

## Loop-mediated Isothermal Amplification Assay to Rapidly Detect *Wheat Streak Mosaic Virus* in Quarantined Plants

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**We developed a loop-mediated isothermal amplification (LAMP) method to rapidly diagnose *Wheat streak mosaic virus* (WSMV) during quarantine inspections of imported wheat, corn, oats, and millet. The LAMP method was developed as a plant quarantine inspection method for the first time, and its simplicity, quickness, specificity and sensitivity were verified compared to current reverse transcription-polymerase chain reaction (RT-PCR) and nested PCR quarantine methods. We were able to quickly screen for WSMV at quarantine sites with many test samples; thus, this method is expected to contribute to plant quarantine inspections.**

**Keywords :** LAMP, loop-mediated isothermal amplification, quarantine, wheat streak mosaic virus, WSMV

*Wheat streak mosaic virus* (WSMV) is an unreported virus in Korea (Animal, Plant and Fisheries Quarantine and Inspection Agency, 2013) and quarantine has raised the possibility that WSMV can cause serious food loss and economic damage (French and Robertson, 1994) by infecting wheat, corn, oats, and millet (Lee et al., 2013a). Quarantine inspections have used reverse transcription-polymerase chain reaction (RT-PCR) and nested PCR to detect pathogens since 2012 in Korea (Lee, 2013; Lee et al., 2013b; Lee et al., 2013c). However, these methods require 10 hrs for results, so they are inconvenient for field

use where rapid inspection and subsequent quarantine are needed. Therefore, in this study, we developed a LAMP assay as an inspection method to detect WSMV in a one-step quick screen during quarantine inspection.

Samples of WSMV and reference viruses [*Cereal chlorotic mottle virus* (CCMV) and *Cucumber mosaic virus* (CMV)] were collected through import approval of prohibited goods. Nucleic acids were extracted and cDNA was synthesized using a method reported previously (Lee and Shin, 2014; Lee et al., 2014; Lee et al., 2015; Shin and Rho, 2014). RNA of 170–200 ng/μl was extracted from the samples, and 100 μl cDNA was synthesized from the RNA.

The WSMA complete genome (NC\_001886), 15 WSMV strains and their base sequences, and the base sequences of 20 reference virus strains that could infect the same host were collected from the National Center for Biotechnology Information. Two sets of WSMV-specific LAMP primers were designed based on the base sequences collected using the LAMP primer designing software PrimerExplorer (Table 1).

Three WSMV template cDNAs were chosen and reacted for 1 hr at three different temperatures (60, 62, and 65°C) to select the best LAMP conditions for detecting WSMV after 10 min at 95°C and 1 min at 4°C. As a result, condition 3 using an 11 μl total volume and no distilled water showed the most specific reaction. The analysis indicated that the specific reaction occurred at all temperatures, so the most stable conditions at 62°C were selected (Fig. 1). In addition, set 1 of the cDNAs for the CCMV and CMV reference viruses indicated nonspecific amplification for both the reference virus and the negative control, so it finally selected as the LAMP primer combination for detecting WSMV (Supplementary Fig. 1).

The LAMP method developed here can detect WSMV in imported samples within 1.5 hrs (30 min for cDNA synthesis and 60 min for LAMP) after RNA extraction. Thus,

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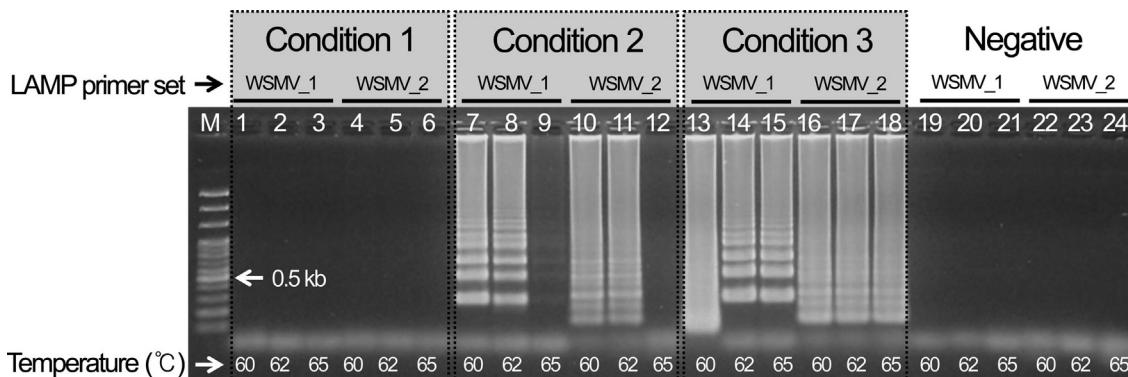
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**Table 1.** Loop-mediated isothermal amplification primer sequence of the amplified poly-protein coding gene to rapidly detect *Wheat streak mosaic virus*

Set	Primer	Sequence (5'→3')	Length (nt)	G+C (%)	Tm (°C)
WSMV_1	F3	GAGTTGGAGGAAAGGAAA	18	44.4	49.0
	B3	TCCGAAAACGGTAGAAAG	18	44.4	49.3
	FIP	GGACCAGAATATGTGAGTCGTATCTTCACACCGCAGAGT	39	-	-
	BIP	AGGAAGGTTACTCACCTTGCAGTATAACTGCAAGACG	41	-	-
WSMV_2	F3	TACGACTCACATATTCTGGT	20	40.0	50.2
	B3	GCCTATGAGTATAACTTGCA	20	40.0	49.4
	FIP	TCGGTCCTCGGAGATAGCGGAAGAAAGGTTGGAGAA	36	-	-
	BIP	ACAAAGGGAGTCCTGAGGATTTCCGAAAGGTGAGTAA	41	-	-

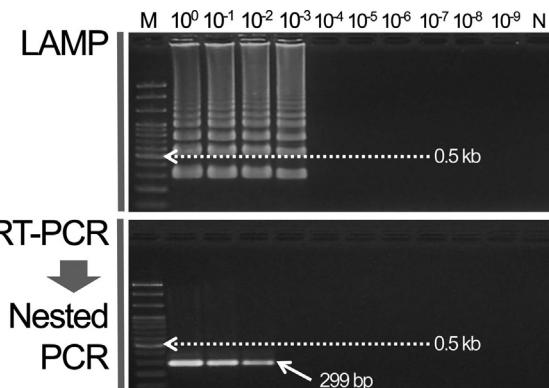


**Fig. 1.** Optimal selection of LAMP conditions for detecting *Wheat streak mosaic virus*.

this method reduces the time to detect WSMV from the previous 10 hrs using the RT-PCR, electrophoresis, nested PCR, and additional electrophoresis steps. Furthermore, it is much simpler and easier to use as the standard quarantine method. Additionally, specificity improved compared to that of the PCR method.

The WSMV cDNA template for LAMP was diluted up to  $10^{-9}$ . The RT-PCR primer combination [forward: 5'-TGG CGA TGA AGA TGT CAG-3', reverse: 5'-CCA TTT CTG TGA AGG CTT T-3' (834 bp)] was used previously to diagnose WSMV. The PCR template result was used for the nested PCR reaction to amplify the specific 299 bp gene (Lee et al., 2013a). The detection sensitivities of the two methods were compared. As a result, a specific band was formed at up to a  $10^{-2}$  dilution in the nested PCR but the specific ladder from the LAMP assay was analyzed at up to a  $10^{-3}$  dilution (Fig. 2).

After LAMP assay, the prepared 10  $\mu$ l of LAMP products were digested with 10 U of restriction enzyme *Bfa*I (New England Biolabs, US) at 37°C for 2 hrs, then they were subjected to electrophoresis in a 1.5% agarose



**Fig. 2.** Compare sensitivity of LAMP and nested PCR assay for detecting *Wheat streak mosaic virus*.

gel and visualized (digestion fragments [194 + 156 bp]) (Supplementary Fig. 2).

The LAMP assay was applied to quickly screen for WSMV during quarantine inspection of imported wheat, corn, oats, and millet. Instead of using Taq DNA polymerase, LAMP amplifies denatures, anneals, and extends at

**Table 2.** Optimal loop-mediated isothermal amplification conditions

Components	Condition (μl)		
	1	2	3
Template	1.5	1.5	1.5
Bst. 10× reaction buffer	2	2	2
dNTP (10 mM)	2	2	2
Primers			
F3 (10 pmol)	0.6	0.6	0.6
B3 (10 pmol)	0.6	0.6	0.6
FIP (10 pmol)	1.4	1.4	1.4
BIP (10 pmol)	1.4	1.4	1.4
Bst. polymerase (12 U)	1.5	1.5	1.5
ddH <sub>2</sub> O	9	4	0
Total (μl)	20	15	11

the same temperature by using Bst polymerase, resulting in quicker detection than that of the existing PCR method. In addition, the 5'→3' exonuclease feature provides high detection sensitivity and a quick result (Ahn et al., 2010; Cho et al., 2013). The LAMP developed in this study is simple and more specific rather than the current two-step quarantine methods using RT-PCR and nested PCR and reduced processing time by approximately 8 hrs. Furthermore, the LAMP method had 10 fold higher detection sensitivity than the current two-step PCR method. We expect that this LAMP method will be applied for use during quarantine inspections in the future.

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