis

The morphological observation of the two unrecorded species was performed using previously described methods [7] on three different media: Czapek yeast autolysate agar (CYA; Difco, Sparks, MD, USA), malt extract agar (MEA; Oxoid, Hampshire, UK), and yeast extract sucrose agar (YES; Difco, Sparks, MD, USA). The Methuen Handbook of Color was used for the color names and alphanumeric codes for macromorphological characteristics [27]. The microscopic features were observed under a light microscope (Eclipse 80i, Nikon, Tokyo, Japan) using colonies grown on MEA at 25°C for seven days.

RESULTS

A total of 19 *Penicillium* strains were isolated from terrestrial (6 strains) and marine (13 strains) environments. They were grouped into eight groups in section *Exilicaulis* based on *BenA* sequences. For accurate identification of each strain, all strains were used for the combined dataset of *BenA*, *CaM*, and *RPB2*. These were confirmed as 8 species: *P. citreonigrum*, *P. citreosulfuratum*, *P. corylophilum*, *P. menonorum*, *P. rubefaciens*, *P. velutinum*, *Penicillium* sp. 1, and *Penicillium* sp. 2 (Fig. 1).

Five strains formed a monophyletic group with *P. citreonigrum* NRRL 761 (type strain), CBS 321.59, and NRRL 1187 (sequence similarity for *BenA*=98.8-100%, *CaM*=98.0-100%, and *RPB*2=98.5-99.2%; bootstrap support=89%). Three strains grouped with *P. citreosulfuratum* DTO 290-I4 (type strain), CV 2015, and NRRL 31271 (sequence similarity for *BenA*=100% and *CaM*=99.8-100%, *RPB*2=99.9; bootstrap support=100%). Two strains formed a monophyletic group with the type strain (CBS 312.48) of *P. corylophilum*, CBS 231.38 and CBS127808 (sequence similarity for *BenA*=98.8-99.0%, *CaM*=99.8%, *RPB*2=99.7; bootstrap support=100%). SFC20200821-M04 and SFC20200821-M05 grouped with *P. menonorum* NRRL 50410 (type strain) (sequence similarity for *BenA*=100% and *CaM*=99.8-100%, *RPB*2=99.7; bootstrap support=100%). SFC20140101-M799 formed a monophyletic group with *P. rubefaciens* CBS 1450.83 (type strain) and CV0597 (sequence similarity for *BenA*=99.2-99.3% and *CaM*=99.4-99.5%, *RPB*2=100; bootstrap support=99%). Three strains grouped with *P. velutinum* NRRL 2069 (type strain) (sequence similarity for *BenA*=100%, *RPB*2=99.9; bootstrap support=100%). The remaining two groups formed distinct group with previously reported species. These groups were designated as *Penicillium* sp. 1 and *Penicillium* sp. 2.

Taxonomy

Penicillium citreonigrum Dierckx (1901)

Description: Colony diam, 7 d, in mm: CYA 25-28; CYA 30°C 24-27; CYA 37°C no growth; MEA 24-26; YES 30-31 (Fig. 2).

Colony characters: CYA, 25°C, 7 d: Colonies low to moderately deep, radially sulcate; margins low, wide, entire; mycelia white to grayish yellow (2B4); texture velvety; sporulation sparse; conidia greyish green (30B4); exudates clear at center; soluble pigments yellow; reverse color deep yellow (4A8). MEA, 25°C, 7 d: Colonies low to moderately deep, radially sulcate; margins low, narrow, entire; mycelia white; texture velvety, floccose at center; sporulation moderate; conidia dull green (25D3); exudates absent; soluble pigments absent; reverse color brownish yellow (5C8). YES, 25°C, 7 d: Colonies low to moderately deep, radially sulcate; margins low, narrow, entire; mycelia white; texture velvety; sporulation sparse to moderate; conidia greyish green (30B3) at margin, light green (30A5) elsewhere; exudates absent; soluble pigments absent; reverse color deep yellow (4A8), greyish yellow (4B5) at center.

Conidiophores monoverticillate, occasionally biverticillate; stipes smooth walls; phialides ampulliform, $6.0-9.0 \times 2.0-3.0 \mu m$; conidia smooth walls, globose to subglobose, $1.8-2.6 \mu m$ diam; sclerotia absent; asci and ascospores not observed.

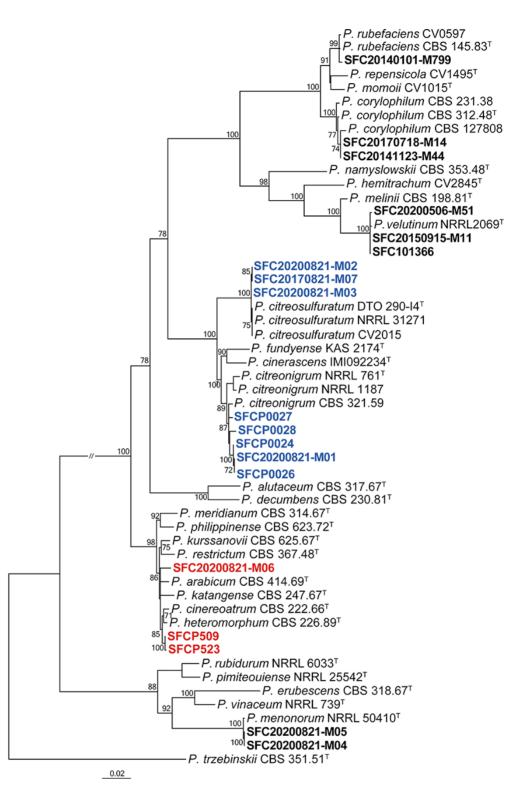


Fig. 1. Maximum likelihood phylogenetic tree of the combined data set of *BenA*, *CaM*, and *RPB2* gene sequences used to identify strains to the species level in *Penicillium* section *Exilicaulis*. Bootstrap scores of >70 are presented at the nodes. The scale bar indicates the number of nucleotide substitutions per site. "T" indicates the ex-type strains. Strains reported in the current study are represented in bold. The species labeled in blue represent previously unrecorded species in South Korea. The names in red are potential new species.

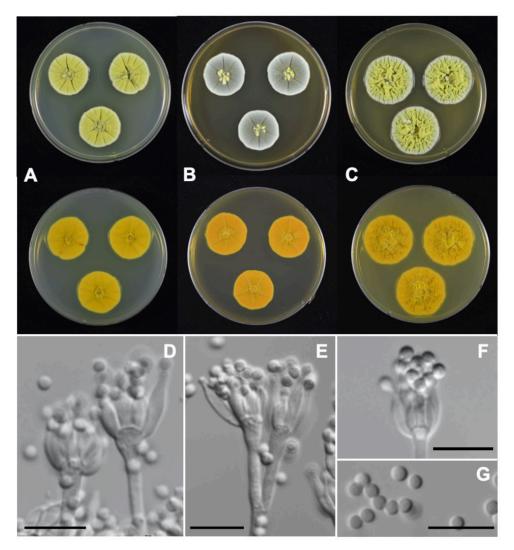


Fig. 2. *Penicillium citreonigrum* SFCP0024 in 7-day-old cultures at 25°C. (A-C) Colonies grown on Czapek yeast autolysate agar (CYA), malt extract agar (MEA), and yeast extract sucrose agar (YES) from left to right (top=obverse, bottom=reverse). (D-F) Conidiophores; (G) Conidia (scale bar: D-G=10 µm).

Strain examined: SFCP0024

Note: — *Penicillium citreonigrum* is morphologically similar to *P. citreosulfuratum*, *P. cinerascens*, and *P. fundyense*. *Penicillium citreonigrum* and *P. cinerascens* can be distinguished from *P. citreosulfuratum* by no growth on CYA at 37°C [8,9]. *Penicillium citreonigrum*, *P. cinerascens*, and *P. fundyense* are difficult to identify based on morphological characteristics. They are accurately identified based on ITS or *BenA* sequences.

Penicillium citreosulfuratum Biourge (1923)

Description: Colony diam, 7 d, in mm: CYA 27-28; CYA 30°C 28-30; CYA 37°C 7-10; MEA 23-25; YES 28-30 (Fig. 3).

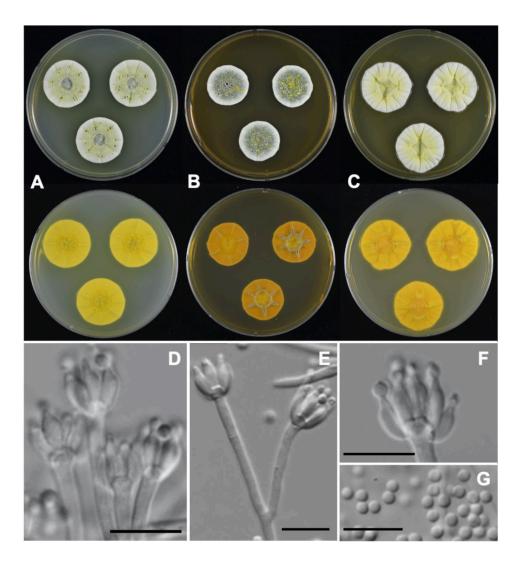


Fig. 3. *Penicillium citreosulfuratum* SFC20170821-M07 in 7-day-old cultures at 25°C. (A-C) Colonies grown on Czapek yeast autolysate agar (CYA), malt extract agar (MEA), and yeast extract sucrose agar (YES) from left to right (top=obverse, bottom=reverse). (D-F) Conidiophores; (G) Conidia (scale bar: D-G=10 μ m).

Colony characters: CYA, 25°C, 7 d: Colonies low to moderately deep, radially sulcate; margins moderately deep, entire; mycelia white at margins, and light green (30A4) elsewhere; texture velvety, floccose at center; sporulation sparse; conidia greenish grey (28D3); exudates pale yellow (1A3), scattered overall; soluble pigments yellow; reverse color grayish yellow (3B5). MEA, 25°C, 7 d: Colonies low to moderately deep, radially sulcate; margins white, entire; mycelia white; texture velvety, floccose at center; sporulation moderate; conidia greyish green (26C3); exudates abundant, pale yellow (1A3); soluble pigments absent; reverse color brownish yellow (5C7). YES, 25°C, 7 d: Colonies low to deep, sulcate; margins low, narrow, entire; mycelia white; texture velvety, floccose at center; sporulation sparse; conidia greyish green (28B4); exudates absent; soluble pigments absent; reverse color reddish yellow (4A7), light orange (5A5) at center.

Conidiophores monoverticillate, occasionally biverticillate, stipes smooth walls; phialides ampulliform, $6.0-8.0 \times 2.0-3.0 \mu m$; conidia smooth walls, globose to subglobose, $2.3-3.2 \mu m$ diam; sclerotia absent; asci and ascospores not observed.

Strain examined: SFC20170821-M07

Note: *Penicillium citreosulfuratum* is morphologically similar to *P. cinerascens*, *P. citreonigrum*, and *P. fundyense*. *P. citreosulfuratum* can be distinguished from them by growth on CYA at 37°C [8,9].

DISCUSSION

We confirmed that eight species in sect. *Exilicaulis* exist in Korea. Four species of them were previously reported in Korea: *P. corylophilum, P. menonorum, P. rubefaciens*, and *P. velutinum. Penicillium citreonigrum* and *P. citreosulfuratum* have been recorded for the first time in Korea. Although other two species were clearly separated from the previously described species based on phylogenies of *BenA*, *CaM*, and *RPB2*, they were designated as *Penicillium* sp. due to the minor morphological differences. Additional examination for a more detailed morphological comparison with phylogenetically similar species will be required to identify these species.

Sect. *Exilicaulis* is divided into six clades based on phylogenies of ITS, *BenA*, *CaM*, and *RPB2*. *Penicillium citreonigrum* clade consists of four species: *P. cinerascens*, *P. citreonigrum*, *P. citreosulfuratum*, and *P. fundyense*. Although *P. citreosulfuratum* can be distinguished from the rest by its ability to grow at 37°C [8,9], these species are difficult to identify based on morphological characteristics due to only a few significant or consistent morphological differences [9]. Phylogeny based on ITS or *BenA* sequences have been proposed for species identification in *P. citreonigrum* clade [9]. The morphological and phylogenetic characteristics of two unrecorded species were consistent with those of the respective type species. *Penicillium citreonigrum* and *P. citreosulfuratum* were isolated from plant and soil in terrestrial environments [9] and produced citreoviridin [28] correlated with yellow rice disease [29,30]. These two species are reported for the first time in Korea as well as from marine environment.

Nine species in section *Exilicaulis* have been previously reported in Korea [31]. Five species of them were not found in this study: *P. decumbens*, *P. erubescens*, *P. melinii*, *P. restrictum* and *P. rubidurum*. Two species (*P. erubescens* and *P. melinii*) were recently reported as unrecorded species in Korea based on morphological characteristics and *CaM* sequence [32]. *Penicillium rubidurum* KNU14-12 was reported from soil based on the ITS sequence (accession no. KP055596) [14]. However, *P. rubidurum* KNU14-12 formed a phylogenetically distinct group with the type strain (CBS 609.73) of *P. rubidurum* and showed the highest similarity of ITS sequence with *P. parvum* CBS 570.73 (sequence similarity for ITS=99.8%). Based on these results, *P. rubidurum* KNU14-12 might be a new species rather than *P. rubidurum*. Penicillium decumbens and *P. restrictum* were identified by morphological characteristics [17]. There are no stored strains of *P. decumbens* and *P. restrictum*, so the identity cannot be verified.

In conclusion, we found eight species including two new species candidates and two unrecorded species in sect. *Exilicaulis* in this study. Three of nine species previously reported in Korea were confirmed by sequence analysis. As a result, there are 11 *Penicillium* species in section *Exilicaulis* in Korea.

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REFERENCES

- 1. Pitt JI. The genus *Penicillium* and its teleomorphic states *Eupenicillium* and *Talaromyces*. London: Academic Press; 1979.
- Samson RA, Houbraken J, Thrane U, Frisvad JC, Andersen B. Food and indoor fungi. Netherlands: CBS-Fungal Biodiversity Centre Utrecht; 2010.
- Park MS, Fong JJ, Oh SY, Kwon KK, Sohn JH, Lim YW. Marine-derived *Penicillium* in Korea: diversity, enzyme activity, and antifungal properties. Antonie van Leeuwenhoek 2014;106:331-45.
- Frisvad JC, Samson RA. Polyphasic taxonomy of *Penicillium* subgenus *Penicillium*. A guide to identification of food and air-borne terverticillate penicillia and their mycotoxins. Stud Mycol 2004;49:1-174.
- Altaf MM, Imran M, Abulreesh HH, Khan MS, Ahmad I. Diversity and applications of *Penicillium* spp. in plant-growth promotion. New and future developments in microbial biotechnology and bioengineering: Penicillum system properties and applications. In: Gupta VK, Rodriguez-Couto S editors. Amsterdam: Elsevier; 2017. p. 261-76.
- Visagie CM, Hirooka Y, Tanney JB, Whitfield E, Mwange K, Meijer M, Amend AS, Seifert KA, Samson RA. *Aspergillus, Penicillium* and *Talaromyces* isolated from house dust samples collected around the world. Stud Mycol 2014;78:63-139.
- Visagie CM, Houbraken J, Frisvad JC, Hong SB, Klaassen CHW, Perrone G, Seifert KA, Varga J, Yaguchi T, Samson RA. Identification and nomenclature of the genus *Penicillium*. Stud Mycol 2014;78:343-71.
- Visagie CM, Renaud JB, Burgess KM, Malloch DW, Clark D, Ketch L, Urb M, Louis-Seize G, Assabgui R, Sumarah MW, et al. Fifteen new species of *Penicillium*. Persoonia 2016;36:247.
- Visagie CM, Seifert KA, Houbraken J, Samson RA, Jacobs K. A phylogenetic revision of *Penicillium* sect. *Exilicaulis*, including nine new species from fynbos in South Africa. IMA fungus 2016;7:75-117.
- Houbraken J, Wang L, Lee HB, Frisvad JC. New sections in *Penicillium* containing novel species producing patulin, pyripyropens or other bioactive compounds. Persoonia 2016;36:299.
- Unoura K, Miyazaki Y, Sumi Y, Tamaoka M, Sugita T, Inase N. Identication of fungal DNA in BALF from patients with homerelated hypersensitivity pneumonitis. Respir Med 2011;105:1696-703.

- McMullin DR, Nsiama TK, Miller JD. Secondary metabolites from *Penicillium corylophilum* isolated from damp buildings. Mycologia 2014;106:621-8.
- Houbraken J, Samson RA. Phylogeny of *Penicillium* and the segregation of Trichocomaceae into three families. Stud Mycol 2011;70:1-51.
- Adhikari M, Kim S, Kim HS, Lee HB, Lee YS. Sixteen new records of Ascomycetes from crop field soil in Korea. Kor J Mycol 2016;44:271-88.
- Babu AG, Kim SW, Yadav DR, Hyum U, Adhikari M, Lee YS. *Penicillium menonorum*: A novel fungus to promote growth and nutrient management in cucumber plants. Mycobiology 2015;43:49-56.
- Hwang HJ, Mun HY, Hwang BS, Nam YH, Chung EJ. Optimal culture conditions for *Penicillium rubefaciens* NNIBRFG5039 possessing antimicrobial activity. Kor J Mycol 2020;48:15-27.
- Lee S, Hong SB, Kim CY. Contribution to the checklist of soil-inhabiting fungi in Korea. Mycobiology 2003;31:9-18.
- Park MS, Eom JE, Fong JJ, Lim YW. New record and enzyme activity of four species in Penicillium section Citrina from marine environments in Korea. J Microbiol 2015;53:219-25.
- 19. Park MS, Lee S, Lim YW. A New record of four *Penicillium* species isolated from *Agarum clathratum* in Korea. J Microbiol 2017;55:237-46.
- Park MS, Oh SY, Lee S, Eimes JA, Lim YW. Fungal diversity and enzyme activity associated with sailfin sandfish egg masses in Korea. Fungal Ecol 2018;34:1-9.
- Rogers SO, Bendich AJ. Extraction of total cellular DNA from plants, algae and fungi. Plant molecular biology manual. In: Gelvin S and Schilperoort R editors. Dordrecht: Kluwer Academic; 1994.
- Park MS, Fong JJ, Oh SY, Houbraken J, Sohn JH, Hong SB, Lim YW. *Penicillium jejuense* sp. nov., isolated from the marine environments of Jeju Island, Korea. Mycologia 2015;107:209-16.
- Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S. MEGA5: Molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. Mol Biol Evol 2011;28:2731-39.
- 24. Katoh K, Standley DM. MAFFT multiple sequence alignment software version 7: Improvements in performance and usability. Mol Biol Evol 2013;30:772-80.
- 25. Stamatakis A. RAxML-VI-HPC: Maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. Bioinformatics 2006;22:2688-90.
- Miller MA, Pfeiffer W, Schwartz T. Creating the CIPRES science gateway for inference of large phylogenetic trees. SC10 workshop on gateway computing environments (GCE10); 2010 Nov 13-19; New Orleans (LA): IEEE Computer Society; 2010. p. 1-8.
- 27. Kornerup A, Wanscher JH. Methuen handbook of colour. 3rd ed. London: Methuen Publishing; 1978.
- Peterson SW, Jurjević Ž, Frisvad JC. Expanding the species and chemical diversity of Penicillium section Cinnamopurpurea. PloS One 10 2015:e0121987.
- 29. Miyake I. Studies on toxin production by a saprophyte growing on stored rice. Report of the Rice Utilization Laboratories. Hôkoku, Japan:1940;1: 1-30.
- Udagawa S, Tatsuno T. Safety of rice grains and mycotoxins- a historical review of yellow rice mycotoxicoses. Yakushigaku Zasshi 2004;39:321-42.

- 31. Lee YS, Jung HY, Lee HB, Kim SH, Shin KS, Eom AH, Kim C, Lee SY, Koo YB, Moon KH, et al. National list of species of Korea. Ascomycota, Glomeromycota, Zygomycota, Myxomycota, Oomycota. Incheon: National Institute of Biological Resources; 2015.
- Ahn GR. A report of eighteen unrecorded fungal species in Korea. Kor J Mycol 2017;45:292-303.