

ORIGINAL ARTICLE

Effects of Phosphorus and Iron on the Phytotoxicity of Lettuce (Lactuca sativa L.) in Arsenic-contaminated Soil

Min-Suk Kim¹, Hyun-Gi Min² and Jeong-Gyu Kim^{1,2}*

¹OJeong Eco-Resilience Institute, Korea University, Seoul, 02841, Korea

²Division of Environmental Science and Ecological Engineering, College of Life and Environmental Sciences, Korea University, Seoul, 02841, Korea

Received 29 November 2017, revised 23 January 2018, accepted 2 February 2018, published online 31 March 2018

ABSTRACT: We examined the effect of simultaneous application of phosphorus (P) and iron (Fe) on the phytotoxicity of lettuce in arsenic (As) contaminated soil using response surface methodology (RSM). To stabilize As and supply nutrient into soil, Fe and P were treated, respectively. Water soluble As and P was decreased by Fe application but increased by P application. Through phytotoxicity test, the result showed that only the addition of P affected lettuce root elongation even though both P and Fe were added. The correlation coefficients between root elongation and other indices indicated that the As content in the roots seemed to be the main reason that root growth was impeded. We could verify that the former result was not a passing phenomenon and Fe was necessarily needed to protect secondary pollution by exclusive usage of P fertilizer.

KEYWORDS: Arsenic, Iron, Mine tailing, Phosphorus, Response surface model

1. Introduction

Tailings containing high levels of arsenic (As) that originated from mining activities have been recognized as a principal anthropogenic source of As contamination of soil (Bruce et al. 2003; Seidel et al. 2005). Many other studies related to the remediation of As-contaminated soil have reported that chemical stabilization using various types of amendments has been identified as a lower cost and lower input method than other physical and chemical methods (Lee et al. 2009; Koo et al. 2013). Kumpiene et al. (2008) also reported that the type of amendments and those of mechanisms of As stabilization. Among the amendments for As stabilization in soil, iron (Fe) sources have been known to be effective and common agents and have been

more widely used than other amendments (Bowell 1994; Warren and Alloway 2003; Lee et al. 2011; Koo et al. 2012).

For real remediation, not only should As be stabilized but re-vegetation should also be accomplished. Vegetation could promote the coverage of the tailing area, resulting in a decrease in soil erosion and As hazards to humans and an improved landscape and pollution management (Tordoff et al. 2000; Conesa et al. 2007). Soils contaminated with mine tailings are not suitable for plant growth because of macronutrient deficiencies (Tordoff et al. 2000; Koo et al. 2013). Hence, many agents, such as pH control agents, carbon sources, and phosphorus (P) have been applied to enhance soil fertility (Chen et al. 1998; Heeraman et al. 2001; Juwarkar et al. 2008).

^{*}Corresponding author: lemonkim@korea.ac.kr, ORCID 0000-0002-5734-1311

[©] Korean Society of Ecology and Infrastructure Engineering. All rights reserved.

This is an open-access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/3.0/), which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Because As and P have similar chemical characteristics in soil, competition for adsorption sites occurs (e.g., Fe - (hydro) oxide and organic matters); thus, the mobility of As in soil has been increased by the input of P-rich agents (Woolson et al. 1973; Cao et al. 2003). Cao et al. (2003) reported that P amendments significantly increased soil water-soluble As, resulting in the enhancement of As uptake by plants. Furthermore, Cao and Ma (2004) found that the incorporation of phosphate in soil increased As accumulation in carrots and lettuce resulting from an increase in soil watersoluble As because of the replacement of As by phosphate in soil. Additionally, Davenport and Peryea (1991), via a column experiment, found that the addition of P fertilizer significantly increased As leaching from soil. To identify specific interactions among As, P, and Fe, Koo et al. (2013) demonstrated the effects of P and Fe in As-spiked soils via a sequential incorporation method for Fe and P using response surface methodology. They found that the P content of soil affected the reduction of As toxicity towards the root elongation of lettuce by providing a nutrient source rather than by suppressing the uptake of As. The soil Fe content also caused root elongation by reducing the mobility of As. Nevertheless, adding P and Fe to As-contaminated soils stimulated plant

growth during the early stage of root growth; the experiment was limited to the use of artificial soil. Unlike artificial soil, various factors in As-contaminated field soil affect As mobility (e.g., the pH value, redox potential, metal oxides, organic matter, and clay minerals) but the related studies are insufficient (Bissen and Frimmel 2003; Signes-Pastor et al. 2007; Hossain et al. 2008).

Thus, the purpose of this study was to investigate the effects of P and Fe on the availability of As and the root elongation of lettuce (*Lactuca sativa* L.) in As-contaminated soil using response surface methodology. The model was used for the interpretation of the effects of P and Fe on As mobility and As accumulation in lettuce.

2. Materials and Methods

2.1 Experimental design and set-up

In the present study, the second-order central composite rotate design (CCRD) was adopted and the values of parameters selected for CCRD are summarized in Table 1. The studied model (3 experimental points and 11 experiments) required for two factors and five levels in the CCRD, which were determined as

| | Treatment | Code levels | | Actual leve | els (g kg ⁻¹) | WS-As ^a | WS-P ^b |
|---------|-----------|-------------|-------|-------------|---------------------------|---------------------|-------------------|
| | | Р | Fe | Р | Fe | mg kg ⁻¹ | mg kg⁻¹ |
| Cube | T1 | -1 | -1 | 0.155 | 0.279 | 11.29±0.59 | 29±2.9 |
| | T2 | -1 | 1 | 0.155 | 0.838 | 7.83±0.34 | 20±1.9 |
| | Т3 | 1 | -1 | 0.465 | 0.279 | 19.71±1.29 | 286±47.4 |
| | T4 | 1 | 1 | 0.465 | 0.838 | 18.31±1.14 | 247±21.8 |
| Star | T5 | 0 | -1.41 | 0.310 | 0.165 | 15.54±0.79 | 126±11.8 |
| | Т6 | 0 | 1.41 | 0.310 | 0.952 | 13.59±1.15 | 129±6.3 |
| | T7 | -1.41 | 0 | 0.091 | 0.559 | 5.47±0.24 | 14±1.1 |
| | T8 | 1.41 | 0 | 0.528 | 0.559 | 10.41±0.88 | 59±6.3 |
| Control | Т9 | 0 | 0 | 0.310 | 0.559 | 14.43±1.69 | 131±13.2 |
| | T10 | 0 | 0 | 0.310 | 0.559 | 14.40±1.27 | 115±17.9 |
| | T11 | 0 | 0 | 0.310 | 0.559 | 11.39±0.61 | 116±3.6 |

Table 1. Experimental set-up used in the central composite rotate design (CCRD) experiment and the effects of phosphorus (P) and iron (Fe) on arsenic (As) and phosphorus (P) extractability

^aWater soluble As extracted by deionized water; ^bWater soluble P extracted by deionized water.

follows: $2^{n} (2^{2} = 4$: cube points) + $2n (2 \times 2$: star point) + 3 (central points: three replicates) (Son et al. 2009, Koo et al. 2011). Low and high factors (cube points) are coded as -1 and 1, the mid factor settings (central points) coded as 0, and the center points of the tube (star points) are coded as -1.41 and 1.41 (Hao et al. 2006).

Using multiple regression analysis of the experimental results, the following response *Y* was calculated through the second-order polynomial model:

$$Y = \beta_0 + \beta_1 \chi_1 + \beta_2 \chi_2 + \beta_{12} \chi_1 \chi_2 + \beta_{11} \chi_1^2 + \beta_{22} \chi_2^2$$
(Eq. 1)

where *Y* contains the dependent variables (As mobility and bioavailability using chemical extraction methods, the root elongation of lettuce, and the amount of As and P in lettuce root and shoot), β_0 is the intercept, β_i the linear coefficient, β_{ij} the interaction coefficient, β_{jj} the quadratic coefficient, and χ_1 and χ_2 (for P and Fe contents, respectively) the coded independent variables. The experimental design of the present study developed mathematical models for explanation of the results. The generated model also was used in simulations for providing useful information on the effects P and Fe on As mobility and toxicity towards lettuce growth.

2.2 Characterization of soil sample and treatment of P and Fe

Surface soil contaminated with mine tailing was taken from the Gangwon mine, a former gold mine site at Gangwon province, the Republic of Korea $(37^{\circ}19'19.6", 128^{\circ}48'47.4")$ at November, 2014. The soil sample was air dried and passed through a 2 mm sieve. The soil pH and electrical conductivity (EC) were determined using 1:5 soil:water suspension with a combination pH-EC meter (Thermo Orion 920A, USA). The total phosphorus was determined followed by HClO₄ digestion methods and the concentration of P in filtrates was determined using an ultraviolet (UV) spectrophotometer (UV-1650PC, Shimadzu, Japan), via a modified molybdenum blue method, which removed the arsenate interference by reducing the arsenate (V) to arsenite (III) using dithionite ($S_2O_4^{2-}$) (Tsang et al. 2007). The available phosphorus in soil was measured using the Mehlich-3 solution (Mehlich 1984). The concentration of P in filtrate was also determined with the same previous method. The total As concentration was determined by digesting samples with aqua regia, a mixture of HNO₃/HCl (v:v = 1:3), using an inductively coupled plasma-optical emission spectrometer (ICP-OES) (Vista Pro, Varian USA). The Fe, Al, and Mn concentrations were determined followed by ammonium oxalate extraction methods (Loeppert and Inskeep 1996).

Eleven different soil compositions were provided by adjusting the P and Fe contents to levels determined by the CCRD (Table 1) considering the results from former study (Koo et al. 2013). P (ranging from 0.091 to 0.465 g kg⁻¹) and Fe (ranging from 0.165 to 0.952 g kg⁻¹) were added in the forms of Na₂HPO₄ (Sigma-Aldrich, USA) and zero-valent iron (ZVI, obtained from Sigma-Aldrich), respectively. After treatment, the soils were equilibrated at room temperature and 60% of the water holding capacity (WHC) for 4 weeks prior to the chemical extraction and phytotoxicity tests. In addition, five confirmation experiments were subsequently performed to support the findings resulting from the CCRD study (Table 2).

2.3 Chemical assay with single extraction method

The effect of P and Fe on As extractability was evaluated using deionized water (DW). For water soluble-As, 1 g soil was added to 20 mL DW in 50 mL polypropylene centrifuge tubes (Falcon, London, UK), which were then shaken for 1 hour on a wristaction shaker, centrifuged for 20 min at 5100 rpm, and then filtered with a PTFE 0.45 um pore size syringe filter (Rodriquez et al. 2003). All the remaining As and P contents in the filtrates were determined using ICP-OES after acidification (Sikora et al. 2005).

| Treatment | Code | levels | Actual leve | els (g kg ⁻¹) | WS-Asa | WS-Pb |
|-----------|-------|--------|-------------|---------------------------|---------------------|---------------------|
| | Р | Fe | Р | Fe | mg kg ⁻¹ | mg kg ⁻¹ |
| T21 | -1 | -2 | 0.155 | 0.000 | 19±0.8 | 47±3.9 |
| T23 | 1 | -2 | 0.465 | 0.000 | 40±1.1 | 346±19.1 |
| T25 | 0 | -2 | 0.310 | 0.000 | 33±0.9 | 148±11.8 |
| T27 | -1.41 | -2 | 0.091 | 0.000 | 11±0.6 | 16±1.4 |
| T28 | 1.41 | -2 | 0.528 | 0.000 | 44±0.4 | 382±13.5 |

Table 2. Experimental set-up used in the confirmation experiment and the effects of phosphorus (P) on arsenic (As) and phosphorus (P) extractability

^aWater soluble As extracted by deionized water; ^bWater soluble P extracted by deionized water.

2.4 Biological assay with lettuce

The root elongation tests using lettuce (Lactuca sativa L.) were conducted according to the OECD Guidelines for the Testing of Chemicals (OECD 208 2006). Prior to sowing, the seeds were sterilized with 10% H₂O₂ and subsequently 12 seeds were placed in 100×20 mm plastic Petri-dishes which containing 40 g of the treated soils. The dishes were placed in a growth chamber randomly. The moisture content was controlled at approximately 60% of the WHC of the soils, with the light conditions of 16 hours of daylight and 8 hours of darkness per day, at $20 \pm 2^{\circ}$ C. Four weeks after sowing, the plants were harvested to determine their root growth using an image analyzer program (WinRhizo 5.0a, Regent, Canada). The root elongation was calculated for each Petri-dish based on the each root length of plant and expressed as mm seedling⁻¹. After root elongation determining, the plants were separated into roots, shoots and dried in an oven at 70°C. The dried samples were digested with HNO₃ and H₂O₂ by hot-block digestion procedure, and immediately filtered. The filtrates were used to determine the concentrations of As using ICE-OES. The concentration of P in filtrates was determined using same method for total phosphorus analysis (section 2.2).

2.5 Statistical analysis

The statistical analysis of the results was performed using the SAS program (SAS 9.2, USA). The relationships among the results were evaluated using a Pearson's correlation analysis at p<0.05. The parameters and their significances in the second-order polynomial models were tested using the multiple regression procedure and fitted to a second-order polynomial model.

3. Results and Discussion

3.1 Soil properties

An As analysis indicated that the soil was highly contaminated by As (1,854 mg kg⁻¹), resulting in the exceedance of the 'regulatory level' for As (75 mg kg⁻¹) described in the Soil Environmental Conservation Act of South Korea (KMoE 2011). The pH and EC of soil were 8.41 and 0.76 ds m⁻¹, respectively, a slightly basic pH. The total and available phosphorus (P) concentrations of 1.10 and 0.07 g kg⁻¹, respectively indicated that the available form of P was low (6.36%), so the effect of available P on As mobility in the soil was also low. The oxalate extractable A1 (Al_{ox}), Fe (Fe_{ox}), and Mn (Mn_{ox}) concentrations were 1.18, 13.4, and 2.0 mg g⁻¹, respectively. The highest Fe_{ox} was observed in a soil sample, indicating that the main adsorption sites for As is in soil.

3.2 Effects of Fe and P on As extractability

The application of P and Fe had an effect on the As extractability in soil and water soluble-As and P (Table 1). A model fitting the results from Table 1 was

| | Code levels | | REa | A p b | A C | Proot ^d | D e |
|-----|-------------|-------|----------|---------------------------------|----------------------------------|--------------------|---------------------------------|
| | Р | Fe | R⊑a | As _{root} ^b | As _{shoot} ^c | Froot | P _{shoot} ^e |
| T1 | -1 | -1 | 26.0±3.7 | 0.62±0.09 | 25±1.9 | 9.8±0.95 | 8.59±0.46 |
| T2 | -1 | 1 | 29.2±2.7 | 0.45±0.06 | 22±1.1 | 7.6±0.88 | 7.15±0.49 |
| Т3 | 1 | -1 | 26.1±2.2 | 0.43±0.06 | 18±0.9 | 24.6±0.24 | 12.1±0.61 |
| T4 | 1 | 1 | 30.0±5.9 | 0.43±0.04 | 45±4.7 | 27.3±1.65 | 16.2±1.78 |
| T5 | 0 | -1.41 | 28.4±3.9 | 0.46±0.06 | 36±6.7 | 19.6±3.35 | 13.1±1.59 |
| T6 | 0 | 1.41 | 25.8±0.9 | 0.45±0.04 | 16±3.9 | 13.0±0.29 | 10.0±1.09 |
| T7 | -1.41 | 0 | 25.7±3.7 | 0.56±0.03 | 18±4.3 | 6.2±0.78 | 7.1±0.53 |
| T8 | 1.41 | 0 | 32.2±1.7 | 0.48±0.05 | 22±4.4 | 11.9±0.37 | 9.9±0.93 |
| Т9 | 0 | 0 | 29.6±2.9 | 0.44±0.02 | 23±0.3 | 15.2±0.49 | 11.1±0.31 |
| T10 | 0 | 0 | 27.4±4.6 | 0.46±0.03 | 24±1.1 | 14.6±2.51 | 12.0±1.26 |
| T11 | 0 | 0 | 36.8±1.3 | 0.41±0.09 | 26±2.3 | 19.8±3.31 | 11.4±0.27 |

Table 3. Effect of phosphorus (P) and iron (Fe) on the root elongation of lettuce (Lactuca sativa L.) seeds and the arsenic (As) and phosphorus (P) uptakes

^aRoot elongation of germinated lettuce seeds (mm seedling⁻¹); ^bAs concentrations in lettuce roots (mg g⁻¹); ^cAs concentrations in lettuce shoots (mg kg⁻¹); ^dP concentrations in lettuce roots (mg g⁻¹); ^eP concentrations in lettuce shoots (mg g⁻¹).

constructed with a regression analysis. The *p* value in analysis of variance (ANOVA) was less than 5% of the estimated *F* value (7.2) obtained for the water soluble-As extractability, indicating that the model was significant at a high confidence level (95%). The correlation coefficient, r^2 , was 0.474, and the model applied to the data yielded the following equation:

The water soluble As $(mg kg^{-1}) = 13.10 + 3.24 \times P$ -0.95 × Fe (Eq. 2)

The model provided significant information for deciding which soil parameter was important for As mobility. As expected, a significant and positive coefficient was obtained for the P content and the Fe content had different results than that of P. In addition, the results showed that the effect of P on the changes in As mobility was greater than that of Fe by comparing the estimated coefficients for P (3.24) and Fe (0.95) (Eq. 2). According Cao and Ma (2004), the application of phosphate in soil increased the water-soluble As because of the replacement of As by phosphate in the soil. Additionally, Fe is well known to reduce As mobility in soil via the adsorption of As onto its surface through previous studies (Bowell 1994; Warren and Alloway 2003; Lee et al. 2011; Koo et al. 2012). In this regard, the changes in As mobility were expected to directly affect plant As uptake and toxicity.

3.3 Effects of Fe and P on root elongation in lettuce

The simultaneous application of P and Fe also affected root elongation in lettuce (Table 3). The *p* value in ANOVA for the root elongation of lettuce was not less than 5% of the estimated *F* value (3.80), indicating that the result was not significant, and the calculated correlation coefficient, $r^2 = 0.3221$, was poor. Although the data did not show significant results, the developed model yielded the following equation:

The root elongation of lettuce (mm seedling⁻¹)
=
$$28.40 + 2.18 \times P$$
 (Eq. 3)

There was no doubt that Fe affected both the As mobility and lettuce root growth. Nevertheless, the model provided information that only the addition of P affected lettuce root elongation even though both P and Fe were added. According to a similar study by Koo et al. (2013), both P and Fe affected lettuce root growth significantly, and the estimated coefficient for P (185.81) was higher than that for Fe (92.91). This result might be explained because the effectiveness of P was much greater than that of Fe. Moreover, the present study used As-contaminated soil unlike the previous study (Koo et al. 2013), so other factors (e.g., macro/micro nutrients, dissolved organic carbon, or clay content, other metal hydroxide/oxides) could have affected As mobility and phytoavailability (Cai et al. 2002; Fitz and Wenzel 2002; Koo et al. 2011). Additionally, this result contradicted our earlier assumption that the addition of P increases As mobility, which causes an increase in phytotoxicity. On the contrary, the addition of P seemed to promote root growth by acting as a nutrient rather than a toxin to increase As mobility. From these results, it was concluded that in As-contaminated soil, the application of P increases soil nutrient levels, promoting lettuce root growth. To interpret and demonstrate the results, more data will be discussed below.

3.4 Correlation analysis for lettuce growth and its As and P uptake

The relationships among the results were evaluated using Pearson's correlation analysis (Table 4). As expected, a negative coefficient (r = 0.4698) was found only between the effect on the lettuce root elongation and the As content in lettuce roots. It is known that the toxic effects of the absorbed As on root growth are caused by oxidative stress induced by an increase in the production of reactive oxygen species (ROS). ROS might cause the inhibition of root elongation by disrupting cellular activity or the interference with plant cell metabolism (Gratão et al. 2005; Koo et al. 2011). The correlation coefficients between root elongation and other indices indicated that the As content in the roots seemed to be the main reason that root growth was impeded, even though the result was not significant. This statistical result resulted from the complexity and variety of the examined field soil compared to that in the previous study using artificial soil (Koo et al. 2013).

The absorbed As content in lettuce roots was decreased by increasing the amount of P (r = -0.5918) and Fe (r = -0.3293) (Table 4). In general, Fe (hydro) oxides have a high affinity for the adsorption of As; hence, the As mobility in soils is reduced with the presence of Fe on the soil surface (Codling and Dao 2007; Koo et al. 2013). Rahman et al. (2013) reported that Fe²⁺ input decreased the As concentration in the roots of rice seedlings in paddy soil despite the uptake of As by the rice roots via a phosphate uptake pathway due to the similar physicochemical characteristics of Fe²⁺ and As (Liu et al. 2004). In addition, the use of Fe⁰ in As contaminated soil also reduced the accumulation

| | RE ^a | As _{root} ^b | As _{shoot} ^c | Proot ^d | P _{shoot} ^e | FW _{root} ^f | FW _{shoot} ^g |
|--------------------------------|-----------------|---------------------------------|----------------------------------|--------------------|---------------------------------|---------------------------------|----------------------------------|
| P _{trt} ^h | 0.3883 | -0.5918 | 0.2926 | 0.7049* | 0.6892* | 0.5632 | 0.6485* |
| Fe _{trt} ⁱ | 0.0737 | -0.3293 | -0.0538 | -0.1466 | -0.0721 | 0.4926 | 0.5114 |
| As _{root} | -0.4698 | | -0.2056 | -0.6436* | -0.5745 | -0.7675** | -0.7296* |
| As _{shoot} | 0.3649 | | | 0.5986 | 0.7460** | -0.1049 | 0.4253 |
| Proot | 0.3068 | | | | 0.9251*** | 0.3219 | 0.5497 |
| P _{shoot} | 0.2773 | | | | | 0.1779 | 0.5762 |

Table 4. Correlation coefficients (r) among the experimental results of the central composite rotate design (CCRD) experiment

Asteriske (*,**, and ***) indicate P<0.05, 0.01, and 0.001, respectively

^aRoot elongation of lettuce; ^bAs concentrations in lettuce roots; ^cAs concentrations in lettuce shoots; ^dP concentrations in lettuce shoots; ^fFreash weights of lettuce roots; ^gFreash weights of lettuce shoots; ^hP treatment contents in soils; ⁱFe treatment contents in soils.

of As in Panax notoginseng (Burk.) roots (Yan et al. 2013). The As concentration in radish roots was significantly decreased by approximately 50% by the addition of steel-making slag (SMS) compared to the control (Qutierrez et al. 2010). For P, as might not be expected, the As concentration in lettuce roots also decreased with an increase in P. This result agreed with a previous study (Koo et al. 2013) that showed that the As content in lettuce roots was negatively related to the P concentration in artificial soil (r =-0.459). These similar results were obtained because of competition between As and P for the same P transporters in plasma membrane of root cells (Ullrich-Eberius et al. 1989; Meharg and Hartly-Whitaker 2002; Gunes et al. 2009). This interpretation was also applicable to another correlation between the As and P concentration in lettuce roots (p = -0.6434). Because As and P competed when uptake occurred near the root surface, both As and P interfered with each other.

The P concentration in roots and shoots increased significantly (p = 0.7049 and p = 0.6892, respectively), and a high correlation coefficient was found between roots and shoots (p = 0.9251) (Table 4). P is a plant growth macronutrient and is involved in key functions, such as structural cell components (e.g., phospholipids and nucleic acids), and in plant metabolism (e.g., energy transfer, photosynthesis, nutrient movement within plants) (Cao et al. 2003; Knudson et al. 2003; Rufyikiri et al. 2006). Thus, the fresh weights of roots and shoots were positively related to the P treatment

concentration (r = 0.5632 and r = 0.6485, respectively). Conversely, the absorbed As content in roots decreased the fresh weight of roots and shoots, significantly (r =-0.7675 and r = -0.7296, respectively), indicating that the uptake of As by roots might suppress root elongation, leading to a decrease in the weight of the lettuce.

3.5 Confirmation study

Additional experiments (Table 5) were conducted to verify the effectiveness of Fe followed by P treatment. The treatment concentration of P in T21, T23, T25, T27, and T28 were same as T1, T3, T5, T7 and T8, respectively, without any Fe treatment. Therefore, the only effects of P in the absence of Fe occurred for As and P extractability and As and P absorption into lettuce. Compared to table 1 (T1, T3, T5, T7, and T8), table 2 (T21, T23, T25, T27, T28) showed that the concentration of water soluble As and P increased in the absence of Fe and that the degree of increase was higher for P than for As, indicating that the available P was increased. The changes in the extractability increased the absorption of P, but decreased that of As by lettuce roots, leading to an increase in root elongation in lettuce (Table 5) compared to Table 3. It seems that the absorption of As by lettuce roots was decreased due to the suppression of P in plasma membrane of root cells, which was same as earlier results. Conversely, these results indicated that although the addition of

| Table 5. Effect of phosphorus (P) on the root elongation | of lettuce (Lactuca sativa L.) seeds and the their arsenic |
|--|--|
| (As) and phosphorus (P) uptakes in confirmation study | , |

| Treatment | Code levels | | RE ^a | Asroot ^b | Asshoot ^c | Proot ^d | Pshoot ^e |
|-----------|-------------|----|-----------------|---------------------|----------------------|--------------------|---------------------|
| | Р | Fe | | ASIOOL | ASSHOOL | FIOOL | FSHOOL |
| T21 | -1 | -2 | 38.6±6.3 | 0.44±0.08 | 29±6.0 | 17.3±3.22 | 9.97±1.85 |
| T23 | 1 | -2 | 40.6±1.1 | 0.37±0.05 | 17±1.0 | 23.5±1.10 | 11.37±0.25 |
| T25 | 0 | -2 | 38.2±4.5 | 0.40±0.07 | 30±7.9 | 23.1±1.45 | 11.31±0.71 |
| T27 | -1.41 | -2 | 38.1±2.9 | 0.44±0.04 | 27±6.2 | 6.6±0.64 | 6.20±0.35 |
| T28 | 1.41 | -2 | 38.4±0.4 | 0.42±0.04 | 23±5.3 | 24.1±3.13 | 12.30±1.76 |

^aRoot elongation of germinated lettuce seeds (mm seedling⁻¹); ^bAs concentrations in lettuce roots (mg g⁻¹); ^cAs concentrations in lettuce shoots (mg kg⁻¹); ^dP concentrations in lettuce roots (mg g⁻¹); ^eP concentrations in lettuce shoots (mg g⁻¹).

Fe decreased the extractability of both As and P, the phytoavailability and phytotoxicity were greatly affected by the relative available amounts of As and P in soil.

4. Conclusion

This study evaluated the effects of P and Fe on the extractability of As and P and phytotoxicity in soil highly contaminated by As. Under an incorporation of both P and Fe, the P treatment increased root elongation of lettuce by supplying nutrient source and decreased As absorption in lettuce root by suppressing the uptake of As. Under only the presence of P condition, the P treatment content increased the contents of available P more than that of available As, which resulted in the deceases in As toxicity toward lettuce, more effectively. Nevertheless, only the application of P could cause increases in As mobility, leading deterioration of environmental health. Also this study used limited environmental soil sample and chemical reagents. Thus, further study seems to be needed for the application of P and Fe to field scale using various adaptable P fertilizers and Fe sources.

Acknowledgement

This work was supported by the Basic Science Research Program through the National Research Foundation of Korea (NRF) [2015R1D1A1A01057594] and by a grant from Korea University.

References

- Bissen, M. and Frimmel, F.H. 2003. Arsenic-a Review: Part I: Occurrence, toxicity, speciation, mobility. Acta Hydrochimica et Hydrobiologica 31(1): 9-18.
- Bowell, R.J. 1994. Sorption of arsenic by iron oxides and oxyhydroxides in soils. Applied Geochemistry 9: 279-286.
- Bruce, S.L., Noller, B.N., Grigg, A.H., Mullen, B.F., Mulligan, D.R., Ritchie, P.J., Currey, N. and Ng. J.C. 2003. A field study conducted at Kidston gold mine to evaluate the impact of arsenic and zinc from mine tailing to grazing

cattle. Toxicology Letters 137: 23-34.

- Cai, L., Xu, Z., Ren, M., Guo, Q., Hu, X., Hu, G., Wan, H. and Peng, P. 2012. Source identification of eight hazardous heavy metals in agricultural soils of Huizhou, Guangdong Province, China. Ecotoxicology and Environmental Safety 78: 2-8.
- Cao, X., Ma, L.Q. and Shiralipour, A. 2003. Effects of compost and phosphate amendments on arsenic mobility in soils and arsenic uptake by the hyperaccumulator, *Pteris vittata* L. Environmental Pollution 126: 157-167.
- Cao, X. and Ma, L.Q. 2004. Effects of compost and phosphate on plant arsenic accumulation from soils near pressure-treated wood. Environmental Pollution 132: 435-442.
- Chen, Y., Chefetz, B., Adani, F., Genevini, P.L. and Hadar, Y. 1998. Organic matter transformation during composting of municipal solid waste. In, Drozd, J., Gonet, S.S., Senesi, N. and Weber, J. (eds.), The role of humic substances in the ecosystems and in environmental protection, PTSH, Wroclaw, Poland. pp. 155-182.
- Codling, E.E. and Dao, T.H. 2007. Short-term effect of lime, phosphorus, and iron amendments on water-extractable lead and arsenic in orchard soils. Communications in Soil Science and Plant Analysis. 38: 903-919.
- Conesa, H.M., Brett, H.R., Schulin, R. and Nowack, B. 2007. Growth of *Lygeum spartum* in acid mine tailings: response of plants developed from seedlings, rhizomes and at field conditions. Environmental Pollution 145: 700-707.
- Davenport, J.R. and Peryea, F.J. 1991. Phosphate fertilizers influence leaching of lead and arsenic in a soil contaminated with lead arsenate. Water Air and Soil Pollution 57(1): 101-110.
- Fitz, W.J. and Wenzel, W.W. 2002. Arsenic transformations in the soil-rhizosphere-plant system: fundamentals and potential application to phytoremediation. Journal of Biotechnology 99(3): 259-278.
- Gratão, P.L., Polle, A., Lea, P.J. and Azevedo, R.A. 2005. Making the life of heavy metal-stressed plants a little easier. Functional Plant Biology 32: 481-494.
- Gunes, A., Pilbeam, D.J. and Inal, A. 2009. Effect of arsenic-phosphorus interaction on arsenic-induced oxidative stress in chickpea plants. Plant and Soil 314: 211-220.
- Gutierrez, J., Hong, C.O., Lee, B.H. and Kim, P.J. 2010. Effect of steel-making slag as a soil amendment on arsenic uptake by radish (*Raphanus sativa L.*) in an upland soil. Biology and Fertility of Soils 46: 617-623.
- Hao, D.C., Zhu, P.H., Yang, S.L. and Yang, L. 2006. Optimization of recombinant cytochrome P450 2C9 protein production in *Escherichia coli* DH5-α by statisticallybased experimental design. Journal of Microbiology and Biotechnology 22: 1169-1176.
- Heeraman, D.A., Claassen, V.P. and Zasosk, R.J. 2001. Interaction of lime, organic matter, and fertilizer on

growth and uptake of arsenic and mercury by Zorro fescue (*Vulpia myuros* L.). Plant and Soil 234: 215-231.

- Hossain, M.B., Jahiruddin, M., Panaullah, G.M., Loeppert, R.H., Islam, M.R. and Duxbury, J.M. 2008. Spatial variability of arsenic concentration in soils and plants, and its relationship with iron, manganese and phosphorus. Environmental Pollution 156: 739-744.
- Juwarkar, A.A., Yadav, S.K., Kumar, P. and Singh, S.K. 2008. Effect of biosludge and biofertilizer amendment on growth of *Jatropha curcas* in heavy metal-contaminated soils. Environmental Monitoring and Assessment 145: 7-15.
- Knudson, J.A., Meikle, T. and DeLuca, T.H. 2003. Role of mycorrhizal fungi and phosphorus in the arsenic tolerance of basin wildrye. Journal of Environment Quality 32: 2001-2006.
- Koo, N., Jo, H.J., Lee, S.H. and Kim, J.G. 2011. Using response surface methodology to assess the effects of iron and spent mushroom substrate on arsenic phytotoxicity in lettuce (*Lactuca sativa* L.). Journal of Hazardous Materials 192: 381-387.
- Koo, N., Lee, S.H. and Kim, J.G. 2012. Arsenic mobility in the amended mine tailings and its impact on soil enzyme activity. Environmental Geochemistry and Health 34: 337-348.
- Koo, N., Kim, M.S., Hyun, S. and Kim, J.G. 2013. Effects of the incorporation of phosphorus and iron into arsenicspiked artificial soils on root growth of lettuce using response surface methodology. Communications in Soil Science and Plants 44: 1259-1271.
- Korea Ministry of Environment (KMoE). 2011. The soil environment conservation act. Republic of Korea: Korea Legislation Research Institute. (in Korean)
- Kumpiene, J., Lagerkvist, A. and Maurice, C. 2008. Stabilization of As, Cr, Cu, Pb, and Zn in soil using amendments: A review. Waste Management 28: 215-225.
- Lee, S.H., Lee, J.S., Choi, Y.J. and Kim, J.G. 2009. In situ stabilization of cadmium-, lead-, and zinc-contaminated soil using various amendments. Chemosphere 77: 1069-1075.
- Lee, S.H., Kim, E.Y., Park, H., Yun, J. and Kim, J.G. 2011. In situ stabilization of arsenic and metal-contaminated agricultural soil using industrial by-products. Geoderma 161: 1-7.
- Liu, W.J., Zhu, Y.G., Smith, F.A. and Smith, S.E. 2004. Do iron plaque and genotypes affect arsenate uptake and translocation by rice seedlings (*Oryza sativa* L.) grown in solution culture? Journal of Experimental Botany 55(403): 1707-1713.
- Loeppert, R.H. and Inskeep, W.P. 1996. Iron, In, Spark, D.L., Page, A.L., Loeppert, R.H., Johnston, C.T., Sumner, M.E. and Bigham, J.M. (eds.), Methods of soil analysis, Part 3, Soil Science Society of America and American Society of Agronomy, Madison, USA, pp. 639-664.

- Meharg, A.A. and Hartley-Whitaker, J. 2002. Arsenic uptake and metabolism in arsenic resistant and nonresistant plant species. New Phytologist Journal 154: 29-43.
- Mehlich, A. 1984. Mehlich 3 soil test extractant: A modification of Mehlich 2 extractant. Communications in Soil Science and Plant Analysis. 15(2): 1409-1416.
- OECD 2006. Test no. 208: Terrestrial plant test, seedling emergence, and seedling growth test, Paris, France: OECD Press.
- Rahman, M.A., Karmakar, S., Salama, H., Gactha-Bandjun, N. and Noubactep, C. 2013. Optimising the design of Fe⁰-based filtration systems for water treatment: The suitability of porous iron composites. Journal of Applied Solution Chemistry and Modeling 2(3): 165-177.
- Rodriguez, R.R., Basta, N.T., Casteel, S.W., Armstrong, F.P. and Ward, D.C. 2003. Chemical extraction methods to assess bioavailable arsenic in soil and solid media. Journal of Environmental Quality 32(3): 876-884.
- Rufyikin, G., Wannijn, J., Wang, L. and Thiry, Y. 2006. Effects of phosphorus fertilization on the availability and uptake of uranium and nutrients by plants grown on soil derived from uranium mining debris. Environmental Pollution 141(3): 420-427.
- Seidel, H., Gorsch, K., Amstatter, K. and Mattusch, J. 2005. Immobilization of arsenic in a tailings material by ferrous iron treatment. Water Research 39: 4073-4082.
- Signes-Pastor, A., Burlo, F., Mitra, K. and Carbonell-Barrachina, A.A. 2007. Arsenic biogeochemistry as affected by phosphorus fertilizer addition, redox potential and pH in a west Bengal (India) soil. Geoderma 137: 504-510.
- Sikora, F.J., Howe, P.S., Hill, L.E., Reid, D.C. and Harover, D.E. 2005. Comparison of colorimetric and ICP determination of phosphorus in Mehlich 3 soil extracts. Communications in Soil Science and Plant Analysis 36(7-8): 875-887.
- Son, J., Shin, K.I. and Cho, K. 2009. Response surface model for predicting chronic toxicity of cadmium to *Paronychiurus kimi* (Collembola), with a special emphasis on the importance of soil characteristics in the reproduction test. Chemosphere 77: 889-894.
- Tordoff, G.M., Baker, A.J.M. and Willis, A.J. 2000. Current approaches to the revegetation and reclamation of metalliferous mine wastes. Chemosphere 41: 219-228.
- Tsang, S., Phu, F., Baum, M.M. and Poskreby, G.A. 2007. Determination of phosphorus/arsenate by a modified molybdenum blue method and reduction of arsenate by $S_2O_4^{2^2}$. Talanta 71: 1560-1568.
- Ullrich-Eberius, C.I., Sanz, A. and Novacky, A.J. 1989. Evaluation of arsenate-and vanadate-associated changes of electrical membrane potential and phosphate transport in Lemna gibba G1. Journal of Experimental Botany 40(210): 119-128.
- Warren, G.P. and Alloway, B.J. 2003. Reduction of arsenic

uptake by lettuce with ferrous sulfate applied to contaminated soil. Journal of Environmental Quality 32: 767-772.

Woolson, E.A., Axley, J.H. and Kearney, P.C. 1973. The chemistry and phytotoxicity of arsenic in soils: II. Effects of time and phosphorus. Soil Science Society of America Journal 37(2): 254-259.

Yan, X.L., Lin, L.Y., Liao, X.Y., Zhang, W.B. and Wen, Y. 2013. Arsenic stabilization by zero-valent iron, bauxite residue, and zeolite at a contaminated site planting *Panax notoginseng*. Chemopshere 93(4): 661-667.