

# **Diagnosis of Trace Toxic Uranium Ions in Organic Liver Cell**

Suw Young Ly<sup>1</sup>, Eun Chul Pack<sup>2</sup> and Dal Woong Choi<sup>2</sup>

<sup>1</sup>Biosensor Biosensor Research Institute, Seoul National University of Technology, Seoul, Korea <sup>2</sup>Department of Public Health Science, Graduate School, Korea University, Seoul, Korea

(Received March 28, 2014; Revised June 17, 2014; Accepted June 30, 2014)

Uranium is toxic and radioactive traces of it can be found in natural water and soils. High concentrations of it in biological systems cause genetic disorders and diseases. For the *in vivo* diagnosis, micro and nano range detection limits are required. Here, an electrochemical assay for trace toxic uranium was searched using stripping voltammetry. Renewable and simplified graphite pencils electrode (PE) was used in a three-electrode cell system. Seawater was used instead of an electrolyte solution. This setup can yield good results and the detection limit was attained to be at 10  $\mu$ gL<sup>-1</sup>. The developed skill can be applied to organic liver cell.

Key words: Uranium, Voltammetry, Fish's liver, Fluorine

## INTRODUCTION

Uranium presence in nature is widespread, as it is found in minute quantities in all rocks, sand, soil (1) and in animal internal organs (2). As it is radioactive and toxic (3,4), its presence in the environment may affect human health and cause medical disorders such as chemical kidney toxicity, functional and histological damages (5), cancer (6) and sterility (7) and can even affect human body systems. There is therefore a need for a sensitive, accurate and reproducible method that can be used on real samples in the biological, environmental and geological fields (8). For detection purposes, several methods have been carried out, such as kinetic phosphorescence analysis (9), multi-group analysis (10), inductively coupled plasma mass spectrometry (11), extraction chromatography (12), alpha particle spectrometry (13), absorption spectrum analysis (14-16), discharge mass spectrometry (17-19) and electrochemical polarographic (20) mercury (21,22) dropping (23,24) electrode. Herein photometric or mass spectrometric methods are demand of spectric light separation, electric acceleration and high energy pumping techniques are required. Moreover complicated electric mercury electrode is toxic in most voltammetric studies and not usable for *in vivo* or *in vitro* low concentration ranges. In this study, a simple and reasonable fluorine-doped pencil sensor was used as a working electrode. The said sensor does not need much preparation time and is nontoxic. Moreover, a reference and an auxiliary electrode were used with graphite pencils unlike other voltammetric methods. Also, deep seawater was used as the electrolyte solution. With the use of pencil electrodes and clean sea water solution, the experiment's expense was kept at a minimum. Reasonable techniques were applied to biological organic cell systems for medical science.

#### **MATERIALS AND METHODS**

The experiment was carried out using Bioelectronics-2 circuit, which was constructed by the authors' institution. The second version of this device was fabricated as a computerized handheld voltammetric system whose size is similar to that of a typical cellular phone. It can be used for bio assay and sensor techniques for individual and laboratory applications. The fluorine-doped graphite electrode (FPE) was prepared by coating the pencil with fluorine. Also, two pencils served as the Ag/AgCl reference and platinum counterelectrode, respectively. The supporting electrolyte was prepared with clean deep seawater. All the other reagents were of analytical grade. Electrolyte voltammetry was carried out on an open circuit. A uranium standard was obtained from Merck. Highly purified water was prepared through threetime distillation using 18 Mcm<sup>-1</sup> of Milli-Q Ultra-Pure Water System (Millipore, Bedford, USA). Fluorine immobilization was performed using a 10-cycle scan with a 1.0 V initial

Correspondence to: Dal Woong Choi, Department of Public Health Science, Graduate School, College of Health Science, Korea University, Sungbuk-gu, Seoul 136-704, Korea E-mail: dwlove@korea.ac.kr

This is an Open-Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http:// creativecommons.org/licenses/by-nc/3.0) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

potential, a 1.0 V switching potential, and a 0.5 Vs-1 scan rate. The SW stripping voltammogram used optimized parameters. All experiments were performed at room temperature and without oxygen removal.

## **RESULTS AND DISCUSSION**

**Cyclic voltammetry (CV) and stripping voltammetry using FPE.** In this study, clean seawater was used as the electrolyte solution, and the working, reference and auxiliary electrodes were made from inexpensive graphite PE. First, cyclic voltammetry and stripping voltammetry were examined using FPE in high concentration. Fig. 1A shows the cyclic scan in the range of 0, 10, 20, 30, 40, 50, 60, 70 and 80 mgL<sup>-1</sup> U variations, using -2.0 V initial potential, -2.0 V switching potential, 0.1 V/s scan rate. In the blank solution, the peak current was clear. By spiking U variations, it began to increase for a -0.2 V peak potential. It con-



**Fig. 1.** Voltammetric concentration effect. (A): the CV effect using FPE ranging from 10 to  $80 \text{ mgL}^{-1}$  U add. (B): stripping voltammetric concentration effects from 10 to  $80 \text{ mgL}^{-1}$  U using sea water electrolyte as a reference and an auxiliary PE.

tinually increased from  $6.545 \times 10^{-5}$  to  $24.7 \times 10^{-5}$  A. The inset figure shows that the linear equation was  $y = 0.247x + 10^{-5}$ 



**Fig. 2.** Low concentration effect. (A): the microgram concentration ranges from 100 to  $800 \ \mu g L^{-1}$  U using CV. (B): the SW results using FPE ranging from 1 to  $8 \ m g L^{-1}$ , using 30 sec accumulation time. (C): the interference by other metal and analogy ions for Fe, Zn, Cr, Al, Ge, Cd, Pb and Cu based on optimized conditions.

7.460 and precision was  $R^2 = 0.897$ , which can be usable for environmental applications. However, Fig. 1B shows the SW effect of ranges from 0 to 80 mgL<sup>-1</sup> variations. No signal was obtained from the blank solution. The first curve shows the result from the blank solution. The peak current was sharp and sensitive as the concentration of U spiked in order, increasing to  $32.05 \times 10^{-6}$  A. These results show that stripping voltammetry is useful in the detection of U concentration. Thus, optimum SW conditions were established.

Square wave (SW) stripping voltammetry. The low concentration of U was also examined using CV and SW. Fig. 2A shows the cyclic ionic activity in the range of 100, 200, 300, 400, 500, 600, 700 and 800  $\mu g L^{-1} \ U$  concentration. When 100  $\mu$ gL<sup>-1</sup> U was spiked in the clean seawater, it obtained peak currents for  $3.402 \times 10^{-5}$  A, after which the peak current started to increase from  $3.402 \times 10^{-5}$  A to  $7.341 \times 10^{-5}$  A without a decline, with y = 0.0056x + 2.6587, statistics of  $R^2 = 0.9927$ . Then Fig. 2B shows the SW scan ranging from 1 to 8 mgL<sup>-1</sup> U concentration. The greater the concentration of U was spiked, the more sensitive peak current was obtained. It increased from  $2.557 \times 10^{-5}$  A to  $10.37 \times 10^{-5}$  A. The equation of the SW result was y = 1.184x + 1.000 and the precision was  $R^2 = 0.986$  can be useful for trace detection. Fig. 2C shows interference by other ions. Various analytical interference ions were examined by adding other metals and analog ions into the medium containing 10 mgL<sup>-1</sup> of Uranium, for the tenfold spiking of 100 mgL<sup>-1</sup> of Fe, Zn, Cr, Al, Ge, Cd, Pb and Cu ion. This yielded results of 100%, 455.03%, -68.28%, 23.33%, -13.06%, -100.00%, 0.00% 0.56% and 0.57%, respectively. The analytical interferences were effectively corrected using standard addition methods.

Statistics and application. Fig. 3A, showing the statistics for sea water blank, and 10 mgL<sup>-1</sup> U constant was examined for 15 times. Here, standard deviation was 1.9158 (blank) and 1.3889 (10 mg/L U spike). Both are stable and sensitive. After that, more sensitive working range was examined using 200 sec accumulation time. Fig. 3B shows the SW effects in the microgram variations. The concentration of 0, 10, 20, 30, 40, 50, 60, 70 and 80  $\mu/L$  U was spiked and it increased from  $4.954 \times 10^{-6}$  A to  $15.95 \times$  $10^{-6}$  A. The linear equation was y = 0.166x + 3.321 and the precision was  $R^2 = 0.980$ . Despite micro ranges, SW using FPE was sensitive and effective in trace analysis. The analytical application was performed under such conditions for in vivo or ex vivo analysis. Fig. 3C shows the SW results of the application to fish liver. Standard addition method was used. The fish liver was dissolved in 1ml nitrate and this solution was diluted with 10ml distilled water. 1 ml solution was spiked and examined using SW with FPE. It obtained  $1.717 \times 10^{-6}$  A. After that, 2, 3 and 4 ml U standard was spiked, obtaining 2.945, 3.456,  $3.75 \times 10^{-6}$  A, individually.



**Fig. 3.** Statistics and application. (A): the statistics for electrolyte blank (- $\bigcirc$ - × 10<sup>-8</sup> A) and 10 mg/L U constant (- $\bigcirc$ - × 10<sup>-6</sup> A) with 15th repetition using optimized parameters. (B): the SW working ranges from 10 to 80 µg/L variation. (C): the standard addition methods for fish liver using SW in the seawater blank, unknown fish liver and standard U was spiked three times.

These results were obtained for  $1.887 \,\mu\text{g/ml}$  U. These results show that the sensor can be used for diagnosis in the human body or in animals.

After the comparison of CV and SW, it was found that the SW was effective in detecting trace U ions in the milli or micro ranges. Under optimized conditions, the analytical detection limit was  $8.0 \ \mu g L^{-1}$ , this despite inexpensive and eco-friendly materials were used for the reference and auxiliary electrodes and the electrolyte solution. The working range was from 10 to  $80 \ \mu g/L$ . The developed sensor was applied to diagnostic liver assay in fish cells, and the results of the application shows that it can be used in fields that require the detection purposes at an affordable cost.

### ACKNOWLEDGEMENT

This study was supported by the Research Program funded by the Seoul National University of Science and Technology.

#### REFERENCES

- Jaspal, S., Lakhwant, S. and Sanjay, K. (2003) A comparison of fission track and laser fluorometry techniques for uranium analysis in water samples. *Radiat. Meas.*, 36, 517-519.
- Becker, J.S., Dobrowolska, J., Zoriy, M. and Matusch, A. (2008) Imaging of uranium on rat brain sections using laser ablation inductively coupled plasma mass spectrometry: a new tool for the study of critical substructures affined to heavy metals in tissues. *Rapid Commun. Mass Spectrom.*, 22, 2768-2772.
- Metilda, P., Prasad, K., Kala, R., Gladis, J.M., Rao, T.P. and Naidu, G.R. (2007) Ion imprinted polymer based sensor for monitoring toxic uranium in environmental samples. *Anal. Chim. Acta*, 582, 147-153.
- Bellis, D., Ma, R., Bramall, N., McLeod, C.W., Chapman, N. and Satake, K. (2001) Airborne uranium contamination - as revealed through elemental and isotopic analysis of tree bark. *Environ. Pollut.*, **114**, 383-387.
- Kurttio, P., Harmoinen, A., Saha, H., Salonen, L., Karpas, Z., Komulainen, H. and Auvinen, A. (2006) Kidney toxicity of ingested uranium from drinking water. *Am. J. Kidney Dis.*, 47, 972-982.
- Burkart, W., Danesi, P.R. and Hendry, J.H. (2005) Properties, use and health effects of depleted uranium. *Int. Congr. Ser.*, 1276, 133-136.
- Linares, V., Albina, M.L., Bellés, M., Mayayo, E., Sánchez, D.J. and Domingo, J.L. (2005) Combined action of uranium and stress in the rat. II. Effects on male reproduction. *Toxicol. Lett.*, **158**, 186-195.
- Hedaya, M.A., Birkenfeld, H.P. and Kathren, R.L. (1997) A sensitive method for the determination of uranium in biological samples utilizing kinetic phosphorescence analysis (KPA). *J. Pharm. Biomed.*, 15, 1157-1165.
- Ejnik, J.W., Hamilton, M.M., Adams, P.R. and Carmichael, A.J. (2000) Optimal sample preparation conditions for the determination of uranium in biological samples by kinetic phosphorescence analysis (KPA). J. Pharm. Biomed., 24, 227-

235.

- Yücel, H. (2007) The applicability of MGA method for depleted and natural uranium isotopic analysis in the presence of actinides (232Th, 237Np, 233Pa and 241Am). *Appl. Radiat. Isot.*, 65, 1269-1280.
- Degueldre, C., Favarger, P.Y., Rossé, R. and Wold, S. (2006) Uranium colloid analysis by single particle inductively coupled plasma-mass spectrometry. *Talanta*, 68, 623-628.
- Lee, M.H. and Lee, C.W. (2001) Radiochemical analysis of uranium isotopes in soil and sediment samples with extraction chromatography. *Talanta*, 54, 181-186.
- Rubio Montero, M.P., Martín Sánchez, A. and Carrasco Lourtau, A.M. (2004) Isotopic uranium and plutonium analysis by alpha-particle spectrometry. *Nucl. Instrum. Methods B*, 213, 429-433.
- Carriere, M., Gouget, B., Gallien, J.P., Avoscan, L., Gobin, R., Verbavatz, J.M. and Khodja, H. (2005) Cellular distribution of uranium after acute exposure of renal epithelial cells: SEM, TEM and nuclear microscopy analysis. *Nucl. Instrum. Methods B*, 231, 268-273.
- Meinrath, G. (1998) Chemometric analysis: Uranium (VI) hydrolysis by UV-Vis spectroscopy. J. Alloys Compd., 275, 777-781.
- Karbowiak, M., Droødøynski, J. and Gajek, Z. (2001) Absorption spectrum analysis of uranium trichloride heptahydrate. J. Alloys Compd., 323, 678-682.
- Pajo, L., Tamborini, G., Rasmussen, G., Mayer, K. and Koch, L. (2001) A novel isotope analysis of oxygen in uranium oxides: comparison of secondary ion mass spectrometry, glow discharge mass spectrometry and thermal ionization mass spectrometry. *Spectrochim. Acta B*, 56, 541-549.
- Wittmaack, K. (2008) SIMS analysis of xenon and krypton in uranium dioxide: A comparison of two models of gas-phase ionization. *Nucl. Instrum. Methods B*, 266, 5151-5158.
- Soldatov, A.V., Lamoen, D., Konstantinovic, M.J., Van den Berghe, S., Scheinost, A.C. and Verwerft, M. (2007) Local structure and oxidation state of uranium in some ternary oxides: X-ray absorption analysis. *J. Solid State Chem.*, 180, 54-61.
- Pournaghi, M.H. and Zargharian, R. (1996) Adsorptive pulse polarographic determination of uranium(V1) oxinate in chloroform and its use for the analysis of uranium mineral ores. *Anal. Chim. Acta*, **328**, 33-39.
- 21. Sander, S., Wagner, W. and Henze, G. (1997) Possibilities uranium for the automated determination of trace concentrations of in water samples by adsorptive stripping voltammetry. *Anal. Chim. Acta*, **349**, 93-99.
- Sander, S., Wagner, W. and Henze, G. (1995) Direct determination of uranium traces by adsorptive stripping voltammetry. *Anal. Chim. Acta*, 305, 154-158.
- Gholivand, M.B., Nassab, H.R. and Fazeli, H. (2005) Cathodic adsorptive stripping voltammetric determination of uranium (VI) complexed with 2, 6-pyridinedicarboxylic acid. *Talanta*, 65, 62-66.
- Farghaly, O.A. and Ghandour, M.A. (1999) Cathodic adsorptive stripping voltammetric determination of uranium with potassium hydrogen phthalate. *Talanta*, 49, 31-40.