

Differential Expression Analysis of Candidate Genes Related with Growth according to Dietary Supplementation of *Curcuma longa* in Chickens

Sun-Ae Park^a, Lee-Kyung Kim^a, Chang-Min Park, Seung-Chang Kim,
Seung-Hwan Lee, Ji-Woong Lee^{1,*} and Bong-Hwan Choi[†]

Division of Animal Genomics & Bioinformatics, National Institute of Animal Science, RDA, Suwon 441-706, Republic of Korea

¹Division of Animal Science, Insti. of Ag. Sci. and Tech., Chonnam National University, Chonnam 500-757, Republic of Korea

ABSTRACT

This experiment was conducted to investigate the genetic effects of candidate genes on the growth of spleen and liver tissues using dietary *Curcuma longa* (*C. longa*) supplementation. Expression analyses of candidate genes regarding animal growth was performed in order to determine the factors affecting the growth related to immune components of Curcumin, Turmerone, and Zingiberene as the bile secretion *Paratolyl methyl carbinol* (PTMC). The animals were divided into four groups of five chicks supplied with experimental diets of *C. longa* at 0.25, 0.5 and 1% and controls. The 19 growth-related genes were known to cell maturation, differentiation significant expression patterns in this analysis. Expression of growth response-related genes in chicks supplemented with 1% of *C. longa* showed better growth performance than chicks with 0.25 and 0.5% in spleen ($p < 0.05$). The IGF1, MSTN, POU1F1, ADCYAP1 gene were known to central roles in mediating gonadotropin function, regulating steroidogenesis and promoting oocyte growth and maturation. Sex steroids, androgen and estrogen can affect sex differentiation and also can affect muscle development. On the other hand, GHSR and FABP3 gene showed significant expression patterns in this analysis. The results would be used as basic information for the variation of growth-related genes expression on the cell growth, sex cell growth, and sex hormones according to dietary supplementation with *C. longa* in chickens.

(Key words : *Curcuma longa*, Differential expression analysis, Genes related growth, Chicken)

INTRODUCTION

Several quantitative traits for production such as growth, egg-laying, feed conversion, carcass weight and body weight at different day-ages are important in domestic animals.

Several chemical compounds and antibiotics have been identified in herbaceous plants by researchers, which play a key role in human and animal health. The medicinal plant *Curcuma longa* is commonly used as a spice in human food. *Curcuma longa* (*C. longa*), a perennial herb which is known as tumeric, is a member of Zingiberaceae. The plant grows to height of 3 to 5 feet and has oblong pointed leaves, which bears funnel shaped yellow flowers (Durrani *et al.*, 2006). The rhizome is the portion of the plant used medicinally; it is usually boiled, cleaned, and dried, yielding a yellow

powder, tumeric, the ingredient that gives curry powder its characteristic yellow color (AL-Sultan, 2003). *C. longa* is medicinal plant widely used cultivated in tropical regions. Plant extracts were found to have anti-fungal, immunomodulatory, anti-oxidative and anti-mutagenic (Soni *et al.*, 1997) activities. Some of pharmacological activities of *C. longa* as nematocidal (Kiuchi *et al.*, 1993) and anti-inflammatory (Ammon *et al.*, 1993) were demonstrated. Furthermore, the plant was used predominantly for endoparasites as well as internal and external injuries (Lans and Brown, 1998). More over (Soni *et al.*, 1997) proved the protective effects of *C. longa* as food additives on aflatoxin-induced mutagenicity and hepatocarcinogenicity (Durrani *et al.*, 2006). Keeping in view the significant importance of *C. longa*, this research study was conducted to investigate the effect of *C. longa* on the growth of broiler chicks.

The experiment was designed to investigate the pos-

* This work was supported by the "Study of gene expression and useful material for immunity improvement in Hanwoo" project of the National Institute of Animal Science, RDA, Korea.

^a These authors contributed equally to this work.

[†] Corresponding author : Phone: +82-31-290-1592, E-mail: bhchoi@korea.kr, Phone: +82-62-503-2111, E-mail: jwlee@jnu.ac.kr

sible effect of *C. longa* feed additive on the growth performance of 20 broiler chickens. And the experiment was designed to select of 19 growth-related gene associated cell growth and sex cell growth.

MATERIALS AND METHODS

Experimental Design

The experimental chicks were divided at age of day-old into 4 groups, each consisted of 20 chicks and assigned as group 1-4. Birds group 1 were fed basal diet, while groups 2, 3, and 4 were fed basal diet supplemented with 0.25 (A group), 0.5 (B group) and 1% (C group) *C. longa*.

Sample Collection

On 21 days, all birds were weighed and feed consumption was records for each pen. Average feed intake and BW gain were determined. Liver and spleen tissue was collected, with at least 1 bird representing each pen. Collected liver and spleen tissue samples were snap-frozen in liquid nitrogen and stored at -80°C for real-time PCR analysis.

Statistical Analysis

The normal distribution of data was confirmed by the χ^2 goodness of fit test. Data were subjected to repeated-measures ANOVA using the R program. Data in the same tissue sampling time were also compared using student's *t*-test. The threshold of significance was 0.05.

RNA Extraction, Reverse Transcription, and Quantitative Real-Time PCR

Ribonucleic acid was extracted from the liver and spleen samples using an TRIzol Reagent (ambion, USA) and Chloroform (Merk, USA), and concentrated using RNase free water. The quality and integrity of the purified RNA was checked through agarose gel electrophoresis and the quantity was measured using an ND-1000 spectrophotometer (Nanodrop Technologies, Wilmington, DE; Flanagan, 2005). The purified RNA samples were preserved at -80°C until used. Two-step quantitative real-time PCR was used to measure expression patterns of genes involved in growth. From each chick liver and spleen, 10ug of total RNA was reverse-transcribed using Super Script II Reverse Transcriptase (invitrogen, Carlsbad, CA, USA) with oligo dT (200 U/ul, Promega, USA) and random primer (200 U/ul, Promega, USA). Forward and reverse primer final concentrations were 100 nM in the SYBR green assay (Applied

Biosystems, Foster City, CA). Primers were designed using the Primer 3 program (Rozen and Skaletsky, 2000) with an annealing temperature of 60°C and amplification size of less than 200 bp (Table 1). Glyceraldehyde phosphated dehydrogenase (GAPDH) was used as the endogenous control gene in the RT-PCR experiments.

RESULTS AND DISCUSSION

Expression of growth response-related genes in chicken supplemented with 1% of *C. longa* showed better growth performance than chicks with 0.25 and 0.5% in spleen ($p<0.05$). The 19 growth-related genes were known to cell maturation, differentiation significant expression patterns in this analysis.

That dietary *C. longa* supplementation modulates the growth response partly through changes in the expression growth-related genes such as TGFBR1, IGF1, IGF-2, MSTN, GHSR, TGFBR2, PDGFB, IGF1R, CTGF, FGF-2, GH1, POU1F1, GHRH, FABP3, ADCYAP1, BMPR2, CAPN3, IL1B and SST in liver and spleen. As shown in the above experiments dietary *C. longa* supplementation modulates the growth response partly through changes in the expression growth-related genes. Fig. 1 summary the data on mRNA expression of substances related to growth responses in the liver in experiment. The highest expression of IGF1, MSTN, POU1F1 and ADCYAP1 gene was observed 0.25, 0.5 and 1.0% that of fed *C. longa* ($p<0.05$). Especially more highly growth-related genes was observed 0.25% group in liver ($p<0.05$). And GHSR, FABP3, IL1B gene was very low observed 0.5% group in liver ($p<0.01$). Fig. 2 summary the data on mRNA expression of substances related to inflammatory responses in the spleen in experiment. The highest expression of ADCYAP1, IL1B and SST gene was observed 0.25, 0.5 and 1.0% that of fed *C. longa*. Especially more highly growth-related genes were observed 1.0% group in spleen. But growth gene expression in spleen was never significant. The IGF1, MSTN, POU1F1 and ADCYAP1 gene were known as follows. The IGF1 (insulin-like growth factors), isolated from plasma, are structurally and functionally related to insulin but have a much higher growth-promoting activity (provided by RefSeq, Mar 2009). IGF genes also regulate development (Silha *et al.*, 2005) and male sex determination (Nef *et al.*, 2003). At the level of the gonad, IGFs play central roles in mediating gonadotropin function (Yu *et al.*, 2003), regulating steroidogenesis (Mukhejee *et al.*, 2006) and promoting oocyte growth and maturation (Santos *et al.*, 2006). The protein encoded by MSTN (myostatin) gene is a member of the bone morphogenetic protein (BMP) family and the TGF-beta su-

Table 1. Primer sequences used in real-time PCR

Gene	Primer sequence		Product size(bp)
	Forward(5'-3')	Reverse(3'-5')	
TGFBR1	AGCTGCGGACAACAAAGACAATGG	CAATTGCTGGTTTGCCTTGTGTGC	200
IGF1	TGGATGAGTGCTGCTTCCAGAGTT	GAGCGTGCAGATTTAGGTGGCTTT	85
IGF2	ATAACAGGAGGATCAACCGTGGCA	ATGGCTTCTGGAAGCTCTCCTTGT	168
IGFBP2	TGAGAGCATCCTTGCTGAGAACCA	TTTGTGCTCCTCGTGGTTGTGATGG	180
MSTN	ACGGTGTTTGTGCAGATCCTGAGA	ATACCAGTGCCTGGGTTTCATGTCA	104
GHSR	ACATGAGGACCACCACCAACTTCT	AACTGGAAGAGCTTGCAGAGGAGA	145
TGFBR2	ATGCCTACCGCACTCACAAGAAGA	TGTGGTTGATGTTGTTGGCACAGG	156
PDGFB	TCTAGCCTGGACCTGAATGCAACT	TGTCACGGGAGATCTCAAAGACCA	151
IGF1R	ACAACGAGTACAACCTACCGCTGCT	ATTCTGGTCTGTGCAGGCTCTCTT	92
CTGF	TCGACCAGGGTCACCAACGATAAT	ACAGCTCAAACCTGATGGGCTTGG	160
FGF2	AAGAGCGATCCGCACATCAAACCTG	AGCCAGAAAGCGGTTTGCACCTAC	90
GH1	TCAGACATGGAGCTGCTTCGGTIT	TCAAACACTCTGTCTGAGGTGCCA	122
POU1F1	TGTGCCCTCCTGTCACTATGGAAA	ATAGCATCTCTGGAGTTGCAGGCT	83
GHRH	TTGGCAAACGGCTCAGAAACAGTG	TCCAAGAAGTCCCTCAGTACCAT	134
FABP3	AGCTGGATGAGGAGTTCGATGAGA	TGCCCATGGTGAGAGTCAGAATCA	168
ADCYAP1	ACATAGACGGCATCTTCACGGACA	ATACGCTACTCGGCTCCTTGTGT	131
BMPR2	ACACAAGTGACTGGGTGAGCTCTT	CATTGCGGCTGTTCAAGTCACGAT	135
CAPN3	ATGCCGGGAAGAACTCAGGAATGT	AGCCGTCTGTATCCATCAAAGCGA	122
IL1B	AAAGTGAGGCTCAACATTGCGCTG	TGTAGAGCTTGTAGCCCTTGTATGC	106
SST	TCTCAGAGCCAAGCCAGACAGAAA	AGAAGTTCTTGCAGCCCGCTTT	156

IGF1 : Insulin-like growth factor 1, IGF2 : Insulin-like growth factor 2, IGFBP2 : Inulin-like growth factor-binding protein 2, MSTN : Myostatin, GHSR : Growth hormone secretagogue receptor, CAPN3 : Calpain 3, TGFBR2 : Transforming growth factor beta receptor II, PDGFB : Platelet-derived growth factor subunit B, IGF1R : Insulin-like growth factor 1 receptor, CTGF : Connective tissue growth factor, FGF2 : Fibroblast growth factor 2, GH1 : Growth hormone 2, POU1F1 : POU class 1 homeobox 1, GHRH : Growth hormone releasing hormone, FABP3 : Fatty acid binding protein 3, ADCYAP1 : Adenylate cyclase activating polypeptide 1, BMPR2 : Bone morphogenetic protein receptor, type II, IL1B : interleukin 1 beta, and SST : Somatostatin.

perfamily. This group of proteins is characterized by a polybasic proteolytic processing site which is cleaved to produce a mature protein containing seven conserved cysteine residues. The members of this family are regulators of cell growth and differentiation in both embryonic and adult tissues. This gene is thought to encode a secreted protein which negatively regulates skeletal muscle growth (provided by RefSeq, Jul 2008). In addition, the expression of MSTN can be regulated by hormone and environmental factors. Sex steroids, androgen and estrogen can affect sex differentiation and also can affect muscle development (Yuhui *et al.*, 2012). The POU1F1 (POU class 1 homeobox 1) gene encode

by a member of the POU family of transcription factors that regulate mammalian development. The protein regulates expression of several genes involved in pituitary development and hormone expression (provided by RefSeq, Jul 2008). The genes of the POU1F1 pathway play a crucial role in embryonic survival rate (khatib *et al.*, 2009). In bovine embryos the GH transcript has been detected in oocytes and all pre-attachment stages including blastocysts (Kolle *et al.*, 1998; Joudrey *et al.*, 2003). The ADCYAP1 (adenylate cyclase activating polypeptide 1) gene encodes a secreted proprotein that is further processed into multiple mature peptides. These peptides stimulate adenylate cyclase and increase

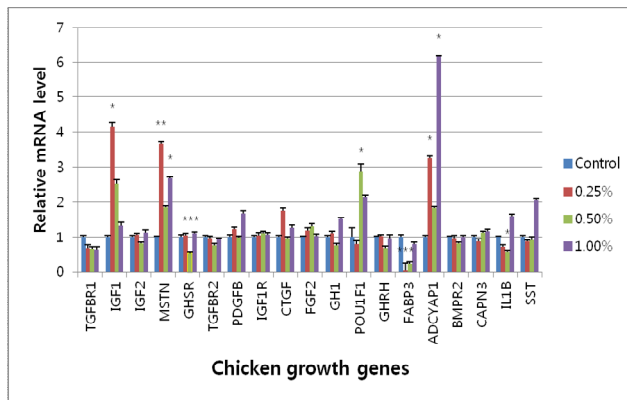


Fig. 1. Effect of dietary supplementation of *C. longa* on mRNA expression of growth response in liver of broiler chickens. * : $p < 0.05$, ** : $p < 0.01$, *** : $p < 0.001$.

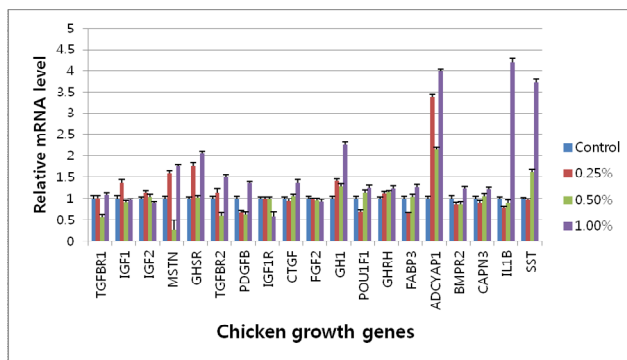


Fig. 2. Effect of dietary supplementation of *C. longa* on mRNA expression of growth response in spleen of broiler chickens. There was no statistically significant difference in the value.

cyclic adenosine monophosphate (cAMP) levels, resulting in the transcriptional activation of target genes (provided by RefSeq, Feb 2013). ADCYAP1 might regulate gonadotropin gene expression in the human fetal pituitary and might play a role in the differential secretion of LH (luteinizing hormone) and FSH (follicle-stimulating hormone) that occurs in normal adolescence, in hypogonadotropic hypogonadism including hyperprolactinemia, during fasting (Vertongen *et al.*, 1995; Stephen *et al.*, 2011).

On the other hand, GHSR and FABP3 gene showed significant expression patterns in this analysis. The GHSR (Growth hormone secretagogue receptor) gene encodes a member of the G-protein coupled receptor family. The encoded protein may play a role in energy homeostasis and regulation of body weight. Mutations in this gene are associated with autosomal idiopathic short stature (provided by RefSeq, Apr 2010). GHSR mRNAs were detected in morula or more advanced embryo stages (Kazuhiro *et al.*, 2003). In addition, the GHSR is expressed in diverse peripheral tissues (Mori *et*

al., 2000; Papotti *et al.*, 2000). In animals with insufficient nutrient intake, the fertility potential is suppressed, probably due to adaptive responses to evade the excess metabolic demands imposed by pregnancy (I'Anson *et al.*, 1991; Wade *et al.*, 1996). The FABP3 (Fatty acid binding protein 3) may also be responsible in the modulation of cell growth and proliferation. Fatty acid-binding protein 3 gene contains four exons and its function is to arrest growth of mammary epithelial cells. This gene is a candidate tumor suppressor gene for human breast cancer (provided by RefSeq, Jul 2008). FABPs are needed for intracellular fatty acid transport and contribute to cell differentiation as shown in embryonic stem cells (Cornelia *et al.*, 2012).

In conclusion, the present results suggest that dietary *C. longa* supplementation above levels recommended for growth increase. Also growth-related genes were known to central roles in mediating gonadotropin function, regulating steroidogenesis and promoting oocyte growth and maturation. The results would be used as basic information for the variation of growth-related genes expression on the cell growth, sex cell growth, and sex hormones according to dietary supplementation with *C. longa* in chickens.

REFERENCES

1. AL-Sultan SI (2003): The effect of *Curcuma longa* (Turmeric) on overall performance of broiler chickens. *International J of Poultry Sci* 25:351-353.
2. Ammon HP, Safayhi H, Mack T, Sabieraj J (1993): Mechanism of anti inflammatory actions of curcumin and boswellic acids. *J Ethnopharmacol* 38:113-119.
3. AOAC (1984): Official Methods of Analysis of the Association of Official Analytical Chemists. 14th edition. AOAC, Arlington, VA, USA.
4. Cornelia AIK, Anne NS, Jliane SS, Sünje F, Bernd F (2012): Expression profile of fatty acid metabolism genes in preimplantation blastocysts of obese and non-obese mice. *Obes Facts* 5:575-586.
5. Durrani FR, Mohammad Ismail, Asad Sultan, Suhail SM, Naila Chand, Durrani Z (2006): Effect of different levels of feed added turmeric (*Curcuma longa*) on the performance of broiler chicks. *J of Agri and Bio Sci* 2:9-10.
6. I'Anson H, Foster DL, Foxcroft GR, Booth PJ (1991): Nutrition and reproduction. *Oxf Rev Reprod Biol* 13:239-311.
7. Joudrey EM, Lechniak D, Petrik J, King WA (2003): Expression of growth hormone and its transcription factor, Pit-1, in early bovine development. *Mol Re-*

- prod Dev 64:275-283.
8. Kazuaki T, Akira A, Testuya T, Yokio A (2008): Dietary supplementation of glycine modulates inflammatory response indicators in broiler chickens. *British J of Nutrition* 100:1019-1028.
 9. Kazuhiro K, Naoki S, Jun F, Hideya K, Jin K, Hideo T, Akira N, Yoko H, Toshiharu S, Toshinobu T (2003): Ghrelin inhibits the development of mouse preimplantation embryos *in vitro*. *Endocrinology* 144 (6):2623-2633.
 10. Khatib H, Huang W, Wang X, Tran AH, Bindrim AB, Schutzkus V, Monson RL, Yandell BS (2009): Single gene and gene interaction effects on fertilization and embryonic survival rates in cattle. *J Dairy Sci* 92:2238-2247.
 11. Kiuchi F, Goto Y, Sugimoto N, Akao N, Kondo K, Tsuda Y (1993): Nematocidal activity of turmeric : synergistic action of curcuminoids. *Chem Pharm Bull (Tokyo)* 41:1640-1643.
 12. Kolle S, Sinowatz F, Boie G, Lincoln D, Palma G, Stojkovic M, Wolf E (1998): Topography of growth hormone receptor expression in the bovine embryo. *Histochem Cell Biol* 109:417-419.
 13. Lans C, Brown G (1998): Ethnoveterinary medicines used for ruminants in Trinidad and Tobago. *Prev Vet Med* 35:149-163.
 14. Lee JW (2013): Analysis of gene expression associated with growth performance in ROSS broiler. *J of Agri & Life Sci* 47(4):129-139.
 15. Mori K, Yoshimoto A, Takaya K, Hosoda K, Ariyasu H, Yahata K, Mukoyama M, Sugawara A, Hosoda H, Kojima M, Kanagawa K, Nakao K (2000): Kidney produces a novel acylated peptide, ghrelin. *FEBS Lett* 486:213-216.
 16. Mukhejee D, Mukherjee D, Sen U, Paul S, Bhattacharyya SP (2006): *In vitro* effects of insulin-like growth factors and insulin on oocyte maturation and maturation-inducing steroid production in ovarian follicles of common carp, *Cyprinus carpio*. *Comp Biochem Physiol A* 144:63-77.
 17. Nef S, Verma-Kurvari S, Merenmies J, Vassalli JD, Efstratiadis A, Accili D, Parada LF (2003): Testis determination requires insulin receptor family function in mice. *Nature* 426:291-295.
 18. Osawa T, Sugiyama Y, Inayoshi M, Kawakishi S (1995): Antioxidative activity of tetrahydrocurcuminoids. *Biosci Biotechnol Biochem* 59:1609-1612.
 19. Papotti M, Ghé C, Cassoni P, Catapano F, Deghenghi R, Ghigo E, Muccioli G (2000): Growth hormone secretagogue binding sites in peripheral human tissues. *J Clin Endocrinol Metab* 85:3803-3807.
 20. Pfaffl MW, Hageleit M (2001): Validities of mRNA quantification using recombinant RNA and recombinant DNA external calibration curves in real-time RT-PCR. *Biotech Lett* 23:273-282.
 21. Santos EM, Workman VL, Paull GC, Filby AL, Van Look KJW, Kille P, Tyler CR (2007): Molecular basis of sex and reproductive status in breeding zebrafish. *Physiol Genomics* 30:111-122.
 22. Silha JV, Murphy LJ (2005) Insulin-like growth factor binding proteins in development. *Adv Exp Med Biol* 567:55-89.
 23. Soni KB, Lahiri L, Chackradeo P, Bhide SV, Kuttan R (1997): Protective effect of food additives on aflatoxin-induced mutagenicity and hepatocarcinogenicity. *Cancer Letters* 115:129-133.
 24. Stephen J, Winters P, Joseph P, Moore JR (2011): PACAP, an autocrine/paracrine regulator of gonadotrophs. *Biology of Reproduction* 84:844-850.
 25. Sunderman FW, Nomoto S (1970): Measurement of human serum ceruloplasmin by its *p*-phenylenediamine oxidase activity. *Clin Chem* 16:903-910.
 26. Vertongen P, d'Haens J, Michotte A, Velkeniers B, van Rempelbergh J, Svoboda M, Robberecht P (1995): Expression of pituitary adenylate cyclase-activating polypeptide and receptors in human brain tumors. *Peptides* 16:713-719.
 27. Wade GN, Schneider JE, Li HY (1996): Control of fertility by metabolic cues. *Am J Physiol* 270:E1-E19.
 28. Yarru LP, Settivari RS, Gowda NKS, Antoniou E, Ledoux DR, Rottinghaus GE (2009): Effects of turmeric (*Cucuma longa*) on the expression of hepatic genes associated with biotransformation, antioxidant, and immune systems in broiler chicks fed aflatoxin. *Poultry Sci* 88:2620-2627.
 29. Yu Y, Li W, Han Z, Luo M, Chang Z, Tan J (2003): The effect of follicle-stimulating hormone on follicular development, granulosa cell apoptosis and steroidogenesis and its mediation by insulin-like growth factor- I in the goat ovary. *Theriogenology* 60: 1691-1704.
 30. Yuhui Y, Kou P, Xiaoyan L, Dongmei Z, Jundan D, Wei H, Haobin Z (2012): Effects of sex steroids on expression of myostatin in rare minnow, *Gobiocypris rarus*. *Aquaculture* 350-353.

(Received: 13 November 2013/ Accepted: 9 December 2013)