

Effects of the mixed formulation of sorafenib and blue honeysuckle on the pharmacokinetics profiles of sorafenib

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Abstract

Objective : This study investigated the effects of concentrated and lyophilized blue honeysuckle powders (BH) on the pharmacokinetics (PK) of sorafenib were observed.

Method : The blood was collected at 0.5 hr before single oral treatment of sorafenib (40 mg/kg) or sorafenib with BH (400, 200 and 100 mg/kg) mixed formulas administration, and 0.5, 1, 2, 3, 4, 6, 8 and 24 hrs after the end of single or mixed formula administration. Plasma concentrations of sorafenib were analyzed using LC-MS/MS methods. Tmax, Cmax, AUC, $t_{1/2}$ and MRT_{inf} of sorafenib were analysis as compared with sorafenib single treatment.

Results : Single oral administration of mixed formulas induced significant increases of plasma sorafenib concentrations from 0.5 hr after end of administration throughout all blood collected time points, as compared with sorafenib single formula treated rats, and significant decreases of sorafenib Tmax with increases of Cmax, AUC_{0-t} and AUC_{0-inf} were detected in sorafenib and BH 400 mg/kg mixed formulation treated rats as compared with sorafenib single formula treated rats, respectively. In addition, sorafenib and BH 200 or 100 mg/kg mixed formula treated rats also showed significant increases of sorafenib Cmax, AUC_{0-t} and AUC_{0-inf} , respectively.

Conclusions : According to these results, mixed formulation of BH with sorafenib increased the bio-availability of sorafenib through the increment of the absorptions.

Key words : Blue honeysuckle, Hepatic cancer, Pharmacokinetics, Drug-drug interactions, Rat, Sorafenib.

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I. Introduction

Sorafenib (NexavarTM) is a kinase inhibitor drug approved for the treatment of primary kidney cancer (advanced renal cell carcinoma), advanced primary liver cancer (hepatocellular carcinoma)^{1,2}. It is a small inhibitor of several tyrosine protein kinases, such as vascular endothelial growth factor receptors (VEGFR), platelet-derived growth factor receptors (PDGFR) and Raf family kinases (more avidly C-Raf than B-Raf)^{3,4}; In many of the molecular pathways, the over activation of protein kinases, such as Raf kinase, platelet-derived growth factor (PDGF), vascular endothelial growth factor (VEGF) receptor 2 and 3 kinases and c Kit the receptor for Stem cell factor, cause cells to become cancerous⁵⁻⁷. However, various side effects were also reported from patients receiving sorafenib, especially include skin rash, swelling, redness, pain and peeling of the skin on the palms of hands and soles of feet, hypertension, diarrhea, fatigue, reversible posterior leukoencephalopathy syndrome and erythrocytosis^{1,8,9}, and hypersensitivity to sorafenib or any ingredient in the formulation were known¹⁰⁻¹². Various drug-drug interactions have been evaluated to improve the side effects of sorafenib or to achieve synergic effects^{1,13-20}. However, interactions of sorafenib with herbal products have not been established except for some restricted natural compounds, interact with warfarin increased risk of bleeding²¹ and St. John's wort (*Hypericum perforatum*) possible decreased plasma concentrations of sorafenib¹. In addition, we have been observed the possible interactions with Korean traditional polyherbal formulas; we observed that marked increases of oral bioavailability of sorafenib after co-administration of *Gongjindan*, a famous Korean tonic mixed herbal formulation within 5 min or

with 3.5hr-intervals co-administration,^{22,23}.

Blue honeysuckle (Berries of *Lonicera caerulea* L., Caprifoliaceae) is a traditional deciduous shrub used in folk medicine in northern Russia, China, and Japan, but its fruits are little known as edible berries in North America and Europe, and also in Korea²⁴. The berries are a rich source of ascorbic acid and polyphenols, particularly anthocyanins, flavonoids and low molecular weight phenolic acids^{24,25}. These compounds have been reported to have multiple biological activities including strong antioxidant activity and anti-inflammatory properties²⁴. Recently, oral administration of blue honeysuckle was reported to protect oxidative damage in mice against ionizing radiation²⁶, ameliorates abnormal lipid and glucose metabolism in rats²⁷, hepatoprotective effects²⁸, anti-inflammatory effects^{29,30}. Especially blue honeysuckle extracts have showed the strongest antioxidant potent among 12 types of colored berries³¹, and phenolic rich extract of BH has been revealed to have anti-inflammatory and wound-healing effects *in vitro and in vivo*²⁹ and skin protective effects to ultraviolet-induced damages^{32,33} with less toxicity³⁴.

In the present study, the effects of blue honeysuckle concentrated and lyophilized powders (BH) on the pharmacokinetics of sorafenib were observed in rats after mixed formulation as sorafenib 40 and BH 400, 200 and 100 mg/kg concentrations.

II. Material and methods

1. Animals and husbandry

Forty male SPF/VAF Outbred CrI:CD [SD] rats (6-wk old upon receipt; OrientBio, Seungnam, Korea) were used after acclimatization for 10 days. Animals were allocated five per

polycarbonate cage in a temperature (20–25°C) and humidity (40–45%) controlled room. Light : dark cycle was 12 hr : 12 hr and feed (Samyang, Korea) and water were supplied free to access. After 10 days of acclimatization, four groups of 5 rats each were selected based on the body weights. All animals were marked by picric acid, and overnight fasted (about 18 hrs; water was not restricted) before treatment, and further fasted during 3 hrs after end of treatment. Animal experiments were conducted according to the national regulations of the usage and welfare of laboratory animals, and approved by the Institutional Animal Care and Use Committee in Daegu Haany University (Gyeongsan, Gyeongbuk, Korea; Approval No. DHU2014–079).

2. Test articles

Deep purple colored solution of concentrated blue honeysuckle, about 63 brix, was supplied by H&K Bioscience Co., Ltd. (Seoul, Korea) and used as same as our previous single oral dose toxicity test³⁴⁾ in this experiment. The brief process for making BH was as follows. 200 g of 63 brix concentrated blue honeysuckle solutions supplied by H&K Bioscience was diluted into 25 brix using distilled water, and then completely lyophilized by programmable freeze dryer (Operon FDB–5503, Kimpo, Korea). Total 124.40 g (yield = 62.2%) of BH was acquired. At proximate analysis of BH by Association of Official Analytical Chemists (AOAC)³⁵⁾ methods, BH contains energy 380 kcal/100 ml, carbohydrate 93 g/100 ml, sugar 41 g/100 ml, protein 2 g/100 ml, sodium 20 mg/100 ml, but it did not contains total lipids (0 mg/100 ml), saturated lipids (0 mg/100 ml), trans-fat (0 mg/100 ml), cholesterol (0 mg/100 ml), respectively. In addition, phytochemical analysis of BH reveals that it contains $4.54 \pm 0.09\%$ of betaine by high performance liquid

chromatography (HPLC), 210.63 ± 23.65 mg gallic acid equivalents (GAE)/g of total phenols by Folin–Ciocalteu colorimetric method³⁶⁾, 159.30 ± 12.51 mg catechin equivalents (CE)/g of total flavonoids by a modified colorimetric method³⁷⁾ and 133.57 ± 4.06 mg malvidin–3–O–glucoside equivalents (M3GE)/g of total antocyanins by a modified pH differential method³⁸⁾, respectively. In addition, white powder of sorafenib (Jeil Pharm. Co., Yongin, Korea) was used as control drug. Both drugs are well dissolved (up to 40 mg/ml solutions in BH and upto 4 mg/ml solutions in sorafenib) in distilled water as vehicle, respectively.

3. Test article formulation and administration

Four groups of 5 rats each were used in this study as follows. The dosages of sorafenib 40 mg/kg were selected, considering the clinical dosage in human and body surface of rats. Single formula of sorafenib was prepared as dissolved in distilled water (4 mg/ml concentration), and each mixed formula consisted of sorafenib and BH was prepared by dissolved of approximate amounts of sorafenib (40 mg/kg) and BH (400, 200 or 100 mg/kg) in distilled water. Sorafenib single formula and all three types of mixed formula consisted of sorafenib and BH were once orally administered, in a volume of 10 ml/kg, respectively.

4. Plasma collections

All rats were anesthetized with 2 to 3% isoflurane (Hana Pharm, Co., Hwasung, Korea) in the mixture of 70% N₂O and 28.5% O₂, and blood samples (0.5 ml) were collected into 50 IU heparinized 0.7 ml Eppendorf tubes through the orbital plexus at 30 min before treatment (as a control), 30 min, 1, 2, 3, 4, 6, 8 and 24 hrs after end of single oral administration of

sorafenib single or mixed formula with three different dosages of BH, respectively. Blood samples were immediately centrifuged for 10 min at 13,000 rpm and about 0.3 ml aliquots of plasma were stored in a $-150\text{ }^{\circ}\text{C}$ deep freezer until analysis of sorafenib.

5. Sample preparation and calibrations

Primary stock solution, 1.0 mg/ml of sorafenib in 90% acetonitrile (Sigma-Aldrich, St. Louise, MO, USA) mixtures with distilled water and internal standard working solution, carbamazepine (Sigma-Aldrich, St. Louise, MO, USA) 500 ng/ml in acetonitrile were prepared. Working standard solutions were prepared by dilution with acetonitrile. All standard solutions were stored at $-20\text{ }^{\circ}\text{C}$ in the dark when not in use, and calibrated the standard samples as 100 μl of blank plasma; working standard solutions and internal standard working solution were mixed with 100 μl of acetonitrile. In addition, 100 μl of sample plasma and internal standard working solution were mixed with 200 μl of acetonitrile. The mixtures were mixed by vortex-mixing and centrifuged at 12,000 rpm for 10 min at $4\text{ }^{\circ}\text{C}$. Clear supernatants (150 μl) were directly transferred to injection vials and the aliquot (5 μl) was injected into the LC-MS/MS system^{22,23}.

6. LC-MS/MS conditions

Concentrations of sorafenib in the rat plasma samples were determined using LC-MS/MS method. Chromatographic analysis was performed using an Agilent 1100 Series HPLC (Agilent Technologies, Santa Clara, CA, USA) equipped with on-line degasser, binary pump, autosampler and column compartment. Separation of the analyte from potentially interfering material was achieved at ambient temperature using Waters

SymmetryTM C₁₈ columns (2.1 \times 50 mm, 3.5 μm) (Waters Corp., Milford, MA, USA). The mobile phase used for the chromatographic separation was composed of 25% distilled water (0.1% formic acid)/75% acetonitrile, and was delivered isocratically at a flow rate of 0.30 ml/min. The column effluent was monitored using an API 2000 triple-quadrupole mass-spectrometric detector (Applied Biosystems, Foster City, CA, USA). The instrument was equipped with an electrospray interface in positive ion mode, and controlled by the Analyst version 1.4.1, Quadratic (1/x², no Iterate) software (Applied Biosystems, Foster City, CA, USA). Samples were introduced to the interface through a Turbo IonSpray with the temperature set at $300\text{ }^{\circ}\text{C}$. A high positive voltage of 5.5 kV was applied to the ion spray. Nitrogen was used as the nebulizer gas, curtain gas, and collision gas with the settings of 20, 20, and 4, respectively. The multiple reaction monitoring (MRM) detection method was employed for the detection of sorafenib; the transitions monitored were carbamazepine (IS): m/z 237>194 (Retention time: 0.56 min), sorafenib: 465>252 (Retention time: 0.85 min). Calibration curves of sorafenib were quadratic over the ranges studied with $r^2 > 0.999$. The lower limit of quantification of the sorafenib in the rat plasma was 40 ng/ml^{22,23}.

7. Pharmacokinetic analysis

The plasma concentration data was analyzed using a noncompartmental method on commercial pharmacokinetics data analyzer programs (PK solutions2.0; Summit, Montrose, CO, USA)^{39,40}. The elimination rate constant (K_{el}) was calculated by the log-linear regression of sorafenib concentration data during the elimination phase, and the terminal half-life ($t_{1/2}$) was calculated by $0.693/K_{el}$. The peak concentration (C_{max}) and time to reach the peak concentration (T_{max}) of sorafenib in the plasma were obtained by visual

inspection of the data in the concentration–time curve. The area under the plasma concentration–time curve (AUC_{0-t}) from time zero to the time of the last measured concentration (C_{last}) was calculated using the linear trapezoidal rule⁴¹⁾. The AUC zero to infinity (AUC_{0-inf}) was obtained by adding AUC_{0-t} and the extrapolated area was determined by C_{last}/K_{el} . The mean residence time infinity (MRT_{inf}) was calculated by dividing the first moment of AUC ($AUMC_{0-inf}$) by AUC_{0-inf} .

8. Statistical analyses

All the means are presented with their standard deviation (SD) of five rats (Mean \pm SD of five rat plasma sorafenib concentrations). The pharmacokinetic parameters were compared using a non–parametric comparison test, Mann–Whitney U (MW) test, on the SPSS for Windows

(Release 14.0K, SPSS Inc., Chicago, IL, USA). A p -value < 0.05 was considered statistically significant.

III. Results

1. Changes on the plasma concentrations of sorafenib

Sorafenib was detected from 30 min to 24 hrs after end of administration in sorafenib single formula and all three different concentrations of mixed formulas with BH administered rats, respectively. Single oral administration of mixed formulas consisted of sorafenib 40 mg/kg and BH 400, 200 or 100 mg/kg induced significant ($p < 0.01$) increases of plasma sorafenib concentrations, from 30 min after end of administration throughout all

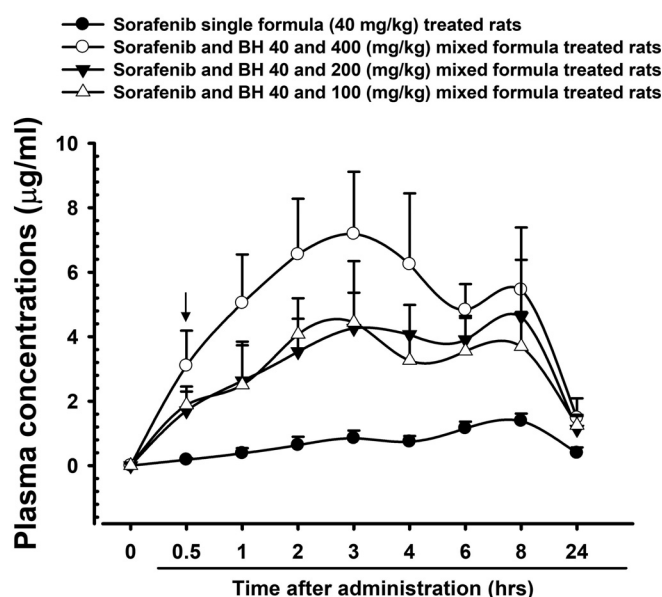


Figure 1. Plasma concentrations of sorafenib after single oral administration of sorafenib single or mixed formulas with BH 400, 200 or 100 mg/kg in male Rats. Sorafenib was detected from 30 min to 24 hrs after end of administration in sorafenib single formula and all three different concentrations of mixed formulas with BH administered rats, respectively. Single oral administration of mixed formulas consisted of sorafenib 40 mg/kg and BH 400, 200 or 100 mg/kg induced significant ($p < 0.01$) increases of plasma sorafenib concentrations, from 30 min after end of administration throughout all blood collected time points, as compared with sorafenib 40 mg/kg single formula treated rats, respectively (arrow). Values are expressed as mean \pm SD of five rats ($\mu\text{g/ml}$). BH = Blue honeysuckle (fruit parts of *Lonicera caerulea* L., Caprifoliaceae) concentrated and lyophilized powder (H&K Bioscience Co., Ltd., Seoul, Korea)

blood collected time points, as compared with sorafenib 40 mg/kg single formula treated rats, respectively (Fig 1).

2. Changes on the Tmax of sorafenib

The Tmax of sorafenib was significantly ($p < 0.01$) decreased as -60.53% points in sorafenib and BH 400 mg/kg mixed formula treated rats (3.00 ± 0.71 hr) as compared with sorafenib single formula treated rats (7.60 ± 0.89 hr). In addition, the Tmax of sorafenib were non-significantly changed as -26.32 and -50.00% points in sorafenib and BH 200 or 100 mg/kg mixed formula treated rats as compared with sorafenib single formula treated rats; they were detected as 5.60 ± 2.51 and 3.80 ± 2.39 hr in sorafenib and BH 200 or 100 mg/kg mixed formula treated rats, respectively (Table 1).

3. Changes on the Cmax of sorafenib

The Cmax of sorafenib were significantly ($p < 0.01$) increased as 444.38 , 262.25 and 250.43% points in sorafenib and BH 400, 200 or 100 mg/kg mixed formula treated rats as compared with sorafenib single formula treated rats,

respectively. They were detected as 7.56 ± 1.82 , 5.03 ± 1.28 and 4.86 ± 1.60 $\mu\text{g/ml}$ in sorafenib and BH 400, 200 or 100 mg/kg mixed formula treated rats, and as 1.39 ± 0.23 $\mu\text{g/ml}$ in sorafenib single formula treated rats, in this experiment (Table 1).

4. Changes on the AUC of sorafenib

The AUC_{0-t} of sorafenib were significantly ($p < 0.01$) increased as 373.45 , 259.20 and 218.34% points in sorafenib and all three different concentrations of BH mixed formula treated rats as compared with sorafenib single formula treated rats, respectively. They were detected as 99.05 ± 28.24 , 75.15 ± 21.43 and 66.60 ± 16.10 $\text{hr} \cdot \mu\text{g/ml}$ in sorafenib and BH 400, 200 or 100 mg/kg mixed formula treated rats, and as 20.92 ± 2.64 $\text{hr} \cdot \mu\text{g/ml}$ in sorafenib single formula treated rats, in this experiment. In addition, the $\text{AUC}_{0-\infty}$ of sorafenib were also significantly ($p < 0.01$) increased as 348.28 , 230.37 and 237.55% points in sorafenib and all three different concentrations of BH mixed formula treated rats as compared with sorafenib single formula treated rats, respectively. They were detected as 122.05 ± 37.24 , 89.95 ± 23.85

Table 1. Pharmacokinetic parameters of sorafenib after single oral administration of sorafenib single or mixed formulas with BH 40, 20 or 10 mg/ml in male rats

Treatment PK Parameters	Sorafenib single formula	Sorafenib mixed formulas with BH		
	40 mg/kg	400 mg/kg	200 mg/kg	100 mg/kg
Tmax (hrs)	7.60 ± 0.89	$3.00 \pm 0.71a$	5.60 ± 2.51	3.80 ± 2.39
Cmax ($\mu\text{g/ml}$)	1.39 ± 0.23	$7.56 \pm 1.82a$	$5.03 \pm 1.28a$	$4.86 \pm 1.60a$
AUC_{0-t} ($\text{hr} \cdot \mu\text{g/ml}$)	20.92 ± 2.64	$99.05 \pm 28.24a$	$75.15 \pm 21.43a$	$66.60 \pm 16.10a$
$\text{AUC}_{0-\infty}$ ($\text{hr} \cdot \mu\text{g/ml}$)	27.23 ± 4.97	$122.05 \pm 37.24a$	$89.95 \pm 23.85a$	$91.90 \pm 14.95a$
$t_{1/2}$ (hr)	9.72 ± 4.08	10.12 ± 2.64	8.87 ± 1.87	13.43 ± 5.95
MRT_{∞} (hr)	16.14 ± 6.60	13.97 ± 2.64	13.40 ± 2.83	19.27 ± 9.18

Values are expressed as mean \pm S.D. of five rats. BH = Blue honeysuckle (fruit parts of *Lonicera caerulea* L., Caprifoliaceae) concentrated and lyophilized powder (H&K Bioscience Co., Ltd., Seoul, Korea). Cmax: The peak plasma concentration, Tmax: Time to reach Cmax, AUC_{0-t} : The total area under the plasma concentration-time curve from time zero to time measured, $\text{AUC}_{0-\infty}$: The total area under the plasma concentration-time curve from time zero to time infinity, $t_{1/2}$: Half life, MRT_{∞} : Mean residence to time infinity. ^a $p < 0.01$ as compared with sorafenib single formula administered rats.

and 91.90 ± 14.95 hr \cdot $\mu\text{g}/\text{ml}$ in sorafenib and BH 400, 200 or 100 mg/kg mixed formula treated rats, and as 27.23 ± 4.97 hr \cdot $\mu\text{g}/\text{ml}$ in sorafenib single formula treated rats, in this experiment, (Table 1).

5. Changes on the $t_{1/2}$ of sorafenib

The $t_{1/2}$ of sorafenib were non-significantly decreased as 4.11, -8.76 and 38.22% points in sorafenib and all three different concentrations of BH mixed formula treated rats as compared with sorafenib single formula treated rats, respectively. They were detected as 10.12 ± 2.64 , 8.87 ± 1.87 and 13.43 ± 5.95 hr in sorafenib and BH 400, 200 or 100 mg/kg mixed formula treated rats, and as 9.72 ± 4.08 hr in sorafenib single formula treated rats, in our experiment (Table 1).

6. Changes on the MRT_{inf} of sorafenib

The MRT_{inf} of sorafenib were non-significantly decreased as -13.44, -16.97 and 19.38% points in sorafenib and all three different concentrations of BH mixed formula treated rats as compared with sorafenib single formula treated rats, respectively. They were detected as 13.97 ± 3.81 , 13.40 ± 2.83 and 19.27 ± 9.18 hr in sorafenib and BH 400, 200 or 100 mg/kg mixed formula treated rats, and as 16.14 ± 6.60 hr in sorafenib single formula treated rats, in our experiment (Table 1).

IV. Discussion

As results of single oral administration of mixed formulas consisted of sorafenib and BH 400, 200 or 100 mg/kg, significant ($p < 0.01$) increases of plasma sorafenib concentrations were demonstrated from 30 min after end of administration

throughout all blood collected time points as compared with those of sorafenib 40 mg/kg single formula treated rats. Significant ($p < 0.01$) decreases of sorafenib T_{max} with increases of C_{max} , AUC_{0-t} and $\text{AUC}_{0-\text{inf}}$ were detected in sorafenib and BH 400 mg/kg mixed formulation treated rats as compared with sorafenib 40 mg/kg single formula treated rats, respectively. In addition, sorafenib and BH 200 or 100 mg/kg mixed formula treatment also showed significant ($p < 0.01$) increases of sorafenib C_{max} , AUC_{0-t} and $\text{AUC}_{0-\text{inf}}$ as compared with sorafenib 40 mg/kg single formula treated rats, respectively. These findings are considered as direct evidences that mixed formulation with BH significantly increased the oral bioavailability of sorafenib through increase of the absorptions, regardless of BH concentrations - 400, 200 and 100 mg/kg, and suggesting the mixed formulation of BH and sorafenib should be avoided as the integrative medicine for hepatic cancer.

The relative oral bioavailability of sorafenib was known as 38-49% after oral administration^{18,42}. The intake of high fat meals with sorafenib can reduce bioavailability by about 29%¹⁸. Sorafenib showed relatively high 99.5% protein bindings^{43,44}. T_{max} of sorafenib is approximately 3 hrs after oral administration^{45,46}, and slowly eliminated through feces (77%) and urine (19%) with relatively long approximately 25-48hr of $t_{1/2}$ ^{47,48}. In Japanese patient, who received sorafenib 400 mg twice daily, AUC of sorafenib reduced by 45% compared with data from phase 1 studies in Caucasian patients⁴⁹⁻⁵¹. In the present study, T_{max} of sorafenib in sorafenib single formula treatment was detected as 7.60 ± 0.89 hr, and C_{max} , AUC_{0-t} , $\text{AUC}_{0-\text{inf}}$, $t_{1/2}$ and MRT_{inf} were observed as 1.39 ± 0.23 $\mu\text{g}/\text{ml}$, 20.92 ± 2.64 hr \cdot $\mu\text{g}/\text{ml}$, 27.23 ± 4.97 hr \cdot $\mu\text{g}/\text{ml}$, 9.72 ± 4.08 hr and 16.14 ± 6.60 hr, respectively. In sorafenib and BH 400 mg/kg mixed formula administration, T_{max} , C_{max} , AUC_{0-t} , $\text{AUC}_{0-\text{inf}}$, $t_{1/2}$ and MRT_{inf} of

sorafenib were detected as 3.00 ± 0.71 hr, 7.56 ± 1.82 $\mu\text{g/ml}$, 99.05 ± 28.24 hr \cdot $\mu\text{g/ml}$, 122.05 ± 37.24 hr \cdot $\mu\text{g/ml}$, 10.12 ± 2.64 hr and 13.97 ± 3.81 hr as changed as -60.53 , 444.38 , 373.45 , 348.28 , 4.11 and -13.44% points as compared with sorafenib 40 mg/kg single formula treated rats, in the present study. They showed significant ($p < 0.01$) decreases of T_{max} with increases of C_{max} , AUC_{0-t} and $AUC_{0-\text{inf}}$ as compared with sorafenib 40 mg/kg single formula treated rats, respectively. Sorafenib and BH 200 mg/kg mixed formula administration also showed significant ($p < 0.01$) increases of C_{max} , AUC_{0-t} and $AUC_{0-\text{inf}}$ as compared with sorafenib 40 mg/kg single formula treated rats, respectively. They showed -26.32 , 262.25 , 259.20 , 230.37 , -8.76 and -16.97% points changes of T_{max} , C_{max} , AUC_{0-t} , $AUC_{0-\text{inf}}$, $t_{1/2}$ and MRT_{inf} as compared with sorafenib 40 mg/kg single formula treated rats, and detected as 5.60 ± 2.51 hr, 5.03 ± 1.28 $\mu\text{g/ml}$, 75.15 ± 21.43 hr \cdot $\mu\text{g/ml}$, 89.95 ± 23.85 hr \cdot $\mu\text{g/ml}$, 8.87 ± 1.87 hr and 13.40 ± 2.83 hr, in our result. In sorafenib and BH 100 mg/kg mixed formula administration, T_{max} , C_{max} , AUC_{0-t} , $AUC_{0-\text{inf}}$, $t_{1/2}$ and MRT_{inf} of sorafenib were detected as 3.80 ± 2.39 hr, 4.86 ± 1.60 $\mu\text{g/ml}$, 66.60 ± 16.10 hr \cdot $\mu\text{g/ml}$, 91.90 ± 14.95 hr \cdot $\mu\text{g/ml}$, 13.43 ± 5.95 hr and 19.27 ± 9.18 hr as changed as -50.00 , 250.43 , 218.34 , 237.55 , 38.22 and 19.38% points as compared with sorafenib 40 mg/kg single formula treated rats, in the present study. They showed significant ($p < 0.01$) increases of C_{max} , AUC_{0-t} and $AUC_{0-\text{inf}}$ as compared with sorafenib 40 mg/kg single formula treated rats, in the current study.

Sorafenib extensively metabolized to 8 metabolites, including an active metabolite, a pyridine N-oxide derivative in the liver through the oxidation by cytochrome P-450 3A4 and the glucuronidation by UGT1A9⁵²⁾ and, therefore, sorafenib can be interacted with other drugs affecting the liver microsomal drug-metabolizing

enzyme system, like dexamethasone, ketoconazole, rifampin and doxorubicin^{1,16-20)}. In addition, interactions with wafarin²¹⁾, St. John's wort¹⁾ and Gonjindan^{22,23)} were also already reported. In the present study, it is demonstrated that mixed formulation of BH with sorafenib significantly increased the oral bioavailability of sorafenib through increase of the absorptions, and therefore, it is considered that the mixed formulation of BH and sorafenib should be avoided in the comprehensive and integrative medicine for hepatic cancer.

V. Conclusions

Based on the results of the present study, mixed formulation of BH with sorafenib significantly increased the oral bioavailability of sorafenib through increase of the absorptions. Hence, the mixed formulation of BH and sorafenib should be avoided in the comprehensive and integrative medicine for hepatic cancer because mixed formulation with BH may be influenced on the pharmacokinetics of sorafenib enough to induce capricious drug-drug interactions, mainly increased the oral bioavailability.

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Reference

1. Bayer. Nexavar (sorafenib tosylate) tablets

- prescribing information. West Haven, CT2007 Feb.
2. Keating GM, Santoro A. Sorafenib: a review of its use in advanced hepatocellular carcinoma. *Drugs*. 2009;69(2):223–240.
 3. Wilhelm SM, Adnane L, Newell P, Villanueva A, Llovet JM, Lynch M. Preclinical overview of sorafenib, a multikinase inhibitor that targets both Raf and VEGF and PDGF receptor tyrosine kinase signaling. *Mol Cancer Ther*. 2008;7(10):3129–3140.
 4. Smalley KS, Xiao M, Villanueva J, Nguyen TK, Flaherty KT, Letrero R, et al. CRAF inhibition induces apoptosis in melanoma cells with non-V600E BRAF mutations. *Oncogene*. 2009;28(1):85–94.
 5. Fabian P, Berkovcov? J. Molecular predictive markers of EGFR-targeted therapy in metastatic colorectal cancer. *Cesk Patol*. 2011;47(4):154–158.
 6. Kang GH. Four molecular subtypes of colorectal cancer and their precursor lesions. *Arch Pathol Lab Med*. 2011;135(6):698–703.
 7. Ranieri G, Gadaleta-Caldarola G, Goffredo V, Patruno R, Mangia A, Rizzo A, et al. Sorafenib (BAY 43–9006) in hepatocellular carcinoma patients: from discovery to clinical development. *Curr Med Chem*. 2012 ;19(7):938–944.
 8. Dasanu CA, Alexandrescu DT, Dutcher J. Yellow skin discoloration associated with sorafenib use for treatment of metastatic renal cell carcinoma. *South Med J*. 2007;100(3):328–330.
 9. Alexandrescu DT, McClure R, Farzanmehr H, Dasanu CA. Secondary erythrocytosis produced by the tyrosine kinase inhibitors sunitinib and sorafenib. *J Clin Oncol*. 2008;26(24):4047–4048.
 10. Franck N, Barete S, Moguelet P, Blanchet B, Carlotti A, Ropert S, et al. Spiny follicular hyperkeratosis eruption: a new cutaneous adverse effect of sorafenib. *J Clin Oncol*. 2010;28(31):e640–642.
 11. Liang CP, Yang CS, Shen JL, Chen YJ. Sorafenib-induced acute localized exanthematous pustulosis in a patient with hepatocellular carcinoma. *Br J Dermatol*. 2011;165(2):443–445.
 12. McLellan B, Kerr H. Cutaneous toxicities of the multikinase inhibitors sorafenib and sunitinib. *Dermatol Ther*. 2011;24(4):396–400.
 13. Siu LL, Awada A, Takimoto CH, Piccart M, Schwartz B, Giannaris T, et al. Phase I trial of sorafenib and gemcitabine in advanced solid tumors with an expanded cohort in advanced pancreatic cancer. *Clin Cancer Res*. 2006;12(1):144–151.
 14. Ricciardi S, Mey V, Nannizzi S, Pasqualetti G, Crea F, Del Tacca M, et al. Synergistic cytotoxicity and molecular interaction on drug targets of sorafenib and gemcitabine in human pancreas cancer cells. *Chemotherapy*. 2010;56(4):303–312.
 15. Kupsch P, Henning BF, Passarge K, Richly H, Wiesemann K, Hilger RA, et al. Results of a phase I trial of sorafenib (BAY 43–9006) in combination with oxaliplatin in patients with refractory solid tumors, including colorectal cancer. *Clin Colorectal Cancer*. 2005;5(3):188–196.
 16. Mross K, Steinbild S, Baas F, Reil M, Buss P, Mersmann S, et al. Drug–drug interaction pharmacokinetic study with the Raf kinase inhibitor (RKI) BAY 43–9006 administered in combination with irinotecan (CPT–11) in patients with solid tumors. *Int J Clin Pharmacol Ther*. 2003;41(12):618–619.
 17. Hu S, Chen Z, Franke R, Orwick S, Zhao M, Rudek MA, et al. Interaction of the multikinase inhibitors sorafenib and sunitinib with solute carriers and ATP-binding cassette transporters. *Clin Cancer Res*. 2009;15(19):6062–6069.

18. Di Gion P, Kanefendt F, Lindauer A, Scheffler M, Doroshenko O, Fuhr U, et al. Clinical pharmacokinetics of tyrosine kinase inhibitors: focus on pyrimidines, pyridines and pyrroles. *Clin Pharmacokinet*. 2011;50(9):551–603.
19. Flaherty KT, Lathia C, Frye RF, Schuchter L, Redlinger M, Rosen M, et al. Interaction of sorafenib and cytochrome P450 isoenzymes in patients with advanced melanoma: a phase I/II pharmacokinetic interaction study. *Cancer Chemother Pharmacol*. 2011;68(5):1111–1118.
20. Infante JR, Jones SF, Bendell JC, Greco FA, Yardley DA, Lane CM, et al. A drug interaction study evaluating the pharmacokinetics and toxicity of sorafenib in combination with capecitabine. *Cancer Chemother Pharmacol*. 2012;69(1):137–144.
21. Moretti LV, Montalvo RO. Elevated International Normalized Ratio associated with concurrent use of sorafenib and warfarin. *Am J Health Syst Pharm*. 2009;66(23):2123–2125.
22. Kim SM, Lee CH, Park SJ, Kang SJ, Song CH, Han CH, et al. Effect of Gongjindan, a Polyherbal Formula on the Pharmacokinetics Profiles of Sorafenib in Male SD Rats (1) – Single Oral Combination Treatment of Sorafenib 50mg/kg with Gongjindan 100mg/kg within 5min. *J Soc Preventive Korean Med*. 2014;18(2):89–100.
23. Lee CH, Kim SM, Kang SJ, Park SJ, Song CH, Han CH, et al. Effect of Gongjindan–gamibang on the Pharmacokinetics Profiles of Sorafenib in Male SD Rats (2) – Single Oral Combination Treatment of Sorafenib 50mg/kg with Gongjindan–gamibang 100 mg/kg, 3.5hr–intervals with 7–day Repeated Treatment. *J Soc Preventive Korean Med*. 2015;19(1):145–159.
24. Svarcova I, Heinrich J, Valentova K. Berry fruits as a source of biologically active compounds: the case of *Lonicera caerulea*. *Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub*. 2007;151(2):163–174.
25. Chaovanalikit A, Thompson MM, Wrolstad RE. Characterization and quantification of anthocyanins and polyphenolics in blueh Honeysuckle (*Lonicera caerulea* L.). *J Agric Food Chem*. 2004;52(4):848–852.
26. Zhao H, Wang Z, Ma F, Yang X, Cheng C, Yao L. Protective effect of anthocyanin from *Lonicera Caerulea* var. *Edulis* on radiation–induced damage in mice. *Int J Mol Sci*. 2012;13(9):11773–11782.
27. Jurgoński A, Juśkiewicz J, Zduńczyk Z. An anthocyanin–rich extract from Kamchatka honeysuckle increases enzymatic activity within the gut and ameliorates abnormal lipid and glucose metabolism in rats. *Nutrition*. 2013;29(6):898–902.
28. Palíková I, Valentová K, Oborná I, Ulrichová J. Protectivity of blue honeysuckle extract against oxidative human endothelial cells and rat hepatocyte damage. *J Agric Food Chem*. 2009;57(15):6584–6589.
29. Jin XH, Ohgami K, Shiratori K, Suzuki Y, Koyama Y, Yoshida K, et al. Effects of blue honeysuckle (*Lonicera caerulea* L.) extract on lipopolysaccharide–induced inflammation in vitro and in vivo. *Exp Eye Res*. 2006;82(5):860–867.
30. Zdarilová A, Rajnochová Svobodová A, Chytilová K, Simánek V, Ulrichová J. Polyphenolic fraction of *Lonicera caerulea* L. fruits reduces oxidative stress and inflammatory markers induced by lipopolysaccharide in gingival fibroblasts. *Food Chem Toxicol*. 2010;48(6):1555–1561.
31. Chen L, Xin X, Yuan Q, Su D, Liu W. Phytochemical properties and antioxidant capacities of various colored berries. *J Sci Food Agric*. 2014;94(2):180–188.

32. Svobodová A, Rambousková J, Walterová D, Vostálová J. Protective effects of phenolic fraction of blue honeysuckle fruits against UVA-induced damage to human keratinocytes. *Arch Dermatol Res*. 2008;300(5):225–233.
33. Vostálová J, Galandáková A, Paliková I, Ulrichová J, Doležal D, Lichnovská R, et al. *Lonicera caerulea* fruits reduce UVA-induced damage in hairless mice. *J Photochem Photobiol B*. 2013;128:1–11.
34. Kim HS, Park SI, Choi SH, Song CH, Park SJ, Shin YK, et al. Single oral dose toxicity test of blue honeysuckle concentrate in mice. *Toxicol Res*. 2015;31(1):61–68.
35. AOAC (Association of Official Analytical Chemists). *Official Methods of Analysis International*. 17th ed. Washington, DC.: AOAC; 2000.
36. Singleton VL, Timberlake CF, Lea AGH. The phenolic cinnamates of white grapes and wine. *J Sci Food Agric*. 1978;29(4):403–410.
37. Yang J, Meyers KJ, van der Heide J, Liu RH. Varietal differences in phenolic content and antioxidant and antiproliferative activities of onions. *J Agric Food Chem*. 2004;52(22):6787–6793.
38. Boyles MJ, Wrolstad RE. Anthocyanin Composition of Red Raspberry Juice: Influences of Cultivar, Processing, and Environmental Factors. *J Food Sci*. 1993;58(5):1135–1141.
39. Gibaldi M, Perrier D. *Pharmacokinetics*. 2nd ed. New York: Marcel–Dekker; 1982.
40. Bailer AJ. Testing for the equality of area under the curves when using destructive measurement techniques. *J Pharmacokinet Biopharm*. 1988;16(3):303–309.
41. Chiou WL. Critical evaluation of the potential error in pharmacokinetic studies of using the linear trapezoidal rule method for the calculation of the area under the plasma level–time curve. *J Pharmacokinet Biopharm*. 1978;6(6):539–546.
42. Wang XQ, Fan JM, Liu YO, Zhao B, Jia ZR, Zhang Q. Bioavailability and pharmacokinetics of sorafenib suspension, nanoparticles and nanomatrix for oral administration to rat. *Int J Pharm*. 2011;419(1–2):339–346.
43. Tod M, Mir O, Bancelin N, Coriat R, Thomas–Schoemann A, Taieb F, et al. Functional and clinical evidence of the influence of sorafenib binding to albumin on sorafenib disposition in adult cancer patients. *Pharm Res*. 2011;28(12):3199–3207.
44. Villarroel MC, Pratz KW, Xu L, Wright JJ, Smith BD, Rudek MA. Plasma protein binding of sorafenib, a multi kinase inhibitor: in vitro and in cancer patients. *Invest New Drugs*. 2012;30(6):2096–2102.
45. Iyer R, Fetterly G, Lugade A, Thanavala Y. Sorafenib: a clinical and pharmacologic review. *Expert Opin Pharmacother*. 2010;11(11):1943–1955.
46. Kennoki T, Kondo T, Kimata N, Murakami J, Ishimori I, Nakazawa H, et al. Clinical results and pharmacokinetics of sorafenib in chronic hemodialysis patients with metastatic renal cell carcinoma in a single center. *Jpn J Clin Oncol*. 2011;41(5):647–655.
47. Clark JW, Eder JP, Ryan D, Lathia C, Lenz HJ. Safety and pharmacokinetics of the dual action Raf kinase and vascular endothelial growth factor receptor inhibitor, BAY 43–9006, in patients with advanced, refractory solid tumors. *Clin Cancer Res*. 2005;11(15):5472–5480.
48. Hornecker M, Blanchet B, Billefont B, Sassi H, Ropert S, Taieb F, et al. Saturable absorption of sorafenib in patients with solid tumors: a population model. *Invest New Drugs*. 2012;30(5):1991–2000.
49. Akaza H, Tsukamoto T, Murai M, Nakajima K, Naito S. Phase II study to investigate the efficacy, safety, and pharmacokinetics of sorafenib in Japanese patients with advanced renal cell carcinoma. *Jpn J Clin Oncol*.

- 2007;37(10):755-762.
50. Furuse J, Ishii H, Nakachi K, Suzuki E, Shimizu S, Nakajima K. Phase I study of sorafenib in Japanese patients with hepatocellular carcinoma. *Cancer Sci.* 2008; 99(1):159-165.
51. Minami H, Kawada K, Ebi H, Kitagawa K, Kim YI, Araki K, et al. Phase I and pharmacokinetic study of sorafenib, an oral multikinase inhibitor, in Japanese patients with advanced refractory solid tumors. *Cancer Sci.* 2008;99(7):1492-1498.
52. Sparidans RW, Vlaming ML, Lagas JS, Schinkel AH, Schellens JH, Beijnen JH. Liquid chromatography-tandem mass spectrometric assay for sorafenib and sorafenib-glucuronide in mouse plasma and liver homogenate and identification of the glucuronide metabolite. *J Chromatogr B Analyt Technol Biomed Life Sci.* 2009;877(3):269-276.