

The Molecular Functions of RalBP1 in Lung Cancer

Seunghyung Lee[†]

*Department of Anatomy & Cell Biology and The Sol Sherry Thrombosis Research Center
Temple University School of Medicine, Philadelphia, PA 19140, U.S.A.*

RalBP1 is an ATP-dependent non-ABC transporter, responsible for the major transport function in many cells including many cancer cell lines, causing efflux of glutathione-electrophile conjugates of both endogenous metabolites and environmental toxins. RalBP1 is expressed in most human tissues, and is over-expressed in non-small cell lung cancer cell lines and in many other tumor types. Blockade of RalBP1 by various approaches has been shown to increase sensitivity to radiation and chemotherapeutic drugs, leading to cell apoptosis. In xenograft tumor models in mice, RalBP1 blockade or depletion results in complete and sustained regression across many cancer cell types including lung cancer cells. In addition to its transport function, RalBP1 has many other cellular and physiological functions, based on its domain structure which includes a unique Ral-binding domain and a RhoGAP catalytic domain, as well as docking sites for multiple signaling proteins. Additionally, RalBP1 is also important for stromal cell function in tumors, as it was recently shown to be required for efficient endothelial cell function and angiogenesis in solid tumors. In this review, we discuss the cellular and physiological functions of RalBP1 in normal and lung cancer cells.

Key Words: RalBP1, Tumorigenesis, Lung cancer, R-Ras, RhoGAP

1. Introduction

2 Ral-binding protein 1 (RalBP1) has many cellular and physiological functions, based on its domain structure which includes a unique Ral-binding domain (RalBD) and a RhoGAP catalytic domain, as well as docking sites for multiple signaling proteins. As a Ral effector, RhoGAP, and adapter protein, RalBP1 has been shown to play important roles in endocytosis, mitochondrial fission, cell spreading and migration, actin dynamics during gastrulation, and Ras-induced tumorigenesis (Fig. 1). Also, RalBP1 is a non-ABC type transporter, which utilizes both of its two ATP binding sites (aa 69-74) and adjacent to the RalBD (aa 418-425) for

ATPase and transport activity (Awasthi et al., 2001).

RalBP1 is expressed in most human tissues including liver, heart, ovary, lung, muscle, and kidney as well in most human tumor cell lines, and plays a crucial role in cancer (Awasthi et al., 1994; Awasthi et al., 1991; Awasthi et al., 2008; Sharma et al., 1990). Also, RalBP1 is over-expressed in multiple cancers, such as lung and ovarian carcinomas and melanomas (Awasthi et al., 1994; Awasthi et al., 1991; Awasthi et al., 2008; Sharma et al., 1990). As a prominent cellular function of RalBP1 is export of chemotherapy agents, it is a major factor in the mechanisms of drug resistance. Moreover, blockade of RalBP1 with targeting antibodies or antisense has been shown to greatly increase sensitivity to radiation and chemotherapy and lead to pronounced tumor regression in multiple types of solid tumors in mice, including xenografted tumors of cancer cells (Singhal et al., 2006; Singhal et al., 2007, Singhal et al., 2009).

In 2010, the most recent year for which detailed lung cancer statistics are available, patient survival was 20~30%

*Received: May 12, 2014 / Revised: June 2, 2014

Accepted: June 2, 2014

[†]Corresponding author: Seunghyung Lee. Temple University School of Medicine 3420 N. Broad Street, MRB 200 Philadelphia, PA 19140.

Tel: +1-215-707-8015, Fax: +1-215-707-6499

e-mail: s.lee@temple.edu

©The Korean Society for Biomedical Laboratory Sciences. All rights reserved.

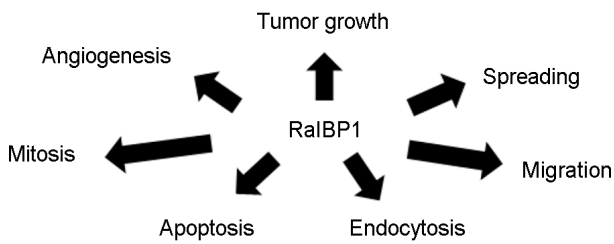


Fig. 1. RalBP1 cellular and physiological functions. RalBP1 regulates tumor growth, angiogenesis, cell spreading and migration, mitosis, apoptosis, and endocytosis in normal cells and tumor cells.

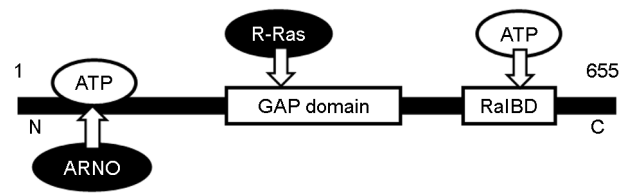


Fig. 2. The schematic diagram of RalBP1 structural domain. RalBP1 structural domains include GTPase-activating protein (GAP) domain and Ral-binding domain (RalBD), ATP binding sites, amino acid 65-80 or 415-448. RalBP1 interacts with ARNO and R-Ras through N-terminal and C-terminal regions, respectively.

(Jemal et al., 2010). Lung tumors have two major types: small cell and non-small cell lung cancer. Roughly 80~85% of lung cancers are non-small cell, and metastatic disease at presentation is common in these patients (Goldstraw et al., 2007; Riaz et al., 2012). However, response rates to chemotherapeutic regimens are low; thus, lung cancer continues to be a major cause of cancer mortality. This review will focus on the mechanisms and regulatory effects of RalBP1 in cancer and its specific roles in lung cancer.

2. RalBP1 protein

RalBP1 is a modular, 655 amino acid protein, harboring an N-terminal putative helical domain of poorly characterized function, a central RhoGAP domain on C-terminal, and a conserved RalBD near the C-terminus (Fig. 2). The RalBD, which bears no homology to classical Ras-binding domains, supports interaction with activated RalA and RalB but not with Ras small G proteins (Bauer et al., 1999; Cantor et al., 1995; Jullien-Flores et al., 1995). Like all Ras superfamily small G proteins, Ral proteins are signal transducers that become activated upon release of guanine diphosphate (GDP) and binding to guanine triphosphate (GTP), upon which Ral undergoes a conformational shift to expose high affinity binding sites for signaling effectors. Thus, RalBP1 is a unique Ral and Ras effectors, connecting upstream activation of Ral to downstream molecular and cellular events.

As shown in Fig. 3, RalBP1 is over-expressed in multiple cancers, such as lung and ovarian carcinomas and melanomas, and is expressed in most human tissues including liver,

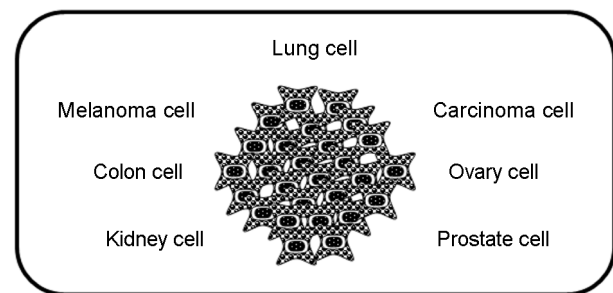


Fig. 3. RalBP1 over-expression in cancer cells. RalBP1 is over-expressed in lung, melanomas, carcinomas, colon, ovary, kidney, and prostate tumor cells.

heart, ovary, lung, muscle, and kidney as well in most human tumor cell lines (Awasthi et al., 1994; Awasthi et al., 2008; Awasthi et al., 1991; Sharma et al., 1990). As a prominent cellular function of RalBP1 is export of chemotherapy agents, it is a major factor in the mechanisms of drug resistance. Also, blockade of RalBP1 with targeting antibodies or antisense has been shown to greatly increase sensitivity to radiation and chemotherapy and lead to pronounced tumor regression in multiple types of solid tumors in mice, including xenografted tumors of lung cancer cells (Singhal et al., 2006; Singhal et al., 2007; Singhal et al., 2009).

3. RalBP1 molecular function

The Ral effector, RalBP1, regulates linking Ral to Rho GTPase pathways through the RhoGAP domain (Awasthi et al., 2000; Jullien-Flores et al., 1995; Park et al., 1995). The Rho subfamily of Ras small G proteins, RhoA, Rac

and Cdc42, are regulated by guanine nucleotide exchange. Signaling by these small G proteins leads to actin remodeling and altered cell morphologies, with Rac being associated with formation of broad lamellipodia protrusions and Cdc42 with filopodia spikes (Burrige et al., 1999). Since its initial characterization as a Ral effector and RhoGAP, RalBP1 has been implicated in various other cellular functions. Also, RLIP76 supports RhoGAP activity towards Rac and Cdc42 (Jullien-Flores et al., 1995), and Ral can be activated downstream of Ras, RalBP1 was proposed to bridge Ras activation. RLIP76 interaction with RalB modulates actin remodeling during gastrulation in *Xenopus* oocytes (Lebreton et al., 2004). Thus, RalBP1 cellular and physiological functions are important with Ral effector function for Rho signaling, actin cytoskeletal remodeling, and cell morphologies.

RalBP1 has implications for the potential ATP-binding site, RalBP1 functions as an ATP-dependent glutathione-conjugate transporter for small molecules (Awasthi et al., 2000), including anticancer drugs and endogenous metabolites (Awasthi et al., 2001; Awasthi et al., 2002; Awasthi et al., 2003), and in endocytosis (Nakashima et al., 1999; Rosse et al., 2003), mitochondrial fission (Kashatus et al., 2011), cell spreading and migration (Goldfinger et al., 2006), and Ras-induced tumorigenesis (Issaq et al., 2010; Lim et al., 2005). Identified binding partners (besides RalA and RalB) include AP2, POB1, HSF-1, R-Ras, and Repl1. Through these interactions, RalBP1 participates in various cellular functions by coupling related molecules in signaling pathways. RalBP1 regulates Ral signaling in the regulation of endocytic recycling of many proteins including growth factor receptors through binding to AP2 and POB1 (Jullien-Flores et al., 2000; Nakashima et al., 1999). During mitosis, RalBP1 is phosphorylated, distributes to centrosomes and the mitotic spindle, and interacts with POB1 and mitotic kinase cdk1, leading to Ral-dependent shut off of endocytosis during mitosis (Fillatre et al., 2012; Kariya et al., 2000; Rosse et al., 2003). Additionally, during mitosis RalBP1 also localizes to mitochondria where it couples Ral phosphorylation by mitotic kinase Aurora A to signaling necessary for mitochondrial fission and proper distribution in the daughter cells (Kashatus et al., 2011). Furthermore,

the role of RalBP1 in endocytosis may be coupled to its transport activity (Singhal et al., 2011). RalBP1 also appears to regulate stress-induced transcriptional activation by complexing with HSF-1 (Heat shock factor-1) (Hu et al., 2003). Moreover, RalBP1 regulates R-Ras signaling leading to cell spreading and migration (Goldfinger et al., 2006; Goldfinger et al., 2007). RalBP1 adapter function is also regulated with a small GTPase guanine exchange factor (ARNO) (Goldfinger et al., 2006), and RalBP1 potentiates Ral-mediated cell spreading, potentially through similar signaling pathways (Takaya et al., 2004). Thus, RalBP1 is regulated in many molecular, cellular and physiological processes, also its function talks to other cells as a molecular adapter.

Since its initial characterization as a Ral effector and RhoGAP, RalBP1 has been implicated in various other cellular functions, at different locations within cells, all of which are likely contributors to its efficacy as a putative cancer therapy target. RalBP1 also functions as an ATP-dependent glutathione-conjugate transporter for small molecules (Awasthi et al., 2000), including anticancer drugs and endogenous metabolites (Awasthi et al., 2001; Awasthi et al., 2002; Awasthi et al., 2003), and in endocytosis (Nakashima et al., 1999; Rosse et al., 2003), mitochondrial fission (Kashatus et al., 2011), cell spreading and migration (Goldfinger et al., 2006), and Ras-induced tumorigenesis (Issaq et al., 2010; Lim et al., 2005). RLIP76 also contains many putative sites of protein phosphorylation by several kinases such as PKCs (a family of Ser/Thr kinases), as well as Ral Interacting Kinase (RIK), and many of the utilized sites in cells are Ser and Thr residues in the N-terminal domain (Herlevsen et al., 2007; Jilkina et al., 2006). PKC-dependent RalBP1 phosphorylation appears to play an important role in its functions in lung cancer (Singhal et al., 2005; Singhal et al., 2006).

4. RalBP1 in lung cancer

RalBP1 relates to cancer progression and initiation, the most thoroughly characterized is as a molecular transporter of glutathione-electrophile conjugates (GS-E). GS-E transport is also essential for protection from xenobiotics (Cheng

et al., 2001; Srivastava et al., 1998). Multi-drug resistance (MDR), particularly for alkylating chemotherapeutic drugs, is very often the result of a failure of transport in the target cells; hence, transporters such as ATP-binding cassette (ABC) type are classified as MDR proteins, which have long been pursued as therapeutic targets to inhibit drug resistance in cancer cells (Ambudkar et al., 1999; Borst et al., 2000). Also, RalBP1 contains many putative sites of protein phosphorylation by several kinases such as Protein kinase Cs (PKCs, a family of Ser/Thr kinases), as well as Ral Interacting Kinase (RIK), and many of the utilized sites in cells are Ser and Thr residues in the N-terminal domain (Herlevsen et al., 2007; Jilkina et al., 2006). PKC-dependent RalBP1 phosphorylation appears to play an important role in its functions in cancer (Singhal et al., 2005; Singhal et al., 2006).

The various cellular functions of RalBP1, and in particularly its ATPase transport activity, have been shown to translate directly to MDR in cancer cells and in tumors of many types. Blockade of RalBP1 by anti-sense or short inhibitory RNA (siRNA) depletion, or with anti-RalBP1 antibodies (presumed to block membrane-targeted RalBP1) cause apoptosis in small cell and non-small cell lung cancer, leukemia, lymphoma, melanoma, colon cancer, and prostate cancer cell lines, and RalBP1 blockade or depletion synergizes with chemotherapeutic agents such as anthracyclines and *Vinca* alkaloids (eg, vinorelbine) to further enhance apoptosis in these cancer cell lines. These *in vitro* effects have translated in every case to pronounced *in vivo* effects in tumor xenografts: blockade of RalBP1 leads to regression of tumors formed by xenografted lung cancer, melanoma, colon cancer, prostate cancer and kidney cancer cells in mice (Sharma et al., 2004; Singhal et al., 2005; Singhal et al., 2006; Singhal et al., 2007; Singhal et al., 2009). In prostate cancer cells, anti-RalBP1 antibodies were equally as effective as siRNA, suggesting that the transport function of RalBP1 is the major driver of MDR in these cells (Singhal et al., 2009).

The role of RalBP1 as a transporter mediating MDR has been most thoroughly studied in the context of lung cancer. Early studies in this area investigated 13 native human lung cancer cell lines, and found that RalBP1 purified from

non-small cell lung cancer (NSCLC) had ~ 2-fold higher ATPase activity than that from small cell lung cancer (SCLC) cell lines, perhaps due to different post-translational modifications, and RalBP1-mediated transport of doxorubicin was similarly enhanced in NSCLC compared to SCLC (Awasthi et al., 2003; Singhal et al., 2003). A recent study found RalBP1 is over-expressed in multiple NSCLC cell lines (Male et al., 2012). Another early study found that blockade of RalBP1 with specific antibodies synergized with doxorubicin to cause apoptosis in NSCLC (Awasthi et al., 2003). Subsequent studies showed that knockdown of RalBP1 by siRNA caused apoptosis in six different NSCLC cell lines, whereas augmenting RalBP1 levels led to MDR and prevention of apoptosis by both endobiotics (4-HNE) and doxorubicin (Singhal et al., 2005). Further support for RalBP1 in mediating chemotherapy resistance in lung cancer came with the finding that depletion or augmentation of RalBP1 had the same corresponding effects in both NSCLC and SCLC cells with respect to vinorelbine, a *Vinca* alkaloid with apparently less resistance in NSCLC than similar drugs (Stuckler et al., 2005). HSF-1 and POB1, binding partners of RalBP1, appear to be inhibitors of RalBP1 transport function at least in NSCLC cells, which may explain why these proteins are associated with drug sensitivity (Singhal et al., 2008; Yadav et al., 2005). However, the most direct evidence to date for a prominent role of RalBP1 in lung cancer, and its putative efficacy as a therapeutic target, was the finding that depletion of RalBP1 either with antisense or with anti-RalBP1 antibodies caused rapid and complete regression, and long-term remission of tumors in mice xenografted with two different NSCLC cell lines. Antibody and antisense treatment yielded similar results, pointing to the dominance of the transport function in the regression, and furthermore RalBP1 blockade enhanced the regressive effects of vinorelbine in this model (Singhal et al., 2007). Thus, RalBP1 is the principal mediator of ATP-dependent transport driving efflux of chemotherapeutic agents in lung cancer cells and tumors, leading to multi-drug resistance, making RalBP1 a prominent target in developing chemotherapeutic approaches to fight lung cancer.

5. Conclusions

RalBP1 is among the most promising candidates for molecular targets in lung cancer treatment. Its multifactorial contributions to tumor growth and survival are reflected in the complete regression of lung cancer cell tumors by blocking or depleting RalBP1 in mice, and drugs targeting RalBP1 will be most useful in combinatorial therapies. Therefore, studying and understanding the molecular mechanisms of RalBP1 function could be the key to developing selective therapeutic approaches in targeting RalBP1 in lung cancer.

REFERENCES

- Ambudkar SV, Dey S, Hrycyna CA, Ramachandra M, Pastan I, Gottesman MM. Biochemical, cellular, and pharmacological aspects of the multidrug transporter. *Annu Rev Pharmacol Toxicol.* 1999. 39: 361-398.
- Awasthi S, Cheng J, Singhal SS, Saini MK, Pandya U, Pikula S, Bandorowicz-Pikula J, Singh SV, Zimniak P, Awasthi YC. Novel function of human RLIP76: ATP-dependent transport of glutathione conjugates and doxorubicin. *Biochemistry.* 2000. 39: 9327-9334.
- Awasthi S, Cheng JZ, Singhal SS, Pandya U, Sharma R, Singh SV, Zimniak P, Awasthi YC. Functional reassembly of ATP-dependent xenobiotic transport by the N- and C-terminal domains of RLIP76 and identification of ATP binding sequences. *Biochemistry.* 2001. 40: 4159-4168.
- Awasthi S, Sharma R, Yang Y, Singhal SS, Pikula S, Bandorowicz-Pikula J, Zimniak P, Awasthi YC. Transport functions and physiological significance of 76 kDa Ral-binding GTPase activating protein (RLIP76). *Acta Biochim Pol.* 2002. 49: 855-867.
- Awasthi S, Singhal SS, Awasthi YC, Martin B, Woo JH, Cunningham CC, Frankel AE. RLIP76 and Cancer. *Clin Cancer Res.* 2008. 14: 4372-4377.
- Awasthi S, Singhal SS, Sharma R, Zimniak P, Awasthi YC. Transport of glutathione conjugates and chemotherapeutic drugs by RLIP76 (RALBP1): a novel link between G-protein and tyrosine kinase signaling and drug resistance. *Int J Cancer.* 2003. 106: 635-646.
- Awasthi S, Singhal SS, Singhal J, Cheng J, Zimniak P, Awasthi YC. Role of RLIP76 in lung cancer doxorubicin resistance: II. Doxorubicin transport in lung cancer by RLIP76. *Int J Oncol.* 2003. 22: 713-720.
- Awasthi S, Singhal SS, Singhal J, Yang Y, Zimniak P, Awasthi YC. Role of RLIP76 in lung cancer doxorubicin resistance: III. Anti-RLIP76 antibodies trigger apoptosis in lung cancer cells and synergistically increase doxorubicin cytotoxicity. *Int J Oncol.* 2003. 22: 721-732.
- Awasthi S, Singhal SS, Srivastava SK, Zimniak P, Bajpai KK, Saxena M, Sharma R, Ziller SA 3rd, Frenkel EP, Singh SV. Adenosine triphosphate-dependent transport of doxorubicin, daunomycin, and vinblastine in human tissues by a mechanism distinct from the P-glycoprotein. *J Clin Invest.* 1994. 93: 958-965.
- Awasthi YC, Singhal SS, Gupta S, Ahmad H, Zimniak P, Radomska A, Lester R, Sharma R. Purification and characterization of an ATPase from human liver which catalyzes ATP hydrolysis in the presence of the conjugates of bilirubin bile acids and glutathione. *Biochem Biophys Res Commun.* 1991. 175: 1090-1096.
- Bauer B, Mirey G, Vetter IR, Garcia-Ranea JA, Valencia A, Wittinghofer A, Camonis JH, Cool RH. Effector recognition by the small GTP-binding proteins Ras and Ral. *J Biol Chem.* 1999. 274: 17763-17770.
- Borst P, Evers R, Kool M, Wijnholds J. A family of drug transporters: the multidrug resistance-associated proteins. *J Natl Cancer Inst.* 2000. 92: 1295-1302.
- Burridge K. Crosstalk between Rac and Rho. *Science.* 1999. 283: 2028-2029.
- Cantor SB, Urano T, Feig LA. Identification and characterization of Ral-binding protein 1, a potential downstream target of Ral GTPases. *Mol Cell Biol.* 1995. 15: 4578-4584.
- Cheng JZ, Sharma R, Yang Y, Singhal SS, Sharma A, Saini MK, Singh SV, Zimniak P, Awasthi S, Awasthi YC. Accelerated metabolism and exclusion of 4-hydroxynonenal through induction of RLIP76 and hGST5.8 is an early adaptive response of cells to heat and oxidative stress. *J Biol Chem.* 2001. 276: 41213-41223.
- Fillatre J, Delacour D, Van Hove L, Bagarre T, Houssin N, Soulika M, Veitaia RA, Moreau J. Dynamics of the sub-cellular localization of RalBP1/RLIP through the cell cycle: the role of targeting signals and of protein-protein interactions. *FASEB J.* 2012. 26: 2164-2174.
- Goldstraw P, Crowley J, Chansky K, Giroux DJ, Groome PA, Rami-Porta R, Postmus PE, Rusch V, Sobin L. The IASLC

- Lung Cancer Staging Project: proposals for the revision of the TNM stage groupings in the forthcoming (seventh) edition of the TNM Classification of malignant tumours. *J Thorac Oncol.* 2007. 2: 706-714.
- Goldfinger LE, Ptak C, Jeffery ED, Shabanowitz J, Hunt DF, Ginsberg MH. RLIP76 (RalBP1) is an R-Ras effector that mediates adhesion-dependent Rac activation and cell migration. *J Cell Biol.* 2006. 174: 877-888.
- Goldfinger LE, Ptak C, Jeffery ED, Shabanowitz J, Han J, Haling JR, Sherman NE, Fox JW, Hunt DF, Ginsberg MH. An experimentally derived database of candidate Ras-interacting proteins. *J Proteome Res.* 2007. 6: 1806-1811.
- Herlevsen MC, Theodorescu D. Mass spectroscopic phosphoprotein mapping of Ral binding protein 1 (RalBP1/Rip1/RLIP76). *Biochem Biophys Res Commun.* 2007. 362: 56-62.
- Hu Y, Mivechi NF. HSF-1 interacts with Ral-binding protein 1 in a stress-responsive, multiprotein complex with HSP90 *in vivo*. *J Biol Chem.* 2003. 278: 17299-17306.
- Issaq SH, Lim KH, Counter CM. Sec5 and Exo84 foster oncogenic ras-mediated tumorigenesis. *Mol Cancer Res.* Feb 2010. 8: 223-231.
- Jemal A, Siegel R, Xu J, Ward E. Cancer statistics, 2010. *CA Cancer J Clin.* Sep-Oct 2010. 60: 277-300.
- Jilkina O, Bhullar RP. A serine kinase associates with 1 the RAL GTPase and phosphorylates RAL-interacting protein 1. *Biochim Biophys Acta.* 2006. 1763: 948-957.
- Jullien-Flores V, Dorseuil O, Romero F, Letourneur F, Saragosti S, Berger R, Tavittian A, Gacon G, Camonis JH. Bridging Ral GTPase to Rho pathways. RLIP76, a Ral effector with CDC42/Rac GTPase-activating protein activity. *J Biol Chem.* 1995. 270: 22473-22477.
- Jullien-Flores V, Mahe Y, Mirey G, Leprince C, Meunier-Bisceuil B, Sorokin A, Camonis JH. RLIP76, an effector of the GTPase Ral, interacts with the AP2 complex: involvement of the Ral pathway in receptor endocytosis. *J Cell Sci.* 2000. 113: 2837-2844.
- Kariya K, Koyama S, Nakashima S, Oshiro T, Morinaka K, Kikuchi A. Regulation of complex formation of POB1/epsin/adaptor protein complex 2 by mitotic phosphorylation. *J Biol Chem.* 2000. 275: 18399-18406.
- Kashatus DF, Lim KH, Brady DC, Pershing NL, Cox AD, Counter CM. RALA and RALBP1 regulate mitochondrial fission at mitosis. *Nat Cell Biol.* 2011. 13: 1108-1115.
- Lebreton S, Boissel L, Iouzalén N, Moreau J. RLIP mediates downstream signalling from RalB to the actin cytoskeleton during *Xenopus* early development. *Mech Dev.* 2004. 121: 1481-1494.
- Lim KH, Baines AT, Fiordalisi JJ, Shipitsin M, Feig LA, Cox AD, Der CJ, Counter CM. Activation of RalA is critical for Ras-induced tumorigenesis of human cells. *Cancer Cell.* 2005. 7: 533-545.
- Male H, Patel V, Jacob MA, Borrego-Diza E, Wang K, Young DA, Wise AL, Huang C, Van Veldhuizen P, O'Brien-Ladner A, Williamson SK, Taylor SA, Tawfik O, Esfandyari T, Farassati F. Inhibition of RalA signaling pathway in treatment of non-small cell lung cancer. *Lung Cancer.* 2012. 77: 252-259.
- Nakashima S, Morinaka K, Koyama S, Ikeda M, Kishida M, Okawa K, Iwamatsu A, Kishida S, Kikuchi A. Small G protein Ral and its downstream molecules regulate endocytosis of EGF and insulin receptors. *EMBO J.* 1999. 18: 3629-3642.
- Park SH, Weinberg RA. A putative effector of Ral has homology to Rho/Rac GTPase activating proteins. *Oncogene.* 1995. 11: 2349-2355.
- Riaz SP, Luchtenborg M, Coupland VH, Spicer J, Peake MD, Moller H. Trends in incidence of small cell lung cancer and all lung cancer. *Lung Cancer.* 2012. 75: 280-284.
- Rosse C, L'Hoste S, Offner N, Picard A, Camonis J. RLIP, an effector of the Ral GTPases, is a platform for Cdk1 to phosphorylate epsin during the switch off of endocytosis in mitosis. *J Biol Chem.* 2003. 278: 30597-30604.
- Sharma R, Gupta S, Singh SV, Medh RD, Ahmad H, LaBelle EF, Awasthi YC. Purification and characterization of dinitrophenylglutathione ATPase of human erythrocytes and its expression in other tissues. *Biochem Biophys Res Commun.* 1990. 171: 155-161.
- Sharma R, Singhal SS, Wickramarachchi D, Awasthi YC, Awasthi S. RLIP76 (RALBP1)-mediated transport of leukotriene C4 (LTC4) 1 in cancer cells: implications in drug resistance. *Int J Cancer.* 2004. 112: 934-942.
- Singhal SS, Awasthi YC, Awasthi S. Regression of melanoma in a murine model by RLIP76 depletion. *Cancer Res.* 2006. 66: 2354-2360.
- Singhal SS, Roth C, Leake K, Singhal J, Yadav S, Awasthi S. Regression of prostate cancer xenografts by RLIP76 depletion. *Biochem Pharmacol.* 2009. 77: 1074-1083.
- Singhal SS, Singhal J, Sharma R, Singh SV, Zimniak P, Awasthi YC, Awasthi S. Role of RLIP76 in lung cancer doxorubicin resistance: I. The ATPase activity of RLIP76 correlates with doxorubicin and 4-hydroxynonenal resistance in lung cancer cells. *Int J Oncol.* 2003. 22: 365-375.

- Singhal SS, Singhal J, Yadav S, Dwivedi S, Boor PJ, Awasthi YC, Awasthi S. Regression of lung and colon cancer xenografts by depleting or inhibiting RLIP76 (Ral-binding protein 1). *Cancer Res.* 2007. 67: 4382-4389.
- Singhal SS, Singhal J, Yadav S, Sahu M, Awasthi YC, Awasthi S. RLIP76: a target for kidney cancer therapy. *Cancer Res.* 2009. 69: 4244-4251.
- Singhal SS, Yadav S, Drake K, Singhal J, Awasthi S. Hsf-1 and POB1 induce drug sensitivity and apoptosis by inhibiting Ralbp1. *J Biol Chem.* 2008. 283: 19714-19729.
- Singhal SS, Yadav S, Singhal J, Awasthi YC, Awasthi S. Mitogenic and drug-resistance mediating effects of PKCalpha require RLIP76. *Biochem Biophys Res Commun.* 2006. 348: 722-727.
- Singhal SS, Yadav S, Singhal J, Drake K, Awasthi YC, Awasthi S. The role of PKCalpha and RLIP76 in transport-mediated doxorubicin-resistance in lung cancer. *FEBS Lett.* 2005. 579: 4635-4641.
- Singhal SS, Yadav S, Singhal J, Zajac E, Awasthi YC, Awasthi S. Depletion of RLIP76 sensitizes lung cancer cells to doxorubicin. *Biochem Pharmacol.* 2005. 70: 481-488.
- Singhal SS, Wickramarachchi D, Yadav S, Singhal J, Leake K, Vatsyayan R, Chaudhary P, Lelsani P, Suzuki S, Yang S, Awasthi YC, Awasthi S. Glutathione-conjugate transport by RLIP76 is required for clathrin-dependent endocytosis and chemical carcinogenesis. *Mol Cancer Ther.* 2011. 10: 16-28.
- Srivastava SK, Hu X, Xia H, Bleicher RJ, Zaren HA, Orchard JL, Awasthi S, Singh SV. ATP-dependent transport of glutathione conjugate of 7beta, alpha-dihydroxy-9alpha,10alpha-oxy-7,8,9,10-tetrahydrobenzo[a]pyrene in murine hepatic canalicular plasma membrane vesicles. *Biochem J.* 1998. 332: 799-805.
- Stuckler D, Singhal J, Singhal SS, Yadav S, Awasthi YC, Awasthi S. RLIP76 transports vinorelbine and mediates drug resistance in non-small cell lung cancer. *Cancer Res.* 2005. 65: 991-998.
- Takaya A, Ohba Y, Kurokawa K, Matsuda M. RalA activation at nascent lamellipodia of epidermal growth factor-stimulated Cos7 cells and migrating Madin-Darby canine kidney cells. *Mol Biol Cell.* 2004. 15: 2549-2557.
-