Tumor Immunology and Immune Checkpoint Inhibitors in Non-Small Cell Lung Cancer



Chi Young Jung, M.D., Ph.D.¹ and Scott J. Antonia, M.D., Ph.D.²

¹Division of Pulmonary and Critical Care Medicine, Department of Internal Medicine, Daegu Catholic University School of Medicine, Daegu, Korea, ²Department of Thoracic Oncology, H. Lee Moffitt Cancer Center and Research Institute, Tampa, FL, USA

Lung cancer is one of the most commonly diagnosed cancers and the leading cause of cancer-related deaths worldwide. Although progress in the treatment of advanced non-small cell lung cancer (NSCLC) has been made over the past decade, the 5-year survival rate in patients with lung cancer remains only 10%–20%. Obviously, new therapeutic options are required for patients with advanced NSCLC and unmet medical needs. Cancer immunotherapy is an evolving treatment modality that uses a patient's own immune systems to fight cancer. Theoretically, cancer immunotherapy can result in long-term cancer remission and may not cause the same side effects as chemotherapy and radiation. Immuno-oncology has become an important focus of basic research as well as clinical trials for the treatment of NSCLC. Immune checkpoint inhibitors are the most promising approach for cancer immunotherapy and they have become the standard of care for patients with advanced NSCLC. This review summarizes basic tumor immunology and the relevant clinical data on immunotherapeutic approaches, especially immune checkpoint inhibitors in NSCLC.

Keywords: Carcinoma, Non-Small-Cell Lung; Immunotherapy; Cell Cycle Checkpoints

Introduction

Lung cancer is one of the most commonly diagnosed cancers and the leading cause of cancer-related deaths world-wide¹. In Korea, lung cancer was the fourth most common cancer in 2013, with 23,177 newly diagnosed patients, and the most common cause of cancer-related deaths, accounting for 22.8% of all cancer-related deaths in 2014²³. Non-small cell lung cancer (NSCLC) is diagnosed in approximately 85% of patients with lung cancer, but most of these patients are diag-

Address for correspondence: Scott J. Antonia, M.D., Ph.D.

Department of Thoracic Oncology, H. Lee Moffitt Cancer Center and Research Institute, 12902 Magnolia Dr., MRC 3-E, Tampa, FL 33612, USA Phone: 1-813-745-8470, Fax: 1-813-745-6847 E-mail: scott.antonia@moffitt.org Received: Nov. 1, 2017 Revised: Nov. 13, 2017 Accepted: Nov. 26, 2017

©It is identical to the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/4.0/).



Copyright © 2018

The Korean Academy of Tuberculosis and Respiratory Diseases. All rights reserved. nosed at an inoperable or advanced stage⁴.

Progress in the treatment of advanced NSCLC over the past decade includes the introduction of the cytotoxic chemotherapy agent pemetrexed for tumors of non-squamous histology and the development of molecularly targeted agents, including epidermal growth factor receptor–tyrosine kinase inhibitors (EGFR-TKI) and anaplastic lymphoma kinase (ALK) inhibitors, for tumors with activating mutations, most of adenocarcinoma histology⁴⁵. These agents have resulted in better survival outcomes^{4,5}. The frequency of *EGFR* mutations in tumors of adenocarcinoma is highest in patients from Asia-Pacific countries, at 47% (range, 20%–76%)⁶. Moreover, the incidence of adenocarcinoma is steadily increasing, making it the most common subtype of lung cancer in Korea⁷.

These advanced treatments, however, are unavailable to patients with squamous cell histology and those without appropriate molecular alterations, making their prognosis as poor as ever. Moreover, acquired resistance to targeted agents is now a challenging problem in patients who progress on these therapies^{4.5}. Therefore, the 5-year survival rate in patients with lung cancer ranges remains only 10%–20% in both developed and developing countries, despite improving up to 10% in most countries⁸. Obviously, new therapeutic options are required for patients with advanced NSCLC and unmet medical needs. Immunotherapy, which uses a patient's own immune system, has recently appeared as another modality for cancer treatment. Immuno-oncology has become an important focus of basic research and clinical trials for the treatment of NSCLC⁹⁻¹⁵. This review summarizes basic tumor immunology and clinical data on immunotherapeutic approaches, especially immune checkpoint inhibitors in NSCLC.

Immune System and Tumor Immunology

Historically, immunity has signified a defense mechanism against infectious diseases. However, noninfectious foreign substances can also elicit immune responses. Therefore, the immune system reacts not only to infectious microbes but to cancer cells, and has the potential to kill cancer cells¹⁶.

The immune system consists of the innate immune system, which reacts initially to foreign substances, and the adaptive immune system, which responds subsequently. The innate immune system includes complement proteins and cellular components, including natural killer cells (NKs), dendritic cells (DCs), polymorphonuclear leukocytes, mast cells, and macrophages. The adaptive immune system includes humoral immunity mediated by antibodies produced by B lymphocytes, and cellular immunity mediated by T lymphocytes¹⁶⁻¹⁸. Natural killer T (NKT) cells and $\gamma\delta$ T cells are involved in both

innate and adaptive immunity¹⁷. The innate immune system is ready to respond rapidly, in the absence of prior exposure, and is antigen-nonspecific. By contrast, the adaptive immune response is slower to develop, educated to recall prior exposure, known as memory, and is antigen-specific¹⁶⁻¹⁸.

Cancer immunotherapy is an evolving treatment modality that uses a patient's own immune system to fight cancer. Theoretically, cancer immunotherapy can result in long-term cancer remission and may not cause the same side effects as chemotherapy and radiation^{19,20}.

Classically, cancer immunosurveillance hypothesizes that the immune system can recognize and eliminate nascent transformed cells²¹. However, tumors that developed in immunocompetent mice were found to be less immunogenic than tumors that developed in immunodeficient mice²². These findings indicated that, paradoxically, the immune system assists in the eventual outgrowth of cancers that are better able to escape immune detection. Thus, tumors are imprinted by the immunologic environment in which they form^{22,23}.

1. Cancer immunoediting

Because the immune system can promote as well as suppress cancer growth, the broader term cancer immunoediting was proposed to describe better these activities, in place of the term cancer immunosurveillance²²⁻²⁴.

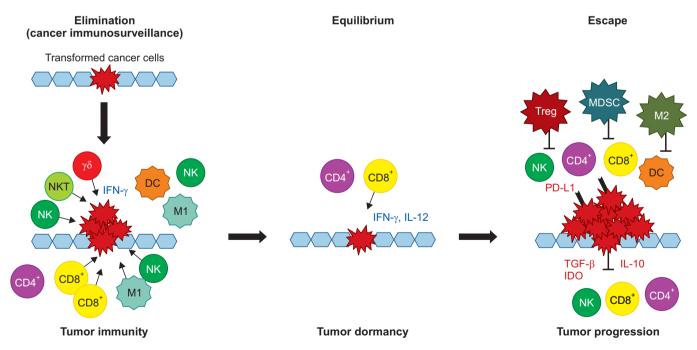


Figure 1. The three phases of the cancer immunoediting process: elimination, equilibrium, and escape. DC: dendritic cell; $\gamma\delta$: $\gamma\delta$ T cell; IDO: indoleamine 2,3-dioxygenase; IFN- γ : interferon γ ; IL: interleukin; M1: M1 macrophage; M2: M2 macrophage; MDSC: myeloid-derived suppressor cell; NK: natural killer cell; NKT: natural killer T cell; PD-L1: programmed death ligand 1; TGF- β : transforming growth factor β ; Treg: regulatory T cell. Modified from Schreiber et al. Science 2011;331:1565-70, with permission of The American Association for the Advancement of Science²⁴.

Cancer immunoediting consists of three phases of relations between cancer cells and the immune system: elimination, equilibrium, and escape (Figure 1). In the elimination phase, the immune system detects and destroys transformed cancer cells from normal tissue before they become clinically detectable tumors. This phase corresponds to the concept of cancer immunosurveillance, in which the innate and adaptive immune systems work together. A proposed model for the elimination phase consists of four steps. In the first step, innate immune cells, such as NKs, NKT cells, and γδ T cells, recognize transformed cells and produce interferon γ (IFN- γ). In the second step, IFN-y induces the death of a limited number of tumor cells and some chemokines recruit NKs, DCs, and macrophages. DCs ingest dead tumor cells and migrate to the draining lymph node. In the third step, NK cells and macrophages produce interleukin (IL)-12 and IFN-y, which kill additional tumor cells while tumor antigen-specific T cells develop in the draining lymph nodes. In the fourth step, tumor antigen-specific T cells home to the tumor site and destroy tumor cells^{23,24}

During the equilibrium phase, any cancer cell variant that has survived the elimination phase has increased resistance to immune recognition. The immune system also holds cancer cells in a functionally dormant state using adaptive immune cells such as T cells. Thus, the immune system prevents cancer cell outgrowth and sculpts the immunogenicity of these cancer cells^{23,24}.

During the escape phase, the immune system can no longer block tumor cells outgrowth, resulting in the emergence of cancer cells and the development of clinically observable malignant disease. Cancer cell escape can occur through many different mechanisms, including the loss of tumor antigens, resulting in the evasion of immune recognition, increased resistance to the cytotoxic effects of the immune system, and establishment of an immunosuppressive tumor microenvironment^{23,24}.

2. Cancer-immunity cycle

An anticancer immune response can be divided into several steps, known as the cancer-immunity cycle (Figure 2)²⁵. First, tumors release tumor cell antigens, which are captured by antigen presenting cells (APCs) such as DCs. The APCs subsequently process and present these captured antigens on major histocompatibility complex (MHC) molecules, and migrate to draining lymph nodes. In the lymph nodes, T cell receptors on the surface of T cells recognize the antigenic peptides presented by the MHC. However, T-cell activation also requires the interaction of co-stimulatory signals, such as between proteins of the B7 family (CD80 or CD86) on APCs and CD28 on T cells. Finally, activated T cells migrate to and infiltrate the tumor bed, binding to tumor cells and killing them²⁵²⁶.

Each step of the cancer-immunity cycle may be a potential target for immunotherapy. Vaccines can promote tumor antigen presentation by APCs, cytotoxic T-lymphocyte-associated antigen 4 (CTLA-4) blockade can promote the priming phase, and programmed cell death protein 1 (PD-1) blockade or programmed death ligand 1 (PD-L1) blockade can promote the effector phase, consisting of the killing of cancer cells by activated T cells^{25,26}.

3. Immunosuppressive mechanism

After T cells are activated, a checkpoint system is triggered to inhibit further T-cell activation. Originally this mechanism helps to regulate immune responses and maintain immune balance, thereby preventing autoimmune reactions. Tumors can also protect themselves from the immune system using checkpoints as immune resistance mechanisms^{9,15,27}.

Cancer cells can induce and recruit immunosuppressive cells in the tumor microenvironment. These immunosuppressive cells include regulatory T cells (Tregs), myeloid-derived suppressor cells (MDSCs), and M2 tumor-associated macrophages (TAMs)^{9,15,28}. Immunosuppressive molecules such as IL-10, transforming growth factor β , and indoleamine 2,3-dioxygenase (IDO) are also secreted by cancer cells or immunosuppressive cells^{15,28}.

4. Immune response in NSCLC

Although lung cancer has been regarded classically as a non-immunogenic malignancy, recent understanding of the immune system suggests that antitumor immune responses to lung cancer can be induced, that their magnitude may correlate with clinical outcomes, and that immunotherapy may

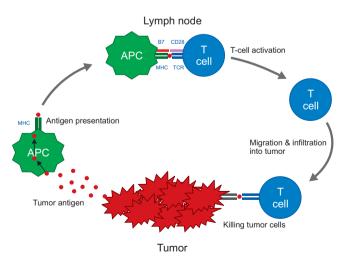


Figure 2. The cancer-immunity cycle. APC: antigen presenting cell; MHC: major histocompatibility complex; TCR: T cell receptors.

be a therapeutic option in lung cancer^{9,11,15}.

Several retrospective, immunohistochemical (IHC) analyses of lung cancer specimens have shown that cellular immune responses may be associated with clinical prognosis. For example, the survival rate of patients with squamous cell carcinoma of the lung, particularly those with early stage tumors, was found to be significantly higher in the presence than in the absence of tumor-infiltrating lymphocytes (TILs), which were almost entirely CD8⁺ T cells²⁹. High levels of infiltration of both CD8⁺ T cells and CD4⁺ T cells were also associated with significantly higher survival rates in patients with NSCLC³⁰. By contrast, a high ratio of Tregs to TILs correlated with a significantly higher risk of recurrence in patients with stage I NSCLC³¹.

Immunotherapy for NSCLC

The goal of immunotherapy is to potentiate the immune system's response to cancer cells. The adaptive immune system, especially T cells, plays an important role in anticancer immune responses¹⁵. Studies to date have focused on two immunotherapeutic strategies in NSCLC: cancer vaccines and immune checkpoint inhibitors. Cancer vaccines are antigenspecific immunotherapies that augment tumor recognition by the immune system, whereas immune checkpoint inhibitors are antigen non-specific therapies that overcome tumor immunosuppression^{9,11,12,15}.

Vaccines

Cancer vaccines stimulate the immune system to recognize tumor antigens. Injection of a cancer vaccine, composed of tumor-associated antigens (TAA) and adjuvant, into the skin (usually intradermally) results in the uptake of TAAs by APCs such as DCs. These APCs process these TAAs and migrate to draining lymph nodes, where the activated APCs present antigens to T cells, resulting in their activation^{9,15,32}.

Several cancer vaccines have been studied in NSCLC, including melanoma-associated antigen-A3 (MAGE-A3)^{33,34}, liposomal-BLP25 (tecemotide)^{35,36}, TG4010³⁷, and CIMAvax-EGF³⁸. To date, however, these cancer vaccines remain suboptimal because the induction of the desired immune response is weak, responses are short lived, and memory formation is defective. The tumor microenvironment is also potently immunosuppressive, resulting in the rapid inactivation of vaccine-induced effector lymphocytes. These hurdles must be overcome to develop clinically effective cancer vaccines^{39,40}. Consistently, however, these vaccines have been found more effective in patients with minimal disease burden. Patient selection based on predictive biomarkers will be a major future challenge in the development of cancer vaccines^{12,41,42}.

Immune Checkpoint Inhibitors

Immune checkpoint inhibitors are the most promising approach for cancer immunotherapy. Immune checkpoint inhibitors most investigated in clinical trials of patients with NSCLC include antibodies against CTLA-4, PD-1, and PD-L1. A major advantage of these immune checkpoint inhibitors is their ability to elicit antitumor immune responses regardless of the specific tumor antigens^{12-15,27}.

CTLA-4 is expressed on T cells and initiates inhibitory regulation during the priming phase of a T-cell activation. CTLA-4 competes with T-cell co-stimulatory receptor CD28, which is needed for T-cell activation, by binding to members of the B7 family (CD80 or CD86) on APCs. Binding of CTLA-4 to CD80 or CD86, rather than to CD28, provides an inhibitory signal to the T cell. By contrast, PD-1 is expressed on activated T cells and regulates the effector phase of T-cell responses in the tumor microenvironment. PD-1 binds to one of its ligands, PD-L1 or PD-L2, which is usually expressed on cancer cells, providing an inhibitory signal to the T cell. Therefore, inhibition of CTLA-4, PD-1, or PD-L1 results in the activation of T cells and enhancement of anticancer immune responses (Fig-(12.15,27) ure 3)^{12-15,27}. Immune checkpoint inhibitors for the treatment of NSCLC and the results of clinical trials are summarized in Tables 1 and 2.

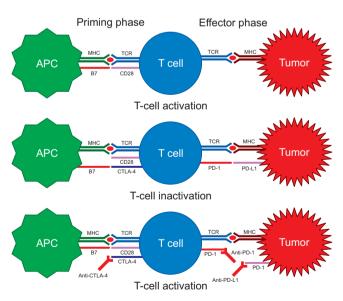


Figure 3. The immune system activation and checkpoint inhibitors. APC: antigen presenting cell; CTLA-4: cytotoxic T-lymphocyte-associated antigen 4; PD-1: programmed cell death protein 1; PD-L1: programmed death ligand 1; TCR: T cell receptors. Modified from Pardoll. Nat Rev Cancer 2012;12:252-64, with permission of Springer Nature²⁷.

MedImmune/Astrazeneca

Table 1. Infinitune che	exponet minorers for the treatment of non-sman centurg cancer	
Agent	Description	Company
Anti-PD-1		
Nivolumab	Fully human IgG4 monoclonal antibody directed against PD-1 on T cells	Bristol-Myers Squibb
Pembrolizumab	Humanized IgG4 monoclonal antibody directed against PD-1 on T cells	Merck
Anti-PD-L1		
Atezolizumab	Human IgG1 monoclonal antibody directed against PD-L1 on tumor cells	Genetech/Roche
Durvalumab	Fully human IgG1 monoclonal antibody directed against PD-L1 on tumor cells	Astrazeneca
Anti-CTLA-4		
Ipilimumab	Fully human IgG1 monoclonal antibody directed against CTLA-4 on T cells	Bristol-Myers Squibb

Table 1. Immune checkpoint inhibitors for the treatment of non-small cell lung cancer

Fully human IgG2 monoclonal antibody directed against CTLA-4 on T cells PD-1: programmed cell death protein 1; PD-L1: programmed death ligand 1; CTLA-4: cytotoxic T-lymphocyte-associated antigen 4.

1. Anti-PD-1

1) Nivolumab

Tremelimumab

Nivolumab is the first anti-PD-1 antibody in clinical development. Two phase III trials compared nivolumab with docetaxel in patients with stage IIIB or IV squamous and nonsquamous NSCLC who experienced disease progression during or after one prior platinum-based chemotherapy regimen^{43,44}. In the first phase III trial, CheckMate 017, 272 patients with squamous NSCLC were randomly assigned to receive nivolumab (3 mg/kg every 2 weeks) or docetaxel (75 mg/m² every 3 weeks). Median overall survival (OS, 9.2 months vs. 6.0 months; hazard ratio [HR], 0.59; 95% confidence interval [CI], 0.44-0.79; p<0.001) and median progression-free survival (PFS, 3.5 months vs. 2.8 months; HR, 0.62; 95% CI, 0.47-0.81; p<0.001) were significantly longer in the nivolumab than in the docetaxel group. In addition, the 1-year OS rate (42% vs. 24%) and the objective response rate (ORR, 20% vs. 9%; p=0.008) were higher in patients treated with nivolumab than with docetaxel. Across the prespecified expression levels (1%, 5%, and 10%), the expression of the ligand for PD-1 (PD-L1) was neither prognostic nor predictive of any of the efficacy end points. Treatment-related adverse events (AEs) of grades 3 and 4 occurred less frequently with nivolumab than with docetaxel $(7\% \text{ vs. } 55\%)^{43}$.

In the second phase III trial, CheckMate 057, 582 patients with non-squamous NSCLC were randomly assigned to receive nivolumab or docetaxel, in a design similar to that of the CheckMate 017 trial. Patients with known EGFR mutation or ALK translocation were allowed to receive additional TKI therapy. Nivolumab was associated with significantly longer median OS (OS, 12.2 months vs. 9.4 months; HR, 0.73; 95% CI, 0.59–0.89; p=0.002) and higher ORR (19% vs. 12%, p=0.02). Although median PFS was shorter in the nivolumab than in the docetaxel group (median PFS, 2.3 months vs. 4.2 months), the difference was not statistically significant (HR, 0.92; 95% CI, 0.77-1.11; p<0.39). In contrast to the CheckMate 017 trial, involving patients with squamous cell NSCLC, the CheckMate 057 trial found that PD-L1 protein expression was associated with improved OS and PFS in patients treated with nivolumab. However, PD-L1 protein expression was evaluated retrospectively in prospectively collected, pretreatment (archival or recent) tumor-biopsy specimens. Grade 3 and 4 treatmentrelated AEs were again less frequent in patients treated with nivolumab than with docetaxel $(10\% \text{ vs. } 54\%)^{44}$.

The phase III CheckMate 026 trial compared nivolumab with platinum-based agents as first-line therapy in patients with stage IV or recurrent NSCLC. Among the 423 patients with a PD-L1 expression level $\geq 5\%$, nivolumab did not improve median PFS compared with chemotherapy (PFS, 4.2 months vs. 5.9 months; HR, 1.15; 95% CI, 0.91–1.45; p=0.25). Median OS (OS, 14.4 months vs. 13.2 months; HR,1.02; 95% CI, 0.80-1.30) and ORR (26% vs. 33%) were similar in the nivolumab and chemotherapy groups. Rates of grades 3 and 4 treatment-related AEs were lower with nivolumab than with chemotherapy $(18\% \text{ vs. } 51\%)^{45}$.

2) Pembrolizumab

The phase I KEYNOTE-001 trial found that higher PD-L1 expression in at least 50% of tumor cells (proportion score $[PS] \ge 50\%$) was associated with higher ORR and longer PFS and OS than lower levels (PS <1% and 1%-49%) in patients with advanced NSCLC⁴⁶. In the subsequent phase II/III KEYNOTE-010 trial, 1,034 patients with previously treated NSCLC and PD-L1 expression (PS $\ge 1\%$) were randomly assigned to receive pembrolizumab (2 mg/kg or 10 mg/kg) or docetaxel (75 mg/m²) every 3 weeks. In the total population (PS $\geq 1\%$), median OS was significantly longer with pembrolizumab 2 mg/kg (10.4 months vs. 8.5 months; HR, 0.71; 95% CI, 0.58–0.88; p=0.0008) and with pembrolizumab 10 mg/ kg (12.7 months vs. 8.5 months; HR, 0.61; 95% CI, 0.49-0.75; p<0.0001) than with docetaxel. Among patients with high PD-L1 expression (PS \geq 50%), median OS was significantly longer with pembrolizumab 2 mg/kg (14.9 months vs. 8.2 months;

Table 2. Selected clinical trials of immune checkpoint inhibitors for treatment of non-small cell lung cancer	inical tri	als of immune ch	seckpoint inhibite	ors for	treatme	ant of nor	1-small cell lung (cancer				
Study	Phase	Histology	Treatment arms	No.	ORR (%)	Median PFS (mo)	HR (95% CI)	Median OS (mo)	HR (95% CI)	1-Year OS (%)	Median DoR (mo)	TRAEs grade ≥3 (%)
Subsequent treatment	ıt											
CheckMate 017 ⁴³	III	Squamous	Nivolumab	135	20	3.5	$0.62\left(0.47 - 0.81 ight)$	9.2	0.59(0.44 - 0.79)	42	NR	2
			Docetaxel	137	6	2.8	p<0.001	6.0	p<0.001	24	8.4	55
CheckMate 057 ⁴⁴	III	Non-squamous	Nivolumab	287	19	2.3	$0.92\left(0.77{-}1.11 ight)$	12.2	0.73(0.59 - 0.89)	51	17.2	10
			Docetaxel	268	12	4.2	p<0.39	9.4	p=0.002	39	5.6	54
KEYNOTE-010 ⁴⁷	III/III	NSCLC	Pembrolizumab	344	18	3.9	$0.88\left(0.74 {-}1.05 ight)$	10.4	$0.71\ (0.58-0.88)$	43.2	NR	13
		PD-L1 ≥1%	$(2 \mathrm{mg/kg})$				p=0.07		p=0.0008			
			Pembrolizumab	346	18	4.0	0.79(0.66-0.94)	12.7	0.61 (0.49 - 0.75)	52.3	NR	16
			$(10 \mathrm{mg/kg})$				p=0.004		p<0.0001			
			Docetaxel	343	6	4.0	I	8.5	I	34.6	9	35
		PD-L1 ≥50%	Pembrolizumab	139	30	5.0	0.59(0.44 - 0.78)	14.9	0.54(0.38-0.77)	I	NR	,
			$(2 \mathrm{mg/kg})$				p=0.0001		p=0.0002			
			Pembrolizumab	151	29	5.2	0.59(0.45 - 0.78)	17.3	$0.50\ (0.36-0.70)$	I	NR	I
			$(10 \mathrm{mg/kg})$				p<0.0001		p<0.0001			
			Docetaxel	152	8	4.1	I	8.2	I	ı	8	ı
POPLAR ⁵⁰	Π	NSCLC	Atezolizumab	144	14.6	2.7	0.94(0.72 - 1.23)	12.6	0.73(0.53-0.99)	I	14.3	11
			Docetaxel	143	14.7	3.0	p=0.645	9.7	p=0.04	I	7.2	39
OAK^{51}	III	NSCLC	Atezolizumab	425	14	2.8	$0.95(0.82{-}1.10)$	13.8	$0.73\ (0.62 - 0.87)$	55	16.3	15
			Docetaxel	425	13	4.0	p=0.4928	9.6	p=0.0003	41	6.2	43
		PD-L1 ≥1%	Atezolizumab	241	18	2.8	$0.91(0.74{-}1.12)$	15.7	0.74(0.58 - 0.93)	58	16.0	
			Docetaxel	222	16	4.1	p=0.38	10.3	p=0.0102	43	6.2	39
ATLANTIC ⁵²	Π	NSCLC										
		PDL1 <25%	Durvalumab	93	7.5	1.9	I	9.3	I	34.5	NR	8.3
		PD-L1 ≥25%	Durvalumab	146	16.4	3.3	I	10.9	I	47.7	12.3	
		PD-L1 ≥90%	Durvalumab	68	30.9	2.4	I	NR	I	50.8	NR	17.6
First-line treatment												
KEYNOTE-024 ^{48,49}	III	NSCLC	Pembrolizumab	154	44.8	10.3	$0.50(0.37{-}0.68)$	NR	0.63(0.46 - 0.88)	70.3	NR	26.6
		PD-L1 ≥50%	Chemotherapy	151	27.8	6.0	p<0.001	14.5	p=0.003	54.8	6.3	53.3

			Turotmont		aao	Median		Median		" Noou	Median	TRAES
Study	Phase	Histology	arms	No.	(%)	PFS (mo)	HR (95% CI)	(ou)	HR (95% CI)	1-rear OS (%)	DoR (mo)	grade ≥3 (%)
CheckMate 026 ⁴⁵	III	NSCLC	Nivolumab	211	26	4.2	1.15(0.91 - 1.45)	14.4	1.02(0.80-1.30)	56	12.1	18
		PD-L1 ≥5%	Chemotherapy	212	33	5.9	p=0.25	13.2		54	5.7	51
BIRCH (cohort1) ⁵³	Π	NSCLC	Atezolizumab	139	22	5.4	I	23.5	I	66.4	9.8	6
		PD-L1 ≥5%										
		TC: PD-L1 ≥50% Atezolizumab	Atezolizumab	65	31	5.6	I	26.9	I	61.5	10.0	ı
		IC: PD-L1 ≥10%										
Adjuvant treatment												
PACIFIC ⁵⁴	III	III NSCLC	Durvalumab	473	28.4	16.8	0.52(0.42 - 0.65)	NR	I	I	NR	29.9
			Placebo	236	16.0	5.6	p<0.001	NR	I	ı	13.8	26.1

HR. 0.54; 95% CI. 0.38-0.77; p=0.0002) and pembrolizumab 10 mg/kg (17.3 months vs. 8.2 months; HR, 0.50; 95% CI, 0.36-0.70; p<0.0001) than with docetaxel. In this latter group (PS ≥50%), median PFS was also significantly longer with pembrolizumab 2 mg/kg (5.0 months vs. 4.1 months; HR, 0.59; 95% CI, 0.44–0.78; p=0.0001) and pembrolizumab 10 mg/kg (5.2 months vs. 4.1 months; HR, 0.59; 95% CI, 0.45-0.78; p<0.0001) than with docetaxel. In the total population, however, median PFS did not differ significantly in the pembrolizumab 2 mg/ kg and 10 mg/kg and docetaxel groups (3.9 months vs. 4.0 months vs. 4.0 months). ORR was significantly higher in the total population of patients treated with pembrolizumab 2 mg/kg (18% vs. 9%, p=0.0005) and pembrolizumab 10 mg/kg (18% vs. 9%, p=0.0002) than with docetaxel. Similarly, ORR in patients with high PD-L1 expression (PS \geq 50%) was significantly higher following treatment with pembrolizumab 2 mg/ kg (30% vs. 8%, p<0.0001) and pembrolizumab 10 mg/kg (29% vs. 8%, p<0.0001) than with docetaxel. Treatment-related AEs of grades 3 to 5 were less frequent in both pembrolizumab groups than in the docectaxel group $(13\% \text{ vs. } 16\% \text{ vs. } 35\%)^{47}$.

The phase III KEYNOTE-024 trial compared pembrolizumab (200 mg every 3 weeks) with platinum-based chemotherapy as first-line treatment in 305 patients with stage IV NSCLC and high PD-L1 expression (PS \geq 50%). Median PFS (10.3 months vs. 6.0 months; HR, 0.50; 95% CI, 0.37–0.68; p<0.001)⁴⁸ and median OS (not reached vs. 14.5 months; HR, 0.63; 95% CI, 0.46–0.88; p=0.003)⁴⁹ were significantly longer with pembrolizumab than with chemotherapy. ORR was higher (44.8% vs. 27.8%) and treatment-related AEs of grades 3 to 5 were less frequent (26.6% vs. 53.3%) with pembrolizumab than with chemotherapy⁴⁸.

2. Anti-PD-L1

1) Atezolizumab

In the phase II POPLAR trial, 287 patients with NSCLC who progressed on post-platinum chemotherapy were randomly assigned to receive atezolizumab 1,200 mg or docetaxel 75 mg/m^2 every 3 weeks. In this study, PD-L1 expression was prospectively determined by IHC in tumor cells (TCs) and tumor-infiltrating immune cells (ICs). Median OS was significantly longer in patients treated with atezolizumab than with docetaxel (12.6 months vs. 9.7 months; HR, 0.73; 95% CI, 0.53–0.99; p=0.04). The OS benefit from atezolizumab increased with increasing PD-L1 expression on TCs and/or ICs, but OS was similar with atezolizumab and doectaxel in patients without PD-L1 expression. Median PFS was similar in patients treated with atezolizumab and docetaxel (2.7 months vs. 3.0 months; HR, 0.94; 95% CI, 0.72-1.23; p=0.645). ORR was similar in these two groups (14.6% vs. 14.7%), but the median duration of response was longer with atezolizumab than with docetaxel (14.3 months vs. 7.2 months; HR, 0.41; 95% CI, 0.18–0.96; p=0.034). Treatment-related AEs of grades 3 and 4

CY Jung et al.

were less frequent in patients treated with a tezolizumab than with docetaxel (11% vs. 39%)⁵⁰.

In the phase III OAK trial, 850 patients with stage IIIB or IV NSCLC who had received 1-2 previous lines of cytotoxic chemotherapy, including at least one platinum-based combination regimen, were randomly assigned to receive atezolizumab 1,200 mg or docetaxel 75 mg/m² every 3 weeks. Median OS was significantly longer with atezolizumab than with docetaxel in the intention-to-treat (ITT) (13.8 months vs. 9.6 months; HR, 0.73; 95% CI, 0.62–0.87; p=0.0003) and the TC1/2/3 or IC1/2/3 (PD-L1 expression \geq 1% on TCs or ICs) (15.7 months vs. 10.3 months; HR, 0.74; 95% CI, 0.58-0.93; p=0.0102) populations. The OS benefit from atezolizumab was the highest in patients with high levels of PD-L1, TC3 or IC3 (PD-L1 expression \geq 50% on TCs or \geq 10% on ICs) (20.5 months vs. 8.9 months; HR, 0.41; 95% CI, 0.27-0.64; p<0.0001). However, OS was also better with atezolizumab than with docetaxel in patients with TC0 and IC0 (PD-L1 expression <1% on TCs and ICs). Median PFS was similar in patients treated with atezolizumab and docetaxel in the ITT population (2.8 months vs. 4.0 months; HR, 0.95; 95% CI, 0.82-1.10; p=0.4928) and in TC1/2/3 or IC1/2/3 populations. However, median PFS was significantly longer with atezolizumab than with docetaxel in the TC3 or IC3 population (4.2 months vs. 3.3 months; HR, 0.63; 95% CI, 0.43-0.91; p=0.0123). Although ORR was similar in the two groups in the ITT (14% vs. 13%) and TC1/2/3 or IC1/2/3 populations, ORR was higher with atezolizumab than with docetaxel in patients with TC3 or IC3 (31% vs. 11%). Rates of grades 3 and 4 treatment-related AEs were lower in patients treated with atezolizumab than with docetaxel $(15\% \text{ vs. } 43\%)^{51}$.

The phase II, single-arm trial BIRCH study included 667 patients with stage IIIB or IV NSCLC and PD-L1 expression $\geq 5\%$ on TCs or ICs (TC2/3 or IC2/3) who received atezolizumab as first-line, second-line, and third-line or higher treatment. ORR in the 139 patients treated with first-line atezolizumab was 22% (31% in those with TC3 or IC3). Median OS was 23.5 months (26.9 months in those with TC3 or IC3) and the 12-month OS rate was 66.4% (61.5% in patients with TC3 or IC3). Median PFS was 5.4 months (5.6 months in those with TC3 or IC3). Grade 3 and 4 treatment-related AEs occurred in 9% of these patients⁵³.

2) Durvalumab

In the phase II, single-arm ATLANTIC study, 333 patients with stage IIIB or IV and *EGFR/ALK* wild-type NSCLC who had received at least two prior lines of chemotherapy were treated with durvalumab 10 mg/kg every 2 weeks. ORRs in patients with PD-L1 expression \geq 25% and \geq 90% were 16.4% and 30.9%, respectively. Median OS was 10.9 months in patients with PD-L1 \geq 25% and was not reached in patients with PD-L1 \geq 90%. One-year OS rates in patients with PD-L1 expression \geq 25% and \geq 90% were 47.7% and 50.8%, respectively⁵².

In the phase III PACIFIC trial, 709 patients with stage III NSCLC who had not progressed after platinum-based chemoradiotherapy were randomly assigned 2:1 to receive durvalumab 10 mg/kg or placebo every 2 weeks for up to 12 months. Median PFS was significantly longer with durvalumab than with placebo (16.8 months vs. 5.6 months; HR, 0.52; 95% CI, 0.42–0.65; p<0.001) and was independent of PD-L1 expression. ORR was significantly higher with durvalumab than with placebo (28.4% vs. 16.0%, p<0.001), whereas rates of grades 3 and 4 treatment-related AEs were similar in the two groups (29.9% vs. 26.1%)⁵⁴.

3. Anti-CTLA-4

1) Ipilimumab

Ipilimumab has been combined with cytotoxic chemotherapy or another immunotherapy agent in patients with NSCLC. In a phase II trial, 204 patients with stage IIIB or IV NSCLC were randomly assigned 1:1:1 to receive firstline paclitaxel and carboplatin with two different schedules (concurrent or phased arm) of ipilimumab or placebo. The concurrent arm consisted of four doses of ipilimumab plus chemotherapy followed by two doses of placebo plus chemotherapy. The phased arm consisted of two doses of placebo plus chemotherapy followed by four doses of ipilimumab plus chemotherapy. The median immune-related PFS (irPFS) was longer with phased treatment than with placebo (5.7 months vs. 4.6 months; HR, 0.72; 95% CI, 0.50–1.06; p=0.05), whereas concurrent treatment did not improve irPFS⁵⁵.

By contrast, a subsequent phase III trial did not show that the combination of ipilimumab plus first-line chemotherapy enhanced median OS compared with chemotherapy alone in patients with squamous NSCLC (13.4 months vs. 12.4 months; HR, 0.91; 95% CI, 0.77–1.07; p=0.25)⁵⁶.

4. Combination therapies

The combination of immune checkpoint inhibitors and conventional therapy with different mechanisms of action may have a synergistic effect and result in clinical benefit. Conventional therapies, especially targeted therapy, can lead to a rapid initial response, but most responders will later acquire resistance and develop progressive disease. Conversely, immune checkpoint inhibitors can lead to a durable response in a relatively small percentage of responders⁵⁷. Conventional therapies can also modulate the immune system, thereby affecting immunotherapy^{57,58}.

The combination of two immune checkpoint inhibitors with distinct targets, particularly anti–PD-1/anti–PD-L1 and anti–CTLA-4, may also improve response rate and survival benefit compared with monotherapy⁵⁸. Many ongoing clinical trials are testing combination immunotherapies, and we look forward to successful results from phase III studies.

As part of the phase II KEYNOTE-021 trial, 123 patients with non-squamous, stage IIIB or IV, chemotherapy-naive NSCLC were randomly assigned to treatment with pembrolizumab plus carboplatin and pemetrexed or to carboplatin and pemetrexed alone. The ORR was significantly higher in patients who received pembrolizumab plus chemotherapy than chemotherapy alone (55% vs. 29%, p=0.0016). In the combination group, the ORR was 80% in patients with PS \geq 50%, although there was no difference between those with PS <1% and PS $\geq 1\%$ (57% vs. 54%). Median PFS was significantly longer in the combination than in the chemotherapy alone (13.0 months vs. 8.9 months; HR, 0.53; 95% CI, 0.31–0.91; p=0.010)⁵⁹. The ORR was higher and PFS was longer in patients treated with pembrolizumab plus chemotherapy in this trial than in those treated with pembrolizumab alone in the KEYNOTE-024 trial. Median OS was also significantly longer in the combination group than in the chemotherapy alone group (not reached vs. 20.9 months; HR, 0.59; 95% CI, 0.34–1.05; p=0.0344)⁶⁰. Treatment-related AEs of grades 3 to 5 were reported in 39% of patients in the combination group compared with 26% in the chemotherapy alone group⁵⁹.

In the phase I, multicohort CheckMate 012 trial, patients with stage IIIB or IV chemotherapy-naive NSCLC were assigned to receive nivolumab alone or combination therapies. In one cohort, 56 patients were assigned to receive nivolumab 10 mg/kg plus three platinum-based doublet chemotherapy agents every 3 weeks or nivolumab 5 mg/kg plus paclitaxel-carboplatin. ORRs ranged from 33% to 47% irrespective of PD-L1 expression. The 2-year OS rate in patients treated with nivolumab 5 mg/kg plus paclitaxel-carboplatin was 62%. Treatment-related grades 3 and 4 AEs were reported in 45% of patients, and the discontinuation rate was 21%⁶¹.

In another cohort of the CheckMate 012 trial, 78 patients were randomly assigned to receive nivolumab 3 mg/kg every 2 weeks plus ipilimumab 1 mg/kg every 6 or 12 weeks. The ORRs in these groups were 38% and 47%, respectively. After pooling these two groups, ORR was found to correlate with PD-L1 expression, being 92% and 57% in patients with PD-L1 expression \geq 50% and \geq 1%, respectively. ORR was higher in patients treated with nivolumab plus ipilimuab in this trial than with nivolumab monotherapy in another cohort of the CheckMate 012 trial. Median PFS was longer in patients treated with ipilimumab every 12 than every 6 weeks (8.1 months vs. 3.9 months). Treatment-related grade 3 and 4 AEs in these two groups were 37% and 33%, respectively⁶².

In a phase Ib trial (study 006), 102 patients with immunotherapy-naive, locally advanced, or metastatic NSCLC were enrolled into the dose-escalation phase and received durvalumab and tremelimumab, a selective human IgG2 monoclonal antibody against CTLA-4. Based on safety and clinical activity, durvalumab 20 mg/kg every 4 weeks plus tremelimumab 1 mg/kg were chosen as the expansion phase doses. This dose cohort showed a manageable safety profile, with a 17% rate of grades 3 and 4 treatment-related AEs, and antitumor activity (ORR 23%) irrespective of PD-L1 status⁶³.

5. Predictive biomarkers for immune checkpoint inhibitors

The immune checkpoint inhibitors showed durable clinical responses and enhanced long-term survival in patients who benefited from these agents. However, clinical trials of unselected patients with NSCLC showed that these agents had low ORRs, of approximately 20%, as monotherapy⁶⁴. Moreover, these agents have immune-related AEs, which may be life-threatening, and their costs might result in financial burdens on patients. Therefore, biomarkers predicting the benefits of these agents are required^{64,65}.

1) PD-L1 expression

PD-L1 expression, as determined by IHC, is the most commonly used predictive biomarker for anti-PD-1 and anti-PD-L1 agents in clinical trials, with tests currently available in clinical practice⁶⁴⁻⁶⁶. PD-L1 expression on TCs has been associated with immunosuppression, inhibiting the antitumor activity of T cells, as PD-L1 binding to PD-1 on activated T cells inhibits T-cell signaling and blocks antitumor immune responses²⁷. PD-L1 expression alone may not be representative of the entire tumor microenvironment, as other concurrent immunosuppressive mechanisms, involving Treg, MDSC, TAM, and IDO, can be present^{64,65}. Higher ORRs have been observed in patients positive than negative for PD-L1, although up to 17% of PD-L1 negative patients responded^{43,65-68}. Moreover, about 50% of patients with high PD-L1 expression did not respond^{48,65}. High PD-L1 expression has also been associated with longer PFS and OS, although the results varied among clinical trials⁶⁴⁻⁶⁸. Taken together, these findings show that PD-L1 IHC is insufficient as a predictive biomarker and has limitations. For example, multiple antibodies have been utilized in IHC assays and the interpretation of their results has not been standardized. Other limitations include tumor heterogeneity among different sections of the same sample or at different tumor sites, and dynamic changes in PD-L1 expression over time⁶⁸. Several studies have compared different antibodies and IHC systems for PD-L1 tests. The Blueprint PD-L1 IHC assay comparison project revealed high concordance among the 28-8, 22C3, and SP263 assays, whereas fewer TCs were stained with the SP142 assay⁶⁹. Another study found that the 28-8 and E1L3N assays were comparable, whereas fewer cells were stained using the 22C3 assay and the SP142 assay detected significantly lower PD-L1 expression in TCs⁷⁰. High similarity among the different IHC assays suggests their potential interchangeability in clinical practice⁷¹. The U.S. Food and Drug Administration (FDA) has approved PD-L1 IHC 22C3 pharmDx as a companion diagnostic assay for pembrolizumab treatment in patients with NSCLC⁷².

2) Alternative biomarkers

Clinical trials have shown that melanomas and NSCLCs respond better to immune checkpoint inhibitors than other tumor types⁶⁸. One study showed that melanomas and NSCLCs have the highest somatic mutation burden among tumor types⁷³, suggesting that tumor mutation burden may be a predictive biomarker for treatment with immune checkpoint inhibitors. In patients with NSCLC who were treated with pembrolizumab, a higher nonsynonymous mutation burden was associated with higher ORR, longer PFS, and more durable clinical benefits⁷⁴.

Many ongoing studies are attempting to identify new predictive biomarkers, including TILs, immune gene signatures, and multiplex IHC assays⁶⁷. Understanding the tumor microenvironment and characterizing TILs and concurrent immunosuppressive mechanisms are therefore important.

Conclusion

Better understanding of the immune system in the tumor microenvironment has resulted in the development of new immunotherapy agents such as immune checkpoint inhibitors. Clinician familiarity with tumor immunology can help in understanding the process of immunotherapy and developing optimal treatment strategies for patients.

Immune checkpoint inhibitors, especially anti–PD-1 and anti–PD-L1, have become the standard of care in patients with advanced NSCLC. Nivolumab, pembrolizumab, and atezolizumab are safer and more effective than docetaxel. Pembrolizumab should be considered a first-line treatment in NSCLC with PD-L1 expression \geq 50%, but without *EGFR* mutation and *ALK* rearrangement. Durvalumab may be an effective adjuvant treatment in stage III NSCLC after chemoradiotherapy.

Despite durable long-term survival, immune checkpoint inhibitors alone have a relatively low response rate. Therefore, combining immune checkpoint inhibitor with other immunotherapy or conventional therapy agents may improve response rates and provide clinical benefits to more patients. Predictive biomarkers are also essential in selecting patients who would benefit from treatment, reducing the unnecessary cost burden to patients who would not benefit from these agents. Although IHC assays of PD-L1 expression have drawbacks, they are the only currently available tests in clinical practice. Ultimately, biomarker-driven combination therapy will become a standard strategy in future immunotherapy.

Conflicts of Interest

No potential conflict of interest relevant to this article was reported

References

- 1. Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. Int J Cancer 2015;136:E359-86.
- 2. Korea Central Cancer Registry. Annual report of cancer statistics in Korea in 2013 [Internet]. Goyang: National Cancer Information Center; 2016 [cited 2016 Jun 20]. Available from: http://www.cancer.go.kr/.
- 3. Korean Statistical Information Service. Statistics Korea [Internet]. Daejeon: Statistics Korea; 2016 [cited 2016 Jun 20]. Available from: http://kosis.kr/.
- 4. Reck M, Heigener DF, Mok T, Soria JC, Rabe KF. Management of non-small-cell lung cancer: recent developments. Lancet 2013;382:709-19.
- National Comprehensive Cancer Network. NCCN Clinical practice guidelines in oncology: non-small cell lung cancer. Version 9. 2017 [Internet]. Fort Washington: National Comprehensive Cancer Network; 2017 [cited 2017 Sep 28]. Available from: https://www.nccn.org/professionals/physician_ gls/pdf/nscl.pdf.
- Midha A, Dearden S, McCormack R. *EGFR* mutation incidence in non-small-cell lung cancer of adenocarcinoma histology: a systematic review and global map by ethnicity (mutMapII). Am J Cancer Res 2015;5:2892-911.
- 7. Park JY, Jang SH. Epidemiology of lung cancer in Korea: recent trends. Tuberc Respir Dis 2016;79:58-69.
- Allemani C, Weir HK, Carreira H, Harewood R, Spika D, Wang XS, et al. Global surveillance of cancer survival 1995-2009: analysis of individual data for 25,676,887 patients from 279 population-based registries in 67 countries (CONCORD-2). Lancet 2015;385:977-1010.
- 9. Shepherd FA, Douillard JY, Blumenschein GR Jr. Immunotherapy for non-small cell lung cancer: novel approaches to improve patient outcome. J Thorac Oncol 2011;6:1763-73.
- Thomas A, Hassan R. Immunotherapies for non-small-cell lung cancer and mesothelioma. Lancet Oncol 2012;13:e301-10.
- 11. Reck M. What future opportunities may immuno-oncology provide for improving the treatment of patients with lung cancer? Ann Oncol 2012;23 Suppl 8:viii28-34.
- 12. Brahmer JR. Harnessing the immune system for the treatment of non-small-cell lung cancer. J Clin Oncol 2013;31:1021-8.
- 13. Massarelli E, Papadimitrakopoulou V, Welsh J, Tang C, Tsao AS. Immunotherapy in lung cancer. Transl Lung Cancer Res 2014;3:53-63.
- 14. Anagnostou VK, Brahmer JR. Cancer immunotherapy: a future paradigm shift in the treatment of non-small cell lung cancer. Clin Cancer Res 2015;21:976-84.
- Carbone DP, Gandara DR, Antonia SJ, Zielinski C, Paz-Ares L. Non-small-cell lung cancer: role of the immune system and potential for immunotherapy. J Thorac Oncol 2015;10:974-84.

- 16. Abbas AK, Lichtman AH, Pillai S. Cellular and molecular immunology. 9th ed. Philadelphia: Saunders/Elsevier; 2018.
- Liu Y, Zeng G. Cancer and innate immune system interactions: translational potentials for cancer immunotherapy. J Immunother 2012;35:299-308.
- Vesely MD, Kershaw MH, Schreiber RD, Smyth MJ. Natural innate and adaptive immunity to cancer. Annu Rev Immunol 2011;29:235-71.
- 19. Finn OJ. Immuno-oncology: understanding the function and dysfunction of the immune system in cancer. Ann Oncol 2012;23 Suppl 8:viii6-9.
- 20. Eggermont AM. Can immuno-oncology offer a truly pantumour approach to therapy? Ann Oncol 2012;23 Suppl 8:viii53-7.
- 21. Burnet M. Cancer: a biological approach. I. The processes of control. Br Med J 1957;1:779-86.
- 22. Shankaran V, Ikeda H, Bruce AT, White JM, Swanson PE, Old LJ, et al. IFNgamma and lymphocytes prevent primary tumour development and shape tumour immunogenicity. Nature 2001;410:1107-11.
- 23. Dunn GP, Bruce AT, Ikeda H, Old LJ, Schreiber RD. Cancer immunoediting: from immunosurveillance to tumor escape. Nat Immunol 2002;3:991-8.
- 24. Schreiber RD, Old LJ, Smyth MJ. Cancer immunoediting: integrating immunity's roles in cancer suppression and promotion. Science 2011;331:1565-70.
- 25. Chen DS, Mellman I. Oncology meets immunology: the cancer-immunity cycle. Immunity 2013;39:1-10.
- 26. Mellman I, Coukos G, Dranoff G. Cancer immunotherapy comes of age. Nature 2011;480:480-9.
- 27. Pardoll DM. The blockade of immune checkpoints in cancer immunotherapy. Nat Rev Cancer 2012;12:252-64.
- 28. Butt AQ, Mills KH. Immunosuppressive networks and checkpoints controlling antitumor immunity and their blockade in the development of cancer immunotherapeutics and vaccines. Oncogene 2014;33:4623-31.
- 29. Ruffini E, Asioli S, Filosso PL, Lyberis P, Bruna MC, Macri L, et al. Clinical significance of tumor-infiltrating lymphocytes in lung neoplasms. Ann Thorac Surg 2009;87:365-71.
- Hiraoka K, Miyamoto M, Cho Y, Suzuoki M, Oshikiri T, Nakakubo Y, et al. Concurrent infiltration by CD8+ T cells and CD4+ T cells is a favourable prognostic factor in non-smallcell lung carcinoma. Br J Cancer 2006;94:275-80.
- 31. Petersen RP, Campa MJ, Sperlazza J, Conlon D, Joshi MB, Harpole DH Jr, et al. Tumor infiltrating Foxp3+ regulatory T-cells are associated with recurrence in pathologic stage I NSCLC patients. Cancer 2006;107:2866-72.
- 32. Drake CG, Lipson EJ, Brahmer JR. Breathing new life into immunotherapy: review of melanoma, lung and kidney cancer. Nat Rev Clin Oncol 2014;11:24-37.
- 33. Vansteenkiste J, Zielinski M, Linder A, Dahabreh J, Gonzalez EE, Malinowski W, et al. Adjuvant MAGE-A3 immunotherapy in resected non-small-cell lung cancer: phase II randomized

study results. J Clin Oncol 2013;31:2396-403.

- 34. Vansteenkiste JF, Cho BC, Vanakesa T, De Pas T, Zielinski M, Kim MS, et al. Efficacy of the MAGE-A3 cancer immunotherapeutic as adjuvant therapy in patients with resected MAGE-A3-positive non-small-cell lung cancer (MAGRIT): a randomised, double-blind, placebo-controlled, phase 3 trial. Lancet Oncol 2016;17:822-35.
- 35. Butts C, Socinski MA, Mitchell PL, Thatcher N, Havel L, Krzakowski M, et al. Tecemotide (L-BLP25) versus placebo after chemoradiotherapy for stage III non-small-cell lung cancer (START): a randomised, double-blind, phase 3 trial. Lancet Oncol 2014;15:59-68.
- 36. Wu YL, Park K, Soo RA, Sun Y, Tyroller K, Wages D, et al. IN-SPIRE: a phase III study of the BLP25 liposome vaccine (L-BLP25) in Asian patients with unresectable stage III nonsmall cell lung cancer. BMC Cancer 2011;11:430.
- 37. Quoix E, Lena H, Losonczy G, Forget F, Chouaid C, Papai Z, et al. TG4010 immunotherapy and first-line chemotherapy for advanced non-small-cell lung cancer (TIME): results from the phase 2b part of a randomised, double-blind, placebo-controlled, phase 2b/3 trial. Lancet Oncol 2016;17:212-23.
- 38. Rodriguez PC, Popa X, Martinez O, Mendoza S, Santiesteban E, Crespo T, et al. A phase III clinical trial of the epidermal growth factor vaccine CIMAvax-EGF as switch maintenance therapy in advanced non-small cell lung cancer patients. Clin Cancer Res 2016;22:3782-90.
- 39. Sasada T, Komatsu N, Suekane S, Yamada A, Noguchi M, Itoh K. Overcoming the hurdles of randomised clinical trials of therapeutic cancer vaccines. Eur J Cancer 2010;46:1514-9.
- 40. Mellstedt H, Vansteenkiste J, Thatcher N. Vaccines for the treatment of non-small cell lung cancer: investigational approaches and clinical experience. Lung Cancer 2011;73:11-7.
- 41. Cuppens K, Vansteenkiste J. Vaccination therapy for nonsmall-cell lung cancer. Curr Opin Oncol 2014;26:165-70.
- 42. Makkouk A, Weiner GJ. Cancer immunotherapy and breaking immune tolerance: new approaches to an old challenge. Cancer Res 2015;75:5-10.
- 43. Brahmer J, Reckamp KL, Baas P, Crino L, Eberhardt WE, Poddubskaya E, et al. Nivolumab versus docetaxel in advanced squamous-cell non-small-cell lung cancer. N Engl J Med 2015;373:123-35.
- 44. Borghaei H, Paz-Ares L, Horn L, Spigel DR, Steins M, Ready NE, et al. Nivolumab versus docetaxel in advanced nonsquamous non-small-cell lung cancer. N Engl J Med 2015;373:1627-39.
- 45. Carbone DP, Reck M, Paz-Ares L, Creelan B, Horn L, Steins M, et al. First-Line Nivolumab in stage IV or recurrent non-smallcell lung cancer. N Engl J Med 2017;376:2415-26.
- 46. Garon EB, Rizvi NA, Hui R, Leighl N, Balmanoukian AS, Eder JP, et al. Pembrolizumab for the treatment of non-small-cell lung cancer. N Engl J Med 2015;372:2018-28.
- 47. Herbst RS, Baas P, Kim DW, Felip E, Perez-Gracia JL, Han JY, et al. Pembrolizumab versus docetaxel for previously

treated, PD-L1-positive, advanced non-small-cell lung cancer (KEYNOTE-010): a randomised controlled trial. Lancet 2016;387:1540-50.

- Reck M, Rodriguez-Abreu D, Robinson AG, Hui R, Csoszi T, Fulop A, et al. Pembrolizumab versus chemotherapy for PD-L1-positive non-small-cell lung cancer. N Engl J Med 2016;375:1823-33.
- 49. Brahmer JR, Rodriguez-Abreu D, Robinson AG, Hui R, Csoszi T, Fulop A, et al. Progression after the next line of therapy (PFS2) and updated OS among patients (pts) with advanced NSCLC and PD-L1 tumor proportion score (TPS) ≥50% enrolled in KEYNOTE-024. J Clin Oncol 2017;35(15 Suppl):9000.
- 50. Fehrenbacher L, Spira A, Ballinger M, Kowanetz M, Vansteenkiste J, Mazieres J, et al. Atezolizumab versus docetaxel for patients with previously treated non-small-cell lung cancer (POPLAR): a multicentre, open-label, phase 2 randomised controlled trial. Lancet 2016;387:1837-46.
- 51. Rittmeyer A, Barlesi F, Waterkamp D, Park K, Ciardiello F, von Pawel J, et al. Atezolizumab versus docetaxel in patients with previously treated non-small-cell lung cancer (OAK): a phase 3, open-label, multicentre randomised controlled trial. Lancet 2017;389:255-65.
- 52. Garassino M, Vansteenkiste J, Kim JH, Lena H, Mazieres J, Powderly J, et al. Durvalumab in ≥3rd-line locally advanced or metastatic, EGFR/ALK wild-type NSCLC: results from the phase 2 ATLANTIC study. J Thorac Oncol 2017;12(Suppl):S10-1.
- 53. Peters S, Gettinger S, Johnson ML, Janne PA, Garassino MC, Christoph D, et al. Phase II trial of atezolizumab as first-line or subsequent therapy for patients with programmed deathligand 1-selected advanced non-small-cell lung cancer (BIRCH). J Clin Oncol 2017;35:2781-9.
- 54. Antonia SJ, Villegas A, Daniel D, Vicente D, Murakami S, Hui R, et al. Durvalumab after chemoradiotherapy in stage III nonsmall-cell lung cancer. N Engl J Med 2017;377:1919-29.
- 55. Lynch TJ, Bondarenko I, Luft A, Serwatowski P, Barlesi F, Chacko R, et al. Ipilimumab in combination with paclitaxel and carboplatin as first-line treatment in stage IIIB/IV nonsmall-cell lung cancer: results from a randomized, doubleblind, multicenter phase II study. J Clin Oncol 2012;30:2046-54.
- 56. Govindan R, Szczesna A, Ahn MJ, Schneider CP, Gonzalez Mella PF, Barlesi F, et al. Phase III trial of ipilimumab combined with paclitaxel and carboplatin in advanced squamous non-small-cell lung cancer. J Clin Oncol 2017;35:3449-57.
- 57. Champiat S, Ileana E, Giaccone G, Besse B, Mountzios G, Eggermont A, et al. Incorporating immune-checkpoint inhibitors into systemic therapy of NSCLC. J Thorac Oncol 2014;9:144-53.
- 58. Antonia SJ, Larkin J, Ascierto PA. Immuno-oncology combinations: a review of clinical experience and future prospects. Clin Cancer Res 2014;20:6258-68.

- 59. Langer CJ, Gadgeel SM, Borghaei H, Papadimitrakopoulou VA, Patnaik A, Powell SF, et al. Carboplatin and pemetrexed with or without pembrolizumab for advanced, non-squamous nonsmall-cell lung cancer: a randomised, phase 2 cohort of the open-label KEYNOTE-021 study. Lancet Oncol 2016;17:1497-508.
- 60. Borghaei H, Langer CJ, Gadgeel S, Papadimitrakopoulou VA, Patnaik A, Powell SF, et al. Updated results from KEY-NOTE-021 cohort G: a randomized, phase 2 study of pemetrexed and carboplatin (PC) with or without pembrolizumab (pembro) as first-line therapy for advanced nonsquamous NSCLC. Ann Oncol 2017;28(Suppl_5):636-7.
- 61. Rizvi NA, Hellmann MD, Brahmer JR, Juergens RA, Borghaei H, Gettinger S, et al. Nivolumab in combination with platinum-based doublet chemotherapy for first-line treatment of advanced non-small-cell lung cancer. J Clin Oncol 2016;34:2969-79.
- 62. Hellmann MD, Rizvi NA, Goldman JW, Gettinger SN, Borghaei H, Brahmer JR, et al. Nivolumab plus ipilimumab as first-line treatment for advanced non-small-cell lung cancer (CheckMate 012): results of an open-label, phase 1, multicohort study. Lancet Oncol 2017;18:31-41.
- 63. Antonia S, Goldberg SB, Balmanoukian A, Chaft JE, Sanborn RE, Gupta A, et al. Safety and antitumour activity of durvalumab plus tremelimumab in non-small cell lung cancer: a multicentre, phase 1b study. Lancet Oncol 2016;17:299-308.
- 64. Remon J, Chaput N, Planchard D. Predictive biomarkers for programmed death-1/programmed death ligand immune checkpoint inhibitors in nonsmall cell lung cancer. Curr Opin Oncol 2016;28:122-9.
- 65. Gridelli C, Ardizzoni A, Barberis M, Cappuzzo F, Casaluce F, Danesi R, et al. Predictive biomarkers of immunotherapy for non-small cell lung cancer: results from an Experts Panel Meeting of the Italian Association of Thoracic Oncology. Transl Lung Cancer Res 2017;6:373-86.
- 66. Chae YK, Pan A, Davis AA, Raparia K, Mohindra NA, Matsangou M, et al. Biomarkers for PD-1/PD-L1 blockade therapy in non-small-cell lung cancer: is PD-L1 expression a good marker for patient selection? Clin Lung Cancer 2016;17:350-61.
- 67. Gibney GT, Weiner LM, Atkins MB. Predictive biomarkers for checkpoint inhibitor-based immunotherapy. Lancet Oncol 2016;17:e542-51.
- 68. Sacher AG, Gandhi L. Biomarkers for the clinical use of PD-1/ PD-L1 inhibitors in non-small-cell lung cancer: a review. JAMA Oncol 2016;2:1217-22.
- 69. Hirsch FR, McElhinny A, Stanforth D, Ranger-Moore J, Jansson M, Kulangara K, et al. PD-L1 immunohistochemistry assays for lung cancer: results from phase 1 of the blueprint PD-L1 IHC assay comparison project. J Thorac Oncol 2017;12:208-22.
- 70. Rimm DL, Han G, Taube JM, Yi ES, Bridge JA, Flieder DB, et al.

A prospective, multi-institutional, pathologist-based assessment of 4 immunohistochemistry assays for PD-L1 expression in non-small cell lung cancer. JAMA Oncol 2017;3:1051-8.

- 71. Buttner R, Gosney JR, Skov BG, Adam J, Motoi N, Bloom KJ, et al. Programmed death-ligand 1 immunohistochemistry testing: a review of analytical assays and clinical implementation in non-small-cell lung cancer. J Clin Oncol 2017;35:3867-76.
- 72. Scheerens H, Malong A, Bassett K, Boyd Z, Gupta V, Harris J, et al. Current status of companion and complementary diag-

nostics: strategic considerations for development and launch. Clin Transl Sci 2017;10:84-92.

- 73. Lawrence MS, Stojanov P, Polak P, Kryukov GV, Cibulskis K, Sivachenko A, et al. Mutational heterogeneity in cancer and the search for new cancer-associated genes. Nature 2013;499:214-8.
- 74. Rizvi NA, Hellmann MD, Snyder A, Kvistborg P, Makarov V, Havel JJ, et al. Cancer immunology: mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. Science 2015;348:124-8.