

Virus-associated Rhabdomyolysis in Children

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Purpose: Virus-associated rhabdomyolysis is very rare. We report 15 patients with rhabdomyolysis caused by various viruses.

Methods: Fifteen patients who were diagnosed with rhabdomyolysis and a viral infection were included in this study. Clinical, laboratory, and radiologic findings were evaluated through retrospective chart reviews.

Results: Chief complaints were severe bilateral lower leg pain and leg weakness. The median age was 5.7 years. The male:female ratio was 2:5. The viral infections were caused by influenza virus B, parainfluenza virus, and rhinovirus. One patient with influenza virus B had coinfection with coronavirus. Median initial laboratory values and ranges were as follows: serum creatinine, 0.4 (0.1-0.5) mg/dL; serum aspartate transaminase, 124 (48-1,098) IU/L; serum alanine transaminase, 30 (16-1,455) IU/L; serum creatine kinase, 2,965 (672-16,594) IU; serum lactate dehydrogenase, 400 (269-7,394) IU/L; serum myoglobin, 644 (314-3,867) ng/mL; urine myoglobin, 3 (3-10,431) ng/mL. All patients recovered without complications.

Conclusion: This is the first report of the simultaneous occurrence of rhabdomyolysis caused by various viruses. This is also the first report of rhinovirus-associated rhabdomyolysis.

Key words: Rhabdomyolysis, Influenza B virus, Paramyxoviridae infection, Coronavirus, Rhinovirus, Children

Introduction

Rhabdomyolysis means the dissolution of striped (skeletal) muscle, which is characterized by the leakage of muscle cell contents, such as electrolytes, myoglobin, and other sarcoplasmic proteins (e.g., creatine kinase (CK), aldolase, lactate dehydrogenase, alanine aminotransferase, and aspartate aminotransferase) into the circulation¹. The clinical presentation of rhabdomyolysis varies from an asymptomatic increase in CK to severe acute renal failure and hypovolemic shock². Rhabdomyolysis is commonly caused by crush injuries, medications, seizures and electrolyte disorders³.

Virus-associated rhabdomyolysis is very rare in adults³. However, rhabdomyolysis can also occur after a viral infection especially in children. A few viruses have been reported to be associated with rhabdomyolysis: influenza A/B²⁻⁵, parainfluenza⁶⁻⁸, coxsackie⁹, Epstein-Barr¹⁰, herpes simplex¹¹, adenovirus¹², and cytomegalovirus¹³. We have experienced the clusters of patients with rhabdomyolysis due to those several viral etiologies during their epidemic periods. Here, we report fifteen pediatric cases of virus-associated

rhabdomyolysis.

Materials and methods

A retrospective chart review was conducted based on medical records filed between January 2010 and December 2016 at Asan medical center children's hospital. Patients were included in the study if they were aged below 18 years, and had a diagnosis of rhabdomyolysis. Patients who had an underlying disease or previous history of exercise, trauma, drug intoxication or recurrent rhabdomyolysis were excluded. Diagnostic criteria of rhabdomyolysis were based on the increased level of serum myoglobin above the normal range. Patients who had serum creatine kinase (CK) level was above 1,500 IU/L, which is six times the upper limit of normal, were included in this study. Acute kidney injury was diagnosed when estimated creatinine clearance is under 35 mL/min/1.73m^{2.14}. All children had increased level of other muscle contents including serum lactate dehydrogenase (LD), alanine aminotransferase (ALT), and aspartate aminotransferase (AST). A multiplex reverse transcriptase polymerase chain reaction for respiratory viruses from a nasopharyngeal aspiration sample was also performed in all patients with the history of viral illness such as fever, cough, rhinorrhea and myalgia, and only those with positive results were finally included in this study.

Whole-body planar bone scintigraphy was performed 4 hours after injection of 1,110×(Body weight in kilograms/70 kg) MBq 99mTc-dicarboxypropane diphosphonate (DPD) in 3 patients to evaluate the severity and to detect the sites of the rhabdomyolysis. Anterior and posterior whole body scans were obtained, respectively using a dual-head gamma camera (Bodyscan; Siemens, Hoffman Estates, IL, or BiadXLT; Trionix, Twinsburg, OH) equipped with high resolution or ultra-high-resolution collimators.

Electrospray tandem mass aminoacid and acylcarnitine analyses was performed in 4 patients for a differential diagnosis of very rare cases of metabolic myopathies¹⁵.

All data were analyzed using the SPSS statistical software version 21 (SPSS Inc., Chicago, IL, USA). Wilcoxon signed-rank test was performed to compare the results of laboratory test between onset and the final follow-up.

Results

Fifteen patients were admitted due to rhabdomyolysis between 2010 and 2016. Chief complaints on admission were suddenly developed severe both calf pain causing progressive difficulty in walking (9 patients, 60.0%) and leg weakness (5 patient, 33.3%). The median age was 5.0 years (range 3-9 years). Male to female ratio was 11 to 4. All patients had a history of a recent viral infection from 3 to 10 days (median 5 days) prior to the presentation of rhabdomyolysis. The month of onsets of viral infections were shown in Fig. 1. Clinical symptoms of the infections were mostly fever (15 patients, 100%) with symptoms of upper respiratory infection (13 patients, 86.7%).

All patients received all their routine immunizations. No patient showed red to brown-colored or dark urine. Their causal viruses were influenza virus B (10 patients, 66.7%), influenza virus A (2 patients, 13.3%), boca virus+ rhinovirus, parainfluenza virus type 1+enterovirus or respiratory syncytial virus (1 patient each, 6.7%).

Their median values and range of laboratory findings were shown in Table 1. Urine myoglobin was checked, but only two of them (13.3%) had urine myoglobin level above 100 mcg/L. Initial GFR was 157.4 (20.8-275.0) ml/min/1.73 m². 99mTc-DPD bone scintigraphy was done in 5 patients which revealed increased soft tissue radiotracer uptake in both lower legs and one or both thighs in all of them.

Only one patient (6.7%) had acute kidney injury due to rhabdomyolysis. As initial symptom of rhabdomyolysis, drowsy mentality was presented in this patient. Initial GFR was decreased as 20.8 ml/min/1.73m². He was treated with

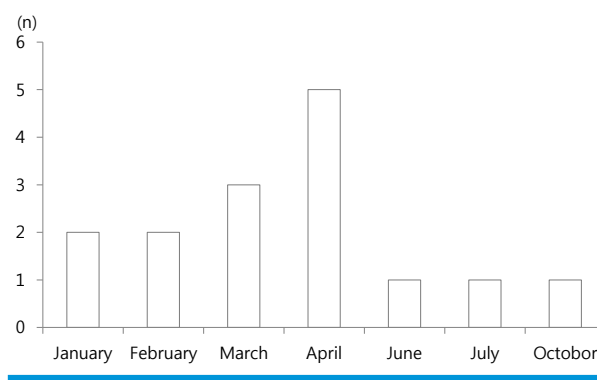


Fig. 1. The months of occurrence of viral infections. The months of onsets of viral infections of patients of virus-associated rhabdomyolysis were shown.

dialysis and recovered to normal GFR (138.1 ml/min/1.73 m²) without complication. Influenza A was detected in this patients. Other patients were treated with saline hydration and alkalization without dialysis. The median hospital days were 3 days (1-10 days) and all patients were discharged within 65 days except 1 patient who had prolonged admission due to acute kidney injury and prolonged increased CK level. All patients were recovered without any complications.

Discussion

The most common cause of rhabdomyolysis was influenza type B in this study. Most reports of influenza associated rhabdomyolysis are about type A^{3-5,16}. There are some reports about benign acute childhood myositis associated with influenza B¹⁷, but reports of influenza B-associated rhabdomyolysis are very rare¹⁸⁻²⁰. There was a recent report by Wu et al² about Taiwanese children with rhabdomyolysis due to influenza B infection. All 24 patients complained of calf pain and recovered without complication during the study periods of 8 years², which are similar to our cases.

We had two patients with rhabdomyolysis due to parainfluenza type 1 infection. There are only 4 case reports of rhabdomyolysis associated with parainfluenza virus on a medical online library search in Pubmed (<http://www.ncbi.nlm.nih.gov/>) so far^{6-8,21}. All patients were children, three boys and one girl with ages between 4 and 10 years old. Three of them had parainfluenza type 1^{6,7,21} and the other one had parainfluenza type 3 infection⁸. Three patients

developed acute renal failure, and one of them died due to respiratory failure⁸. Parainfluenza virus induced rhabdomyolysis might be more common than expected according to the number of case reports. It usually seems to have a benign clinical course based on our clinical experiences, but sometimes progresses to fatal outcome depending on the circumstances. This discordance of presentation might result from underdiagnosis. If patients have benign clinical course, PCR of CK may not be performed.

We had one patient with rhabdomyolysis associated with bocavirus and rhinovirus coinfection. There was only one report of rhabdomyolysis caused by rhinovirus²² and there was no report of rhabdomyolysis related to bocavirus so far. This is the first report of the patient with rhabdomyolysis caused by bocavirus.

No patient in this study showed red to brown-colored urine. All the patients had increased serum myoglobin level, but only 2 of them (13.3%) had clinically significant myoglobinuria. Only those with serum myoglobin level above 2,000 ng/mL had myoglobinuria and positive urine occult blood without microscopic hematuria. The classic triad of rhabdomyolysis includes myalgia, red-to-brown or dark urine and muscle weakness¹⁶. However, only less than 10% of patients with rhabdomyolysis show all three classic features and only 3.6% have dark urine implicating a potentially insidious onset^{1,23}. The test for myoglobin in urine might be negative because myoglobin has a short half-life (2-3 hours) and could be rapidly and unpredictably eliminated by hepatic metabolism and renal excretion²⁴. So even in patients without red-to-brown or dark urine, we should suspect the possibility of rhabdomyolysis as in our

Table 1. The Median Values and Range of Laboratory Findings of Patients

	Initial lab finding (median values and range)	Follow-up lab finding (median values and range)	Change	P-value of change
Serum calcium (mg/dL)	9.0 (7.4-9.6)	9.3 (8.9-9.8)	0.4 (-0.1-1.9) [4.4 (-1.1-25.7) %]	0.001
Serum albumin (g/dL)	3.8 (3.2-4.2)	4.3 (3.3-4.7)	0.6 (-0.5-1.3) [16.7 (-13.2-40.6) %]	0.007
Serum sodium (mmol/L)	137 (134-143)	140 (137-143)	3 (-2-6) [2.2 (-1.4-4.4) %]	0.036
Serum AST (IU/L)	137 (69-5,029)	33 (23-96)	109 (-4997--13) [-77.9 (-99.4-18.8) %]	0.001
Serum ALT (IU/L)	38 (18-3,843)	20 (12-126)	11 (-3,825-27) [-40.0 (-99.5-77.1) %]	0.083
Serum CK (IU/L)	4,022 (1,840 ~30,255)	152 (95-1,887)	-3,870 (-2,8368--1462) [-95.4 (-98.7-79.5) %]	0.001
Serum LD (IU/L)	388 (277-4651)	293 (193-617)	84 (-4,408-170) [-23.1 (-94.8-49.6) %]	0.023
Serum BUN (mg/dL)	6 (4-42)	9 (4-13)	2 (-5-7) [33.3 (-50.0-175.0) %]	0.174
Serum Cr (mg/dL)	0.37 (0.2-2.99)	0.41 (0.28-0.55)	-0.02 (-2.54-0.22) [-5.4 (-84.9-100.0) %]	0.608
Serum myoglobin (ng/mL)	766.5 (140-5,589)	30.5 (17-185)	-688.5 (-5,568--84) [-95.8 (99.6--60.0) %]	0.001

cases.

There was a male predominance in virus associated rhabdomyolysis in this study. All patients were between 3-9 years old. Patients who had a previous history of exercise or trauma were excluded. In general, it was reported that male patient is usually more prevalent in rhabdomyolysis even in non-exertional, non-traumatic cases such as virus associated rhabdomyolysis^{2,25}. This study shows that male sex hormone or any kinds of overactivities or masculine activities which might be expected in boys seem to be related to virus associated rhabdomyolysis.

^{99m}Tc-DPD bone scintigraphy revealed increased soft tissue radiotracer uptake in both legs in all five patients who had the test in this study. Phosphate derivative tracers such as ^{99m}Tc-DPD, used for bone imaging, are taken up by some soft tissue lesions²⁶. Proposed mechanisms for the tracer uptake by these soft tissue lesions are increased calcium concentration and accumulation through the defects in cell membrane after tissue injury, and the tracer binding to hydroxyapatite and calcium phosphate crystals in these tissues²⁶. There were relatively clear differences in the amount of uptake and extent of lesions between the patients with higher CK and myoglobin levels and those with lower levels, whereas the scintigraphic findings looked alike between those groups with similar levels regardless of their difference in ages. It seems that ^{99m}Tc-DPD bone scintigraphy is very useful in clinically suspected rhabdomyolysis as a sensitive and reliable diagnostic test which enables to localize and quantify muscular involvement.

We did not perform muscle biopsy nor cytokine analysis for the pathogenic evaluations because clinical courses in all patients in this study were benign and recovered without complication. The mechanism of rhabdomyolysis due to viral infection has not been established with certainty yet. The following possibilities have been proposed: First, direct viral invasion of muscle tissue is one possibility; however, the identification of virus or viral particles in affected muscles has been difficult to demonstrate consistently²⁷. Second, myotoxic cytokines released in response to viral infection might be another possibility^{9,28}. Third, immunologic processes induced by the viral infection could result in muscle damage²⁷. It appears that the time from viral illness to the onset of myositis varies, from coincidental to a 3-week delay. In this study, the intervals between viral

infection and rhabdomyolysis were from 3 to 10 days. This time course variation seems to support the possibility of different pathophysiologic mechanisms.

There are some limitations in this study. First, not all patients conducted Whole-body planar bone scintigraphy since this study is retrospective study. Second, as this study was conducted in the single center, the selection bias may exist. At last, since sample size is not enough to represent all Korean children, the larger study will be required.

In conclusion, we have experienced patients with rhabdomyolysis due to different kinds of respiratory viral infections, mostly influenza B, followed by parainfluenza and rhinovirus during their outbreaks in the spring of 2010. This is the first report of the simultaneous occurrence of rhabdomyolysis due to multiple viral etiologies. We are not sure if these findings are just coincidental findings by chance or if there are any reciprocal actions among different viruses which favor rhabdomyolysis. This is also the first report of rhinovirus associated rhabdomyolysis.

Conflicts of interest

No potential conflict of interest relevant to this article was reported.

References

1. Bosch X, Poch E, Grau JM. Rhabdomyolysis and acute kidney injury. *N Engl J Med* 2009;361:62-72.
2. Wu CT, Hsia SH, Huang JL. Influenza B-associated rhabdomyolysis in Taiwanese children. *Acta Paediatr* 2010;99:1701-4.
3. Sato E, Nakamura T, Koide H. Rhabdomyolysis induced by influenza a infection: case report and review of literature. *Ther Apher Dial* 2011;15:208-9.
4. Unverdi S, Akay H, Ceri M, Inal S, Altay M, Demiroz AP, Duranay M. Acute Kidney Injury due to Rhabdomyolysis in H1N1 Influenza Infection. *Ren Fail*. 2011.
5. Lai CC, Wang CY, Lin HI. Rhabdomyolysis and acute kidney injury associated with 2009 pandemic influenza A(H1N1). *Am J Kidney Dis* 2010;55:615.
6. Pana ZD, Tragiannidis A, Douma S, Chrisa K. Rhabdomyolysis in an adolescent associated with parainfluenza type 1 virus. *Pediatr Infect Dis J* 2011;30:450.
7. Ebbeson RL, De Kock MJ, Penny N, Kollman TR. Rhabdomyolysis,

- acute renal failure, and compartment syndrome in a child with parainfluenza type 1 infection. *Pediatr Infect Dis J* 2009;28:850-2.
8. Ueda K, Robbins DA, Iitaka K, Linnemann CC, Jr. Fatal rhabdomyolysis associated with parainfluenza type 3 infection. *Hiroshima J Med Sci* 1978;27:99-103.
 9. Fodili F, van Bommel EF. Severe rhabdomyolysis and acute renal failure following recent Coxsackie B virus infection. *Neth J Med* 2003;61:177-9.
 10. Osamah H, Finkelstein R, Brook JG. Rhabdomyolysis complicating acute Epstein-Barr virus infection. *Infection* 1995;23:119-20.
 11. Koizumi K, Yokoyama K, Nishio M, Shibata S, Ozutusmi K, Yamaguchi M, Sato N, Yasukouchi T, Sawada K, Koike T. A case of virus-associated hemophagocytic syndrome (VAHS) complicated by rhabdomyolysis which were associated with herpes-simplex virus infection. *Rinsho Ketsueki* 1996;37:40-5.
 12. Meshkinpour H, Vaziri ND. Acute rhabdomyolysis associated with adenovirus infection. *J Infect Dis* 1981;143:133.
 13. Hirohama D, Shimizu T, Hashimura K, Yamaguchi M, Takamori M, Awatsu Y, Tsujino M, Mizutani T. Reversible respiratory failure due to rhabdomyolysis associated with cytomegalovirus infection. *Intern Med* 2008;47:1743-6.
 14. Akcan-Arikan A, Zappitelli M, Loftis LL, Washburn KK, Jefferson LS, Goldstein SL. Modified RIFLE criteria in critically ill children with acute kidney injury. *Kidney International* 2007;71(10):1028-35.
 15. Tonin P, Lewis P, Servidei S, DiMauro S. Metabolic causes of myoglobinuria. *Ann Neurol* 1990;27:181-5.
 16. Chen SC, Liu KS, Chang HR, Lee YT, Chen CC, Lee MC. Rhabdomyolysis following pandemic influenza A (H1N1) infection. *Neth J Med* 2010;68:317-9.
 17. Agyeman P, Duppenhaler A, Heining U, Aebi C. Influenza-associated myositis in children. *Infection* 2004;32:199-203.
 18. Paletta CE, Lynch R, Knutsen AP. Rhabdomyolysis and lower extremity compartment syndrome due to influenza B virus. *Ann Plast Surg* 1993;30:272-3.
 19. Marcos Sanchez F, Celdran Gil J, Duran Perez-Navarro A. Acute rhabdomyolysis as a complication of the influenza B virus. *An Med Interna* 1992;9:519.
 20. Berbegal J, Robles A, de Gracia C, Lopez M. Rhabdomyolysis associated with influenza B virus. *Rev Clin Esp* 1992;190:158.
 21. Vrsalovic R, Tesovic G, Mise B. Rhabdomyolysis and acute renal failure in a child with para-influenza type 1 infection. *Pediatr Nephrol* 2007;22:1369-71.
 22. Tan LO, Thoon KC, Chong CY, Tan NW. Rhabdomyolysis caused by rhinovirus. *Glob Pediatr Health* 2016;3:1-3.
 23. Luck RP, Verbin S. Rhabdomyolysis: a review of clinical presentation, etiology, diagnosis, and management. *Pediatr Emerg Care* 2008;24:262-8.
 24. Vanholder R, Sever MS, Ereke E, Lameire N. Rhabdomyolysis. *J Am Soc Nephrol* 2000;11:1553-61.
 25. Mannix R, Tan ML, Wright R, Baskin M. Acute pediatric rhabdomyolysis: causes and rates of renal failure. *Pediatrics* 2006;118:2119-25.
 26. Lim ST, Sohn MH, Park SA. Tc-99m MDP three-phase bone scintigraphy in disciplinary exercise-induced rhabdomyolysis. *Clin Nucl Med* 2000;25:558-9.
 27. Nauss MD, Schmidt EL, Pancioli AM. Viral myositis leading to rhabdomyolysis: a case report and literature review. *Am J Emerg Med* 2009;27:372 e375-6.
 28. Konrad RJ, Goodman DB, Davis WL. Tumor necrosis factor and coxsackie B4 rhabdomyolysis. *Ann Intern Med* 1993;119:861.