

A New Saponin from the Seeds of *Zizyphus jujuba* var. *spinosa*So Young Lee, Ju Sun Kim, Je-Hyun Lee,[†] Yeong Shik Kim, and Sam Sik Kang*

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Sour jujube (*Zizyphus jujuba* Miller var. *spinosa* Hu ex H. F. Chou) is a thorny, rhamnaceous deciduous plant distributed widely throughout tropical and subtropical regions. The dried seeds are named sanjoin (*Zizyphi Spinosi Semen*) in Korean and have a thousand-year history of use in traditional medicine as a sedative for treating mild anxiety, nervousness and sleep-related problems.¹ Previous phytochemical investigations on the seeds of this plant have resulted in the isolation of saponins,²⁻⁶ flavone C-glycosides,⁷⁻¹² cyclopeptide alkaloids,¹ triterpenoids,^{13,14} and others.⁶ These compounds exhibit multiple activities, such as hypnotic,¹⁵ histamine-release inhibitory,⁴ anti-inflammatory¹⁶ and sedative,¹⁶⁻¹⁸ inhibition of foam cell formation,¹⁴ adjuvant,⁴ and cardiotoxic¹⁹ effects. Our previous studies focused on isolating the flavonoid-C-glycosides^{7,8} and the sedative activities of these flavonoids.^{17,18} In the continuing search for new chemicals and biomarkers from *Z. jujuba* var. *spinosa* for quality control studies of related herbal medicines, 8 flavonoids, along with 3 other known components, were isolated.²⁰ In the present investigation, we report the isolation and structural elucidation of a new saponin, named jujuboside A₂ (**1**), together with three known saponins, zizyphus saponin III (**2**),²¹ jujuboside B (**3**),^{2,21} jujuboside A (**4**),² and a known sesquiterpene glucoside, dihydrophaseic acid 3-O-β-D-glucopyranoside (**5**),²² from the seeds of *Z. jujuba* var. *spinosa*.

Compound **1** was obtained as a colorless amorphous powder. Its molecular formula was established as C₅₈H₉₄O₂₆ from the [M-H]⁻ peak at *m/z* 1205.5961 (Calcd for C₅₈H₉₃O₂₆: 1205.5955) in the HR-FAB-MS. The IR absorption bands at 3414, 1647, and 1043 cm⁻¹ implied the presence of hydroxyl, double bond, and glycosidic C-O functionalities. Acid hydrolysis of compound **1** with 5% HCl afforded a mixture of ebelin lactone and 17(*Z*)-ebelin lactone as the aglycon³ and D-glucose, 6-deoxy-L-talose, D-xylose, and L-arabinose as the sugar components identified on GLC analysis of the thiazolidine derivatives.²³ The ¹H- and ¹³C-NMR spectra of **1** suggested the presence of a jujubogenin moiety [δ_{H} 0.72, 0.97, 1.09, 1.10, 1.38, 1.67, and 1.70 (all s, CH₃-19, CH₃-29, CH₃-18, CH₃-28, CH₃-21, CH₃-26, and CH₃-27), 3.18 (dd, *J* = 4.0, 11.7 Hz, H-3), 5.20 (br t, *J* = 9.5 Hz, H-23), and 5.53 (br d, *J* = 7.9 Hz, H-24)] and five monosaccharide units through easily identifiable signals for anomeric protons and carbons [δ_{H} 4.86 (d, *J* = 5.2 Hz), 4.90 (d, *J* = 7.7 Hz), 5.00

(d, *J* = 7.6 Hz), 5.43 (d, *J* = 7.3 Hz), and 6.11 (br s); δ_{C} 104.4, 105.2, 103.6, 106.0, and 101.7]. The (-)-FAB-MS showed an [M-H]⁻ ion at *m/z* 1205, which was consistent with a pentasaccharide glycoside carrying two glucose, one 6-deoxy-L-talose, two pentose (xylose and arabinose) units, and an aglycon with a molecular mass of 472. The presence of other

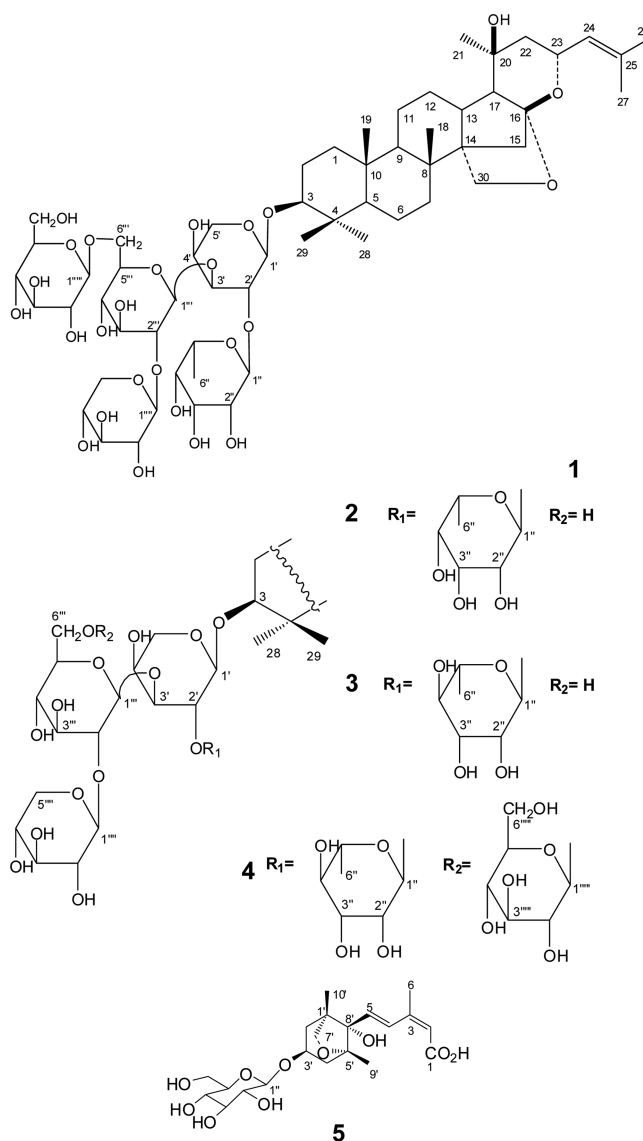


Figure 1. Structural formulas of compounds 1–5.

fragment ions at m/z 1073 [(M-H)-132]⁻, 1059 [(M-H)-146]⁻, 1043 [(M-H)-162]⁻, 911 [(M-H)-132-162]⁻, 749 [(M-H)-132-162-162]⁻, 603 [(M-H)-132-162-162-146]⁻, and 471 [(M-H)-132-162-162-146-132]⁻, supported this suggestion. Complete assignments of the ¹H- and ¹³C-NMR signals of compound **1**, assigned by extensive 2D-NMR experiments, led to the identification of three terminal sugars, β-glucopyranosyl, β-xylopyranosyl, and 6-deoxy-α-talopyranosyl, 2,6-disubstituted β-glucopyranosyl, and 2,3-disubstituted α-arabinopyranosyl unit. The linkage sites, and sequences of the five saccharides and of the aglycon were confirmed by the HMBC experiments. The HMBC exhibited cross peaks between H-1' of the arabinose and C-3 of aglycon, H-1 of 6-deoxy-talose and C-2' of arabinose, H-1'' of the inner glucose and C-3' of arabinose, H-1'''' of xylose

and C-2''' of the inner glucose, and H-1'''' of glucose and C-6''' of the inner glucose. Thus, **1** was elucidated as jujubogenin 3-*O*-β-D-glucopyranosyl(1→6)-[β-D-xylopyranosyl(1→2)]-β-D-glucopyranosyl(1→3)-[6-deoxy-α-L-talopyranosyl(1→2)]-α-L-arabinopyranoside, named jujuboside A₂ (**1**), which had close similarity to jujuboside A₁, except that D-fucose in jujuboside A₁ was replaced by 6-deoxy-L-talose in **1**. Additionally, by comparing its NMR spectral data with those in the literature,³ the ¹H- and ¹³C-NMR spectral features of **1** and jujuboside A₁ were identical to each other as indicated in Table 2. The only difference was the ¹³C-NMR chemical shift assignment of the ¹³C-NMR chemical signals arising from C-2'', C-3'' and C-5'' of D-fucose in jujuboside A₁ and 6-deoxy-L-talose in **1**. The known compounds were identified as zizyphus saponin III (**2**),²¹ jujuboside B (**3**),^{2,5,21}

Table 1. ¹H-NMR data^a for sugar moieties of **1–4** in pyridine-*d*₅. δ in ppm, *J* in Hz

No.	1 ^b	2 ^b	3 ^c	4 ^c
	Arabinose	Arabinose	Arabinose	Arabinose
1'	4.86 (d, <i>J</i> = 5.2)	4.81 (d, <i>J</i> = 5.1)	4.88 (d, <i>J</i> = 3.8)	4.88 (d, <i>J</i> = 3.5)
2'	4.74 (t, <i>J</i> = 6.4)	4.74 (dd, <i>J</i> = 5.1, 6.9)	4.77 (t, <i>J</i> = 5.3)	4.72 (t, <i>J</i> = 5.0)
3'	4.36 (dd, <i>J</i> = 2.3, 9.5)	4.30 (dd, <i>J</i> = 2.7, 7.2)	4.30–4.34 (m)	4.30 (t, <i>J</i> = 4.4)
4'	4.49–4.54 (m)	4.43–4.48 (m)	4.44–4.50 (m)	4.43–4.52 (m)
5'	3.99 (br d, <i>J</i> = 10.6)	3.70 (dd, <i>J</i> = 2.3, 12.1)	3.71 (br d, <i>J</i> = 11.8)	3.91 (br d, <i>J</i> = 10.7)
	4.26 (dd, <i>J</i> = 3.9, 10.6)	4.23 (dd, <i>J</i> = 5.2, 12.1)	4.22–4.28 (m)	4.32 (dd, <i>J</i> = 4.4, 10.7)
	6-Deoxy-talose	6-Deoxy-talose	Rhamnose	Rhamnose
1''	6.11 (br s)	6.13 (br s)	5.96 (br s)	5.89 (br s)
2''	4.48 (br s)	4.49 (br s)	4.65 (br s)	4.60 (br s)
3''	4.42 (br s)	4.42 (br s)	4.53 (dd, <i>J</i> = 2.9, 9.1)	4.50 (dd, <i>J</i> = 3.3, 8.5)
4''	4.13 (br s)	4.11 (br s)	4.27 (t, <i>J</i> = 8.5)	4.27 (t, <i>J</i> = 8.5)
5''	4.66–4.70 (m)	4.67 (br q, <i>J</i> = 6.3)	4.49–4.55 (m)	4.44–4.53 (m)
6''-CH ₃	1.56 (d, <i>J</i> = 6.4)	1.54 (d, <i>J</i> = 6.4)	1.63 (d, <i>J</i> = 6.0)	1.61 (d, <i>J</i> = 6.0)
	Glucose	Glucose	Glucose	Glucose
1'''	5.00 (d, <i>J</i> = 7.6)	5.11 (d, <i>J</i> = 7.6)	5.10 (d, <i>J</i> = 7.4)	4.95 (d, <i>J</i> = 7.6)
2'''	4.14 (t, <i>J</i> = 8.6)	4.11–4.18 (m)	4.10–4.17 (m)	4.09 (t, <i>J</i> = 8.4)
3'''	4.23 (t, <i>J</i> = 9.0)	4.26–4.33 (m)	4.14 (t, <i>J</i> = 8.3)	4.17–4.22 (m)
4'''	4.21 (t, <i>J</i> = 9.0)	4.05–4.12 (m)	4.07 (t, <i>J</i> = 9.0)	4.19 (t, <i>J</i> = 8.7)
5'''	4.03–4.07 (m)	3.91 (ddd, <i>J</i> = 2.5, 5.9, 9.5)	3.87–3.94 (m)	3.99–4.04 (m)
6'''	3.97 (br d, <i>J</i> = 10.5)	4.23 (dd, <i>J</i> = 5.2, 12.1)	4.17–4.21 (m)	3.88 (br d, <i>J</i> = 10.6)
	4.87 (br d, <i>J</i> = 10.5)	4.48 (br d, <i>J</i> = 12.1)	4.45–4.51 (m)	4.84 (br d, <i>J</i> = 10.6)
	Xylose	Xylose	Xylose	Xylose
1''''	5.43 (d, <i>J</i> = 7.3)	5.40 (d, <i>J</i> = 7.5)	5.35 (d, <i>J</i> = 7.1)	5.33 (d, <i>J</i> = 6.4)
2''''	4.18 (t, <i>J</i> = 7.5)	4.11–4.20 (m)	4.17 (t, <i>J</i> = 8.4)	4.13 (t, <i>J</i> = 7.0)
3''''	4.14 (t, <i>J</i> = 8.6)	4.06–4.17 (m)	4.29 (t, <i>J</i> = 10.2)	4.07–4.17 (m)
4''''	4.21 (t, <i>J</i> = 9.0)	4.14–4.21 (m)	4.13–4.22 (m)	4.12–4.22 (m)
5''''	3.78 (t, <i>J</i> = 10.8)	3.73 (t, <i>J</i> = 11.0)	3.76 (t, <i>J</i> = 10.8)	3.77 (t, <i>J</i> = 11.3)
	4.49 (dd, <i>J</i> = 2.3, 11.9)	4.45 (dd, <i>J</i> = 5.5, 11.8)	4.44–4.50 (m)	4.46 (br d, <i>J</i> = 11.3)
	Glucose			Glucose
1'''''	4.90 (d, <i>J</i> = 7.7)			4.85 (d, <i>J</i> = 7.8)
2'''''	4.05 (t, <i>J</i> = 8.4)			3.98–4.04 (m)
3'''''	4.25 (t, <i>J</i> = 8.8)			4.21 (t, <i>J</i> = 8.6)
4'''''	3.84 (t, <i>J</i> = 9.1)			3.80 (t, <i>J</i> = 9.4)
5'''''	3.87–3.92 (m)			3.81–3.89 (m)
6'''''	4.37 (br d, <i>J</i> = 11.9)			4.29–4.34 (m)
	4.49 (dd, <i>J</i> = 2.3, 11.9)			4.46 (br d, <i>J</i> = 12.1)

^aAll δ_H assignments are based on 2D-NMR (¹H-¹H COSY, DEPT, HMQC, HMBC). ^b500 MHz. ^c400 MHz.

jujuboside A (**4**),^{2,5} and (1*R*,3*S*,5*R*,8*S*,2*Z*,4*E*)-dihydrophaseic acid 3'-*O*- β -D-glucopyranoside (**5**)²² by detailed NMR spectroscopic analysis and by comparing the physical and spectral data with those reported in the literature. Although partial NMR spectral data have been reported for some of the isolated compounds (**2–4**), the complete ¹H- and ¹³C-NMR spectral assignments of all the isolated compounds are reported herewith for the first time based on ¹H-¹H COSY, HMQC, HMBC, and NOESY spectroscopic data as indicated in Tables 1 and 2 and supplementary Table, which led to the revision of chemical shifts for some carbons such as C-18, C-27, C-3, and C-2" in **2–4** (Table 2).^{5,21} The small *J* value for the H-1 of the arabinopyranosyl moiety of **3** and **4** (3.5–3.8 Hz) suggested that the arabinose ring predominantly adopts ¹C₄ conformation in **3** and **4**, whereas in **1** and **2**, the arabinose ring predominantly adopts ⁴C₁ conformation.^{23–25}

Selective 1D NOESY data obtained by irradiating the H-1' signal showed the sub-spectrum of the arabinosyl residue, which showed NOEs from H-1' to H-2', H-3', and H-5' in **3**

and **4**, and from H-1' to H-3' and H-4' in **1** and **2**, respectively. These results suggest that the rhamnose moiety at C-2' in arabinose increases the population of the ¹C₄ conformation.^{23–25} (1*R*,3'*S*,5'*R*,8'*S*,2*Z*,4*E*)-Dihydrophaseic acid 3'-*O*- β -D-glucopyranoside (**5**) has been isolated from several plant species,²² and our study seems to be the first instance of its isolation from the genus *Zizyphus*. Ten dammarane-type triterpene glycosides such as jujubosides A,² B,^{2,21} C,³ E,⁵ A₁ (= D),^{3,6} B₁,³ acetyljujubosides B,³ protojujubosides A,⁴ B⁴ and B₁⁴ have been isolated as the saponin constituents from the seeds of this plant. Among these saponins, jujubosides A, B, and A₁ have been isolated as the major components of *Zizyphi Spinosi Semen*,³ but we could not isolate jujuboside A₁ from our sample.

In view of the previous report on the reducing the risk of cardiovascular disease,²⁶ three major saponins (**1**, **3** and **4**) were evaluated for their antiplatelet effects. Only jujuboside B (**3**) showed a significant inhibitory effect on collagen-, thrombin- and arachidonic acid-induced platelet aggregation.²⁷

Table 2. ¹³C-NMR data^a of **1–4** in pyridine-*d*₅

No.	A ₁ ⁵	1 ^b	2 ^b	3 ^c	4 ^c	No.	A ₁	1 ^b	2 ^b	3 ^c	4 ^c
1	38.8	38.8	38.8	38.8	38.8	Arabinose					
2	26.6	26.5	26.6	26.5	26.4	1'	104.5	104.4	104.5	104.2	103.9
3	88.1	88.1	88.0	88.3	88.3	2'	74.6	74.6	74.5	74.9	75.1
4	39.5	39.5	39.6	39.7	39.6	3'	83.1	83.1	82.66	82.2	82.8
5	56.2	56.2	56.2	56.2	56.1	4'	68.5	68.3	68.2	67.8	67.8
6	18.3	18.3	18.3	18.4	18.3	5'	64.5	64.3	64.5	63.6	63.2
7	36.0	36.0	36.0	36.0	36.0	6-Deoxyhexose					
8	37.5	37.5	37.5	37.5	37.5	1"	101.7	101.7	101.8	101.6	101.6
9	53.0	52.9	53.0	53.0	52.9	2"	67.7	72.1	72.1	72.3	72.3
10	37.2	37.2	37.2	37.2	37.2	3"	72.1	67.0	67.1	72.5	72.5
11	21.7	21.7	21.7	21.7	21.7	4"	74.2	74.2	74.2	73.9	73.9
12	28.5	28.5	28.5	28.5	28.5	5"	67.0	67.8	67.8	70.0	70.0
13	37.1	37.1	37.1	37.1	37.1	6"	17.3	17.3	17.3	18.5	18.5
14	53.7	53.7	53.7	53.7	53.7	Glucose					
15	36.9	36.8	36.9	36.8	36.8	1'''	103.6	103.6	103.4	103.7	103.9
16	110.6	110.5	110.5	110.6	110.5	2'''	82.1	82.3	82.74	83.4	83.2
17	54.0	53.9	54.0	54.0	53.9	3'''	78.1	78.1	78.47	78.1	78.0
18	18.9	18.8	18.9	18.9	18.8	4'''	71.4	71.5	71.2	71.2	71.4
19	16.3	16.3	16.3	16.4	16.3	5'''	76.7	76.7	78.5	78.5	76.5
20	68.5	68.4	68.5	68.5	68.4	6'''	70.3	70.3	62.4	62.4	70.3
21	30.0	30.0	30.0	30.0	30.0	Xylose					
22	45.4	45.4	45.5	45.4	45.4	1''''	105.9	106.0	106.0	106.3	106.4
23	68.6	68.5	68.6	68.6	68.5	2''''	75.9	76.0	75.9	76.1	76.2
24	127.1	127.1	127.1	127.1	127.0	3''''	78.1	78.0	78.2	78.3	78.1
25	134.2	134.1	134.1	134.1	134.1	4''''	70.8	70.8	70.9	70.8	70.8
26	25.6	25.5	25.5	25.5	25.5	5''''	67.7	67.7	67.7	67.8	67.8
27	18.3	18.3	18.3	18.3	18.3	Glucose					
28	28.0	28.0	28.0	28.1	28.1	1'''''	105.2	105.2			105.1
29	16.7	16.7	16.7	16.9	16.9	2'''''	75.3	75.3			75.3
30	65.8	65.8	65.8	65.8	65.8	3'''''	78.4	78.4			78.0
						4'''''	71.4	71.5			71.4
						5'''''	78.4	78.4			78.4
						6'''''	62.5	62.5			62.5

^aAll δ assignments are based on 2D-NMR (¹H-¹H COSY, DEPT, HMQC, HMBC). ^b125 MHz. ^c100 MHz.

Experimental Section

Plant Material. The seeds of *Z. jujuba* var. *spinosa* were cultivated at Jinan, Shandong province, China, harvested in October 2009, and authenticated by Dr. J.-H. Lee, one of the authors. A voucher specimen was deposited in the Herbarium of the Traditional Herb Research Center, Korea Food and Drug Administration [no. 11E-1001].

Jujuboside A₂ (1): $[\alpha]_{\text{D}}^{28} = -55.4^{\circ}$ (c 0.55, MeOH); IR (KBr) cm^{-1} : 3414, 1647, 1043, 980, 813; (–)-FAB-MS m/z : 1205 [M–H][–], 1073 [M–132][–], 1059 [M–146][–], 1043 [M–162][–], 911 [M–132–162][–], 749 [(M–H)–132–162–162][–], 603 [(M–H)–132–162–162–146][–]; HR-FAB-MS m/z : 1205.5961 [M–H][–] (Calcd for C₅₈H₉₃O₂₆: 1205.5955); ¹H- and ¹³C-NMR: see Tables 1 and 2.

6-Deoxy-L-talopyranose: $[\alpha]_{\text{D}}^{24} = -54.0^{\circ}$ (c 0.3, MeOH); FAB-MS m/z : 187 [M+Na]⁺, 165 [M+H]⁺; ¹H-NMR (pyridine-*d*₅) δ 8.36 (1H, d, $J = 3.6$ Hz, 1-HO), 6.64 (1H, br s, 2-HO), 6.23 (1H, br s, 3-HO), 6.29 (1H, br s, 4-HO), 5.91 (1H, br s, H-1), 4.49 (1H, br s, H-2), 4.55 (1H, br s, H-3), 4.11 (1H, br s, H-4), 4.61 (1H, q, $J = 6.5$ Hz, H-5), 1.56 (3H, d, $J = 6.5$ Hz, 6-CH₃); ¹³C-NMR (pyridine-*d*₅) δ 96.5 (C-1), 73.4 (C-2), 67.3 (C-3), 74.5 (C-4), 67.1 (C-5), 17.8 (C-6) for major 6-deoxy- α -L-talopyranose. ¹H-NMR (500 MHz, pyridine-*d*₅) δ 5.07 (1H, br s, H-1), 4.37 (1H, br s, H-2), 3.95 (1H, br s, H-3), 3.91 (1H, br s, H-4), 3.67 (1H, q, $J = 6.5$ Hz, H-5), 1.53 (3H, d, $J = 6.5$ Hz, 6-CH₃); ¹³C-NMR (pyridine-*d*₅) δ 96.2 (C-1), 73.9 (C-2), 70.8 (C-3), 73.2 (C-4), 71.8 (C-5), 17.8 (C-6) for minor 6-deoxy- β -L-talopyranose. The integrals and the ¹³C signal intensities provided 6-deoxy- α -L-talopyranose: 6-deoxy- β -L-talopyranose anomer ratio of about 81:19.

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Supporting Informations. General procedures, Extraction and Isolation, Acid Hydrolysis, ¹H-NMR data for jujubogenin moiety of 1–4 (Table) and NMR and MS spectra of 1 are available as Supporting Information.

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