

Gut Microbiota-Derived Short-Chain Fatty Acids, T Cells, and Inflammation

Chang H. Kim*, Jeongho Park and Myunghoo Kim

Laboratory of Immunology and Hematopoiesis, Department of Comparative Pathobiology, Purdue Veterinary Medicine; Weldon School of Biomedical Engineering; Center for Cancer Research, Purdue University, West Lafayette, IN 47907, USA

T cells are central players in the regulation of adaptive immunity and immune tolerance. In the periphery, T cell differentiation for maturation and effector function is regulated by a number of factors. Various factors such as antigens, co-stimulation signals, and cytokines regulate T cell differentiation into functionally specialized effector and regulatory T cells. Other factors such as nutrients, micronutrients, nuclear hormones and microbial products provide important environmental cues for T cell differentiation. A mounting body of evidence indicates that the microbial metabolites short-chain fatty acids (SCFAs) have profound effects on T cells and directly and indirectly regulate their differentiation. We review the current status of our understanding of SCFA functions in regulation of peripheral T cell activity and discuss their impact on tissue inflammation.

[Immune Network 2014;14(6):277-288]

Keywords: Short-chain fatty acids, Th1, Th17, IL-10, FoxP3, Microbiota, Inflammation, Colitis, Microbial metabolites

INTRODUCTION

Commensal microbiota functions not only to serve as targets of host immunity but also as active players in regulation of host physiology and immunity as a result of long-term co-

evolution of the host and microbes. T cells play central roles in the regulation of anti-microbial immunity and tissue inflammation. Most major T cell types are made in the thymus, although extrathymic generation of some T cell subsets has been described (1,2). T cells are divided into the major TCR- $\alpha\beta$ and minor $\gamma\delta$ T cell groups. $\alpha\beta$ T cells are highly heterogeneous and grouped into CD4⁺ conventional T cells, CD8⁺ conventional T cells, NKT cells, and other innate TCR α -expressing T cells such as mucosal-associated invariant T (MAIT) cells (3-6). CD4⁺ conventional T cells are further divided into FoxP3⁺ regulatory and FoxP3⁻ T cells (7,8). FoxP3⁻ CD4⁺ T cells include various effector and regulatory T cells based on their cytokine phenotype (IFN γ , IL-17, IL-22, IL-4, IL-9, IL-10, IL-35, and/or LAP-TGF β 1) (6,9). These T helper cells include IFN γ ⁺ Th1 cells, IL-17/IL-22⁺ Th17 cells, IL-4⁺ Th2 cells, IL-9⁺ Th9 cells, IL-21⁺ T-FH cells, and IL-10/IL-35/TGF β 1⁺ Tregs (9-12). All of these T helper cell subsets are generated mainly in the periphery from naïve T cells made in the thymus. TCR repertoire and antigen specificity/affinity greatly influence T cell differentiation in the thymus and periphery (13,14). Co-stimulation signals such as CD28, ICOS, CTLA4, OX-40, and PD-1 signaling reciprocally regulate T cell differentiation and effector function (15-17). Cytokine milieu during T cell activation is crucial to generate specialized effector versus regulatory T cell subsets (6,9). A mounting body

Received on October 16, 2014. Revised on November 19, 2014. Accepted on November 28, 2014.

© This is an open access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

*Corresponding Author. Chang H. Kim, Laboratory of Immunology and Hematopoiesis, Department of Comparative Pathobiology, Purdue Veterinary Medicine; Weldon School of Biomedical Engineering; Center for Cancer Research, Purdue University, VPTH 126, 725 Harrison Street, Purdue University, West Lafayette, IN 47907, USA. Tel: 1-765-494-0976; Fax: 1-765-494-9830; E-mail: chkim@purdue.edu

Abbreviations: C2, acetic acid; C3, propionic acid; C4, butyric acid; C5, valeric acid; DCs, dendritic cells; HDAC, histone deacetylase; SCFA, short-chain fatty acid; MCT1, monocarboxylate transporter 1 (SLC16a1); SMCT1, sodium-coupled monocarboxylate transporter 1 (SLC5a8)

of evidence indicates that nutrients and metabolites provide significant regulatory signals for T cell differentiation (18-23). Potentially important roles of gut microbial products such as short-chain fatty acids (SCFAs) have been recently documented (24-26). In this review, we will review the recent progress in our understanding of the roles of SCFAs in regulating CD4⁺ T helper cell differentiation and the impact of this process on tissue inflammation.

ORIGIN, PRODUCTION, TRANSPORT, AND RECEPTORS OF SCFAs

SCFAs refer to free fatty acids containing fewer than 6 carbons and therefore they have short aliphatic carbon-chains. Formic acid (C1), acetic acid (C2), propionic acid (C3), butyric acid (C4), and valeric acid (C5) belong to the SCFA group. These metabolites are distinguished from longer fatty acids such as medium-chain (6-12 carbons) and long-chain free fatty acids. Because they have relatively shorter hydrophobic chains as well as the hydrophilic carboxyl group, SCFAs are water soluble and readily absorbed or transported into cells. SCFAs are produced by gut microbiota as fermentation products, meaning that they are partially oxidized from sugar molecules under anaerobic conditions in the colon. Carbohydrates are good sources of SCFAs but SCFAs can be made from other nutrients such as proteins and peptides albeit at low levels (27). These SCFA precursors, however, are easily degraded by host digestive enzymes in the upper alimentary tract and don't reach the microbiota in the colon in significant amounts for SCFA production. In contrast, digestion-resistant oligosaccharides and fibers (e.g. oligofructose, inulin, pectin, and arabinoxylan) are good sources of SCFAs. Insoluble fibers including cellulose and chitin, however, are not readily fermented by the microbiota and thus do not produce SCFAs at significant levels.

While it is yet to be determined clearly through extensive bacterial isolation and metagenomics studies, available information indicates that bacteria species greatly differ in their genetic make-up of enzymes involved in SCFA production (28,29). Among SCFAs, C2 is relatively more readily produced than C3 and C4 by most enteric and acetogenic bacteria (30). Propionate can be produced by three pathways (i.e. succinate, acrylate, and propanediol) from various sugar molecules such as pentoses, hexoses, and rhamnose (31). Bacteroidetes and some Firmicutes are good producers of C3 mainly through the succinate pathway. Production of C4 requires ad-

ditional enzymatic processes that extend acetyl-CoA with butyryl-CoA:acetate CoA-transferase, which is active in some bacteria including *Roseburia*, *Eubacterium* and *Anaerostipes species* and *Faecalibacterium prausnitzii* (27,32).

The combined concentrations of SCFAs produced in the colon reach ~150 mM, making SCFAs the most abundant anions in the colon. SCFAs are absorbed in the colon and either utilized in colonocytes or transported via the portal vein to reach the blood circulation and other organs. The liver and muscle are major systemic organs for SCFA metabolism and consumption. SCFAs enter cells through passive diffusion and carrier-mediated transportation through SMCT1/SLC5a8 and MCT1/SLC16a1 (33-35). SMCT1 is a sodium-coupled monocarboxylate transporter 1 for cell intake of SCFAs and related organic acids such as lactate and pyruvate (34). SMCT1 belongs to the SLC5 Na⁺/glucose cotransporter gene family (33). MCT1 is an H⁺-coupled transporter for SCFAs and related organic acids and it transports these molecules depending on the net chemical gradients for H⁺ and monocarboxylates across the membrane (36). Expression of these transporters in the apical membrane of colonocytes, DCs, kidney cells, and/or brain cells has been documented (Table I).

SCFAs activate several G-protein-coupled cell surface receptors (GPCR). GPR41 and GPR43 are major receptors that can be activated by most SCFAs (37). Gut enteroendocrine cells highly express GPR41 and GPR43 (38,39). Other regular enterocytes express these receptors also at functional levels (38-41). GPR41 is also expressed in adipocytes, renal smooth muscle cells, enteric neuronal cells, and pancreatic cells (Table I) (42,43). The expression of GPR41 is co-regulated with GPR40, a receptor for medium and long-chain fatty acids, because their gene transcription is regulated by the same promoter (44). GPR43 is expressed by granulocytes and some myeloid cells (45-47). GPR109a, a receptor for niacin (also called nicotinic acid and vitamin B3), is a receptor also for C4 (48). GPR109a is expressed by gut epithelial cells, adipocytes, macrophages and dendritic cells (Table I). Olfir78 is expressed in the kidney juxtaglomerular apparatus and is activated by C2 and C3 (49). However, T cells do not express these receptors at functionally significant levels (unpublished results) (24). Major cell types expressing these receptors are listed in Table I.

BASIC FUNCTIONS OF SCFAs IN THE BODY

SCFAs, also called volatile fatty acids because of their rela-

Table I. Expression of SCFA transporters and cell surface receptors

Transporter/ receptor	Cell types	Known functions	References
MCT1 (SLC16a1)	Colonic epithelial cells	A H ⁺ -coupled transporter for SCFAs and related organic acids	(107-109)
SMCT1 (SLC5a8)	Apical membrane of colonocytes, dendritic cells, kidney, retina, and brain	A Na(+)-coupled transport of monocarboxylates and ketone bodies into various cell types	(34, 86, 110-115)
GPR41 (FFAR3)	Enteroendocrine, enteroneuronal cells, sympathetic ganglia, adipocytes, pancreas, renal smooth muscle cells	A receptor for SCFAs. Regulation of gut hormone, leptin production, and sympathetic activation, Epithelial innate immunity	(39, 40, 42, 44, 46, 72)
GPR43 (FFAR2)	Enteroendocrine L cells, adipocytes, gut epithelial cells, leukocytes (eosinophils, basophils, neutrophils, monocytes, dendritic cells), mucosal mast cells, vascular endothelium in the myometrium	A receptor for SCFAs. Secretion of PYY and GLP-1, adipocyte development, adipogenesis, suppression of lipolysis, epithelial innate immunity, antitumor activity, anti-inflammatory effect, and Treg differentiation	(39, 40, 43, 46, 47, 58, 116-121)
GPR109a (NIACR1)	Adipocytes, dendritic cells, intestinal epithelial cells, macrophages, hepatocytes, epidermis in squamous carcinoma	A receptor for C4 and niacin. cAMP regulation, suppression of adipocyte lipolysis, HDL metabolism, DC trafficking, antitumor activity, and HDL metabolism	(48, 122-126)
Olf78 (MOL2.3, MOR18-2, PSGR, RA1c)	Large renal vessels, renal afferent arterioles, extrarenal vascular beds, prostate cancer, cells of the autonomic nervous system	A receptor for C2 and C3. Renin production from kidney juxtaglomerular apparatus for regulation of blood pressure	(49, 127, 128)

tively more volatile nature compared to longer fatty acids, have been studied for more than a century (50,51). These early observations linked SCFAs to diarrhea and ion balance in the intestine. SCFAs are physiologically important in the intestine as they regulate ion absorption and gut motility. Because SCFAs are absorbed first into colonic epithelial cells and can be metabolized in these cells, they profoundly affect the basic biology of intestinal epithelial cells. SCFAs, particularly C4, are used as the major energy source for colonic epithelial cells and regulate their gene expression, proliferation, differentiation, and apoptosis (52). For example, SCFAs promote the production of mucin and gastrointestinal peptide (e.g. LL-37) (53), molecules important for gut barrier function.

SCFAs condition intestinal epithelial cells to make them more readily respond to bacterial products (40). This function is important to prepare epithelial cells for mounting optimal innate immune responses to invading pathogens and commensal bacteria, and therefore helps prevent chronic intestinal inflammatory responses to microbes and their products. In this regard, SCFAs have anti-inflammatory activity in regulating intestinal inflammation (54). Intestinal epithelial cells express GPR41, GPR43, and GPR109a, which mediate a significant portion of the SCFA function (48,55-57).

These GPCRs activate signaling processes such as RAS, protein kinase A, PI3K, and ERK1/2 for activation of transcription factors such as ATF2 (40,47,48,58,59). Activation of this pathway is important for expression of key immune and inflammatory mediators such as IL-1, IL-6, TNF- α , CXCL1, and CXCL2. Another function of SCFAs is to activate GPR41 and GPR43 on secretory epithelial cells to produce glucagon-like peptide (GLP)-1 (60). Enteric neurons express GPR41 to sense SCFAs for regulation of gut motility (39). To support this, there is a high correlation in expression sites between SCFA receptors and gut endocrine hormones such as GLP-1, PYY, and neurotensin. Another major mechanism for the SCFA regulation of epithelial cells is mediated through inhibition of HDACs by SCFAs (61,62).

Other cell types are also regulated by SCFAs. SCFAs induce the chemotaxis of neutrophils via activation of GPR43 (58,59) and regulate neutrophil degranulation (63,64). SCFAs also regulate macrophages and dendritic cells (DCs) (65,66). SCFAs suppresses NF-kB and the production of inflammatory cytokines such as IL-6 and TNF- α but increases IL-10 secretion from macrophages (67). In contrast, increased C2 levels in alcoholism can increase the expression of inflammatory cytokines in macrophages and even exacerbate the inflammatory response in the liver (68). Thus, the SCFA func-

tion in regulation of immune responses may be altered in pathological conditions. SCFAs increase satiety and reciprocally regulate adipogenesis and lipolysis (69-71). Adipocytes express GPR41 and are activated by SCFAs to produce leptin (42,43,72). Olf1r8 activation promotes renin production from the kidney to regulate blood pressure (49).

COMPLEX ROLES OF SCFAs IN REGULATION OF EFFECTOR VERSUS REGULATORY T CELLS

Early work on C4 revealed its regulatory effect on cytokine production by lymphocytes (73,74). C4 had regulatory effects on production of cytokines such as IL-2, IL-4, IL-5, IL-6, and IL-10 (75). Others observed that C4 induced Fas-upregulation and apoptosis in T cells (76). Smith et al. reported that mice fed with SCFAs had increased numbers of IL-10-producing FoxP3⁺ T cells in the colon (25). The effect was specific for colonic FoxP3⁺ T cells, and FoxP3⁺ T cells in other organs were not expanded after SCFA administration. A mechanism provided by this group for the expanded colonic FoxP3⁺ T cells was decreased HDAC expression and activity by SCFAs in a GPR43-dependent manner. SCFAs can enter cells through diffusion or carrier-mediated transport and thus do not necessarily go through cell surface receptors. Moreover, T cells do not express GPR43 at significant levels and thus this mechanism remains to be verified. Another group reported that C2 and C3 can directly suppress HDACs and increase histone acetylation at the *FoxP3* gene locus for increased transcription (20). Similarly, it was reported that Treg generation was increased by SCFAs as a result of HDAC inhibition by SCFAs and histone H3 acetylation in key regulatory regions of the *Foxp3* locus (77). In relation to these reports, inoculation of germ-free mice with SCFA-producing *Clostridia* groups induced IL-10 and ICOS-expressing FoxP3⁺ T cells (78). Overall, these studies suggest that SCFAs expand colonic Tregs for immune tolerance.

Our group found that SCFAs can increase IL-10, but not necessarily FoxP3, expression in T cells (24). Interestingly, SCFA either positively or negatively regulate induced FoxP3⁺ cells depending on the strength of T cell activation *in vitro*. In high T cell activation conditions, SCFAs can even suppress FoxP3⁺ cell induction promoted by TGFβ1 and T cell activation. In contrast, SCFAs enhance FoxP3⁺ cell induction at low T cell activation conditions. Independent of FoxP3 regulation, SCFAs increased IL-10 production in all T cell activation conditions (24). We observed that the FoxP3⁺ T cells

even in the colon were not reproducibly regulated by SCFA administration *in vivo* (unpublished results). These results imply that FoxP3 induction by SCFAs may be regulated by indirect mechanisms through non-T cells. A rather surprising finding was that SCFAs facilitated naïve T cell differentiation into Th1 and Th17 cells in appropriate T cell polarization conditions. Thus, SCFAs can enhance both effector and regulatory T cells depending on the immunological milieu. In support of this, C2 administration via drinking water increased Th1 and Th17 cells in the intestine and secondary lymphoid tissues during *C. rodentium* infection (40). In the absence of infection, SCFAs increased gut IL-10⁺ T cells *in vivo*, which would promote immune tolerance. It appears that SCFAs se-

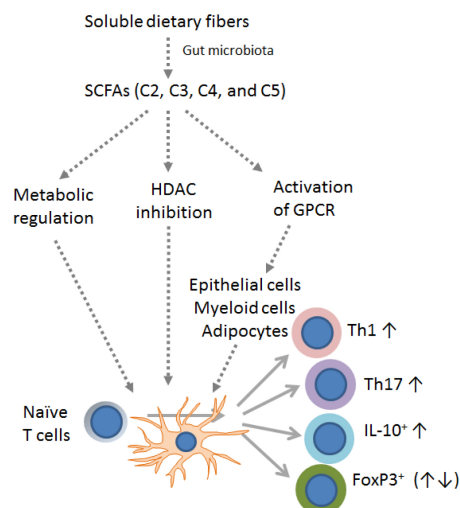


Figure 1. Regulation of T cells by SCFAs. SCFAs are actively produced by anaerobic microbiota in the colon as fermentation products of dietary materials. Most carbohydrates and proteins are completely digested and absorbed in the stomach and small intestine, and don't efficiently make SCFAs. SCFAs are mainly produced from digestion-resistant dietary fibers that reach the colon to be processed by the microbiota. SCFAs are absorbed or transported into colonocytes. They are metabolized in colonocytes or transported into blood circulation to reach other organs such as the liver and muscles. SCFAs exert their regulatory effects on epithelial cells, antigen presenting cells and T cells. Multiple mechanisms are involved including metabolic regulation, HDAC inhibition, and GPCR activation by SCFAs. These activation signals are combined to regulate T cell differentiation directly or indirectly. The direct effect of SCFAs on T cells enhances the generation of Th1 and Th17 cells in appropriate cytokine conditions, which is important to boost immunity to fight pathogens. SCFAs efficiently promote T cell production of IL-10, which is important to prevent inflammatory responses. It has been reported that SCFAs can expand FoxP3⁺ T cells in certain activation conditions. SCFAs may exert their regulatory effects on developing DCs to generate DCs that are limited in their ability to present antigens and cytokines to make effector T cells. These effects are combined to create the overall tolerogenic gut environment with a strong barrier function.

lectively promote only the right types of T cells required to handle specific immunological conditions (Fig. 1).

POTENTIAL MECHANISMS FOR THE REGULATORY EFFECT OF SCFAs ON T CELLS

Cells can be regulated by SCFAs largely in three different mechanisms (Fig. 1). The first mechanism involves the activation of SCFA-binding G-protein-coupled receptors (GPCRs) such as GPR41, GPR43, GPR109A, and Olf78. GPCR signaling can regulate cell activation, proliferation, and differentiation. However, T cells do not express any of these receptors at significant levels, according to published information and unpublished microarray data. Thus, SCFA receptors are not likely to be important for direct regulation of T cells by SCFAs. Another pathway is to regulate cell energy status and relevant signaling processes through integration of SCFAs into cellular metabolism. SCFAs can be converted to Acetyl-CoA and integrated into the citric acid cycle (Krebs cycle). Acetyl-CoA is a central molecule that stores energy in the molecule, which is eventually oxidized to CO₂ for energy production. As the result, the cellular energy [ATP/ADP] level increases, and this change boosts mTOR activation (79). In T cells, activation of mTOR skews T cell differentiation into effector T cells such as Th1 and Th17 cells at the expense of FoxP3⁺ T cells (80). mTOR activation also promotes the generation of IL-10⁺ cells (81). Thus, the SCFA-regulation of cell metabolism and mTOR accounts for the increased generation of Th1, Th17 cells, and IL-10⁺ cells. The third mechanism is mediated through the HDAC inhibitor activity of SCFAs (Fig. 1). All major SCFAs such as C2, C3, C4 and C5 have HDAC inhibitor activity (24,82,83). Some regarded that C2 does not have the HDAC inhibitor activity but it has clear HDAC inhibitor activity at concentrations (~10 mM) higher than C3 and C4 (~1 mM) (24). Moreover, C2 is maintained at relatively high concentrations in blood (~1 mM). This HDAC inhibitor activity requires the transport of SCFAs into cells and enzymatic inhibition of HDACs. Class I/II HDACs are major targets of SCFA inhibition. While SCFAs do not suppress class III HDAC such as Sirt1, down-regulation of Sirt1 expression by SCFAs was reported (84). Thus, SCFAs may affect a broad range of HDACs for their regulatory effects. Because HDAC inhibition increases the acetylation of histone and other proteins, the impact of this activity is far reaching and affecting a number of genes and proteins. Physical interaction between HDACs and S6K has been reported (85), and S6K is a downstream

effector molecule of the mTOR pathway. P70-S6 Kinase 1 (S6K) is hyper-acetylated by SCFAs in T cells, leading to increased mTOR activity in T cells (24).

SCFAs can indirectly affect T cells through their effects on other cells that control T cell differentiation such as DCs (Fig. 1). SCFAs suppress the development of bone marrow progenitors into myeloid DCs *in vitro* (86). It has been observed that SCFAs also inhibit functional maturation of DCs *in vitro* (66,86-90). For example, C4 suppressed the maturation of bone marrow-derived DCs and production of IL-12 but increased the expression of IL-23p19 (89). Valproic acid, a branched short-chain fatty acid and potent HDAC inhibitor, suppressed the maturation of human DCs *in vitro*, inhibiting the up-regulation of T-cell activating molecules such as MHC II, CD80, CD86 and IL-12 (90). While the functional importance is yet to be determined, a report indicates that C4 increased CD1d at the expense of CD1a expression on developing human DCs (88). GPR109a activation affects colonic macrophages and DCs for generation of Tregs and IL-10-producing T cells (91). This effect, however, is not solely due to C4, because GPR109a is a receptor for niacin as well. In this regard, niacin treatment suppressed colitis and colon cancer in a Gpr109a-dependent manner. Moreover, Gpr109a^{-/-} colonic epithelial cells were defective in producing IL-18 in response to C4. More studies are required to separate the niacin from SCFA effect in regulation of GPR109a. Overall, the published results indicate that the regulatory effects of SCFAs have the potential to steer DC development into tolerogenic DCs for promotion of immune tolerance. A caveat is that it remains to be fully determined if SCFAs would exert the same inhibitory effect on DCs *in vivo*.

REGULATION OF TISSUE INFLAMMATION BY SCFAs

The intestine is the first organ that encounters gut commensal bacteria-derived SCFAs. Therefore, SCFAs have been studied for decades for their effects on inflammatory bowel diseases (IBD). Despite some conflicting reports, high SCFA-producing conditions formed with high levels of dietary fibers are linked to decreased tissue inflammation in the intestine (92-94). Oral administration of C4 ameliorated T cell-induced colitis in lymphopenic mice (26). C4 administration attenuated inflammation and mucosal lesions in dextran sodium sulfate (DSS)-induced colitis, an experimental model frequently used for ulcerative colitis (95). However, there is a conflicting report that C4 administration via drinking water worsened the colitis in-

duced by DSS (89). SCFAs also failed to regulate the acute colitis induced with 2,4,6-Trinitrobenzenesulfonic acid (TNBS) (96). These conflicting results may have been obtained due to differences in methods to induce inflammation and regimens to treat the heterogeneous inflammation. To make the function of SCFAs even more complicated, both increased and decreased DSS-induced inflammation in GPR43-deficient mice has been reported (45,97). GPR43-deficient mice had exacerbated inflammation in animal models of colitis, arthritis and asthma (97). GPR43 may modulate gut inflammation, in part, through cytokine production by mononuclear cells (98). GPR43 and GPR41, expressed by tissue cells such as epithelial cells, are also important to prevent chronic inflammation in the intestine following *C. rodentium* infection (40). Thus, the available information suggests that SCFA receptors play an overall beneficial role in prevention of inflammation (Fig. 2). More work is required to identify the cell types and mecha-

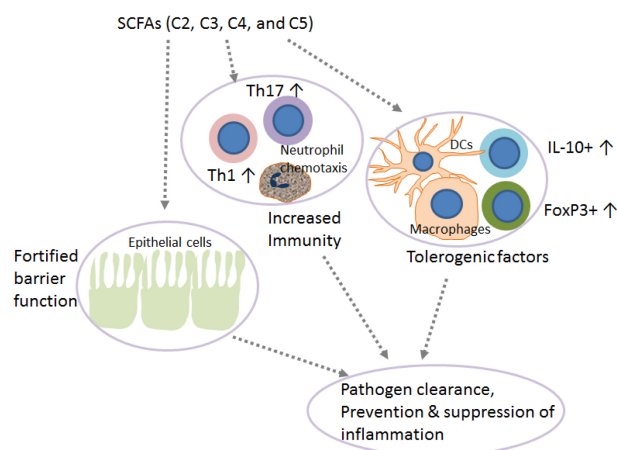


Figure 2. Regulation of tissue inflammation by SCFAs. SCFAs have the potential to regulate tissue inflammation through their effects on multiple cell types. The first cell type that is regulated by SCFAs is intestinal epithelial cells. SCFAs condition these cells to produce immune mediators that enhance the gut barrier function. Also, the response to pathogens and commensal bacteria is heightened by SCFAs during inflammatory responses. The next cell type that is affected is antigen presenting cells. SCFAs act on DCs to limit the expression of T cell-activating molecules such as MHC II molecules, co-stimulatory molecules, CCR7 and cytokines, leading to generation of tolerogenic T cells rather than inflammatory T cells. The tolerogenic effect of SCFAs on DCs can lower inflammatory responses. SCFAs can directly affect naïve T cells to steer their differentiation into both effector and IL-10-producing T cells. Moreover, SCFAs attract neutrophils to the gut during immune responses. Together, the enhanced barrier function, T cell immunity, and neutrophil recruitment help prevent infection by pathogens and invasion by commensal bacteria. By the same token, the activating activity of SCFAs for the immune cells and epithelial cells may boost inflammatory responses, if not properly regulated.

nisms that mediate the beneficial effect of SCFAs in a SCFA receptor-dependent manner.

In humans, C4 enemas had a small ameliorating effect on human colitis patients (99). Treatment of patients with distal ulcerative colitis with C4 (100 mM) was effective in ameliorating disease activity (100). Moreover, treatment of patients with mild to moderate distal ulcerative colitis with combined SCFA enemas (100 mL, twice daily enemas of sodium acetate 80 mM, sodium propionate 30 mM, and sodium butyrate 40 mM) were effective in ameliorating colitis (101). A similar therapeutic effect was observed in ~50% of ulcerative colitis patients who were refractory to a rectal and oral therapy with 5-aminosalicylic acid and corticosteroid (101). SCFAs improved the efficacy of other treatments such as oral mesalazine therapy (102). There is a report that patients with mild to moderate ileocolonic Crohn's disease who were treated with 4 g/day C4 tablets for 8 weeks had decreased clinical activity (103). A caveat is that several large randomized studies found no significant effects of SCFA therapies on ulcerative colitis patients (104,105). These mixed results indicate that SCFAs and their receptors may regulate inflammatory responses only in certain pathological conditions, ameliorating certain types of inflammatory responses while exacerbating other types of responses. Beyond inflammatory bowel diseases, high fiber diets and SCFAs have suppressive effects on respiratory allergic diseases (106). Overall, SCFAs have the potential to work through multiple cell types, including T cells, to exert their regulatory effects on tissue inflammation (Fig. 2).

CONCLUDING REMARKS

The gut microbial metabolites SCFAs profoundly regulate T cell differentiation in the body. Because these metabolites are produced at high levels in the gut, the T cells in the intestine and gut-associated lymphoid tissues are an important cell target for regulation by SCFAs. SCFAs can be transported into the blood and have the potential to regulate T cell activity in systemic tissue sites as well. Beyond T cells, SCFAs regulate the function and phenotype of a number of immunologically important cell types such as epithelial cells, neutrophils, and antigen presenting cells. While the anti-inflammatory activity of SCFAs has been emphasized, SCFAs can also promote the generation of effector T cells and enhance gut barrier function and innate immunity. All of these effects of SCFAs are important to maintain a healthy immune system and to

prevent inflammatory diseases. More studies are required to sort out the detailed mechanism of SCFA-mediated regulation of T cells and other immune cells. The current body of literature indicates that SCFAs are not a panacea for inflammatory diseases and may exacerbate certain types of tissue inflammation. Therefore, it is important to identify the types of cells, immune responses, tissue inflammation, and diseases that are highly responsive to SCFA-based therapies.

ACKNOWLEDGEMENTS

The authors thank Kim's laboratory members for their input and assistance in preparation of this review. This study was supported, in part, from grants from NIH (R01AI074745, R01DK076616, 1R01AI080769, R21AI105620, and 1S10RR028293), USDA-NIFA, and the National Multiple Sclerosis Society to CHK.

CONFLICTS OF INTEREST

The authors have no financial conflict of interest.

REFERENCES

1. Heinonen, K. M., and C. Perreault. 2008. Development and functional properties of thymic and extrathymic T lymphocytes. *Crit. Rev. Immunol.* 28: 441-466.
2. Bhandoola, A., B. H. von, H. T. Petrie, and J. C. Zuniga-Pflucker. 2007. Commitment and developmental potential of extrathymic and intrathymic T cell precursors: plenty to choose from. *Immunity* 26: 678-689.
3. Gapin, L. 2014. Check MAIT. *J. Immunol.* 192: 4475-4480.
4. Rossjohn, J., D. G. Pellicci, O. Patel, L. Gapin, and D. I. Godfrey. 2012. Recognition of CD1d-restricted antigens by natural killer T cells. *Nat. Rev. Immunol.* 12: 845-857.
5. Wan, Y. Y. 2010. Multi-tasking of helper T cells. *Immunology* 130: 166-171.
6. Li, P., R. Spolski, W. Liao, and W. J. Leonard. 2014. Complex interactions of transcription factors in mediating cytokine biology in T cells. *Immunol. Rev.* 261: 141-156.
7. Gratz, I. K., and D. J. Campbell. 2014. Organ-specific and memory treg cells: specificity, development, function, and maintenance. *Front Immunol.* 5: 333.
8. Liston, A., and D. H. Gray. 2014. Homeostatic control of regulatory T cell diversity. *Nat. Rev. Immunol.* 14: 154-165.
9. Liu, X., R. I. Nurieva, and C. Dong. 2013. Transcriptional regulation of follicular T-helper (Tfh) cells. *Immunol. Rev.* 252: 139-145.
10. Tripathi, S. K., and R. Lahesmaa. 2014. Transcriptional and epigenetic regulation of T-helper lineage specification. *Immunol. Rev.* 261: 62-83.
11. Bonelli, M., H. Y. Shih, K. Hirahara, K. Singelton, A. Laurence, A. Poholek, T. Hand, Y. Mikami, G. Vahedi, Y. Kanno, and J. J. O'Shea. 2014. Helper T cell plasticity: impact of extrinsic and intrinsic signals on transcriptomes and epigenomes. *Curr. Top. Microbiol. Immunol.* 381: 279-326.
12. Kara, E. E., I. Comerford, K. A. Fenix, C. R. Bastow, C. E. Gregor, D. R. McKenzie, and S. R. McColl. 2014. Tailored immune responses: novel effector helper T cell subsets in protective immunity. *PLoS. Pathog.* 10: e1003905.
13. Man, K., M. Miasari, W. Shi, A. Xin, D. C. Henstridge, S. Preston, M. Pellegrini, G. T. Belz, G. K. Smyth, M. A. Febbraio, S. L. Nutt, and A. Kallies. 2013. The transcription factor IRF4 is essential for TCR affinity-mediated metabolic programming and clonal expansion of T cells. *Nat. Immunol.* 14: 1155-1165.
14. Nakayama, T., and M. Yamashita. 2010. The TCR-mediated signaling pathways that control the direction of helper T cell differentiation. *Semin. Immunol.* 22: 303-309.
15. Nurieva, R. I., X. Liu, and C. Dong. 2009. Yin-Yang of costimulation: crucial controls of immune tolerance and function. *Immunol. Rev.* 229: 88-100.
16. Ishii N., T. Takahashi, P. Soroosh, and K. Sugamura. 2010. OX40-OX40 ligand interaction in T-cell-mediated immunity and immunopathology. *Adv. Immunol.* 105: 63-98.
17. Ford, M. L., and C. P. Larsen. 2009. Translating costimulation blockade to the clinic: lessons learned from three pathways. *Immunol. Rev.* 229: 294-306.
18. Mace, T. A., S. A. King, Z. Ameen, O. Elnaggar, G. Young, K. M. Riedl, S. J. Schwartz, S. K. Clinton, T. J. Knobloch, C. M. Weghorst, and G. B. Lesinski. 2014. Bioactive compounds or metabolites from black raspberries modulate T lymphocyte proliferation, myeloid cell differentiation and Jak/STAT signaling. *Cancer Immunol. Immunother.* 63: 889-900.
19. Nicolaou, A., C. Mauro, P. Urquhart, and F. Marelli-Berg. 2014. Polyunsaturated Fatty Acid-derived lipid mediators and T cell function. *Front Immunol.* 5: 75.
20. Arpaia, N., C. Campbell, X. Fan, S. Dikiy, d. van, V. P. deRoos, H. Liu, J. R. Cross, K. Pfeffer, P. J. Coffey, and A. Y. Rudensky. 2013. Metabolites produced by commensal bacteria promote peripheral regulatory T-cell generation. *Nature* 504: 451-455.
21. Benson, M. J., K. Pino-Lagos, M. Roseblatt, and R. J. Noelle. 2007. All-trans retinoic acid mediates enhanced T reg cell growth, differentiation, and gut homing in the face of high levels of co-stimulation. *J. Exp. Med.* 204: 1765-1774.
22. Kang, S. G., H. W. Lim, O. M. Andrisani, H. E. Broxmeyer, and C. H. Kim. 2007. Vitamin A metabolites induce gut-homing FoxP3⁺ regulatory T cells. *J. Immunol.* 179: 3724-3733.
23. Mucida, D., Y. Park, G. Kim, O. Turovskaya, I. Scott, M. Kronenberg, and H. Cheroutre. 2007. Reciprocal TH17 and regulatory T cell differentiation mediated by retinoic acid. *Science* 317: 256-260.
24. Park, J., M. Kim, S. G. Kang, A. H. Jannasch, B. Cooper, J. Patterson, and C. H. Kim. 2014. Short-chain fatty acids induce both effector and regulatory T cells by suppression

- of histone deacetylases and regulation of the mTOR-S6K pathway. *Mucosal Immunol*. doi: 10.1038/mi.2014.44.
25. Smith, P. M., M. R. Howitt, N. Panikov, M. Michaud, C. A. Gallini, Y. Bohlooly, J. N. Glickman, and W. S. Garrett. 2013. The microbial metabolites, short-chain fatty acids, regulate colonic Treg cell homeostasis. *Science* 341: 569-573.
 26. Furusawa, Y., Y. Obata, S. Fukuda, T. A. Endo, G. Nakato, D. Takahashi, Y. Nakanishi, C. Uetake, K. Kato, T. Kato, M. Takahashi, N. N. Fukuda, S. Murakami, E. Miyauchi, S. Hino, K. Atarashi, S. Onawa, Y. Fujimura, T. Lockett, J. M. Clarke, D. L. Topping, M. Tomita, S. Hori, O. Ohara, T. Morita, H. Koseki, J. Kikuchi, K. Honda, K. Hase, and H. Ohno. 2013. Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells. *Nature* 504: 446-450.
 27. Macfarlane, S., and G. T. Macfarlane. 2003. Regulation of short-chain fatty acid production. *Proc. Nutr. Soc.* 62: 67-72.
 28. Barcenilla, A., S. E. Pryde, J. C. Martin, S. H. Duncan, C. S. Stewart, C. Henderson, and H. J. Flint. 2000. Phylogenetic relationships of butyrate-producing bacteria from the human gut. *Appl. Environ. Microbiol.* 66: 1654-1661.
 29. Charrier, C., G. J. Duncan, M. D. Reid, G. J. Rucklidge, D. Henderson, P. Young, V. J. Russell, R. I. Aminov, H. J. Flint, and P. Louis. 2006. A novel class of CoA-transferase involved in short-chain fatty acid metabolism in butyrate-producing human colonic bacteria. *Microbiology* 152: 179-185.
 30. Miller, T. L., and M. J. Wolin. 1996. Pathways of acetate, propionate, and butyrate formation by the human fecal microbial flora. *Appl. Environ. Microbiol.* 62: 1589-1592.
 31. Reichardt, N., S. H. Duncan, P. Young, A. Belenguer, L. C. McWilliam, K. P. Scott, H. J. Flint, and P. Louis. 2014. Phylogenetic distribution of three pathways for propionate production within the human gut microbiota. *ISME, J.* 8: 1323-1335.
 32. Louis, P., G. L. Hold, and H. J. Flint. 2014. The gut microbiota, bacterial metabolites and colorectal cancer. *Nat. Rev. Microbiol.* 12: 661-672.
 33. Li, H., L. Myeroff, D. Smiraglia, M. F. Romero, T. P. Pretlow, L. Kasturi, J. Lutterbaugh, R. M. Rerko, G. Casey, J. P. Issa, J. Willis, J. K. Willson, C. Plass, and S. D. Markowitz. 2003. SLC5A8, a sodium transporter, is a tumor suppressor gene silenced by methylation in human colon aberrant crypt foci and cancers. *Proc. Natl. Acad. Sci. U. S. A.* 100: 8412-8417.
 34. Miyauchi, S., E. Gopal, Y. J. Fei, and V. Ganapathy. 2004. Functional identification of SLC5A8, a tumor suppressor down-regulated in colon cancer, as a Na(+)-coupled transporter for short-chain fatty acids. *J. Biol. Chem.* 279: 13293-13296.
 35. Yanase, H., K. Takebe, J. Nio-Kobayashi, H. Takahashi-Iwanaga, and T. Iwanaga. 2008. Cellular expression of a sodium-dependent monocarboxylate transporter (Slc5a8) and the MCT family in the mouse kidney. *Histochem. Cell Biol.* 130: 957-966.
 36. Halestrap, A. P., X. Wang, R. C. Poole, V. N. Jackson, and N. T. Price. 1997. Lactate transport in heart in relation to myocardial ischemia. *Am. J. Cardiol.* 80: 17A-25A.
 37. Eberle, J. A., P. Widmayer, and H. Breer. 2014. Receptors for short-chain fatty acids in brush cells at the "gastric groove". *Front Physiol.* 5: 152.
 38. Tazoe, H., Y. Otomo, S. Karaki, I. Kato, Y. Fukami, M. Terasaki, and A. Kuwahara. 2009. Expression of short-chain fatty acid receptor GPR41 in the human colon. *Biomed. Res.* 30: 149-156.
 39. Nohr, M. K., M. H. Pedersen, A. Gille, K. L. Egerod, M. S. Engelstoft, A. S. Husted, R. M. Sichlau, K. V. Grunddal, S. S. Poulsen, S. Han, R. M. Jones, S. Offermanns, and T. W. Schwartz. 2013. GPR41/FFAR3 and GPR43/FFAR2 as co-sensors for short-chain fatty acids in enteroendocrine cells vs FFAR3 in enteric neurons and FFAR2 in enteric leukocytes. *Endocrinology* 154: 3552-3564.
 40. Kim, M. H., S. G. Kang, J. H. Park, M. Yanagisawa, and C. H. Kim. 2013. Short-chain fatty acids activate GPR41 and GPR43 on intestinal epithelial cells to promote inflammatory responses in mice. *Gastroenterology* 145: 396-406.
 41. Wang, A., R. M. Akers, and H. Jiang. 2012. Short communication: Presence of G protein-coupled receptor 43 in rumen epithelium but not in the islets of Langerhans in cattle. *J. Dairy Sci.* 95: 1371-1375.
 42. Xiong, Y., N. Miyamoto, K. Shibata, M. A. Valasek, T. Motoike, R. M. Kedzierski, and M. Yanagisawa. 2004. Short-chain fatty acids stimulate leptin production in adipocytes through the G protein-coupled receptor GPR41. *Proc. Natl. Acad. Sci. U. S. A.* 101: 1045-1050.
 43. Zaibi, M. S., C. J. Stocker, J. O'Dowd, A. Davies, M. Bellahcene, M. A. Cawthorne, A. J. Brown, D. M. Smith, and J. R. Arch. 2010. Roles of GPR41 and GPR43 in leptin secretory responses of murine adipocytes to short chain fatty acids. *FEBS Lett.* 584: 2381-2386.
 44. Bahar, H. K., A. Veprik, N. Rubins, O. Naaman, and M. D. Walker. 2012. GPR41 gene expression is mediated by internal ribosome entry site (IRES)-dependent translation of bicistronic mRNA encoding GPR40 and GPR41 proteins. *J. Biol. Chem.* 287: 20154-20163.
 45. Sina, C., O. Gavrilova, M. Forster, A. Till, S. Derer, F. Hildebrand, B. Raabe, A. Chalaris, J. Scheller, A. Rehmann, A. Franke, S. Ott, R. Hasler, S. Nikolaus, U. R. Folsch, S. Rose-John, H. P. Jiang, J. Li, S. Schreiber, and P. Rosenstiel. 2009. G protein-coupled receptor 43 is essential for neutrophil recruitment during intestinal inflammation. *J. Immunol.* 183: 7514-7522.
 46. Brown, A. J., S. M. Goldsworthy, A. A. Barnes, M. M. Eilert, L. Tcheang, D. Daniels, A. I. Muir, M. J. Wigglesworth, I. Kinghorn, N. J. Fraser, N. B. Pike, J. C. Strum, K. M. Steplewski, P. R. Murdock, J. C. Holder, F. H. Marshall, P. G. Szekeres, S. Wilson, D. M. Ignar, S. M. Foord, A. Wise, and S. J. Dowell. 2003. The Orphan G protein-coupled receptors GPR41 and GPR43 are activated by propionate and other short chain carboxylic acids. *J. Biol. Chem.* 278: 11312-11319.
 47. Voltolini, C., S. Battersby, S. L. Etherington, F. Petraglia, J. E. Norman, and H. N. Jabbour. 2012. A novel anti-inflammatory role for the short-chain fatty acids in human labor. *Endocrinology* 153: 395-403.
 48. Thangaraju, M., G. A. Cresci, K. Liu, S. Ananth, J. P.

- Gnanaprakasam, D. D., Browning, J. D., Mellinger, S. B., Smith, G. J., Digby, N. A., Lambert, P. D., Prasad, and V. Ganapathy. 2009. GPR109A is a G-protein-coupled receptor for the bacterial fermentation product butyrate and functions as a tumor suppressor in colon. *Cancer Res.* 69: 2826-2832.
49. Pluznick, J. L., R. J. Protzko, H. Gevorgyan, Z. Peterlin, A. Sipos, J. Han, I. Brunet, L. X. Wan, F. Rey, T. Wang, S. J. Firestein, M. Yanagisawa, J. I. Gordon, A. Eichmann, J. Peti-Peterdi, and M. J. Caplan. 2013. Olfactory receptor responding to gut microbiota-derived signals plays a role in renin secretion and blood pressure regulation. *Proc. Natl. Acad. Sci. U. S. A.* 110: 4410-4415.
50. McCrudden, F. H., and H. L. Fales. 1913. The cause of the excessive calcium excretion through the feces in infantilism. *J. Exp. Med.* 17: 24-28.
51. Zoller, H. F., and W. M. Clark. 1921. The production of volatile fatty acids by bacteria of the dysentery group. *J. Gen. Physiol.* 3: 325-330.
52. Topping, D. L., and P. M. Clifton. 2001. Short-chain fatty acids and human colonic function: roles of resistant starch and nonstarch polysaccharides. *Physiol. Rev.* 81: 1031-1064.
53. Finnie, I. A., A. D. Dwarakanath, B. A. Taylor, and J. M. Rhodes. 1995. Colonic mucin synthesis is increased by sodium butyrate. *Gut* 36: 93-99.
54. Tan, J., C. McKenzie, M. Potamitis, A. N. Thorburn, C. R. Mackay, and L. Macia. 2014. The role of short-chain fatty acids in health and disease. *Adv. Immunol.* 121: 91-119.
55. Tazoe, H., Y. Otomo, I. Kaji, R. Tanaka, S. I. Karaki, and A. Kuwahara. 2008. Roles of short-chain fatty acids receptors, GPR41 and GPR43 on colonic functions. *J. Physiol. Pharmacol.* 59 Suppl 2: 251-262.
56. Wang, A., Z. Gu, B. Heid, R. M. Akers, and H. Jiang. 2009. Identification and characterization of the bovine G protein-coupled receptor GPR41 and GPR43 genes. *J. Dairy Sci.* 92: 2696-2705.
57. Tazoe, H., Y. Otomo, S. Karaki, I. Kato, Y. Fukami, M. Terasaki, and A. Kuwahara. 2009. Expression of short-chain fatty acid receptor GPR41 in the human colon. *Biomed. Res.* 30: 149-156.
58. Le, P. E., C. Loison, S. Struyf, J. Y. Springael, V. Lannoy, M. E. Decobecq, S. Brezillon, V. Dupriez, G. Vassart, D. J. Van, M. Parmentier, and M. Detheux. 2003. Functional characterization of human receptors for short chain fatty acids and their role in polymorphonuclear cell activation. *J. Biol. Chem.* 278: 25481-25489.
59. Vinolo, M. A., G. J. Ferguson, S. Kulkarni, G. Damoulakis, K. Anderson, Y. Bohlooly, L. Stephens, P. T. Hawkins, and R. Curi. 2011. SCFAs induce mouse neutrophil chemotaxis through the GPR43 receptor. *PLoS. One* 6: e21205.
60. Cani, P. D., A. Everard, and T. Duparc. 2013. Gut microbiota, enteroendocrine functions and metabolism. *Curr. Opin. Pharmacol.* 13: 935-940.
61. Licciardi, P. V., K. Ververis, and T. C. Karagiannis. 2011. Histone deacetylase inhibition and dietary short-chain Fatty acids. *ISRN. Allergy* 2011: 869647.
62. Yin, L., G. Laevsky, and C. Giardina. 2001. Butyrate suppression of colonocyte NF-kappa B activation and cellular proteasome activity. *J. Biol. Chem.* 276: 44641-44646.
63. Eftimiadi, C., E. Buzzi, M. Tonetti, P. Buffa, D. Buffa, M. T. van Steenberg, G. J. de, and G. A. Botta. 1987. Short-chain fatty acids produced by anaerobic bacteria alter the physiological responses of human neutrophils to chemotactic peptide. *J. Infect.* 14: 43-53.
64. Carretta, M. D., I. Conejeros, M. A. Hidalgo, and R. A. Burgos. 2013. Propionate induces the release of granules from bovine neutrophils. *J. Dairy Sci.* 96: 2507-2520.
65. Luhrs, H., T. Gerke, J. G. Muller, R. Melcher, J. Schaubert, F. Boxberge, W. Scheppach, and T. Menzel. 2002. Butyrate inhibits NF-kappaB activation in lamina propria macrophages of patients with ulcerative colitis. *Scand. J. Gastroenterol.* 37: 458-466.
66. Millard, A. L., P. M. Mertes, D. Ittelet, F. Villard, P. Jeannesson, and J. Bernard. 2002. Butyrate affects differentiation, maturation and function of human monocyte-derived dendritic cells and macrophages. *Clin. Exp. Immunol.* 130: 245-255.
67. Park, J. S., E. J. Lee, J. C. Lee, W. K. Kim, and H. S. Kim. 2007. Anti-inflammatory effects of short chain fatty acids in IFN-gamma-stimulated RAW 264.7 murine macrophage cells: involvement of NF-kappaB and ERK signaling pathways. *Int. Immunopharmacol.* 7: 70-77.
68. Kendrick, S. F., G. O'Boyle, J. Mann, M. Zeybel, J. Palmer, D. E. Jones, and C. P. Day. 2010. Acetate, the key modulator of inflammatory responses in acute alcoholic hepatitis. *Hepatology* 51: 1988-1997.
69. Arora, T., R. Sharma, and G. Frost. 2011. Propionate. Anti-obesity and satiety enhancing factor? *Appetite* 56: 511-515.
70. Hong, Y. H., Y. Nishimura, D. Hishikawa, H. Tsuzuki, H. Miyahara, C. Gotoh, K. C. Choi, D. D. Feng, C. Chen, H. G. Lee, K. Katoh, S. G. Roh, and S. Sasaki. 2005. Acetate and propionate short chain fatty acids stimulate adipogenesis via GPCR43. *Endocrinology* 146: 5092-5099.
71. Ge, H., X. Li, J. Weiszmann, P. Wang, H. Baribault, J. L. Chen, H. Tian, and Y. Li. 2008. Activation of G protein-coupled receptor 43 in adipocytes leads to inhibition of lipolysis and suppression of plasma free fatty acids. *Endocrinology* 149: 4519-4526.
72. Kimura, I., D. Inoue, T. Maeda, T. Hara, A. Ichimura, S. Miyauchi, M. Kobayashi, A. Hirasawa, and G. Tsujimoto. 2011. Short-chain fatty acids and ketones directly regulate sympathetic nervous system via G protein-coupled receptor 41 (GPR41). *Proc. Natl. Acad. Sci. U. S. A.* 108: 8030-8035.
73. Nancey, S., J. Bienvenu, B. Coffin, F. Andre, L. Descos, and B. Flourie. 2002. Butyrate strongly inhibits *in vitro* stimulated release of cytokines in blood. *Dig. Dis. Sci.* 47: 921-928.
74. Cavaglieri, C. R., A. Nishiyama, L. C. Fernandes, R. Curi, E. A. Miles, and P. C. Calder. 2003. Differential effects of short-chain fatty acids on proliferation and production of pro- and anti-inflammatory cytokines by cultured lymphocytes. *Life Sci.* 73: 1683-1690.
75. Kurita-Ochiai, T., K. Fukushima, and K. Ochiai. 1995. Volatile fatty acids, metabolic by-products of periodontopathic bacteria, inhibit lymphocyte proliferation and cytokine production. *J. Dent. Res.* 74: 1367-1373.

76. Zimmerman, M. A., N. Singh, P. M. Martin, M. Thangaraju, V. Ganapathy, J. L. Waller, H. Shi, K. D. Robertson, D. H. Munn, and K. Liu. 2012. Butyrate suppresses colonic inflammation through HDAC1-dependent Fas upregulation and Fas-mediated apoptosis of T cells. *Am. J. Physiol. Gastrointest. Liver Physiol.* 302: G1405-G1415.
77. Furusawa, Y., Y. Obata, S. Fukuda, T. A. Endo, G. Nakato, D. Takahashi, Y. Nakanishi, C. Uetake, K. Kato, T. Kato, M. Takahashi, N. N. Fukuda, S. Murakami, E. Miyauchi, S. Hino, K. Atarashi, S. Onawa, Y. Fujimura, T. Lockett, J. M. Clarke, D. L. Topping, M. Tomita, S. Hori, O. Ohara, T. Morita, H. Koseki, J. Kikuchi, K. Honda, K. Hase, and H. Ohno. 2013. Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells. *Nature* 504: 446-450.
78. Atarashi, K., T. Tanoue, K. Oshima, W. Suda, Y. Nagano, H. Nishikawa, S. Fukuda, T. Saito, S. Narushima, K. Hase, S. Kim, J. V. Fritz, P. Wilmes, S. Ueha, K. Matsushima, H. Ohno, B. Olle, S. Sakaguchi, T. Taniguchi, H. Morita, M. Hattori, and K. Honda. 2013. Treg induction by a rationally selected mixture of Clostridia strains from the human microbiota. *Nature* 500: 232-236.
79. Dennis, P. B., A. Jaeschke, M. Saitoh, B. Fowler, S. C. Kozma, and G. Thomas. 2001. Mammalian TOR: a homeostatic ATP sensor. *Science* 294: 1102-1105.
80. Delgoffe, G. M., T. P. Kole, Y. Zheng, P. E. Zarek, K. L. Matthews, B. Xiao, P. F. Worley, S. C. Kozma, and J. D. Powell. 2009. The mTOR kinase differentially regulates effector and regulatory T cell lineage commitment. *Immunity* 30: 832-844.
81. Chen, S., D. Liu, J. Wu, B. Xu, K. Lu, W. Zhu, and M. Chen. 2014. Effect of inhibiting the signal of mammalian target of rapamycin on memory T cells. *Transplant. Proc.* 46: 1642-1648.
82. Hinnebusch, B. F., S. Meng, J. T. Wu, S. Y. Archer, and R. A. Hodin. 2002. The effects of short-chain fatty acids on human colon cancer cell phenotype are associated with histone hyperacetylation. *J. Nutr.* 132: 1012-1017.
83. Haberland, M., R. L. Montgomery, and E. N. Olson. 2009. The many roles of histone deacetylases in development and physiology: implications for disease and therapy. *Nat. Rev. Genet.* 10: 32-42.
84. Yu, X., A. M. Shahir, J. Sha, Z. Feng, B. Eapen, S. Nithianantham, B. Das, J. Karn, A. Weinberg, N. F. Bissada, and F. Ye. 2014. Short-chain fatty acids from periodontal pathogens suppress histone deacetylases, EZH2, and SUV39H1 to promote Kaposi's sarcoma-associated herpesvirus replication. *J. Virol.* 88: 4466-4479.
85. Fenton, T. R., J. Gwalter, J. Ericsson, and I. T. Gout. 2010. Histone acetyltransferases interact with and acetylate p70 ribosomal S6 kinases *in vitro* and *in vivo*. *Int. J. Biochem. Cell Biol.* 42: 359-366.
86. Singh, N., M. Thangaraju, P. D. Prasad, P. M. Martin, N. A. Lambert, T. Boettger, S. Offermanns, and V. Ganapathy. 2010. Blockade of dendritic cell development by bacterial fermentation products butyrate and propionate through a transporter (Slc5a8)-dependent inhibition of histone deacetylases. *J. Biol. Chem.* 285: 27601-27608.
87. Wang, B., A. Morinobu, M. Horiuchi, J. Liu, and S. Kumagai. 2008. Butyrate inhibits functional differentiation of human monocyte-derived dendritic cells. *Cell Immunol.* 253: 54-58.
88. Nascimento, C. R., C. G. Freire-de-Lima, O. A. da Silva de, F. D. Rumjanek, and V. M. Rumjanek. 2011. The short chain fatty acid sodium butyrate regulates the induction of CD1a in developing dendritic cells. *Immunobiology* 216: 275-284.
89. Berndt, B. E., M. Zhang, S. Y. Owyang, T. S. Cole, T. W. Wang, J. Luther, N. A. Veniaminova, J. L. Merchant, C. C. Chen, G. B. Huffnagle, and J. Y. Kao. 2012. Butyrate increases IL-23 production by stimulated dendritic cells. *Am. J. Physiol. Gastrointest. Liver Physiol.* 303: G1384-G1392.
90. Frikeche, J., T. Simon, E. Brissot, M. Gregoire, B. Gaugler, and M. Mohty. 2012. Impact of valproic acid on dendritic cells function. *Immunobiology* 217: 704-710.
91. Singh, N., A. Gurav, S. Sivaprakasam, E. Brady, R. Padia, H. Shi, M. Thangaraju, P. D. Prasad, S. Manicassamy, D. H. Munn, J. R. Lee, S. Offermanns, and V. Ganapathy. 2014. Activation of Gpr109a, receptor for niacin and the commensal metabolite butyrate, suppresses colonic inflammation and carcinogenesis. *Immunity* 40: 128-139.
92. Ananthkrishnan, A. N., H. Khalili, G. G. Konijeti, L. M. Higuchi, S. P. de, J. R. Korzenik, C. S. Fuchs, W. C. Willett, J. M. Richter, and A. T. Chan. 2013. A prospective study of long-term intake of dietary fiber and risk of Crohn's disease and ulcerative colitis. *Gastroenterology* 145: 970-977.
93. Amre, D. K., S. D'Souza, K. Morgan, G. Seidman, P. Lambrette, G. Grimard, D. Israel, D. Mack, P. Ghadirian, C. Deslandres, V. Chotard, B. Budai, L. Law, E. Levy, and E. G. Seidman. 2007. Imbalances in dietary consumption of fatty acids, vegetables, and fruits are associated with risk for Crohn's disease in children. *Am. J. Gastroenterol.* 102: 2016-2025.
94. Hou, J. K., B. Abraham, and H. El-Serag. 2011. Dietary intake and risk of developing inflammatory bowel disease: a systematic review of the literature. *Am. J. Gastroenterol.* 106: 563-573.
95. Vieira, E. L., A. J. Leonel, A. P. Sad, N. R. Beltrao, T. F. Costa, T. M. Ferreira, A. C. Gomes-Santos, A. M. Faria, M. C. Peluzio, D. C. Cara, and J. I. varez-Leite. 2012. Oral administration of sodium butyrate attenuates inflammation and mucosal lesion in experimental acute ulcerative colitis. *J. Nutr. Biochem.* 23: 430-436.
96. Tarrerias, A. L., M. Millecamps, A. Alloui, C. Beaughard, J. L. Kemeny, S. Bourdu, G. Bommelaer, A. Eschalier, M. Dapoigny, and D. Ardid. 2002. Short-chain fatty acid enemas fail to decrease colonic hypersensitivity and inflammation in TNBS-induced colonic inflammation in rats. *Pain* 100: 91-97.
97. Maslowski, K. M., A. T. Vieira, A. Ng, J. Kranich, F. Sierro, D. Yu, H. C. Schilter, M. S. Rolph, F. Mackay, D. Artis, R. J. Xavier, M. M. Teixeira, and C. R. Mackay. 2009. Regulation of inflammatory responses by gut microbiota and chemoattractant receptor GPR43. *Nature* 461: 1282-1286.
98. Masui, R., M. Sasaki, Y. Funaki, N. Ogasawara, M. Mizuno, A. Iida, S. Izawa, Y. Kondo, Y. Ito, Y. Tamura, K. Yanamoto,

- H. Noda, A. Tanabe, N. Okaniwa, Y. Yamaguchi, T. Iwamoto, and K. Kasugai. 2013. G protein-coupled receptor 43 moderates gut inflammation through cytokine regulation from mononuclear cells. *Inflamm. Bowel. Dis.* 19: 2848-2856.
99. Hamer, H. M., D. M. Jonkers, S. A. Vanhoutvin, F. J. Troost, G. Rijkers, B. A. de, A. Bast, K. Venema, and R. J. Brummer. 2010. Effect of butyrate enemas on inflammation and antioxidant status in the colonic mucosa of patients with ulcerative colitis in remission. *Clin. Nutr.* 29: 738-744.
100. Scheppach, W., H. Sommer, T. Kirchner, G. M. Paganelli, P. Bartram, S. Christl, F. Richter, G. Dusel, and H. Kasper. 1992. Effect of butyrate enemas on the colonic mucosa in distal ulcerative colitis. *Gastroenterology* 103: 51-56.
101. Vernia, P., A. Marcheggiano, R. Caprilli, G. Frieri, G. Corrao, D. Valpiani, M. C. Di Paolo, P. Paoluzi, and A. Torsoli. 1995. Short-chain fatty acid topical treatment in distal ulcerative colitis. *Aliment. Pharmacol. Ther.* 9: 309-313.
102. Vernia, P., G. Monteleone, G. Grandinetti, G. Villotti, G. E. Di, G. Frieri, A. Marcheggiano, F. Pallone, R. Caprilli, and A. Torsoli. 2000. Combined oral sodium butyrate and mesalazine treatment compared to oral mesalazine alone in ulcerative colitis: randomized, double-blind, placebo-controlled pilot study. *Dig. Dis. Sci.* 45: 976-981.
103. Di, S. A., R. Morera, R. Ciccocioppo, P. Cazzola, S. Gotti, F. P. Tinozzi, S. Tinozzi, and G. R. Corazza. 2005. Oral butyrate for mildly to moderately active Crohn's disease. *Aliment. Pharmacol. Ther.* 22: 789-794.
104. Steinhart, A. H., T. Hiruki, A. Brzezinski, and J. P. Baker. 1996. Treatment of left-sided ulcerative colitis with butyrate enemas: a controlled trial. *Aliment. Pharmacol. Ther.* 10: 729-736.
105. Breuer, R. I., K. H. Soergel, B. A. Lashner, M. L. Christ, S. B. Hanauer, A. Vanaganas, J. M. Harig, A. Keshavarzian, M. Robinson, J. H. Sellin, D. Weinberg, D. E. Vidican, K. L. Flemal, and A. W. Rademaker. 1997. Short chain fatty acid rectal irrigation for left-sided ulcerative colitis: a randomized, placebo controlled trial. *Gut* 40: 485-491.
106. Trompette, A., E. S. Gollwitzer, K. Yadava, A. K. Sichelstiel, N. Sprenger, C. Ngom-Bru, C. Blanchard, T. Junt, L. P. Nicod, N. L. Harris, and B. J. Marsland. 2014. Gut microbiota metabolism of dietary fiber influences allergic airway disease and hematopoiesis. *Nat. Med.* 20: 159-166.
107. Hadjiagapiou, C., L. Schmidt, P. K. Dudeja, T. J. Layden, and K. Ramaswamy. 2000. Mechanism(s) of butyrate transport in Caco-2 cells: role of monocarboxylate transporter 1. *Am. J. Physiol. Gastrointest. Liver Physiol.* 279: G775-G780.
108. Alrefai, W. A., S. Tyagi, R. Gill, S. Saksena, C. Hadjiagapiou, F. Mansour, K. Ramaswamy, and P. K. Dudeja. 2004. Regulation of butyrate uptake in Caco-2 cells by phorbol 12-myristate 13-acetate. *Am. J. Physiol. Gastrointest. Liver Physiol.* 286: G197-G203.
109. Ritzhaupt, A., A. Ellis, K. B. Hosie, and S. P. Shirazi-Beechey. 1998. The characterization of butyrate transport across pig and human colonic luminal membrane. *J. Physiol.* 507(Pt 3): 819-830.
110. Gopal, E., Y. J. Fei, S. Miyauchi, L. Zhuang, P. D. Prasad, and V. Ganapathy. 2005. Sodium-coupled and electrogenic transport of B-complex vitamin nicotinic acid by slc5a8, a member of the Na/glucose co-transporter gene family. *Biochem. J.* 388: 309-316.
111. Miyauchi, S., E. Gopal, E. Babu, S. R. Srinivas, Y. Kubo, N. S. Umapathy, S. V. Thakkar, V. Ganapathy, and P. D. Prasad. 2010. Sodium-coupled electrogenic transport of pyroglutamate (5-oxoproline) via SLC5A8, a monocarboxylate transporter. *Biochim. Biophys. Acta* 1798: 1164-1171.
112. Thangaraju, M., G. Cresci, S. Itagaki, J. Mellinger, D. D. Browning, F. G. Berger, P. D. Prasad, and V. Ganapathy. 2008. Sodium-coupled transport of the short chain fatty acid butyrate by SLC5A8 and its relevance to colon cancer. *J. Gastrointest. Surg.* 12: 1773-1781.
113. Gopal, E., Y. J. Fei, M. Sugawara, S. Miyauchi, L. Zhuang, P. Martin, S. B. Smith, P. D. Prasad, and V. Ganapathy. 2004. Expression of slc5a8 in kidney and its role in Na(+)-coupled transport of lactate. *J. Biol. Chem.* 279: 44522-44532.
114. Martin, P. M., Y. Dun, B. Mysona, S. Ananth, P. Roon, S. B. Smith, and V. Ganapathy. 2007. Expression of the sodium-coupled monocarboxylate transporters SMCT1 (SLC5A8) and SMCT2 (SLC5A12) in retina. *Invest. Ophthalmol. Vis. Sci.* 48: 3356-3363.
115. Martin, P. M., E. Gopal, S. Ananth, L. Zhuang, S. Itagaki, B. M. Prasad, S. B. Smith, P. D. Prasad, and V. Ganapathy. 2006. Identity of SMCT1 (SLC5A8) as a neuron-specific Na⁺-coupled transporter for active uptake of L-lactate and ketone bodies in the brain. *J. Neurochem.* 98: 279-288.
116. Tolhurst, G., H. Heffron, Y. S. Lam, H. E. Parker, A. M. Habib, E. Diakogiannaki, J. Cameron, J. Grosse, F. Reimann, and F. M. Gribble. 2012. Short-chain fatty acids stimulate glucagon-like peptide-1 secretion via the G-protein-coupled receptor FFAR2. *Diabetes* 61: 364-371.
117. Hong, Y. H., Y. Nishimura, D. Hishikawa, H. Tsuzuki, H. Miyahara, C. Gotoh, K. C. Choi, D. D. Feng, C. Chen, H. G. Lee, K. Katoh, S. G. Roh, and S. Sasaki. 2005. Acetate and propionate short chain fatty acids stimulate adipogenesis via GPCR43. *Endocrinology* 146: 5092-5099.
118. Dewulf, E. M., Q. Ge, L. B. Bindels, F. M. Sohet, P. D. Cani, S. M. Brichard, and N. M. Delzenne. 2013. Evaluation of the relationship between GPR43 and adiposity in human. *Nutr. Metab. (Lond)* 10: 11.
119. Tang, Y., Y. Chen, H. Jiang, G. T. Robbins, and D. Nie. 2011. G-protein-coupled receptor for short-chain fatty acids suppresses colon cancer. *Int. J. Cancer* 128: 847-856.
120. Karaki, S., R. Mitsui, H. Hayashi, I. Kato, H. Sugiyama, T. Iwanaga, J. B. Furness, and A. Kuwahara. 2006. Short-chain fatty acid receptor, GPR43, is expressed by enteroendocrine cells and mucosal mast cells in rat intestine. *Cell Tissue Res.* 324: 353-360.
121. Nilsson, N. E., K. Kotarsky, C. Owman, and B. Olde. 2003. Identification of a free fatty acid receptor, FFA2R, expressed on leukocytes and activated by short-chain fatty acids. *Biochem. Biophys. Res. Commun.* 303: 1047-1052.
122. Wanders, D., E. C. Graff, and R. L. Judd. 2012. Effects of high fat diet on GPR109A and GPR81 gene expression. *Biochem. Biophys. Res. Commun.* 425: 278-283.

123. Taggart, A. K., J. Kero, X. Gan, T. Q. Cai, K. Cheng, M. Ippolito, N. Ren, R. Kaplan, K. Wu, T. J. Wu, L. Jin, C. Liaw, R. Chen, J. Richman, D. Connolly, S. Offermanns, S. D. Wright, and M. G. Waters. 2005. (D)-beta-Hydroxybutyrate inhibits adipocyte lipolysis via the nicotinic acid receptor PUMA-G. *J. Biol. Chem.* 280: 26649-26652.
124. Ingersoll, M. A., S. Potteaux, D. Alvarez, S. B. Hutchison, R. N. van, and G. J. Randolph. 2012. Niacin inhibits skin dendritic cell mobilization in a GPR109A independent manner but has no impact on monocyte trafficking in atherosclerosis. *Immunobiology* 217: 548-557.
125. Li, X., J. S. Millar, N. Brownell, F. Briand, and D. J. Rader. 2010. Modulation of HDL metabolism by the niacin receptor GPR109A in mouse hepatocytes. *Biochem. Pharmacol.* 80: 1450-1457.
126. Bermudez, Y., C. A. Benavente, R. G. Meyer, W. R. Coyle, M. K. Jacobson, and E. L. Jacobson. 2011. Nicotinic acid receptor abnormalities in human skin cancer: implications for a role in epidermal differentiation. *PLoS. One* 6: e20487.
127. Xu, L. L., B. G. Stackhouse, K. Florence, W. Zhang, N. Shanmugam, I. A. Sesterhenn, Z. Zou, V. Srikantan, M. Augustus, V. Roschke, K. Carter, D. G. McLeod, J. W. Moul, D. Soppett, and S. Srivastava. 2000. PSGR, a novel prostate-specific gene with homology to a G protein-coupled receptor, is overexpressed in prostate cancer. *Cancer Res.* 60: 6568-6572.
128. Weber, M., U. Pehl, H. Breer, and J. Strotmann. 2002. Olfactory receptor expressed in ganglia of the autonomic nervous system. *J. Neurosci. Res.* 68: 176-184.