

Sex Reversal and Masculinization according to Growth in Longtooth Grouper *Epinephelus bruneus*

Seong-Rip Oh, Hyeong-Cheol Kang, Chi-Hoon Lee, Sang-Woo Hur and †Young-Don Lee

Marine and Environmental Research Institute, Jeju National University, Jeju 695-965, Republic of Korea

ABSTRACT : To understand the sex reversal characteristics in the longtooth grouper (*Epinephelus bruneus*), this study examined the sex reversal and artificial masculinization of wild caught *E. bruneus* reared in indoor rearing tank after a 17 α -methyltestosterone injection. To domesticate a broodstock, 64 wild caught *E. bruneus*, between 47.0 to 110.0 cm in total length and from 1.5 to 21.4 kg in body weight, were reared in indoor rearing tank (4.0 to 5.0 m wide, and 2.5 to 3.0 m depth) for four years. Seven specimens showed sex reversal from female to male during indoor rearing condition, whose total length and body weights were from 63.0 to 99.0 cm and from 4.4 to 13.2 kg, respectively. After inducing artificial masculinization in 14 female *E. bruneus* with a 17 α -methyltestosterone (2.0 mg/kg BW) implants for 3 years, spermiation occurred in 9 specimens (total length: 54.0 to 68.0 cm, body weight: 2.3 to 4.3 kg). Among the female to male sex reversals, two specimens returned back to being female, whose body weights were 2.8 kg (initially 2.6 kg) and 2.7 kg (initially 2.3 kg). Therefore, this study suggested that *E. bruneus* (> 3.0 kg) was more effective in masculinizing by 17 α -methyltestosterone implants.

Key words : *Epinephelus bruneus*, Longtooth grouper, Masculinization, 17 α -Methyltestosterone, Sex reversal

INTRODUCTION

Epinephelinae contains 159 species in 15 genera and generally inhabits rocky or coral subtropical and tropical regions (FAO, 1993). There are 11 species of grouper inhabit the Southern Sea near Jeju Island in Korea, including *Epinephelus bruneus*, *E. septemfasciatus*, *E. akaara* and *E. fasciatus* (Kim et al., 2001). Groupers are commercially important species in Southeast Asian countries including Korea. Recently, grouper resources decreased rapidly due to over-fishing in Jeju coastal waters. Thus, there are many studies on artificial seed production of groupers in several Asian countries including Korea. Previous research on the *E. bruneus* has been conducted as follows: spawning behavior and ontogeny development in indoor rearing tank (Manabe

& Kasuga, 1988; Yang et al., 2007), larvae's environmental tolerance, growth, and morphological development (Okada et al., 1996; Sawada et al., 1999), the effectiveness of water temperature on the growth, feeding of juvenile (Inoue, 2001), seed production (Lee et al., 2008) and pubertal characteristics (Ryu et al., 2013).

Epinephelinae has sex characterization of protogynous hermaphroditism in that all fish sex differentiate and function as a female, and then some fish reverse sex to male as they age and grow. The grouper's natural sex reversal has been found in an *E. tauvina* (Chao & Lim, 1991) showing it took seven years to have natural sex reversal after hatching. Another study on *E. costae* showed 11.0 kg in body weight fish have natural sex reversal (Glamuzina et al., 2000). As for the *E. akaara* (Hamamoto et al., 1986),

Manuscript received 24 April 2013, received in revised form 2 May 2013, accepted 12 May, 2013

† Corresponding Author : Young-Don Lee, Marine and Environmental Research Institute, Jeju National University, 3288 Hamdoek, Jocheon, Jeju 695-965, Republic of Korea. Tel. : +82-64-783-8281, Fax : +82-64-782-8281, E-mail : leemri@jejunu.ac.kr

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License(<http://creativecommons.org/licenses/by-nc/3.0>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

the fish longer than 25.0 cm in total length and heavier than 500.0 g in body weight has an increased ratio of males. Due to their sex characteristics in nature, capture of mature male is difficult and it is the most significant constraint to artificial seed production of grouper. Thus, the sex reversal technique is being used to produce functional males by injecting androgen. Previous experiments attempted masculinization in groupers through hormonal manipulation. In *E. tauvina* (Chen et al., 1977; Chao & Chow, 1990), *E. septemfasciatus* (Tukashima & Kitajima, 1983; Tsuchihashi et al., 2003; Song, 2004), and *E. akaara* (Hwang et al., 1998), functional males were obtained when exogenous androgen or 17 α -methyltestosterone (MT) was administered. In *E. merra* (Alam et al., 2006), an aromatase inhibitor, or an endogenous hormone, was used to induce masculinization. However, more research is needed whether the functional males remained male after the maturation season or if they revert back to be females.

In this study, we investigated induced sex reversal and sex revert of *E. bruneus* reared in indoor artificial rearing condition to obtain functional males for fertilization egg production and seed production via observation of the treatment of 17 α -methyltestosterone (MT).

MATERIALS AND METHOD

1. Specimen

From September 2002 to December 2005, 64 wild caught *E. bruneus* (total length 65.6 \pm 13.1 cm, body weight 4.7 \pm 4.0 kg) were used in this study. Specimens were transported to the Ocean and Fisheries Research Institute in Korea's Jeju Province and maintained in 6 inner concrete rearing tanks (two tanks were 5.0 \times 5.0 \times 3.0 m; 62 m³, and four tanks were 4.0 \times 4.0 \times 2.5 m; 36 m³). Specimens were individually marked via insertion of micro ID tag into the dorsal skin (ID tag, Trovan Ltd. UK). Specimens were fed frozen horse mackerel twice a week and were fed a mixture of frozen horse mackerel and squid 2 to 3 months before the maturation season.

2. Sex reversal and masculinization

1) The sex ratio of broodstock and sex reversal

We examined the sex ratio depending on the body weight and sex reversal of *E. bruneus* with ID tags reared in indoor rearing tanks. The sex division was judged by cannulation between June and August. Female *E. bruneus* were identified when the oocyte diameter in the gonad is larger than 200 μ m, males showing spermiation when pressed on the abdomen, and unidentified when showing neither ovulation nor spermiation. When spermiation occurred to specimens identified as female the previous year, we decided those were naturally sex reversed in the rearing tank.

2) Masculinization

(1) Induced males with the 17 α -methyltestosterone treatment

Sexually immatures *E. bruneus* (n=14) between 50 to 69 cm in total length (61.5 \pm 4.5 cm) and a body weight between 2.3 to 4.3 kg (3.4 \pm 0.5 kg) were used to experiment induced masculinization 4 times from 2003 to 2006. To induce masculinization via the method of Tsuchihashi *et al.* (2003), a silastic capsule with androgen hormone (MT, 17 α -methyltestosterone, Sigma Co. Ltd, USA) was implanted into the abdominal cavity. MT (1 mg MT/10 μ l) in the silastic capsule was produced by adding castor oil (Junsei Chemical Co. Ltd., Japan) 800 μ l to 100 mg of MT dissolved in 200 μ l of 95% ethanol. The lower abdominal underneath the pectoral fin was cut about 0.5 cm to inject the silastic capsule after anesthetizing with 2-phenoxyethanol (Sigma Co. Ltd, USA) 300 ppm, and the wound sealed after-wards. We examined the success of artificial sex reversal with a histological analysis and external observation by a method of seeing if spermiation occurs when pressing the abdomen in 2003. From 2004 to 2006, we judged sex reversal with external observation described above. For the histological analysis, gonads were dissected from abdominal cavity, and fixed in Bouin's solution before the silastic capsule implantand 8 weeks after the implant. The samples were dehydrated in

a graded series of ethanol, embedded in paraffin and then cut into 5 µm cross section. Slides were stained with Hansen's hematoxylin and 0.5% eosin then observed using a light microscope (HBO 50, Carl Zeiss). For external observation, we judged if masculinization was successful through observing spermiation by pressing the abdomen between 8 and 14 weeks after the silastic implant.

(2) Induced males revert to female

Once specimens were identified as functional males, they were tested a year after the MT treatment. The spermiation for specimen was observed between June and August in the maturation season by the abdomen pressing method and the cannulation method. Revert to females among masculinized specimen was determined that oocytes could be obtained by cannulation or ovulation occurred after HCG treatment.

RESULTS

1. The sex ratio and sex reversal of inner rearing broodstock

Body weight between 1.0 to 3.0 kg, had 1 female (14.2%), 0 male (0%), and 6 indiscernible (85.7%) from a sample of 7 fish. Body weight between 3.0 to 5.0 kg, had 9 females (60.0%), 4 males (26.6%) and 2 indiscernible (13.3%) from a sample of 15 fish. Body weight between 5.0 to 8.0

kg, had 7 females (63.6%), 3 males (27.2%), 1 indiscernible (9.0%) from a sample of 11 fish. Body weight between 8.0 to 11.0 kg, had 1 female (20.0%), 3 males (60.0%), 1 indiscernible (20.0%) from a sample of 5 fish. Body weight more than 11.0 kg, had 1 female (50.0%), 1 male (50.0%) from a sample of 2 fish (Fig. 1).

For the sex reversal in indoor rearing condition, we obtained 1 sex-reversed male (total length 99.0 cm, body weight 13.2 kg) from a sample of 5 females bought in 2002. We obtained 5 sex-changed males out of 23 females bought in 2003. It occurred in a different time period, 1 female (total length 63.0 cm, body weight 4.4 kg) in

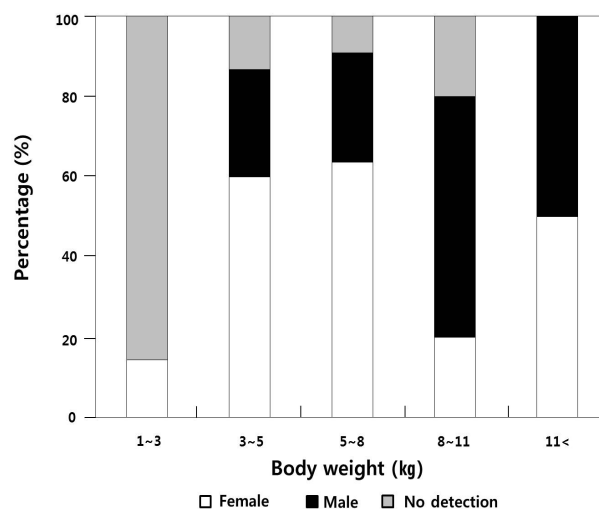


Fig. 1. Sex distribution of the cultured *E. bruneus* with fish size in the rearing tank.

Table 1. Sex reversal of wild caught longtooth grouper *E. bruneus* broodstock in culturing period from year 2002 to 2006 at inner rearing tank

ID number	2003		2004		2005		2006		Starting of inner rearing tank
	BW ¹⁾ (kg)	Sex	BW (kg)	Sex	BW (kg)	Sex	BW (kg)	Sex	
0634-F69B	12.9	F ²⁾	12.9	F	13.2	M	13.6	M	'02. 9
0636-1EF9	4.3	F	4.4	F	5.1	F	7.0	M	'03. 5
0634-DDE0	3.8	F	4.4	M ³⁾	4.5	M	5.2	M	'03. 5
0636-5863	5.1	F	5.3	F	5.3	F	6.7	M	'03. 5
063D-BA84			4.8	F	5.0	M	6.4	M	'03. 9
0633-B2A3			5.7	F	6.2	F	7.3	M	'03. 12
0633-C1AE			7.8	F	7.2	M	6.4	M	'04. 2

¹⁾ BW: body weight, ²⁾ F: female, ³⁾ M: male.

2004, 1 female (total length 72.0 cm, body weight 5.0 kg) in 2005, 3 females (total length 77.0 cm, body weight 6.7 kg; total length 77.0 cm, body weight 7.0 kg; total length 78.0 cm, body weight 7.3 kg) in 2006. In 2005, we obtained 1 sex-reversed male (total length of 70.0 cm, body weight of 7.2 kg) from a sample of 22 females that were bought in 2004. Sex reversal was observed when fish were 63.0 to 99.0 cm total length and 4.4 to 13.2 kg body weight, but most of sex reversal occurred in 5.0 to 8.0 kg body weight.

2. Induced functional males

1) Induced functional males with 17α -methyltestosterone treatment

At the onset to the experiment in March 2003, the gonad of *E. bruneus* mainly contained perinucleolus oocyte (Fig. 2A). 8 weeks after the experiment, the control group had gonad with perinucleolus oocyte (Fig. 2B) but the gonad of MT treated had developed into testes, which the lumen of lobule and efferent duct was filled with sperms (Fig. 2C). 14 weeks after implant silastic capsule, we collected

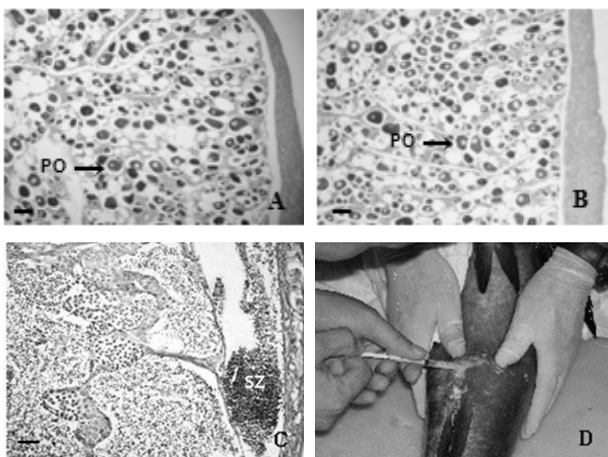


Fig. 2. Histological images of gonads of the sex reversal and spermatogenesis phase in *E. bruneus* by implantation of silastic capsule with 17α -methyltestosterone. A, Initial; B, 8 weeks later on control fish; C, 8 weeks later implantation; D, extraction of semen 12 weeks later implantation. PO, perinucleolus oocyte; SZ, spermatozoa. Scale bar : 200 μ m.

Table 2. Induced spermiation of sex changed male *E. bruneus* implanted with 17α -methyltestosterone (MT 2 mg/kg \cdot BW)

Year ¹⁾	TL ²⁾ (cm)	BW ³⁾ (kg)	MT (mg)	Spermiation	ID number
2003	65	4.2	8	Y ⁴⁾	0633-AAA8
	58	3.2	6	Y	0635-0030
	50	2.6	6	Y	0637-F351
	60	3.6	8	Y	0634-2E83
	66	3.6	8	N ⁵⁾	0634-F4F3
	59	3.1	6	N	0636-586F
2004	60	3.2	6	Y	0636-586F
	68	4.3	8	Y	063D-9358
	54	2.3	5	Y	063D-CD62
	60	3.2	6	N	0642-DFC9
2005	62	3.1	6	N	0633-4997
	69	3.4	6	Y	0642-DFC9
	63	3.4	6	N	063D-F5E4
2006	63	3.5	6	N	063D-AEC2
	59	3.3	6	Y	0634-E476
	60	3.4	6	N	063D-915D

¹⁾ Year: year of MT implantation performed, ²⁾ TL: total length, ³⁾ BW: body weight, ⁴⁾ Y: occurred of spermiation, ⁵⁾ N: not occurred of spermiation.

sperm by pressing the abdomen and were able to collect again 17 days later (Fig. 2D). The following is the yearly amount of induced functional males by silastic capsule that showed spermiation: 4 out of 6 in 2003, 3 out of 5 in 2004, 1 out of 3 in 2005, 1 out of 2 in 2006 (Table 2).

2) Revert to females

Two (2.3 and 2.6 kg) out of 8 sex-reversed males (2.3 to 4.3 kg) with artificial masculinization during 2003 to 2005 reverted back to being female and ovulated. One out of 4 sex-reversed males that had artificial masculinization in 2003 has reverted to female in 2004, and ovulated with HCG treatment in 2006. One out of 5 sex-reversed males that had artificial masculinization in 2004 reverted back to being female in 2005, and ovulated with HCG treatment in 2006 (Table 3).

Table 3. Revert to females in sex-changed males *E. bruneus* implanted with 17 α -methyltestosterone (MT 2mg/kg · BW)

Year ¹⁾	2003		2004		2005		2006		ID number
	Sex	BW ²⁾ (kg)	Sex	BW (kg)	Sex	BW (kg)	Sex	BW (kg)	
2003	M ³⁾	2.6	F ⁴⁾	2.8	F	3.1	F	3.7	0637-F351
	M	3.2	M	3.3	M	4.0	M	4.0	0635-0030
	M	3.2	M	3.5	D ⁵⁾	3.5	D	-	0634-2E83
	M	4.2	M	5.2	M	6.6	M	8.3	0633-AAA8
2004			M	2.3	F	2.7	F	2.9	063D-CD62
			M	3.2	M	3.3	ND ⁶⁾	4.2	0636-586F
			M	4.3	M	4.4	ND	5.0	063D-9358
2005					M	3.4	M	3.9	0642-DFC9

¹⁾ Year: year of MT implantation performed, ²⁾ BW: bodyweight, ³⁾ M: male, ⁴⁾ F: female, ⁵⁾ D: died, ⁶⁾ ND: no detection.

DISCUSSION

Most groupers are protogynous hermaphrodites, which change sex depending on the rearing condition such as age, size and social control. In the different size *E. coioides* of the same rearing tank, the bigger fish changed into males, while the smaller ones remained female (Quinitio et al., 1997). Each species had a different period for the sex reversal. *E. morio* had sex reversal when 9 years old and weighing 11.0 kg, *E. marginatus* when 9 to 16 years old and weighing 6.0 to 10.0 kg (Chauvet, 1988; Randall & Heemstra, 1991), *E. septemfasciatus* when weighing more than 6.0 kg (Tsuchihashi et al., 2003), and *E. malabaricus* when 5 years after hatching and weighing more than 6.0 kg (body length 60.0 to 70.0 cm) (Tang et al., 1979; Tsuchihashi et al., 2003). From this study, inner tank reared *E. bruneus* reversed sex from female to male when their body length was 63.0 to 99.0 cm and their weight was 4.4 to 13.2 kg. In the male population according to the body weight, 5 out of 32 (less than 5.0 kg body weight) were male and 10 out of 18 (5.0 kg or more body weight) were male.

Since groupers, such as *E. septemfasciatus* and *E. marginatus* reverse their sex at a large size, it is difficult to obtain sex-reversed males in the rearing tank for those fish. To solve this problem, many studies are being done to induce small sized females to have sex reversal with androgen treatment (Chen et al., 1977; Tukashima & Kitajima,

1983; Chao & Chow, 1990; Tsuchihashi et al., 2003). Sex reversal in groupers induced by androgen administration is more common, but various dose concentrations and methods of hormone administration. In *E. tauvina*, sex-reversed males were obtained when a dose of 145.0 mg MT for a year and MT 120.0 mg/kg · BW for seven months was given orally, and in *E. septemfasciatus* males sex-reversed when a dose of MT 1.0 mg/kg · BW once a day for 73 days was given orally (Chen et al., 1977; Tukashima & Kitajima, 1983; Chao & Chow, 1990). For the abdominal cavity or muscle injection methods, sex reversal occurred when MT 30.0 mg/kg · BW was injected 6 times every other week to *E. coioides* (Chao & Lim, 1991). *E. suillus* were induced to functional males when injected with a MT 0.5 mg/kg · BW silastic capsule over 4 months (Tan-Fermin et al., 1994). *E. septemfasciatus* had spermiation 2 months after a MT 1.0 mg/kg · BW and 4.0 mg/kg · BW silastic capsule was implanted and was induced to have sex reversal when injected with MT 0.5 to 2.0 mg/kg · BW and a MT 1.0 to 2.0 mg/kg · BW silastic capsule implanted into the abdomen (Tsuchihashi et al., 2003; Song, 2004). *E. merra* were induced to functional males after 24 days injecting an aromatase inhibitor fadrozole 1.0 mg/kg · BW with a silastic capsule during the maturation season (Alam et al., 2006). For *E. bruneus*, we obtained functional males from the 12 weeks of implant a MT 2.0 mg/kg · BW silastic capsule into the abdomen and was able to identify spermiation in 9 out 14 fish. In

some specimen, we did not produce sex-reversed males due to the silastic capsules slipping out of the fish.

Two sex-reversed males reverted back to female 2 to 3 years after the hormone treatment. It was due to the aspect of hormone treatment for immature fish (body weight 2.3 to 2.6 kg). We suggested that immature fish had androgen with MT silastic capsule for a while, and they reverted back to female because there is no effect or low concentration level of MT treatment. This result corresponds to the result of an experiment where MT treated sex-reversed males did not remain as males and reverted back to being females during the next maturation period (Marino et al., 2000; Sarter et al., 2006). These results show that masculinization in *E. bruneus* with MT treatment is more effective for fish weighing more than 3.0 kg to have sex-reversed males remain as functional males.

REFERENCES

- Alam MA, Bhandari RK, Kobayashi Y, Soyano K, Nakamura M (2006) Induction of sex change within two full moons during breeding season and spawning in grouper. *Aquaculture* 255:532-535.
- Chao TM, Chow M (1990) Effects of methyltestosterone on gonadal development of *Epinephelus tauvina* (Forsk.) in Singapore. *J Pri Ind* 18:1-14.
- Chao TM, Lim LC (1991) Recent development in the breeding of grouper (*Epinephelus* spp.) in Singapore. *Singapore J Pri Ind* 19:79-33.
- Chauvet C (1988) Study of the growth of the grouper *Epinephelus guaza* (Linnaeus, 1758) from the Tunician coasts. *Aquat Living Resour* 1:277-288.
- Chen FY, Chow M, Chao TM, Lim R (1977) Artificial spawning and larval rearing of the grouper, *Epinephelus tauvina* (Forsk.) in Singapore. *Singapore J Pri Ind* 5: 1-21.
- FAO (1993) FAO Species Catalogue Vol. 16. Groupers of the World. pp. 1-10. FAO Rome.
- Glamuzina B, Glavic N, Tutman P, Kozul V, Skaramuca B (2000) Notes on first attempt at artificial spawning and rearing of early stages with gold blotch grouper, *Epinephelus costae* (Steindacner, 1875). *Aquacult Int* 8:551-555.
- Hamamoto S, Yokogawa K, Tochino M (1986) Several problems on cultivating the parent fish of red spotted grouper, *Epinephelus akaara* (Temminck et Schlegel) and judging the qualities of the eggs obtained from them. *Bull Kagawa Pref Fish Exp Sta* 2:13-22.
- Hwang SI, Lee YD, Song CB, Rho S (1998) Gonadal development and the effect of 17 α -methyltestosterone on sex inversion of the red spotted grouper, *Epinephelus akaara*. *J of Aquaculture* 11:173-182. (in Korean)
- Inoue M (2001) Effects of water temperature on feeding and growth of juvenile kelp grouper, *Epinephelus moara*. *Bull Fish Res Inst Mie* 9:35-38. (in Japanese)
- Kim IS, Choi Y, Kim BJ (2001) Percoid fishes of Korea. Korea Research Institute of Bioscience and Biotechnology Korea. pp. 279.
- Lee CK, Hur SB, Ko TK, Park S (1998) Maturation, sex ratio and sex reversal of red spotted grouper, *Epinephelus akaara*. *J of Aquaculture* 11:573-580. (in Korean)
- Lee YD, Song YB, Lim BS, Oh SR, Kim HB (2008) Grouper aquaculture research in Jeju Island, Korea. *Bull Mar Environ Res Inst* 32:49-60.
- Manabe S, Kasuga A. 1988. Spawning behavior and early life history of *Epinephelus moara* in an aquarium. *J Jpn Assoc Zool Gardens Aquariums* 30:16-24.
- Marino G, Azzurro E, Finioia MG, Messina MT, Massari A, Mandrich A (2000) Recent advances in induced breeding of the dusky grouper *Epinephelus marginatus* (Lowe, 1834). *Cah Options Mediterr* 47:215-225.
- Okada T, Yoneshima H, Mukai Y, Sawada Y (1996) Tolerance limits of young kelp grouper, *Epinephelus moara* for environmental stress. *Bull Fish Lab Kinki Univ* 5:139-146.
- Quinitio GE, Caberoy NB, Reyes Jr DM (1997) Induction of sex change in female *Epinephelus coioides* by social control. *Isr J Aqua Bamidgeh*. 49:77-83.
- Randall JE, Heemstra PC (1991) Revision of Indo-Pacific

- groupers (Perciformes: Serranidae: Epinephelinae). With descriptions of five new species. Indo-Pacific Fishes No. 20. Bernice Pauahi Bishop Museum. Honolulu. Hawaii, pp. 322.
- Ryu YW, Hur SW, Hur SP, Lee CH, Lim BS, Lee YD (2013) Characterization of pubertal development phases in female longtooth grouper, *Epinephelus bruneus* via classification of bodyweight. *Dev Reprod* 17:55-62.
- Sarter K, Papadaki M, Zanuyan S, Mylonas CC (2006) Permanent sex inversion in 1-year-old juveniles of the protogynous dusky grouper (*Epinephelus marginatus*) using controlled-release 17 α -methyltestosterone implants. *Aquaculture* 256:443-456.
- Sawada Y, Kato K, Okada T, Kurata M, Mukkai Y, Miyashita S, Murata O, Kumai H (1999) Growth and morphological development of larval and juvenile *Epinephelus bruneus* (Perciformes:Serranidae). *Ichthyol Res* 46:245-257.
- Song YB (2004) Induction of sexual maturation and early development of the sevenband grouper, *Epinephelus septemfasciatus*. Ph.D. thesis, Jeju National University Kor. pp. 120. (in Korean)
- Tsuchihashi Y, Tanaka H, Kuromiya Y, Kashiwagi M, Yoshioka M (2003) Control of induction of sex reversal in the sevenband grouper, *Epinephelus septemfasciatus*. *Suisanzoshoku* 51(2):189-196.
- Tukashima Y, Kitajima C (1983) Acceleration of sex reversal from immature female into functional males through oral administration of methyltestosterone in sevenband grouper, *Epinephelus septemfasciatus* (Thunberg). *Bull Nagasaki PrefInst Fish* 9:55-57.
- Tan-Fermin JD, Garcia LMB, Castillo AR Jr (1994) Induction of sex inversion in juvenile grouper, *Epinephelus suillus* (Valenciennes) by injection of 17 α -methyltestosterone. *Japan J Ichthyol* 40:413-420.
- Tang HC, Twu JY, Su WC (1979) Preliminary report on artificial propagation of black spotted grouper, *Epinephelus amblycephalus* Bleeker. *Bull Taiwan Fish Res Inst.* 31:511-517.
- Yang MH, Choi YW, Jung MM, Ku HD, Oh BS, Moon TS, Lee CH, Kim KM, Han SJ (2007) Temperature effect in egg development and hatching of longtooth grouper, *Epinephelus bruneus*. *Dev Reprod* 11(2): 105-109.