

Effect of Diet and Water Intake on Aquaporin 2 Function

Jun-Mo Kim, M.D., Ph.D.¹
Tae-Hee Kim, M.D., Ph.D.²
Tong Wang, M.D., Ph.D.³

Department of Urology¹, Department of Obstetrics and Gynecology², School of Medicine, Soonchunhyang University, Bucheon, Republic of Korea, Department of Cellular and Molecular Physiology³, School of Medicine, Yale University, New Haven, USA

Corresponding author:

Tong Wang, M.D., Ph.D.
Department of Cellular and Molecular Physiology, Yale University, School of Medicine, 333 Cedar St, New Haven, CT, USA 06520-8026

Tel: +1-203-785-4074
Fax: +1-203-785-4951
E-mail: tong.wang@yale.edu

Received: 20 March 2016
Revised: 20 March 2016
Accepted: 22 March 2016

This is an open-access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/bync/4.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Copyright © 2016 The Korean Society of Pediatric Nephrology

Appropriate control of diet and water intake is important for maintaining normal blood pressure, fluid and electrolyte homeostasis in the body. It is relatively understood that the amount of sodium and potassium intake directly affects blood pressure and regulates ion transporters; Na and K channel functions in the kidney. However, little is known about whether diet and water intake regulates Aquaporin (AQP) function. AQPs, a family of aquaporin proteins with different types being expressed in different tissues, are important for water absorption by the cell. Water reabsorption is a passive process driven by osmotic gradient and water permeability is critical for this process. In most of the nephron, however, water reabsorption is unregulated and coupled to solute reabsorption, such as AQP1 mediated water absorption in the proximal tubule. AQP2 is the only water channel founded so far that can be regulated by hormones in the kidney. AQP2 expressed in the apical membrane of the principal cells in the collecting tubule can be regulated by vasopressin (antidiuretic hormone) controlling the final volume of urine excretion. When vasopressin binds to its receptor on the collecting duct cells, it stimulates the translocation of AQP2 to the membrane, leading to increased water absorption via this AQP2 water channel. However, some studies also indicated that the AQP2 is also been regulated by vasopressin independent mechanism. This review is focused on the regulation of AQP2 by diet and the amount of water intake on salt and water homeostasis.

Key words: Aquaporin 2, Arginine vasopressin, Diet, Osmolarity, Nocturan enuresis

Introduction

Water is an essential component for life for all plants and animals; however, cell membranes are composed of lipid bilayers that have low permeability to water. Therefore, simple diffusion by osmotic pressure is not sufficient for water to enter cells, and specialized transporters (aquaporins (AQP) that allow cells to absorb large amounts of water are needed to control the level of both extracellular and intercellular fluid (280-295 mOsmol/kgH₂O in humans). AQP was first discovered by Peter Agre in 1992¹⁾. The AQP protein is 30 kDa in weight, and is expressed in plants, bacteria, fungi, and mammals²⁾. Thirteen types of AQP (AQP 0-12) have been identified in mammals. The majority of water absorption is via AQP1, localized in the proximal tubule and AQP2, expressed in the principal cells of the collecting tubule in the kidney (Fig. 1)²⁻⁴⁾. Water reabsorption in the proximal tubule is coupled to solute reabsorption driven by the osmotic gradient⁵⁾. Although 67% of the filtered water is reabs-

orbed in the proximal tubule (since AQP1 is not regulated by anti-diuretic hormone), the amount of water absorption in this segment does not control the final urine volume². In contrast, AQP2 is localized in the principal cells that are also arginine vasopressin receptor target cells in the collecting tubule. AQP2 can be regulated by these hormones and controls the final urine volume². The functional regulation of AQP2 thus becomes more important than AQP1. Indeed, mutations of the AQP2 caused several common diseases, including diabetes insipidus and nocturnal enuresis². AQP2 surface expression and function is activated by arginine vasopressin (AVP). After AVP binds to the arginine vasopressin receptor 2 (V2R) in the cell basolateral membrane, intracellular cAMP is increased, which ultimately leads to the trafficking of AQP2 to the apical plasma membrane of collecting ducts (Fig. 2). Increased intracellular water absorbed by AQP2 is transported to the blood by AQP3 and AQP4 located in the basolateral membrane. The two main factors that regulate AQP2 are environmental osmolality and AVP⁵. Although the cortico-medullary osmolality gradient has a greater influence over AQP2 than AVP, the regulatory mechanisms controlling water balance through AQP2 are complex and are still not well understood. Interestingly, both aging has a significant effect on

the expression of AQPs, and gender affect diurnal variation of plasma vasopressin and effect of desmopressin⁶⁻⁸. V2R expression and its ability to bind to AVP decreased by up to 30% in an aging rat model (F344BN rats)^{6,9}. However, while AQP1 and 4 expressions were unchanged by aging, the expressions of AQP2 and 3 were decreased 80% and

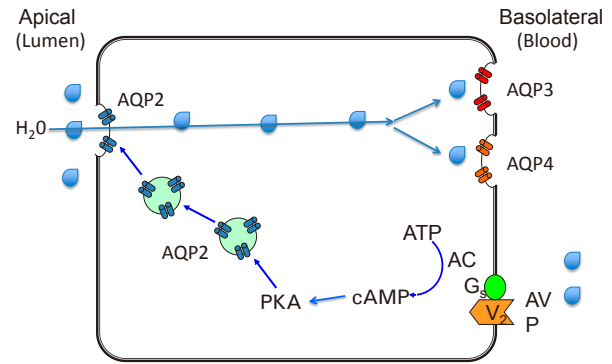


Fig. 2. Regulation of AQP2 trafficking and expression in principal cells of the collecting tubule. AVP acts on V2 receptors (V2R) in the basolateral plasma membrane through the GTP-binding protein Gs adenylylcyase (AC) is activated, which accelerates the production of cAMP from ATP and activation of PKA. PKA phosphorylates AQP2 in intracellular vesicles and accelerates AQP2 trafficking to the apical membrane surface. More water enters the cell from the apical side through AQP2 and AQP 3 and 4 transport intracellular water cross the basolateral membrane into the blood vessel.

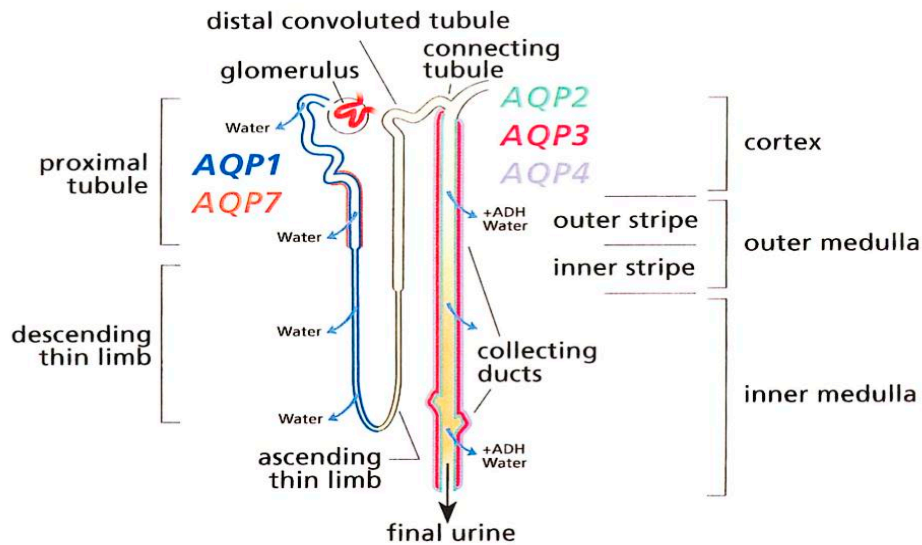


Fig. 1. Diagrammatic representation of the localization of different aquaporins along the nephron. AQP1 (blue) is present in the proximal tubule and descending thin limb. AQP2 (green) is abundant in the apical and subapical part of collecting duct principal cells, whereas AQP3 (red) and AQP4 (purple) are both present in the basolateral plasma membrane of collecting duct principal cells. AQP7 (orange) is confined to the apical brush border of straight proximal tubules. ADH, antidiuretic hormone (The figure is reproduced from Nielsen et al, 2002, with permission).

50%, respectively, in 30-month-old rats compared to 10-month-old rats⁹. The circadian rhythm, which is controlled by the suprachiasmatic nucleus, is also known to decrease in the elderly¹⁰. Plasma levels of AVP are also higher in men compared to women, and the effects of vasopressin on the kidneys are greater in men¹¹.

The aim of this review is to evaluate the effect of different diets on AQP regulation and nighttime urine output. Information obtained from the available literature reports suggests that certain diets could be recommended to children with nocturnal enuresis, and may include avoiding water 2-3 hours before sleeping, a reduction in salt during dinner, and avoidance of high protein diets. This review also summarized the effects of hypercalciuria on AQP expression.

Effect of Water Loading and Deprivation on Urine Volume and AQP2

After a significant amount of water is loaded into the kidneys, plasma and urine osmolality is decreased and AQP2 expression is down-regulated to reduce renal water absorption¹². In a previous study examining five healthy volunteers, urine osmolality decreased from 895±38.5 to 86.3±8.0 mOsm/kgH₂O two hours after acute water loading (20 mL/Kg), urine volume increased from 0.68±0.15 to 8.94±3.14 mL/min, and urinary AQP2 was dramatically down-regulated from 266±28 to 48±17 pmol/mL¹². In contrast, overnight dehydration decreased urine volume (1.0±0.2 to 0.6±0.1 ml/min), increased urine osmolality (888±18 to 1004±17 mOsmol/kg), and doubled urine AQP2

(140±45 to 285±63 fmol AQP2/μmol creatinine)¹³. The results of these studies suggest that preventing water intake two hours before sleep should reduce night urine output.

AQP and a High Salt Diet

Many different studies have investigated the relationship between high and low sodium diets and the expression of AQP2 (Table 1)¹⁴⁻¹⁷. However, since the detection of renal AQP levels is difficult in humans, the studies have typically used urine AQP2 measures. Three percent of renal AQP2 is excreted daily into the urine, and urinary excretion of AQP2 per day was the same in men and women (mean age: 48.4±15.8, 40.7±16.6 years, average excretion: 380 pmol/d)¹⁸. This study suggested that because urine osmolality is positively correlated with urine AQP2 levels, interpreting AQP2 levels through spot urine samples should be carefully conducted¹⁸. Baumgarten et al. also suggested that because there is no simple relationship between urine osmolality and urine AQP2, random or singular measurement of urine AQP2 is not useful for evaluation of renal vasopressin activity¹³. Several previous studies have used a hypertonic saline infusion to mimic high sodium intake, and water deprivation and loading have also been used. Interestingly, even though urinary AQP2 excretion was increased by both water deprivation and hypertonic saline infusion, there were differences in the time it took for these changes to occur. The hypertonic saline infusion-induced change occurred more rapidly, within minutes, while the effects of dehydration took hours¹⁹. In a recent human study on the effect of high and low sodium intake on AQP2, urine

Table 1. Summary of Previously Reported Results of Effects of High Sodium Intake on AQP2 Expression

Authors	Year of Publication	Subjects	Methods of high salt intake	AVP	U-Osm	Urine Vol.	AQP2
Baumgarten et al	2000	Human	Dehydration+ hypertonic saline	13-fold↑	6.4-fold↑	5-fold↓	No change
Pedersen et al	2001	Human	Fluid restriction+ hypertonic saline	1.5-fold↑	5-fold↑	3-fold↓	No change
Roxas et al	2002	SD rats Dahl SS/Jr rat	High salt diet				5-fold↓ 3-fold↑
Song et al	2004	SD rats	2.59% NaCl 65% fructose + NaCl	1.5-fold↓	1.3-fold↑	2-fold↑	↑
Graffe et al	2012	Human	High-low sodium intake Hypertonic saline	2-fold↑	4-fold↑	3-fold↓	No change

Abbreviations: AVP, arginine vasopressin; U-Osm, urine osmolality; Urine Vol., Urine volume; AQP2, aquaporin 2.

AQP2 concentration increased from a baseline of 113 to 144 ng/mmol after a high sodium diet (300 mmol sodium/day; 17.5 g salt/day for 4 days) and hypertonic saline infusion (3%, 7 ml/kg)¹⁶. Urine volume (7.6-8.4 ml/min) was decreased by both a high salt diet (2.7-3.4 ml/min) and a low salt diet (1.5-2.1 ml/sec). Interestingly, urine AQP2-creatinine was slightly increased from 79 to 84-102 ng/mmol by a low sodium diet (30 mmol sodium/day; 1.8 g salt/day for 4 days)¹⁶. While blood pressure was slightly increased by a high sodium diet, angiotensin II and aldosterone, which are the important regulators of reabsorption of sodium in the kidney, decreased from 13.6 to 9.3 pmol/l and 225 to 148 pmol/l, respectively¹⁶. These results suggest that if plasma osmolality is increased by a high salt diet, renal sodium excretion is increased by the down-regulation of angiotensin II and aldosterone, which subsequently increases water reabsorption to reduce blood osmolality. However, another study reported different results and found that a 2.59% NaCl diet increased urine volume (40± to 81±7 ml/day/kg/BW) and AQP2 despite similar AVP levels (0.6±0.3 to 0.4±0.1 pmol/l)²⁰. In addition, Roxas et al. fed either a normosodic diet (0.4% NaCl) or a high-sodium diet (8%) for 10 days to male Sprague Dawley rats and Dahl SS/Jr rats¹⁷. While AQP1 and AQP2, measured by RT-PCR, were significantly decreased (5 fold lower) in Sprague Dawley rats in the high-sodium diet group, AQP2 expression was three-fold higher in Dahl SS/Jr rats treated with a high salt diet. They suggested that the reduction in AQP-2 expression may be a compensatory mechanism for reducing salt and water reabsorption after a high salt diet in Sprague Dawley rats. The increase in AQP2 in Dahl SS/Jr rats may be a mechanism to increase salt and water reabsorption that has arisen through AQP2 mutation or polymorphism of AQP2 in this rat model, and could be linked to hypertension¹⁷.

In a similar study, Penna et al. reported that a high-sodium diet (8% NaCl) increased angiotensin II, TGF-β1, and α-SMA, and decreased AQP-1, AQP-2, and eNOX expression in male Sprague Dawley rats¹⁵. In addition, losartan (40 mg/kg/day), an AT1 receptor blocker, prevented this effect. Unfortunately, the relationship between urine AQP2 and serum AVP during a high salt diet or hypertonic saline infusion is also unclear. The changes in urine AQP2 rate and AVP in healthy humans were positively correlated du-

ring water deprivation and hypertonic saline infusion¹⁹. In contrast, Saito et al. reported that a 5% saline infusion in five young controls resulted in an elevation of plasma AVP (1.0 ±0.2 to 4.8±0.7 pg/mL) due to an increase in plasma osmolality, and a decrease in urine volume and urinary AQP2²¹. Elliot et al. reported different results from previous studies, and found that urine volume increased from 76±15 to 156±29 mL/h in 90 min after a 3% NaCl infusion in healthy volunteers²². The significant discrepancy between these studies raises several questions namely, why does urinary AQP2 decrease while plasma AVP increases, and why does urine volume decrease while urinary AQP2 decreases? Although the exact mechanisms are still unclear, there are several possible explanations. The studies mentioned here have been conducted in multiple different species, including various kinds of rats and humans. The basic mechanism for absorption and excretion of sodium and water is relatively similar between these species, but there are some important differences. Furthermore, the response of AQP2 after water loading or deprivation differs by time. For example, urinary AQP2 levels are lowest two hours after water loading, and then increase for up to four hours, although absolute levels still remain lower than baseline¹². Furthermore, no studies have been conducted in children. In addition, and most importantly, the amount of water given can significantly affect the results because both plasma and urine water/sodium concentration (osmolality) are significantly influenced by the regulation of AVP and AQPs. The plasma concentration of AVP in rats may have decreased because they could freely access water. AQP concentration could have decreased due to an excess of water, and because plasma osmolality was increased by the hypertonic saline infusion and/or high salt diet in the human study. AQP2 levels and renal absorption may be increased to reduce plasma concentration. Therefore, different conditions after a high salt diet can variably influence urine volume and AQP. As previously mentioned, we have to consider the limitations of urine AQP2 sampling in the evaluation of renal AQP2. Finally, many other molecules, including angiotensin II/aldosterone, ENaC, and AQP1, also play important roles in the control of sodium, water and blood pressure.

Effect of Fasting

The renal response to fasting differs from that of water deprivation. Although the effects of acute fasting have not been reported, fasting for 24 hours reduced urine AQP2 (14%) and increased urine output²³. Because fasting increases prostaglandin synthesis, and because the inhibition of prostaglandin can increase the antidiuretic activity of vasopressin, many studies have been conducted using NSAIDs. However, study results have shown inconsistent effects of prostacyclin and NSAIDs on AQP2^{24,25}.

Effect of Low Potassium Diet

The kidney and muscle have a role in compensation for acute effects of low potassium diet on urine volume and AQP2 levels. Amlal et al. reported that urine osmolality decreased in as little as 12 hours after beginning a potassium free diet, and that urine volume increased after 24 hours²⁶. Interestingly, whereas an early urinary concentrating defect was induced from the down-regulation of cortical AQP2, later onset of the same defect resulted from decreased medullary AQP2²⁶. A long-term low potassium diet has been reported to lower urine osmolality by about 50%, to double urine excretion volume, and decrease AQP-2 levels by 50%^{26,27}.

Effect of High Protein Diet

High protein diets are included in this review because protein intake is a factor in osmotic diuresis. In addition to electrolytes, such as sodium and potassium, urea from a high protein diet and glucose are the main solutes responsible for osmotic diuresis^{28,29}. Total solute loss is higher than the loss of water in osmotic diuresis, and serum osmolality decreases²⁹. Previous studies have shown the effects of high and low-protein diets on the expression of AQP2³⁰⁻³². Urine concentration decreased after a low-protein diet in both humans and rats. A decrease in AQP2 protein expression was also accompanied by reduced urine concentrating ability^{30,31}. In contrast, a high-protein diet led to increased absorption of water and increased urine AQP2 (92.5 to

164.0 ng/ μ mol creatinine), but there was no change in urine volume between a normal (2,423 mL/24 h) and protein enriched diet (2,376 mL/24 h)³². Urine sodium excretion increased from 52 on a normal diet to 75 mmol/24 h on a high protein diet³². Urine prostaglandin levels were also reduced by a high protein diet (690 to 543 pg/min); however, while AVP and ANGII up-regulate AQP2, prostaglandin downregulates AQP2 expression³².

Effect of Hypercalciuria

Hypercalciuria is one of the causes of voiding dysfunction in children, including nocturnal enuresis and daytime frequency syndrome. The effect of hypercalciuria on AQP2 was investigated in 80 children with NE and 9 controls, 24 hour urine (day and night) was collected in control and NE groups³³. The NE subjects were divided into three groups (G1: low vasopressin levels and nighttime diuresis, G2: low vasopressin levels and balanced diuresis, G3: normal vasopressin and daytime diuresis). The results showed that the day/night AQP2 ratio was approximately three-fold higher in G1 subjects who displayed hypercalciuria (1.67 ± 0.41) compared to healthy children (0.59 ± 0.11), and the ratio was approximately two-fold higher in normocalciuric patients with NE (1.27 ± 0.24)³³. A study was recently published on how NE can be improved through a low calcium diet using vasopressin in children with NE and hypercalciuria³⁴. The same research group recently reported that there is a relationship between urinary calcium excretion and AQP2³⁵. Hypercalciuria was induced by bone demineralization for 7 days of adaptation and 24 days of bed rest in 10 healthy men³⁵. Urinary calcium excretion increased in the first 7 days, and then gradually decreased over 35 days. AQP2 excretion increased in the first 7 days, decreased through day 14, and then increased again until day 35³⁵. One possible explanation for this result is that the increased urinary calcium excretion may temporarily down-regulate AQP2, and since plasma volume is reduced by increased water excretion, AQP2 levels recover after calcium excretion is normalized³⁵.

Conclusion

The expression of renal AQP2 is mainly regulated by the absorption and excretion of sodium and water. Plasma concentration of other electrolytes, such as potassium and calcium, and nutrients, including protein, ingested through the diet can also regulate AQP2 concentration. Other regulatory factors, including AVP, PGE2, ENaC, and renin/ANGII/aldosterone, are also involved in this regulatory process. It is relatively clear that AQP2 is upregulated by water loading and high protein diet intake and down regulated by water restriction, low protein diet, low K diet and fasting. AQP2 is also down regulated by hypercalciuria (Fig. 3). However, the regulation of AQP2 by Na intake is contradictory as shown in table 1. The contradictory results may be caused by many factors, such as different experimental conditions, and compensatory mechanisms that regulate AQP2. Urine sampling of AQP2 in spot urine samples has also limited ability to reflect renal AQP2 concentration, and results differ significantly depending on the time to evaluation and duration of specific diets. Because drinking a significant amount of water can induce maximum diuresis after 2 hours, reduced water intake could greatly help to reduce nighttime diuresis. The effect of a high salt diet on AQP2 expression was shown to be inconsistent in previous studies, and therefore requires further investigation. If amount of protein intake is not significant such as parenteral nutrition and/or combined with other solute and/or water diuresis, because increased AQP2 after high protein diet can increase water absorption, it may not be an important cause for diuresis in children with NE. Hypercalciuria can reduce AQP2 and increase urine volume, so reducing intake of calcium rich foods may also help reduce nighttime diuresis.

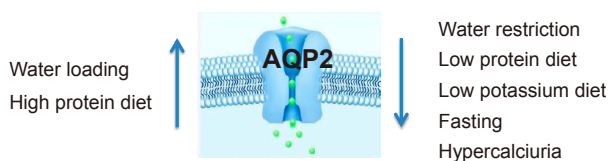


Fig. 3. Effects of various diets on expression of AQP2.

References

1. Preston GM, Carroll TP, Guggino WB, Agre P. Appearance of water channels in *Xenopus* oocytes expressing red cell CHIP28 protein. *Science* 1992;256:385-7.
2. Kortenoeven ML, Fenton RA. Renal aquaporins and water balance disorders. *Biochim Biophys Acta* 2014;1840:1533-49.
3. Nielsen S, Frøkiaer J, Marples D, Kwon TH, Agre P, Knepper MA. Aquaporins in the kidney: from molecules to medicine. *Physiol Rev* 2002;82:205-44.
4. Matsuzaki T, Yaguchi T, Shimizu K, Kita A, Ishibashi K, Takata K. The distribution and function of aquaporins in the kidney: resolved and unresolved questions. *Anat Sci Int* 2016 Jan 21. [Epub ahead of print].
5. Stanton BA, Koeppe BM (2004) Solute and water transport along the nephron: tubular function. In: Bern RM, Levy MN, Koeppe BM, Stanton BA (eds) *Physiology*. Elsevier Mosby, St. Louis, pp 643-58.
6. Tamma G, Goswami N, Reichmuth J, De Santo NG, Valenti G. Aquaporins, vasopressin, and aging: current perspectives. *Endocrinology* 2015;156:777-88.
7. Graugaard-Jensen C, Hvistendahl GM, Frøkiaer J, Bie P, Djurhuus JC. Urinary concentration does not exclusively rely on plasma vasopressin. A study between genders. *Gender and diurnal urine regulation. Acta Physiol (Oxf)* 2014;212:97-105.
8. Juul KV, Klein BM, Sandström R, Erichsen L, Nørgaard JP. Gender difference in antidiuretic response to desmopressin. *Am J Physiol Renal Physiol* 2011;300:F1116-22.
9. Preisser L, Teillet L, Aliotti S, Gobin R, Berthonaud V, Chevalier J, et al. Downregulation of aquaporin-2 and -3 in aging kidney is independent of V(2) vasopressin receptor. *American journal of physiology Renal physiology* 2000;279:F144-52.
10. Hofman MA, Swaab DF. Alterations in circadian rhythmicity of the vasopressin-producing neurons of the human suprachiasmatic nucleus (SCN) with aging. *Brain research* 1994;651:134-42.
11. Stachenfeld NS, Splenser AE, Calzone WL, Taylor MP, Keefe DL. Sex differences in osmotic regulation of AVP and renal sodium handling. *Journal of applied physiology* 2001;91:1893-901.
12. Umenishi F, Summer SN, Cadnapaphornchai M, Schrier RW. Comparison of three methods to quantify urinary aquaporin-2 protein. *Kidney International* 2002;62:2288-93.
13. Baumgarten R, van de Pol MH, Deen PM, van Os CH, Wetzels JF. Dissociation between urine osmolality and urinary excretion of aquaporin-2 in healthy volunteers. *Nephrology, dialysis, transplantation : official publication of the European Dialysis and Transplant Association - European Renal Association* 2000;15:1155-61.
14. Buemi M, Bolignano D, Coppolino G, Di Pasquale G, Cosentini V, Campo S, et al. Aquaporin-2 (AQP2) urinary excretion and assumption of water with different mineral content in healthy subjects. *Ren Fail* 2007;29:567-72.

15. Della Penna SL, Cao G, Fellet A, Balaszczuk AM, Zotta E, Cerrudo C, et al. Salt-induced downregulation of renal aquaporins is prevented by losartan. *Regul Pept* 2012;177:85-91.
16. Graffe CC, Bech JN, Pedersen EB. Effect of high and low sodium intake on urinary aquaporin-2 excretion in healthy humans. *Am J Physiol Renal Physiol* 2012;302:F264-75.
17. Roxas B, Farjah M, Danziger RS. Aquaporin-2 transcript is differentially regulated by dietary salt in Sprague-Dawley and Dahl SS/Jr rats. *Biochem Biophys Res Commun* 2002;296:755-8.
18. Rai T, Sekine K, Kanno K, Hata K, Miura M, Mizushima A, et al. Urinary excretion of aquaporin-2 water channel protein in human and rat. *J Am Soc Nephrol* 1997;8:1357-62.
19. Pedersen RS, Bentzen H, Bech JN, Pedersen EB. Effect of water deprivation and hypertonic saline infusion on urinary AQP2 excretion in healthy humans. *American journal of physiology Renal Physiology* 2001;280:F860-7.
20. Song J, Hu X, Shi M, Knepper MA, Ecelbarger CA. Effects of dietary fat, NaCl, and fructose on renal sodium and water transporter abundances and systemic blood pressure. *Am J Physiol Renal Physiol* 2004;287:F1204-12.
21. Saito T, Ishikawa SE, Sasaki S, Nakamura T, Rokkaku K, Kawakami A, et al. Urinary excretion of aquaporin-2 in the diagnosis of central diabetes insipidus. *The Journal of Clinical Endocrinology and Metabolism* 1997;82:1823-7.
22. Elliot S, Goldsmith P, Knepper M, Haughey M, Olson B. Urinary excretion of aquaporin-2 in humans: a potential marker of collecting duct responsiveness to vasopressin. *Journal of the American Society of Nephrology: JASN* 1996;7:403-9.
23. Starklint J, Bech JN, Pedersen EB. Down-regulation of urinary AQP2 and unaffected response to hypertonic saline after 24 hours of fasting in humans. *Kidney Int* 2005;67:1010-8.
24. Buemi M, Di Pasquale G, Ruello A, Floccari F, Aloisi C, Latassa G, et al. Effect of a prostacyclin analogue, iloprost, on urinary aquaporin-2 excretion in humans. *Nephron* 2002;91:197-202.
25. Kim SW, Kim JW, Choi KC, Ma SK, Oh Y, Jung JY, et al. Indomethacin enhances shuttling of aquaporin-2 despite decreased abundance in rat kidney. *Journal of the American Society of Nephrology: JASN* 2004;15:2998-3005.
26. Amlal H, Krane CM, Chen Q, Soleimani M. Early polyuria and urinary concentrating defect in potassium deprivation. *American Journal of Physiology Renal Physiology* 2000;279:F655-63.
27. Nguyen MT, Yang LE, Fletcher NK, Lee DH, Kocinsky H, Bachmann S, et al. Effects of K⁺-deficient diets with and without NaCl supplementation on Na⁺, K⁺, and H₂O transporters' abundance along the nephron. *American Journal of Physiology Renal Physiology* 2012;303:F92-104.
28. Bhasin B, Velez JC. Evaluation of Polyuria: The Roles of Solute Loading and Water Diuresis. *American journal of kidney diseases : the official journal of the National Kidney Foundation* 2016;67: 507-11.
29. Popli S, Tzamaloukas AH, Ing TS. Osmotic diuresis-induced hypernatremia: better explained by solute-free water clearance or electrolyte-free water clearance? *International Urology and Nephrology* 2014;46:207-10.
30. Sands JM, Naruse M, Jacobs JD, Wilcox JN, Klein JD. Changes in aquaporin-2 protein contribute to the urine concentrating defect in rats fed a low-protein diet. *The Journal of Clinical Investigation* 1996;97:2807-14.
31. Elfers K, Breves G, Muscher-Banse AS. Modulation of aquaporin 2 expression in the kidney of young goats by changes in nitrogen intake. *Journal of comparative physiology B, Biochemical, systemic, and environmental physiology* 2014;184:929-36.
32. Lauridsen TG, Vase H, Starklint J, Bech JN, Pedersen EB. Protein-enriched diet increases water absorption via the aquaporin-2 water channels in healthy humans. *Nephrology, dialysis, transplantation : official publication of the European Dialysis and Transplant Association - European Renal Association* 2010;25: 2502-10.
33. Valenti G, Laera A, Pace G, Aceto G, Lospalluti ML, Penza R, et al. Urinary aquaporin 2 and calciuria correlate with the severity of enuresis in children. *Journal of the American Society of Nephrology: JASN* 2000;11:1873-81.
34. Valenti G, Laera A, Gouraud S, Pace G, Aceto G, Penza R, et al. Low-calcium diet in hypercalciuric enuretic children restores AQP2 excretion and improves clinical symptoms. *American Journal of Physiology Renal physiology* 2002;283:F895-903.
35. Tamma G, Di Mise A, Ranieri M, Svelto M, Pisot R, Bilancio G, et al. A decrease in aquaporin 2 excretion is associated with bed rest induced high calciuria. *Journal of Translational Medicine* 2014; 12:133.