

## RESEARCH ARTICLE

## Characterization of *Rhizodermea veluwensis* Isolated from the Roots of *Rhododendron mucronulatum* in Korea

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### Abstract

A fungal strain was isolated from surface-sterilized roots of *Rhododendron mucronulatum*, a plant species belonging to the Ericaceae family, collected from Mt. Minjujisan, Korea. This fungal strain was identified as *Rhizodermea veluwensis* based on its morphological characteristics and based on phylogenetic analysis of its internal transcribed spacer regions and large-subunit rDNA. *R. veluwensis* has not been previously reported in Korea, and for the first time, we report and describe it herein.

**Keywords:** Endophytic fungi, Ericaceae, Korean rosebay, *Rhizodermea veluwensis*

### Introduction

The genus *Rhizodermea* Verkley & Zijlstra is a new taxon recently discovered by Verkley et al. [1] during evaluation of the unidentified fungi recorded as *Helotiales* sp. Currently, only one species, *Rhizodermea veluwensis* Verkley & Zijlstra, belongs to this genus. The genus *Rhizodermea* belongs to the family Dermateaceae, order Helotiales, class Leotiomycetes, and phylum Ascomycota. Through phylogenetic analysis of DNA sequences, this genus was clustered within the family Dermateaceae as a distinct clade basal to the genus *Pezicula* [2]. The teleomorph has not been found in *R. veluwensis*. It was known that the fungi produce only chlamydospore-like structures in culture and no other reproductive structure was found to date [1, 2].

*R. veluwensis* has been mainly isolated from ericaceous plants. It was first reported from surface-sterilized roots of *Erica tetralix* in Netherlands [1] and isolated from roots of *Empetrum nigrum* and *Vaccinium* spp. In Morocco, this fungus was isolated from roots of *Calluna vulgaris* and *Vaccinium myrtillus* [3]. However, the host range of the endophytic fungus could be plants other than those belonging to the family Ericaceae. It was isolated from roots of *Larix decidua* belonging to the family Pinaceae and *Clethra barbinervis*

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belonging to the family Clethraceae [4]. In addition, the internal transcribed spacer (ITS) sequence of fungi isolated from roots of *Banksia spinulosa* belonging to the family Proteaceae in Australia showed 100% identity to *R. veluwensis* [1].

In this study, a fungal strain was isolated from the surface-sterilized roots of *Rhododendron mucronulatum*, a plant belonging to the family Ericaceae, and the strain was confirmed as *R. veluwensis* based on its morphological characteristics and phylogenetic analysis. To the best of our knowledge, this species has not been previously reported in Korea. Here, we describe the morphological characteristics and phylogenetic analysis of the strain.

## Materials and Methods

### Root sampling and isolation of the fungal strain.

Roots of *R. mucronulatum* were collected from Mt. Minjuji located in Chungbuk, Korea (N 36° 2' 49.88", E 127° 46' 36.7"). These samples were packed in a polyethylene bag and transported to the laboratory. The roots were washed with sterilized distilled water and then treated with 70% ethanol and 3% NaClO solution [5]. The surface-sterilized roots were cut into 0.5 cm length segments. The root segments were placed on water agar (WA) medium and the plates were incubated under dark conditions at 25°C. Mycelia growing out from the root segments were transferred to potato dextrose agar (PDA) and incubated at 25°C for 7 days. The pure isolate, 16E003, was stored in 20% glycerol at -80°C at the Mycology laboratory of Korea National University of Education, Cheongju, Korea, and deposited as glycerol stock at the Culture Collection of National Institute of Biological Resources (NIBR), Incheon, Korea, with an accession number NIBRFG0000499917.

### Morphological characterization

PDA and malt extract agar (MEA) were used for morphological characterization of the fungal strain. Morphological characteristics of the isolate 16E003 were determined after incubation on both the media under dark conditions at 25°C for 7 days. Fungal isolates were mounted using lactophenol solution and observed under a light microscope (AXIO imager A1; Carl Zeiss, Oberkochen, Germany).

### Phylogenetic analysis

Genomic DNA was extracted from the isolate 16E003 using Exgene Plant SV mini kit (GeneAll, Seoul, Korea) according to the manufacturer's protocol. The ITS including 5.8S and the large subunit (LSU) regions of ribosomal DNA were amplified using ITS1F/ITS4 [6] and LR0R/LR16 primers [7], respectively. The amplified PCR product was sequenced by SolGent Co. (Daejeon, Korea). The sequence was deposited in NCBI GenBank (accession number MF042207). Phylogenetic analysis was conducted using neighbor-joining methods

in MEGA 6 software [8]. Support for specific nodes on the tree was estimated by bootstrapping 1,000 replications. The sequence for *Phialocephala fortinii* was used as an outgroup.

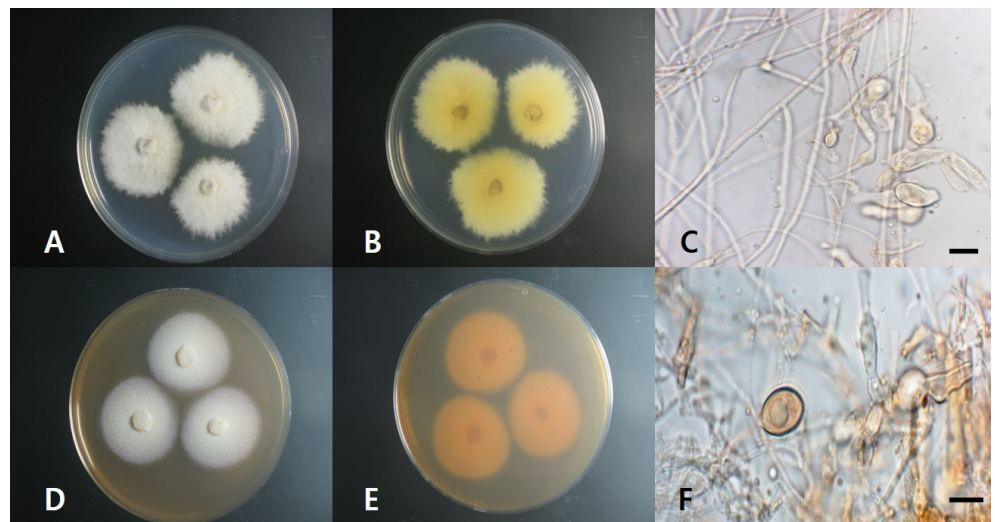
## Results and Discussion

### Taxonomy of isolate 16E003

#### *Rhizodermea veluwensis* Verkley & Zijlstra, *Persoonia* 24: 131. 2010 (Fig. 1, Table 1)

Diameters of the colonies on PDA grown for 7 days were 36–40 mm, and obverse of the colony was observed to be ivory and the reverse was observed to be pale yellow in color. The colonies were flattened and serrated at the margin. Diameters of the colonies on MEA grown for 7 days were 34–37 mm, and obverse of the colony was observed to be beige and reverse was observed to be reddish brown in color. Colonies were flat and blunt-serrated at the margin. At the margin of the colonies, aerial mycelium was formed and its color was hyaline or vinaceous. The hyphae were 1–2  $\mu\text{m}$  wide initially and 3–6  $\mu\text{m}$  wide later, and were isodiametric inflated cells, rarely big gourd-shaped cells about 15  $\mu\text{m}$  wide. Chlamydospores were globose to limoniform, hyaline to yellowish, 1-septate, smooth-walled or warty, 15–20  $\mu\text{m}$  in diameter.

Specimen examined: KOREA; Mt. Minjuji, N 36° 2' 49.88", E 127° 46' 36.7"; isolated from roots of *R. mucronulatum*; September 21, 2016; Culture 16E003 (NIBRFG 0000499917, GenBank MF042207).



**Fig. 1.** Morphological characteristics of *Rhizodermea veluwensis* 16E003. A, obverse colonies on potato dextrose agar (PDA); B, reverse colonies on PDA; C, mycelium; D, obverse colonies on malt extract agar (MEA); E, reverse colonies on MEA; F, chlamydospore (scale bars: C, F =10  $\mu\text{m}$ ).

**Table 1.** Morphological characteristics of fungal strain 16E003 isolated from this study

Characteristics	Strain 16E003 from this study	<i>Rhizodermea veluwensis</i> <sup>a</sup>
Colony	MEA, 18°C, 3 weeks	MEA, 18°C, 3 weeks
Color	beige to pale ivory, pale brown aerial mycelium, reverse sepia, colourless to vinaceous margin	cinnamon, pale honey aerial mycelium, reverse sepia, colourless to vinaceous-buff margin
Size	34~37 mm in diam	45~48 mm in diam
Shape	surface plane, crenulate margin	surface radially creased or plane, crenulate margin
Mycelium	hyaline, 1~2 µm wide hyphae, later also 3~6 µm wide, hypha transformed to isodiametric inflated cells, rarely big gourd-shaped cells about 15 µm wide	hyaline, septate, 1.5~2.5 µm wide hyphae, later also 5.5~8.5 µm wide, hypha transformed to a chain of pigmented, isodiametric inflated cells
chlamydospores	0-septate, smooth-walled, warted, 15~20 µm in diam.	0~1-septate, smooth-walled, warted, 18~25 µm in diam.

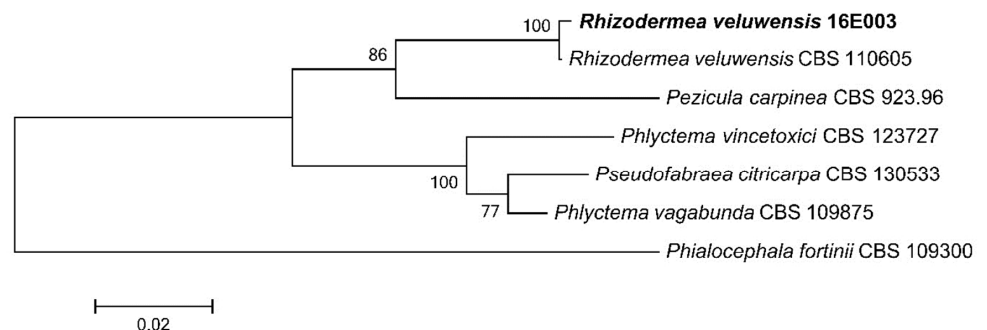
MEA, malt extract agar.

<sup>a</sup>Verkley et al. [1]

### Phylogenetic analysis

BLAST results in NCBI showed that the ITS sequence of this isolate was closely related to that of *R. veluwensis* KR859283.1 with 99% similarity and the LSU region of this isolate was closely related to that of *R. veluwensis* KR859076.1 with 100% similarity. Both the sequences were obtained from fungal strains isolated from roots of species belonging to the family Ericaceae (*Vaccinium myrtillus* and *Erica tetralix*, respectively) inhabiting the Netherlands. Phylogenetic tree using the combined sequence of both ITS and LSU regions in Fig. 2 showed that the sequence from this study formed a monophyletic group with the strain *R. veluwensis* CBS 110605 isolated from roots of *Erica tetralix* in Netherlands, supported by 99% bootstrap value.

The morphological characteristics of fungal strain 16E003 isolated from the roots of *R. mucronulatum*, in this study, were generally consistent with the original description of the species [1]. Additionally, phylogenetic analysis using ITS and LSU sequences strongly



**Fig. 2.** Neighbor-joining phylogenetic tree based on a combined alignment of both internal transcribed spacer (ITS) and large subunit (LSU) sequences. *Phialocephala fortinii* was used as an outgroup. Numbers on branches indicate bootstrap values (1,000 replicates). Fungal strain isolated in this study are in bold.

indicated a close relation with the sequence of *R. veluwensis*.

*R. mucronulatum* is known as an endophytic fungus colonizing roots of plants, mainly belonging to the family Ericaceae. Endophytic fungi are defined as those living inside healthy plant tissues without disease symptoms. Although their relationship is not well understood, they may enhance host growth and improve the tolerance of host against various environmental stresses [9, 10]. For example, a previous study showed that *R. mucronulatum* enhanced heavy metal stress tolerance in a host plant growing in mining sites [4]. Therefore, further studies on relationships between this fungal endophyte as well as roles of the fungus in ecosystems will be needed.

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## REFERENCES

1. Verkley GJ, Hofland-Zijlstra JD, Berendse F. *Rhizodermea veluwensis*. Persoonia 2010; 24:130-1.
2. Chen C, Verkley GJ, Sun G, Groenewald JZ, Crous PW. Redefining common endophytes and plant pathogens in *Neofabraea*, *Pezicula*, and related genera. Fungal Biol 2016;120:1291-322.
3. Hamim A, Miché L, Douaik A, Mrabet R, Ouhammou A, Duponnois R, Hafidi M. Diversity of fungal assemblages in roots of Ericaceae in two Mediterranean contrasting ecosystems. C R Biol 2017;340:226-37.
4. Yamaji K, Watanabe Y, Masuya H, Shigeto A, Yui H, Haruma T. Root fungal endophytes enhance heavy-metal stress tolerance of *Clethra barbinervis* growing naturally at mining sites via growth enhancement, promotion of nutrient uptake and decrease of heavy-metal concentration. PLoS One 2016;11:e0169089.
5. Ahlich K, Sieber TN. The profusion of dark septate endophytic fungi in non-ectomycorrhizal fine roots of forest trees and shrubs. New Phytol 1996;132:259-70.
6. Gardes M, Bruns TD. ITS primers with enhanced specificity for basidiomycetes-application to the identification of mycorrhizae and rusts. Mol Ecol 1993;2:113-8.
7. Moncalvo JM, Lutzoni FM, Rehner SA, Johnson J, Vilgalys R. Phylogenetic relationships of agaric fungi based on nuclear large subunit ribosomal DNA sequences. Syst Biol 2000;49:278-305.
8. Tamura K, Stecher G, Peterson D, Filipowski A, Kumar S. MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. Mol Biol Evol 2013;30:2725-9.
9. Carroll G. Fungal endophytes in stems and leaves: from latent pathogen to mutualistic symbiont. Ecology 1988;69:2-9.
10. Saikkonen K, Faeth SH, Helander M, Sullivan TJ. Fungal endophytes: a continuum of interactions with host plants. Annu Rev Ecol Syst 1998;29:319-43.