## A New Furfural Diglycoside and Other Carbohydrate Derivatives from Fermented Beverage of *Prunus mume* Fruit

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Prunus mume Sieb. et Zucc. (Rosaceae) is a deciduous fruit tree that is distributed widely in East Asian. Its fruit, commonly known as Maesil in Korea, is a traditional raw ingredient used in herbal medicines and healthy foods in Korea, China, and Japan. In particular, a beverage produced by the fermentation of a mixture of green P. mume fruit and sugar is popular in Korea. Recent studies have shown that this fermented beverage, which contains a variety of probiotics, enhances immune activity and suppresses the development of atopic dermatitis-like skin lesions and 7,12-dimethylbenz[ $\alpha$ ]anthracene (DMBA)- and 12-O-tetradecanoyl phorbol-13-acetate (TPA)-induced skin carcinogenesis in mice.<sup>1-3</sup> Despite these findings, the chemical constituents of fermented P. mume beverage have yet to be characterized. In the present study, 19 compounds, including a new furfural diglycoside, 5-[ $\beta$ -D-fructopyranosyl-(2 $\rightarrow$ 6)- $\alpha$ -D-glucopyranosyloxymethyl]-2-furancarboxaldehyde (1), 16 carbohydrate derivatives (2-10 and 12-18), and two additional compounds (11 and 19) were isolated from samples of fermented P. *mume* fruit beverage. This is the first report detailing the chemical composition of fermented P. mume beverage and provides a useful reference for further biological investigation. This report describes the isolation and structural elucidation of compounds 1-19.

The known compounds that were isolated from the

fermented beverage of P. mume fruit were identified by comparisons of their physical and spectroscopic data with those reported in the literature. They were identified as 5hydroxymethyl-2-furfural (2),<sup>4</sup>  $\alpha$ -methoxy-2,5-furandimethanol (3),<sup>5</sup> benzyl- $\beta$ -D-glucopyranoside (4),<sup>6</sup> benzyl- $\beta$ -Dglucopyranosyl- $(1\rightarrow 6)$ - $\beta$ -D-glucopyranoside (5),<sup>7</sup> benzyl- $\beta$ -D-xylopyranosyl- $(1\rightarrow 6)$ - $\beta$ -D-glucopyranoside (6),<sup>8</sup> benzyl- $\alpha$ -L-arabinopyranosyl-(1 $\rightarrow$ 6)- $\beta$ -D-glucopyranoside (7),<sup>7</sup> benzyl- $\alpha$ -L-arabinofuranosyl- $(1\rightarrow 6)$ - $\beta$ -D-glucopyranoside (8),<sup>9</sup> (2*E*)-7-hydroxy-3,7-dimethyl-2-octenyl  $\alpha$ -L-arabinopyranosyl- $(1\rightarrow 6)$ - $\beta$ -D-glucopyranoside (9),<sup>10</sup> (2E)-7-hydroxy-3,7-dimethyl-2-octenyl  $\alpha$ -L-arabinofuranosyl-(1 $\rightarrow$ 6)- $\beta$ -D-glucopyranoside (10),<sup>11</sup> methyl (E)-p-coumarate (11),<sup>12</sup> 3-*O-p*-coumaroylsucrose (12),<sup>13</sup> 3-*O*-feruloylsucrose (13),<sup>14</sup> *n*butyl- $\beta$ -D-glucopyranoside (14),<sup>15</sup> sucrose (15),<sup>16</sup> methyl  $\beta$ -D-fructopyranoside (16),<sup>17</sup> D-glucopyranose (an equilibrium mixture of 64%  $\beta$ -D-glucopyranose and 36%  $\alpha$ -D-glucopyranose in solution, 17),<sup>18</sup> D-fructose (an equilibrium mixture of 70% &-D-fructopyranose and 22% &-D-fructofuranose in solution, 18),<sup>19</sup> and tyrosol  $(19)^{20}$  (Figure 1).

Compound 1 was obtained as a colorless syrup with a positive optical rotation of  $[\alpha]_D^{18}$  +47.2 (c = 1.0, MeOH). Its molecular formula was determined to be C<sub>18</sub>H<sub>26</sub>O<sub>13</sub> by high-resolution electrospray ionization mass spectrometry (HR-ESI-MS) at m/z 473.1295 [M+Na]<sup>+</sup> (calcd for C<sub>18</sub>H<sub>26</sub>O<sub>13</sub>Na,



Figure 1. Chemical structures of compounds 1-19.

Notes

 Table 1. NMR data (600/150 MHz) of compound 1 in CD<sub>3</sub>OD

$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Position	δ <sub>C</sub>	$\delta_{\rm H} (J \text{ in Hz})$	HMBC	COSY
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	1	179.7	9.56, s	C-2	
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2	154.4			
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	3	124.5	7.39, d (3.7)	C-2, 4, 5	H-4
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	4	113.2	6.72, d (3.7)	C-2, 3, 5	Н-3
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	5	159.7			
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	6	62.5	4.67, d (13.8)	C-4, 5, 1'	Η-6β
1'100.24.92, d (3.7)C-6, 2', 3'H-2'2'73.63.42, dd (10.1, 3.7)C-3'H-1', 3'3'75.03.64, mC-2', 4'H-2', 4'4'71.83.30, mC-2', 3', 6'H-3', 5'5'74.33.60, mC-4', 6'H-4', 6'6'62.73.66, mC-4', 5', 2"H-5', 6' $\beta$ 3.78, mC-4', 5', 2"H-5', 6' $\alpha$ 1"63.63.69, d (11.0)C-2", 3"H-1" $\beta$ 3.74, mC-2", 3"H-1" $\alpha$ 2"101.93"70.63.90, d (10.1)C-4"H-4"4"71.73.78, mC-3", 5", 6"H-3", H-5"5"71.33.84, br sC-3"H-4", H-6"6"65.33.66, mC-2", 4", 5"H-5", H-6" $\beta$ 3.75, mC-2", 4", 5"H-5", H-6" $\beta$			4.76, d (13.8)	C-4, 5, 1'	Η-6α
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	1'	100.2	4.92, d (3.7)	C-6, 2', 3'	H-2'
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	2'	73.6	3.42, dd (10.1, 3.7)	C-3'	H-1', 3'
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	3'	75.0	3.64, m	C-2', 4'	H-2', 4'
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	4'	71.8	3.30, m	C-2', 3', 6'	H-3', 5'
	5'	74.3	3.60, m	C-4', 6'	H-4', 6'
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	6'	62.7	3.66, m	C-4', 5', 2"	Η-5', 6'β
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$			3.78, m	C-4', 5', 2"	Η-5', 6'α
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	1"	63.6	3.69, d (11.0)	C-2", 3"	Η-1"β
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$			3.74, m	C-2", 3"	Η-1"α
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	2"	101.9			
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	3"	70.6	3.90, d (10.1)	C-4"	H-4"
5"       71.3       3.84, br s       C-3"       H-4", H-6"         6"       65.3       3.66, m       C-2", 4", 5"       H-5", H-6"β         3.75, m       C-2", 4", 5"       H-5", H-6"α	4"	71.7	3.78, m	C-3", 5", 6"	H-3", H-5"
6" 65.3 3.66, m C-2", 4", 5" H-5", H-6"β 3.75, m C-2", 4", 5" H-5", H-6"β	5"	71.3	3.84, br s	C-3"	H-4", H-6"
3.75, m C-2", 4", 5" H-5", H-6"α	6"	65.3	3.66, m	C-2", 4", 5"	H-5", H-6"β
			3.75, m	C-2", 4", 5"	H-5", H-6"α

473.1266). The IR spectrum contained clear absorption bands corresponding to hydroxyl groups (3362 cm<sup>-1</sup>), a conjugated aldehyde group (2927 cm<sup>-1</sup>), and conjugated double bonds (1666 cm<sup>-1</sup>). The <sup>1</sup>H-NMR spectrum of **1** (Table 1) contained signals corresponding to an aldehyde proton at  $\delta_{\rm H}$  9.56 (1H, s, H-1), two coupled olefinic protons at  $\delta_{\rm H}$  7.39 (1H, d, J = 3.7 Hz, H-3) and 6.72 (1H, d, J = 3.7 Hz, H-4), and two coupled oxygenated protons at  $\delta_{\rm H}$  4.76 and 4.67 (each 1H, d, J = 13.8 Hz, H-6 $\beta$  and 6 $\alpha$ ). The <sup>13</sup>C-NMR spectrum showed signals corresponding to a carbonyl carbon atom at  $\delta_{\rm C}$  179.7, two quarternary carbon atoms at  $\delta_{\rm C}$  159.7 and 154.4, two methine carbon atoms at  $\delta_{\rm C}$  124.5 and 113.2, and an oxygenated methylene carbon atom at  $\delta_{\rm C}$  62.5. These spectral data implied the presence of a 5-hydroxymethylfurfural moiety in the molecule of 1.4 The <sup>13</sup>C-NMR and DEPT spectra of 1 also revealed two glycosyl moieties with two anomeric carbon signals at  $\delta_{\rm C}$  101.9 and 100.2 and ten oxygenated carbon signals at  $\delta_{\rm C}$  62.7-75.0 including three methylene carbon atoms and seven methine carbon atoms, although only one anomeric proton signal was evident in the <sup>1</sup>H-NMR spectrum at  $\delta_{\rm H}$  4.92 (1H, d, J = 3.7 Hz). Analysis of COSY and HMBC data of the disaccharide moiety (Table 1 and Figure 2) suggested the presence of glucopyranosyl and fructopyranosyl units. This was confirmed by a comparison of the NMR data with previously published data.<sup>21-23</sup> The  $\alpha$ -configuration at the anomeric center of the glucopyranosyl unit is supported by a relatively small J value (J =3.7 Hz) and the  $\beta$ -configuration at the anomeric center of the fructopyranosyl unit is supported by a comparison of its  $\delta_{\rm C}$ values with those of  $\alpha$ - and  $\beta$ -configured methyl-D-fructo-



Figure 2. Key HMBC correlations of compound 1.

pyranoside.<sup>24</sup> Acid hydrolysis of **1** yielded D-glucose and D-fructose, which were identified by GC analysis. The HMBC spectrum showed a correlation between H-6' ( $\delta_{\rm H}$  3.66 and 3.78) and C-2" ( $\delta_{\rm C}$  101.9), indicating a 2" $\rightarrow$ 6' linkage in the disaccharide moiety. The linkage between this disaccharide and the 5-hydroxymethylfurfural moiety was determined by HMBC correlations between H-1' ( $\delta_{\rm H}$  4.92) and C-6 ( $\delta_{\rm C}$  62.5) and between H-6 ( $\delta_{\rm H}$  4.67 and 4.76) and C-1' ( $\delta_{\rm C}$  100.2). Thus, the compound **1** was identified as a new furfural diglycoside, 5-[ $\beta$ -D-fructopyranosyl-( $2\rightarrow$ 6)- $\alpha$ -D-glucopyranosyloxymethyl]-2-furancarboxaldehyde.

## Experimental

**Plant Materials.** Fresh green fruits of *P. mume* (10 kg) were purchased from local markets (Yuseong, Daejeon, Korea) in June 2012 and authenticated by one of the authors, Prof. Young Ho Kim. Thick, fermented *P. mume* beverage was obtained by mixing cleaned fruits of *P. mume* (10 kg) with sugar (7 kg) and storing the mixture in the dark at room temperature for 3 months. During this time, the mixture was stirred weekly. To obtain the final beverage, fermentation residue and seeds were filtered from the fermentation broth.

Extraction and Isolation. Thick fermented beverage of P. mume fruit (4.5 L) were firstly diluted with distilled water (10.5 L) to 15 L. The dilution was subjected to adsorptive macroporous resins HP-20 column chromatography (CC) and eluted successively with distilled water, 25% aqueous MeOH, 50% aqueous MeOH, 75% aqueous MeOH, and 100% MeOH to yield 5 fractions, respectively. The 25% MeOH elute (9.4 g) was further subjected to silica gel CC eluted with  $CH_2Cl_2/MeOH/H_2O$  (8:1:0.1 – 2:1:0.1) to afford 12 fractions (Frs. W1-W12). Fr. W1 was separated by repeated RP-C<sub>18</sub> CC (MeOH-H<sub>2</sub>O, 1:10) to yield 2 (11 mg) and 3 (5 mg). Fr. W4 was purified on a RP- $C_{18}$  column (Me<sub>2</sub>CO-H<sub>2</sub>O, 1:20) to yield 14 (110 mg). Fr. W5 was subjected to RP-C<sub>18</sub> CC (MeOH/H<sub>2</sub>O, 1:30) to yield three subfractions (Frs. W5-1-W5-3). Compound 1 (6 mg) was obtained by the separation of Fr. W5-2 on a silica gel column (EtOAc/MeOH, 10:1) and 16 (7 mg) was obtained by the separation of Fr. W5-3 on a silica gel column (CHCl<sub>3</sub>/94% EtOH, 5:2). 18 (426 mg), 17 (266 mg), and 15 (39 mg) were obtained by the separation of Frs. W9, W10, and W12 on a RP-C<sub>18</sub> column (MeOH/H<sub>2</sub>O, 1:25), respectively. The 50% MeOH elute (5.1 g) was subjected to silica gel CC eluted with  $CH_2Cl_2/MeOH/H_2O$  in a gradient (8:1:0.1 - 3:1:0.1) to afford 11 fractions (Frs. M1 - M11). 11 (3 mg) and 19 (5 mg) were obtained by the separation of Fr. M1 on a RP-C<sub>18</sub> column (MeOH/H<sub>2</sub>O, 1:15 - 1:5) and a silica gel column (CHCl<sub>3</sub>/94% EtOH, 30:1 - 10:1). Fr. M4 was subjected to RP-C<sub>18</sub> CC (Me<sub>2</sub>CO /H<sub>2</sub>O, 1:20) to yield **4** (200 mg). Fr. M7 was purified on a silica gel column (CHCl<sub>3</sub>/MeOH, 5:1) and a RP-C<sub>18</sub> column (MeOH/H<sub>2</sub>O, 1:4) to yield **8** (9 mg). Fr. M8 was subjected to silica gel CC (EtOAc/94% EtOH, 5:1 – 5:2) and RP-C<sub>18</sub> CC (MeOH/H<sub>2</sub>O, 1:12) to yield **12** (4 mg), **6** (105 mg), and **7** (135 mg). Fr. M9 was separated on a RP-C<sub>18</sub> column (MeOH/H<sub>2</sub>O, 1:7 – 1:2) to afford **10** (3 mg) and **9** (5 mg). Fr. M10 was subjected to RP-C<sub>18</sub> CC (MeOH/H<sub>2</sub>O, 0.2:1:10 – 0:1:3) and silica gel CC (CHCl<sub>3</sub>/94% EtOH, 4:1 – 3:2) to yield **5** (12 mg) and **13** (2 mg).

**5-**[β-D-Fructopyranosyl-(2→6)-α-D-glucopyranosyloxymethyl]-2-furancarboxaldehyde (1): Colorless syrup;  $[\alpha]_D^{18}$  +47.2 (c = 1.0, MeOH); UV (MeOH)  $\lambda_{max}$  (log $\varepsilon$ ): 278 nm (3.65); IR (KBr)  $\nu_{max}$ : 3362, 2927, 1666, 1027 cm<sup>-1</sup>; HR-ESI-MS (positive mode): m/z 473.1295 [M+Na]<sup>+</sup> (calcd for C<sub>18</sub>H<sub>26</sub>O<sub>13</sub>Na, 473.1266); <sup>1</sup>H- and <sup>13</sup>C-NMR (600/150 MHz, CD<sub>3</sub>OD) data, see Table 1.

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**Supporting Information.** General experimental procedures, hydrolysis procedure and NMR spectra of compound **1** are available as supporting information.

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