# Synthesis of Novel 2'-Fluoro-5'-deoxyphosphonic Acids and Bis(SATE) Adenine Analogue as Potent Antiviral Agents 

Guang Huan Shen and Joon Hee Hong*<br>BK-21 Project Team, College of Pharmacy, Chosun University, Kwangju 501-759, Korea<br>*E-mail: hongjh@chosun.ac.kr

Received August 27, 2013, Accepted September 11, 2013


#### Abstract

Novel 5'-deoxythreosyl purine phosphonic acid analogues containing a 2'-electropositive moiety such as fluorine atom, were designed and synthesized from commercially available 1,3-dihydroxy acetone. Condensation successfully proceeded from a glycosyl donor $\mathbf{6}$ under Vorbrüggen conditions and cross-metathesis gave the desired phosphonate analogues $\mathbf{7 a}, \mathbf{7 b}, \mathbf{1 7 a}$ and $\mathbf{1 7 b}$. The synthesized nucleoside phosphonic acid analogues 13, 16, 23, 26, 28 were subjected to antiviral screening against HIV-1. The bis(SATE) adenine analogue 28 exhibited significant in vitro activities against HIV-1.


Key Words : anti-HIV agents, 2'-Fluoro-5'-deoxyphosphonic acid analogue, Vorbrüggen reaction

## Introduction

As mimics of nucleoside monophosphates, phosphonate analogues exert their antiviral effect following sequential activation by cellular kinases to their diphosphate derivatives (nucleoside triphosphate analogues) which act as potent inhibitors of viral polymerases. ${ }^{1}$ The selective inhibition of viral polymerases, as opposed to host cell DNA polymerases, is critical for the therapeutic use of such compounds. Various attempts to improve selectivity indices have led to nucleoside analogues with a modified furanose ring system due to the introduction of an electropositive moiety at the $2^{\prime}-$ position of the furanose ring, ${ }^{2}$ and recently, a nucleoside analogue with a fluorine atom at this position, GS-9148 (1), was reported to show excellent anti-HIV activity (Figure 1). ${ }^{3}$
Threose phosphonate nucleosides, ${ }^{4}$ such as, PMDTA (2) and PMDTT (3), have been assembled from natural precursor molecules. Furthermore, it has been demonstrated threose nucleic acids (TNA) form thermal stable duplexes with DNA and RNA that are reminiscent of the natural





Figure 1. Synthesis rationale of threosyl 5'-deoxyphosphonic acid nucleosides as potent antiviral agents.
associations of nucleic acids. ${ }^{5}$ Moreover, diphosphates of threose nucleosides are accepted as substrates by several polymerases, and can be enzymatically incorporated into DNA. ${ }^{6}$ In addition, these nucleosides are also accepted as substitutes for ribonucleosides at the catalytic site of hammerhead ribozyme, although the catalytic efficiency of ribozyme is then significantly reduced. ${ }^{7}$ PMDTA has a phosphonomethoxy group at the $3^{\prime}$-position of its furanose ring and no substituent at the 4 -position. ${ }^{8}$ This absence of a 4'-hydroxymethyl group avoids problems of steric hindrance during phosphorylation reactions with kinases. To study the influences of different substituents on anti-HIV activity further, we undertook to synthesize 2 '-fluorinated analogues of PMDTA.

Several 5'-phosphate isosteres have been used to prepare nucleoside phosphonates. As shown in Figure 1, compound $\mathbf{4}^{9}$ is a simple $5^{\prime}$-deoxynucleoside phosphonate, in which the $5^{\prime}$-oxygen of a nucleoside phosphate is replaced by methylene group. Importantly, all resultant phosphonates mimic the overall shape and geometry of nucleoside monophosphates.

Stimulated by the findings that 2 -electropositive nucleoside analogues and $5^{\prime}$-deoxyphosphonic acid nucleosides have excellent antiviral activities, we sought to synthesize a novel class of nucleosides consisting of 2'-fluorinated-5'-deoxythreosylphosphonic acid analogues in order to find therapeutics that are more effective against HIV.

## Results and Discussion

As depicted in Scheme 1, target compounds were prepared from the fluorinated glycosyl donor 6, which was readily prepared from 1,3-dihydroxyacetone 5, as previously described. ${ }^{10}$ The synthesis of adenine nucleoside was carried out by Vorbrüggen condensation ${ }^{11}$ of compound 6 with silylated 6-chloropurine using TMSOTf as a catalyst in DCE to give the protected 6 -chloropurine derivatives $7 \mathbf{a}$ and $\mathbf{7 b}$,


Reagents: i) Silylated 6-chloropurine, TMSOTf, DCE; ii) TBAF, THF; iii) Dess-Martin, $\mathrm{CH}_{2} \mathrm{Cl}_{2}$; iv) $n$ - $\mathrm{BuLi}, \mathrm{Ph}_{3} \mathrm{PCH}_{3} \mathrm{I}, \mathrm{PPh}_{3}$, THF.
Scheme 1. Synthesis of threosyl-2'-fluoro-3'-vinylidene 6-chloropurine analogue.
respectively. Strong NOE ( $0.8 \%$ ) of $\mathrm{H}-1^{\prime} \leftrightarrow \mathrm{CH}-3^{\prime}$, which showed a $1^{\prime}, 3$ '-cis relationship, was observed. According to this result, the 3 '-hydroxymethyl group and the 1 '-purine base of $\mathbf{7 b}$ were located on the $\beta$ face. On the other hand, for 7 a compound, weak NOE ( $0.4 \%$ ) of $\mathrm{H}-1^{\prime} \leftrightarrow \mathrm{CH}-3^{\prime}$, demonstrated a 1',3'-trans relationship (Figure 2).
For the homologation, removal of the silyl protecting group of $7 \mathbf{b}$ using tetra $n$-butylammonium fluoride (TBAF) gave the primary alcohol 8. Dess-Martin oxidation ${ }^{12}$ of the alcohol of $\mathbf{8}$ gave the aldehyde 9 , which was subjected to Wittig olefination ${ }^{13}$ to give compound 10 without loss of the 3 '-stereochemistry. Cross-metathesis ${ }^{14}$ of $\mathbf{1 0}$ with vinyl diethylphosphonate using a $2^{\text {nd }}$ generation Grubbs catalyst ${ }^{15}$ gave the vinylidene phosphonate nucleoside analogue 11 in $57 \%$ yield. The chlorine group of the purine analogue 11 was then converted to amine with methanolic ammonia at $62{ }^{\circ} \mathrm{C}$ to give the corresponding adenosine phosphonate derivative 12 at in $63 \%$ yield. Hydrolysis of the diethyl phosphonate functional groups of $\mathbf{1 2}$ by treatment with bromotrimethylsilane in $\mathrm{CH}_{3} \mathrm{CN}$ in the presence of 2,6 -lutidine then gave the adenosine phosphonic acid derivative $\mathbf{1 3}{ }^{16}$ When the vinylidene phosphonate was saturated under transfer catalytic hydrogenation conditions ${ }^{17}$ it produced the ethyl phos-


7a


7b

Figure 2. NOE differences between the proximal hydrogens of $7 \mathbf{a}$ and $7 \mathbf{b b}$.


Reagents: i) Vinyldiethylphosphonate, Grubbs cat.(II) $\mathrm{CH}_{2} \mathrm{Cl}_{2}$; ii) $\mathrm{NH}_{3}$, $\mathrm{MeOH}, 62{ }^{\circ} \mathrm{C}$; iii) TMSBr, 2,6-lutidine, $\mathrm{CH}_{3} \mathrm{CN}$; iv) $\mathrm{Pd} / \mathrm{C}$, cyclohexene, MeOH .

Scheme 2. Synthesis of threosyl-2'-fluoro-5'-deoxyphosphonic acid adenine analogues.
phonate nucleoside analogue 14 in $74 \%$ yield. The adenine phosphonic acid analogue 16 was prepared using conditions similar to the ammonolysis and hydrolysis described to produce 13 (Scheme 2).

The guanine analogues, 2-fluoro-6-chloropurine ${ }^{18}$ was condensed with the glycosyl donor 6 using conditions similar to those used for the preparation of $7 \mathbf{a}$ and $7 \mathbf{b}$ to give the analogues $\mathbf{1 7 a}$ ( $31 \%$ ) and $\mathbf{1 7 b}$ ( $32 \%$ ) from 6-chloropurine (Scheme 3). A complete NOE study allowed the unambiguous determination of the relative stereochemistries of purine analogues as described for 7a and 7b. Homologation was performed using reactions similar to those used to produce 10, such as desilylation, Dess-Martin oxidation and Wittig olefination. Cross-metathesis of 20 with diethylvinylphos-


Reagents: i) Silylated 2-fluoro-6-chloropurine, TMSOTf, DCE; ii) TBAF, THF; iii) Dess-Martin, $\mathrm{CH}_{2} \mathrm{Cl}_{2}$; iv) $n$ - $\mathrm{BuLi}, \mathrm{Ph}_{3} \mathrm{PCH}_{3} 1, \mathrm{PPh}_{3}$, THF.

Scheme 3. Synthesis of threosyl-2'-fluoro-3'-vinylidene 2-fluoro-6-chloropurine analogue.


Reagents: i) vinyldiethylphosphonate, Grubbs cat.(II) $\mathrm{CH}_{2} \mathrm{Cl}_{2}$; ii) $\mathrm{NH}_{3}$, DME, rt; iii) (a) $\mathrm{TMSBr}, 2,6$-lutidine, $\mathrm{CH}_{3} \mathrm{CN}$; (b) $\mathrm{NaOMe}, \mathrm{HSCH}_{2} \mathrm{CH}_{2} \mathrm{OH}, \mathrm{MeOH}$; iv) $\mathrm{Pd} / \mathrm{C}$, cyclohexene, MeOH .
Scheme 4. Synthesis of threosyl-2'-fluoro-5'-deoxyphosphonic acid guanine analogue.
phonate provided 21 in 60\% yield.
Bubbling ammonia into the compound 21 gave separable 2-fluoro-6-aminopurine ${ }^{19} \mathbf{2 2 a}$ (14\%) and 2-amino-6-chloropurine 22b (45\%) analogues, respectively. The 2-amino-6chloropurine derivative 22b was treated with TMSBr to provide phosphonic acid and sequentially treated with sodium methoxide and 2-mercaptoethanol in methanol to give the desired guanine phosphonic acid 23 (Scheme 4). ${ }^{20}$ Furthermore, the guanine phosphonic acid analogue 26 was synthesized from 21 via transfer catalytic hydrogenation, ammonolysis, and hydrolysis using conditions similar to those described for the synthesis of the adenine 6 , line derivative 16. To synthesize the thioester-protected analogue, compound 16 was reacted with thioester $27^{21}$ in the presence of 1-(2-mesitylenesulfonyl)-3-nitro-1 H -1,2,4-triazole (MSNT) ${ }^{22}$ to provide the bis(SATE) derivative as a target compound 28 (Scheme 5).
Antiviral Activity. The antiviral activities of phosphonate nucleosides are explained by their intracellular metabolism to diphosphates, subsequent incorporation into the viral genome, and chain termination. ${ }^{23}$ MT-4 cells $\left(1 \times 10^{5}\right.$ cell/ mL ) were infected with HIV-1 (HTLV-III ${ }_{\mathrm{B}}$ strain) at a multi-


Reagents: i) thioester, 27, 1-(2-mesitylenesulfonyl)-3-nitro-1H-1,2,4triazole, pyridine.

Scheme 5. Synthesis of target bis(SATE) prodrug of adenine analogue 16.

Table 1. Median effective $\left(\mathrm{EC}_{50}\right)$ and cytotoxic $\left(\mathrm{CC}_{50}\right)$ concentrations of the synthesized nucleoside analogues

| Compound <br> No. | Anti-HIV-1 <br> $\mathrm{EC}_{50}(\mu \mathrm{M})^{c}$ | Cytotoxicity <br> $\mathrm{CC}_{50}(\mu \mathrm{M})^{d}$ |
| :---: | :---: | :---: |
| $\mathbf{1 3}$ | 34.2 | 95 |
| $\mathbf{1 6}$ | 8.8 | 80 |
| $\mathbf{2 3}$ | 66.8 | 98 |
| $\mathbf{2 6}$ | 47.1 | 98 |
| $\mathbf{2 8}$ | $\mathbf{2 . 2}$ | 80 |
| $\mathbf{A Z T}^{a}$ | 0.003 | $>100$ |
| PMEA $^{b}$ | $>10$ | $>10$ |
| bis(SATE)PMEA | 0.81 | $>10$ |

${ }^{a}$ AZT: azidothymidine. ${ }^{b}$ PMEA: 9-[2-(phosphonomethoxy)ethyl]adenine. ${ }^{c} \mathrm{EC}_{50}(\mu \mathrm{M})$ : $\mathrm{EC}_{50}$ values are for $50 \%$ inhibition of virus production as indicated by supernatant RT levels. ${ }^{d} \mathrm{CC}_{50}(\mu \mathrm{M})$ : $\mathrm{CC}_{50}$ values indicate $50 \%$ cytotoxic concentration.
plicity of infection (MOI) of 0.02 , and then cultured in the presence of various concentrations of the test compounds. After a 4-day incubation at $37^{\circ} \mathrm{C}$, numbers of viable cells were determined using the 3-(4,5-di-methylthiazole-2-yl)-2,5-diphenyltetrazolium bromide method. The cytotoxicities of the compounds were evaluated in parallel with their antiviral activities, by determining the viabilities of mockinfected cells. ${ }^{24}$ Compounds 13, 16, 23, 26 and 28 were tested against HIV-1, and the adenine analogue 16 showed moderate antiviral activity (Table 1). However, other three 5'-deoxyphosphonic acid nucleoside analogues showed weak or no anti-HIV activity at concentrations up to 100 $\mu \mathrm{M}$.

In summary, based on the potent anti-HIV activities of 2'electropositive nucleosides and 5'-deoxyphosphonic acid nucleoside analogues, we designed and successfully synthesized novel $2^{\prime}$-fluoro-5'-deoxyphosphonic acid nucleoside analogues starting from 1,3-dihydroxy acetone. The synthesized bis(SATE) adenine analogue 28 showed significant activity in a cell-based assay than the 2 '-modified guanine phosphonic acid analogues 13, 23 and 26. Since 2 '-fluorinated guanine nucleoside derivatives are not perfect mimics of the ribofuranose moiety, mechanisms of virus inhibition, that is, phosphorylation or the inhibition of RNA synthesis, might be impaired for these compounds. For the discovery of improved antiviral nucleoside derivatives, bis(SATE) analogue 28 was synthesized and assayed for anti-HIV activity using an in vitro assay system, It showed much improved



Compound 16


Figure 3. Superimpose of PMDTA and 16.
anti-HIV activity than adenine nucleoside phosphonic acid 16 (Table 1). As shown in the superimposition model of PMDTA (2) and the corresponding analogue 16 (Figure 3), discrepancies of phosphonic acid regions are more pronounced than those of the base moiety. Note the furanose puckering of PMDTA (2) is closer to that of the adenine analogue 16. ${ }^{25}$

## Experimental Section

Uncorrected melting points were determined on a Meltemp II laboratory device. NMR spectra were recorded on a JEOL 300 Fourier transform spectrometer (JEOL, Tokyo, Japan); chemical shifts are reported in parts per million ( $\delta$ ) and signals are reported as $s$ (singlet), $d$ (doublet), $t$ (triplet), q (quartet), m (multiplet), or dd (doublet of doublets). UV spectra were obtained on a Beckman DU-7 spectrophotometer (Beckman, South Pasadena, CA, USA). MS spectra were collected in electrospray ionization (ESI) mode. Elemental analyses were performed using a Perkin-Elmer 2400 analyzer (Perkin-Elmer, Norwalk, CT, USA). TLC was performed on Uniplates (silica gel) purchased from Analtech Co. (7558, Newark, DE, USA). All reactions were carried out in a nitrogen atmosphere unless otherwise specified. Dry dichloromethane, benzene, and pyridine were obtained by distillation from $\mathrm{CaH}_{2}$. Dry THF was obtained by distillation from Na and benzophenone immediately prior to use.
(rel)-(1'S,2'R,3'S)-9-(3'-t-Butyldimethylsilanyloxymethyl-$\mathbf{2}^{\prime}$-fluoro-tetrahydrofuran-1'-yl) 6-chloropurine (7a) and (rel)-(1'R,2'R,3'S)-9-(3'-t-butyldimethylsilanyloxymethyl-2'-fluoro-tetrahydrofuran-1'-yl) 6-chloropurine (7b): 6Chloropurine ( $216 \mathrm{mg}, 1.4 \mathrm{mmol}$ ), anhydrous HMDS ( 10 mL ), and a catalytic amount of ammonium sulfate ( 14 mg ) were refluxed to a clear solution, and the solvent was then distilled off under anhydrous conditions. The residue obtained was dissolved in anhydrous 1,2-dichloroethane (8 $\mathrm{mL})$, and to this mixture, a solution of $\mathbf{6}(175 \mathrm{mg}, 0.6 \mathrm{mmol})$ in dry DCE ( 10 mL ) and TMSOTf ( $311 \mathrm{mg}, 1.4 \mathrm{mmol}$ ) was added, and stirred for 8 h at rt . The reaction mixture was quenched with 5.0 mL of saturated $\mathrm{NaHCO}_{3}$, stirred for 1 h , filtered through a Celite pad, and the filtrate obtained was then extracted twice with $\mathrm{CH}_{2} \mathrm{Cl}_{2}(80 \mathrm{~mL})$. Combined organic layers were dried over anhydrous $\mathrm{MgSO}_{4}$, filtered, and con-
centrated in vacuo. The residue was purified by silica gel column chromatography (EtOAc/hexane/MeOH, 4:1:0.01) to give compounds $7 \mathbf{a}(74 \mathrm{mg}, 32 \%)$ and 7 b ( $79 \mathrm{mg}, 34 \%$ ). Data for 7a: ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}, 300 \mathrm{MHz}\right) \delta 8.72(\mathrm{~s}, 1 \mathrm{H})$, $8.34(\mathrm{~s}, 1 \mathrm{H}), 6.23(\mathrm{dd}, J=18.6,5.8 \mathrm{~Hz}, 1 \mathrm{H}), 3.77-3.67$ (m, $5 \mathrm{H}), 2.39-2.28(\mathrm{~m}, 1 \mathrm{H}), 0.88-0.86(\mathrm{~m}, 9 \mathrm{H}), 0.01(\mathrm{~s}, 6 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}, 75 \mathrm{MHz}\right) \delta 151.7,151.4,151.1,144.8$, 132.5, $92.2(\mathrm{~d}, J=172.0 \mathrm{~Hz}), 88.5(\mathrm{~d}, J=23.2 \mathrm{~Hz}), 60.5$, $57.4,39.3(\mathrm{~d}, J=22.2 \mathrm{~Hz}), 25.4,18.3,-5.1$; Anal. Calc. for $\mathrm{C}_{16} \mathrm{H}_{24} \mathrm{ClFN}_{4} \mathrm{O}_{2} \mathrm{Si}: \mathrm{C}, 49.67$; H, 6.25 ; N, 14.48. Found: C, 49.71; H, 6.23; N, 14.50; MS m/z 387 (M+H) ${ }^{+}$. Data for 7b: ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}, 300 \mathrm{MHz}\right) \delta 8.70(\mathrm{~s}, 1 \mathrm{H}), 8.31(\mathrm{~s}, 1 \mathrm{H})$, 6.19 (dd, $J=18.2,4.8 \mathrm{~Hz}, 1 \mathrm{H}), 3.77-3.63$ (m, 5H), 2.38-2.27 $(\mathrm{m}, 1 \mathrm{H}), 0.83(\mathrm{~m}, 9 \mathrm{H}), 0.01(\mathrm{~s}, 6 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}, 75\right.$ $\mathrm{MHz}) \delta 151.6,151.3,151.0,144.7,132.3,91.6(\mathrm{~d}, J=170.8$ $\mathrm{Hz}), 87.2(\mathrm{~d}, J=21.2 \mathrm{~Hz}), 59.4,56.2,38.2(\mathrm{~d}, J=21.4 \mathrm{~Hz})$, 25.5, 18.4, -4.8 ; Anal. Calc. for $\mathrm{C}_{16} \mathrm{H}_{24} \mathrm{ClFN}_{4} \mathrm{O}_{2} \mathrm{Si}(+0.5$ MeOH): C, 49.18; H, 6.50; N, 13.90. Found: C, 49.21; H, 6.52; N, 13.88; MS m/z $387(\mathrm{M}+\mathrm{H})^{+}$.
(rel)-(1'R,2'R,3'S)-9-(3'-Hydroxymethyl-2'-fluoro-tetra-hydrofuran- $\mathbf{1}^{\prime}$-yl) 6-chloropurine (8). To a solution of 7b $(2.54 \mathrm{~g}, 6.56 \mathrm{mmol})$ in THF $(12 \mathrm{~mL})$, TBAF $(7.7 \mathrm{~mL}, 1.0 \mathrm{M}$ solution in THF) was added at $0^{\circ} \mathrm{C}$. The mixture was stirred overnight at rt and concentrated in vacuo. The residue was purified by silica gel column chromatography (Hexane/ $\mathrm{EtOAc} / \mathrm{MeOH}, 2: 1: 0.05$ ) to give 8 (1.59 g, 89\%): ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}, 300 \mathrm{MHz}\right) \delta 8.69(\mathrm{~s}, 1 \mathrm{H}), 8.29(\mathrm{~s}, 1 \mathrm{H}), 6.15(\mathrm{dd}, J=$ $16.8,5.4 \mathrm{~Hz}, 1 \mathrm{H}), 3.79-3.64(\mathrm{~m}, 5 \mathrm{H}), 2.36-2.24(\mathrm{~m}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR ( $\left.\mathrm{CDCl}_{3}, 75 \mathrm{MHz}\right) \delta 151.8,151.4,151.1,144.5,132.6$, $92.5(\mathrm{~d}, J=166.8 \mathrm{~Hz}), 86.4(\mathrm{~d}, J=20.4 \mathrm{~Hz}), 60.2,56.2,38.2$ $(\mathrm{d}, J=21.4 \mathrm{~Hz})$; Anal. Calc. for $\mathrm{C}_{10} \mathrm{H}_{10} \mathrm{ClFN}_{4} \mathrm{O}_{2}(+1.0$ $\mathrm{MeOH}): \mathrm{C}, 43.36 ; \mathrm{H}, 4.62$; N, 18.39. Found: C, 43.32; H, 4.64; N, 18.41; MS m/z $273(\mathrm{M}+\mathrm{H})^{+}$.
(rel)-(1'R,2'R,3'R)-9-(3'-Carbaldehyde-2'-fluoro-tetra-hydrofuran-1'-yl) 6-chloropurine (9). Compound 8 (290 $\mathrm{mg}, 1.066 \mathrm{mmol}$ ) was dissolved in anhydrous $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ ( 8 mL ), and to this solution was added Dess-Martin reagent ( $588 \mathrm{mg}, 1.38 \mathrm{mmol}$ ). The mixture was stirred for 3 h at ambient temperature, concentrated and the residue was purified by silica gel column chromatography using Hexane/ $\mathrm{EtOAc}(1: 4)$ as eluent. A second column, which was also eluted with EtOAc, was necessary to remove traces of DessMartin reagent-related impurities to give 9 ( $253 \mathrm{mg}, 88 \%$ ): ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}, 300 \mathrm{MHz}\right) \delta 9.69(\mathrm{~s}, 1 \mathrm{H}), 8.70(\mathrm{~s}, 1 \mathrm{H})$, $8.28(\mathrm{~s}, 1 \mathrm{H}), 6.19(\mathrm{dd}, J=18.0,5.6 \mathrm{~Hz}, 1 \mathrm{H}), 4.05-3.94(\mathrm{~m}$, $3 \mathrm{H}), 2.93-2.85(\mathrm{~m}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}, 75 \mathrm{MHz}\right) \delta$ 204.7, 151.5, 151.0, 150.7, 144.1, 133.4, 92.5 (d, $J=18.4$ $\mathrm{Hz}), 88.5(\mathrm{~d}, J=164.4 \mathrm{~Hz}), 55.7,51.1(\mathrm{~d}, J=18.8 \mathrm{~Hz}) ; \mathrm{MS}$ $m / z 271(\mathrm{M}+\mathrm{H})^{+}$.
(rel)-(1'R,2'R,3'S)-9-(3'-Vinyl-2'-fluoro-tetrahydrofuran-1'-yl) 6-chloropurine (10). To ylide solution [methyltriphenylphosphonium iodide ( $188 \mathrm{mg}, 0.462 \mathrm{mmol}$ ), triphenylphosphine ( $14.25 \mathrm{mg}, 0.055 \mathrm{mmol}$ ), $1.6 \mathrm{M} n$-butyllithium solution ( $0.289 \mathrm{~mL}, 0.462 \mathrm{mmol}$ ) in dry tetrahydrofuran ( 5.0 mL ) at $-78^{\circ} \mathrm{C}$, was added dropwise to a solution of olefin 9 $(125 \mathrm{mg}, 0.462 \mathrm{mmol})$ in dry THF $(7 \mathrm{~mL})$. The reaction mixture was warmed to room temperature, stirred for 4 h ,
quenched with saturated sodium bicarbonate solution, and then partitioned between saturated sodium bicarbonate solution and ethyl acetate. The organic layer was separated and the aqueous layer was extracted with ethyl acetate. Combined extracts were dried over anhydrous sodium sulfate, filtered, concentrated in vacuo, and chromatographed (HexaneEtOAc, 1:2) to afford $\mathbf{1 0}(76 \mathrm{mg}, 61 \%)$ as a colorless oil. ${ }^{1} \mathrm{H}$ NMR ( $\left.\mathrm{CDCl}_{3}, 300 \mathrm{MHz}\right) \delta 8.73(\mathrm{~s}, 1 \mathrm{H}), 8.31(\mathrm{~s}, 1 \mathrm{H}), 6.21$ (dd, $J=19.2,5.2 \mathrm{~Hz}, 1 \mathrm{H}), 5.77-5.75(\mathrm{~m}, 1 \mathrm{H}), 5.05-4.96(\mathrm{~m}$, $2 \mathrm{H}), 3.74-3.68(\mathrm{~m}, 3 \mathrm{H}), 2.85-2.81(\mathrm{~m}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}, 75 \mathrm{MHz}\right) \delta 151.7,151.3,150.9,144.6,143.6,132.8$, $112.3,96.1(\mathrm{~d}, J=168.4 \mathrm{~Hz}), 88.3(\mathrm{~d}, J=16.8 \mathrm{~Hz}), 62.1$, 39.4 (d, $J=18.4 \mathrm{~Hz}$ ); Anal. Calc. for $\mathrm{C}_{11} \mathrm{H}_{10} \mathrm{ClFN}_{4} \mathrm{O}$ : C, 49.17; H, 3.75; N, 20.85; Found: C, 49.21; H, 3.78; N, 20.82; MS m/z $269(\mathrm{M}+\mathrm{H})^{+}$.
(rel)-(1'R,2'R,3'S)-Diethyl \{9-(3'-vinyl-2'-fluoro-tetra-hydrofuran-1'-yl) 6-chloropurine\} phosphonate (11). To a solution of the 6 -chloropurine derivative 10 ( $174 \mathrm{mg}, 0.650$ mmol ) and diethyl vinylphosphonate ( $426 \mathrm{mg}, 2.60 \mathrm{mmol}$ ) in $\mathrm{CH}_{2} \mathrm{Cl}_{2}(8.0 \mathrm{~mL}), 2^{\text {nd }}$-generation Grubbs catalyst ( 22.10 $\mathrm{mg}, 0.026 \mathrm{mmol}$ ) was added. The reaction mixture was refluxed for 26 h under dry argon and concentrated under reduced pressure. The residue was purified by silica gel column chromatography ( $\mathrm{EtOAc} / n$-Hexane/MeOH, 2:1:0.05) to give $11(150 \mathrm{mg}, 57 \%)$ as a foam: ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}, 300\right.$ $\mathrm{MHz}) \delta 8.72(\mathrm{~s}, 1 \mathrm{H}), 8.32(\mathrm{~s}, 1 \mathrm{H}), 6.67(\mathrm{dd}, J=17.2,21.4$ $\mathrm{Hz}, 1 \mathrm{H}), 6.20(\mathrm{dd}, J=19.2,5.2 \mathrm{~Hz}, 1 \mathrm{H}), 6.11$ (dd, $J=17.2$, 19.8. Hz, 1H), 4.12-4.06 (m, 4H), 3.73-3.66 (m, 3H), 2.82$2.78(\mathrm{~m}, 1 \mathrm{H}), 1.33-1.30(\mathrm{~m}, 6 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}, 75\right.$ $\mathrm{MHz}) \delta 151.8,151.4,151.0,149.5,144.2,132.6,117.2,95.6$ (d, $J=168.4 \mathrm{~Hz}), 87.6(\mathrm{~d}, J=17.0 \mathrm{~Hz}), 62.4,61.8,60.6$, $40.7(\mathrm{~d}, J=16.4 \mathrm{~Hz}), 15.8$; Anal. Calc. for $\mathrm{C}_{15} \mathrm{H}_{19} \mathrm{ClFN}_{4} \mathrm{O}_{4} \mathrm{P}$ (+0.5 MeOH): C, 44.24; H, 5.03; N, 13.31; Found: C, 44.26; H, 5.02; N, 13.29; MS m/z 405 (M+H) .
(rel)-(1'R,2'R,3'S)-Diethyl \{9-(3'-vinyl-2'-fluoro-tetra-hydrofuran- $\mathbf{1}^{\prime}$-yl) adenine\} phosphonate (12). A solution of $\mathbf{1 1}(188 \mathrm{mg}, 0.464 \mathrm{mmol})$ in saturated methanolic ammonia $(7 \mathrm{~mL})$ was stirred overnight at $66^{\circ} \mathrm{C}$ in a steel bomb, and volatiles were evaporated. The residue was purified by silica gel column chromatography $\left(\mathrm{MeOH} / \mathrm{CH}_{2} \mathrm{Cl}_{2}, 1: 10\right)$ to give 12 (112 mg, $63 \%$ ) as a white solid: UV (MeOH) $\lambda_{\max } 261.5$ $\mathrm{nm} ;{ }^{1} \mathrm{H}$ NMR (DMSO- $\left.d_{6}, 300 \mathrm{MHz}\right) \delta 8.36(\mathrm{~s}, 1 \mathrm{H}), 8.17(\mathrm{~s}$, $1 \mathrm{H}), 6.65$ (dd, $J=17.4,20.8 \mathrm{~Hz}, 1 \mathrm{H}$ ), 6.23 (dd, $J=18.0,5.4$ $\mathrm{Hz}, 1 \mathrm{H}), 6.10(\mathrm{dd}, J=17.2,19.8 . \mathrm{Hz}, 1 \mathrm{H}), 4.10-4.05(\mathrm{~m}$, $4 \mathrm{H}), 3.76-3.68(\mathrm{~m}, 3 \mathrm{H}), 2.83-2.77(\mathrm{~m}, 1 \mathrm{H}), 1.32-1.30(\mathrm{~m}$, 6 H ); ${ }^{13} \mathrm{C}$ NMR (DMSO- $d_{6}, 75 \mathrm{MHz}$ ) $\delta$ 155.5, 152.7, 150.4, 148.8, 141.3, 119.0, 115.6, 94.8 (d, $J=167.8 \mathrm{~Hz}$ ), 88.4 (d, $J$ $=16.6 \mathrm{~Hz}), 63.3,62.7,61.5,41.2(\mathrm{~d}, J=16.6 \mathrm{~Hz}), 15.3$; Anal. Calc. for $\mathrm{C}_{15} \mathrm{H}_{21} \mathrm{FN}_{5} \mathrm{O}_{4} \mathrm{P}(+1.0 \mathrm{MeOH}): \mathrm{C}, 46.04$; H , 6.04; N, 16.78; Found: C, 46.08; H, 6.02; N, 16.80; MS m/z $386(\mathrm{M}+\mathrm{H})^{+}$.
(rel)-(1'R,2'R,3'S)-9-[(3'-Vinyl-2'-fluoro-tetrahydrofuran-$1^{\prime}$-yl) adenine]phosphonic acid (13). To a solution of the phosphonate $\mathbf{1 2}$ ( $161 \mathrm{mg}, 0.419 \mathrm{mmol}$ ) in anhydrous $\mathrm{CH}_{3} \mathrm{CN}$ $(10 \mathrm{~mL})$ and 2,6-lutidine $(0.898 \mathrm{~mL}, 8.38 \mathrm{mmol})$ was added trimethylsilyl bromide ( $641 \mathrm{mg}, 4.19 \mathrm{mmol}$ ). The mixture was heated overnight at $75^{\circ} \mathrm{C}$ under nitrogen and then
concentrated in vacuo to give a brown residue, and then coevaporated from conc-aqueous $\mathrm{NH}_{4} \mathrm{OH}(2 \times 22 \mathrm{~mL})$. The resultant purified by twice triturating the residue in acetone $(8 \mathrm{~mL})$ and removing the acetone by evaporation. The residue so obtained was then purified by preparative reversephase chromatography. Lyophilization of the appropriate fraction provided the phosphonic acid salt $\mathbf{1 3}$ ( 53.65 mg , $37 \%$ ) as a white salt (ammonium salt): mp $157-159{ }^{\circ} \mathrm{C}$; UV $\left(\mathrm{H}_{2} \mathrm{O}\right) \lambda_{\text {max }} 261.0 \mathrm{~nm} ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{D}_{2} \mathrm{O}, 300 \mathrm{MHz}\right) \delta 8.34(\mathrm{~s}$, $1 \mathrm{H}), 8.16(\mathrm{~s}, 1 \mathrm{H}), 6.66(\mathrm{dd}, J=17.6,21.0 \mathrm{~Hz}, 1 \mathrm{H}), 6.21(\mathrm{dd}$, $J=17.6,5.2 \mathrm{~Hz}, 1 \mathrm{H}), 6.12(\mathrm{dd}, J=17.4,19.4 . \mathrm{Hz}, 1 \mathrm{H})$, 3.72-3.65 (m, 3H), 2.85-2.78 (m, 1H); ${ }^{13} \mathrm{C}$ NMR $\left(\mathrm{D}_{2} \mathrm{O}, 75\right.$ $\mathrm{MHz}) \delta 155.3,152.4,150.1,148.5,141.5,118.8,114.7,93.6$ (d, $J=165.8 \mathrm{~Hz}), 89.6(\mathrm{~d}, J=16.8 \mathrm{~Hz}), 61.6,40.6(\mathrm{~d}, J=$ $16.8 \mathrm{~Hz}) ;$ HPLC $t_{\mathrm{R}}=10.67$; HRMS [M-H] ${ }^{+}$req. 328.0678 , found 328.0679 .
(rel)-(1'R,2'R,3'S)-Diethyl \{9-(3'-ethyl-2'-fluoro-tetra-hydrofuran-1'-yl) 6-chloropurine\} phosphonate (14). A solution of vinyl phosphonate nucleoside analogue 13 (265 $\mathrm{mg}, 0.655 \mathrm{mmol}$ ) in methanol ( 8 mL ) was added $10 \% \mathrm{Pd} / \mathrm{C}$ $(10 \mathrm{mg})$ and cyclohexene $(4 \mathrm{~mL})$ under Ar. The reaction mixture was refluxed for 24 h . The reaction mixture was filtered through a pad of Celite, evaporated, and purified by silica gel column chromatography using methanol and methylene chloride (10:1) to give ethyl phosphonate analogue $14(197 \mathrm{mg}, 74 \%)$ as a white solid: ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right.$, $300 \mathrm{MHz}) \delta 8.75(\mathrm{~s}, 1 \mathrm{H}), 8.34(\mathrm{~s}, 1 \mathrm{H}), 6.19(\mathrm{dd}, J=18.6,5.4$ $\mathrm{Hz}, 1 \mathrm{H})$, 4.11-4.06 (m, 4H), 3.76-3.68 (m, 3H), 2.28-1.86 $(\mathrm{m}, 5 \mathrm{H}), 1.31-1.28(\mathrm{~m}, 6 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}, 75 \mathrm{MHz}\right) \delta$ $151.8,151.4,150.0,142.9,132.8,94.8(\mathrm{~d}, J=162.8 \mathrm{~Hz})$, 89.1 (d, $J=16.8 \mathrm{~Hz}$ ), $61.4,38.9$ (d, $J=16.8 \mathrm{~Hz}$ ), 28.7, 18.8, 14.9; Anal. Calc. for $\mathrm{C}_{15} \mathrm{H}_{21} \mathrm{ClFN}_{4} \mathrm{O}_{4} \mathrm{P}(+1.0 \mathrm{MeOH})$ : C, 43.79; H, 5.74; N, 12.77; Found: C, 43.83; H, 5.72; N, 12.79; MS m/z $407(\mathrm{M}+\mathrm{H})^{+}$.
(rel)-(1'R,2'R,3'S)-Diethyl \{9-(3'-ethyl-2'-fluoro-tetra-hydrofuran- $\mathbf{1 '}^{\prime}$-yl) adenine\} phosphonate (15). The adenine derivative 15 was prepared from the 6 -chloropurine analogue 14 using an ammonolysis procedure similar to that described for 12: yield $60 \%$; mp $172-174{ }^{\circ} \mathrm{C}$; UV (MeOH) $\lambda_{\max } 260.5 \mathrm{~nm} ;{ }^{1} \mathrm{H}$ NMR (DMSO- $\left.d_{6}, 300 \mathrm{MHz}\right) \delta 8.38$ (s, $1 \mathrm{H}), 8.19$ (s, 1H), 6.16 (dd, $J=18.8,5.5 \mathrm{~Hz}, 1 \mathrm{H}), 4.15-4.09$ $(\mathrm{m}, 4 \mathrm{H}), 3.73-3.65(\mathrm{~m}, 3 \mathrm{H}), 2.28-2.27(\mathrm{~m}, 2 \mathrm{H}), 2.01-1.88$ (m, 3H), 1.30-1.26 (m, 6H), ${ }^{13} \mathrm{C}$ NMR (DMSO- $d_{6}, 75 \mathrm{MHz}$ ) $\delta 154.9,152.3,150.2,141.4,118.7,93.5(\mathrm{~d}, J=162.4 \mathrm{~Hz})$, 88.4 (d, $J=16.3 \mathrm{~Hz}$ ), $60.6,37.6$ (d, $J=16.2 \mathrm{~Hz}$ ), 28.3, 18.5 , 15.1; Anal. Calc. for $\mathrm{C}_{15} \mathrm{H}_{23} \mathrm{FN}_{5} \mathrm{O}_{4} \mathrm{P}(+1.0 \mathrm{MeOH})$ : C, 45.82; H, 6.49; N, 16.69; Found: C, 45.78; H, 6.51; N, 16.71; MS $m / z 388(\mathrm{M}+\mathrm{H})^{+}$.
(rel)-(1'R,2'R,3'S)-\{9-(3'-Ethyl-2'-fluoro-tetrahydrofuran-1'-yl) adenine\} phosphonic acid (16). Phosphonic acid 16 was synthesized from 15 using hydrolysis conditions identical to that for 13: yield $40 \%, \mathrm{mp} 160-162{ }^{\circ} \mathrm{C}$; UV $\left(\mathrm{H}_{2} \mathrm{O}\right)$ $\lambda_{\text {max }} 262.0 \mathrm{~nm} ;{ }^{1} \mathrm{H} \operatorname{NMR}\left(\mathrm{D}_{2} \mathrm{O}, 300 \mathrm{MHz}\right) \delta 8.36(\mathrm{~s}, 1 \mathrm{H})$, $8.17(\mathrm{~s}, 1 \mathrm{H}), 6.14(\mathrm{dd}, J=17.6,5.4 \mathrm{~Hz}, 1 \mathrm{H}), 3.70-3.64(\mathrm{~m}$, $3 \mathrm{H}), 2.25-2.21(\mathrm{~m}, 2 \mathrm{H}), 2.03-1.89(\mathrm{~m}, 3 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR ( $\mathrm{D}_{2} \mathrm{O}$, $75 \mathrm{MHz}) \delta 155.2,152.5,150.6,141.9,119.4,95.2(\mathrm{~d}, J=$ $160.8 \mathrm{~Hz}), 87.7(\mathrm{~d}, J=16.4 \mathrm{~Hz}), 61.4,38.2(\mathrm{~d}, J=16.5 \mathrm{~Hz})$,
29.1, 18.8; HPLC $t_{\mathrm{R}}=10.92$; HRMS $[\mathrm{M}-\mathrm{H}]^{+}$req. 330.0677, found 330.0679 .
(rel)-(1'S,2'R,3'S)-9-(3'-t-Butyldimethylsilanyloxymethyl-2'-fluoro-tetrahydrofuran-1'-yl) 6-chloropurine (17a) and (rel)-(1'R,2'R,3'S)-9-(3'-t-Butyldimethylsilanyloxy-methyl-2'-fluoro-tetrahydrofuran-1'-yl) 2-fluoro-6-chloropurine (17b). Condensation of 6 with 2-fluoro-6-chloropurine under Vorbruggen condensation conditions similar to those described for $\mathbf{7 a}$ and $\mathbf{7 b}$ gave $\mathbf{1 7 a}$ and $\mathbf{1 7 b}$, respectively. Data for 17a: yield $31 \%$; UV (MeOH) $\lambda_{\text {max }} 267.5 \mathrm{~nm}$; ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}, 300 \mathrm{MHz}\right) \delta 8.47(\mathrm{~s}, 1 \mathrm{H}), 6.21(\mathrm{dd}, J=$ $18.4,5.4 \mathrm{~Hz}, 1 \mathrm{H}), 3.75-3.68(\mathrm{~m}, 5 \mathrm{H}), 2.23-2.19(\mathrm{~m}, 1 \mathrm{H})$, $0.90-0.88(\mathrm{~m}, 9 \mathrm{H}), 0.02-0.01(\mathrm{~m}, 6 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}, 75\right.$ MHz) $\delta 157.1(\mathrm{~d}, J=219 \mathrm{~Hz}), 153.3,145.6,136.2,120.6$, 92.2 (d, $J=166.7 \mathrm{~Hz}), 89.1(\mathrm{~d}, J=16.4 \mathrm{~Hz}), 60.5,57.6$, 38.7 (d, $J=16.2 \mathrm{~Hz}$ ), 25.5, 18.7, -4.6 ; Anal. Calc. for $\mathrm{C}_{16} \mathrm{H}_{23} \mathrm{ClF}_{2} \mathrm{~N}_{4} \mathrm{O}_{2} \mathrm{Si}: \mathrm{C}, 47.46$; H, 5.73 ; N, 13.84; Found: C, 47.48; H, 5.75; N, 13.86; MS m/z $405(\mathrm{M}+\mathrm{H})^{+}$. data for 17b: yield $32 \%$; UV (MeOH) $\lambda_{\text {max }} 268.5 \mathrm{~nm}$; ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right.$, $300 \mathrm{MHz}) \delta 8.45(\mathrm{~s}, 1 \mathrm{H}), 6.19(\mathrm{dd}, J=18.5,5.5 \mathrm{~Hz}, 1 \mathrm{H})$, 3.74-3.67 (m, 5H), 2.22-2.18 (m, 1H), 0.88-0.86 (m, 9H), 0.02-0.01 (m, 6H); ${ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}, 75 \mathrm{MHz}\right) \delta 157.3(\mathrm{~d}, J$ $=219.6 \mathrm{~Hz}), 153.5,145.8,135.8,120.8,91.6(\mathrm{~d}, J=167.8$ $\mathrm{Hz}), 88.5(\mathrm{~d}, J=16.6 \mathrm{~Hz}), 59.6,57.4,38.4(\mathrm{~d}, J=16.4 \mathrm{~Hz})$, 25.4, 18.4, -5.2; Anal. Calc. for $\mathrm{C}_{16} \mathrm{H}_{23} \mathrm{ClF}_{2} \mathrm{~N}_{4} \mathrm{O}_{2} \mathrm{Si}$ : C, 47.46; H, 5.73; N, 13.84; Found: C, 47.42; H, 5.72; N, 13.81; MS m/z $405(\mathrm{M}+\mathrm{H})^{+}$.
(rel)-(1'R,2'R,3'S)-9-(3'-Hydroxymethyl-2'-fluoro-tetra-hydrofuran-1'-yl) 2-fluoro-6-chloropurine (18). Desilylation of $\mathbf{1 7 b}$ was performed using a procedure similar to that described for 8: yield $78 \%$; UV (MeOH) $\lambda_{\text {max }} 269.5 \mathrm{~nm}$; ${ }^{1} \mathrm{H}$ NMR ( $\left.\mathrm{CDCl}_{3}, 300 \mathrm{MHz}\right) \delta 8.46(\mathrm{~s}, 1 \mathrm{H}), 6.21(\mathrm{dd}, J=18.4$, $5.5 \mathrm{~Hz}, 1 \mathrm{H}), 3.74-3.67(\mathrm{~m}, 3 \mathrm{H}), 3.51-3.49(\mathrm{~m}, 2 \mathrm{H}), 2.21-$ $2.15(\mathrm{~m}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}, 75 \mathrm{MHz}\right) \delta 156.9(\mathrm{~d}, J=$ $217.8 \mathrm{~Hz})$, 154.0, 144.6, 134.8, 121.6, $91.6(\mathrm{~d}, J=169.4$ Hz ), 89.4 (d, $J=16.8 \mathrm{~Hz}$ ), $57.6,56.2,37.2(\mathrm{~d}, J=16.6 \mathrm{~Hz})$; Anal. Calc. for $\mathrm{C}_{10} \mathrm{H}_{9} \mathrm{ClF}_{2} \mathrm{~N}_{4} \mathrm{O}_{2}(+1.0 \mathrm{MeOH})$ : C, $40.94 ; \mathrm{H}$, 4.06; N, 17.36; Found: C, 40.97; H, 4.05; N, 17.34; MS m/z $291(\mathrm{M}+\mathrm{H})^{+}$.
(rel)-(1'R,2'R,3'S)-9-(3'-Carbaldehyde-2'-fluoro-tetra-hydrofuran-1'-yl) 2-fluoro-6-chloropurine (19). Oxidation of $\mathbf{1 8}$ was performed using the Dess-Martin reaction conditions described for 9: yield $66 \%$; ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}, 300\right.$ $\mathrm{MHz}) \delta 8.45(\mathrm{~s}, 1 \mathrm{H}), 6.20(\mathrm{dd}, J=18.6,5.6 \mathrm{~Hz}, 1 \mathrm{H}), 3.72-$ $3.68(\mathrm{~m}, 3 \mathrm{H}), 3.48-3.42(\mathrm{~m}, 2 \mathrm{H}), 2.23-2.16(\mathrm{~m}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR ( $\left.\mathrm{CDCl}_{3}, 75 \mathrm{MHz}\right) \delta 204.6,157.4(\mathrm{~d}, J=218.6 \mathrm{~Hz})$, $154.2,144.7,133.9,122.3,91.3$ (d, $J=167.8 \mathrm{~Hz}$ ), 88.5 (d, $J$ $=16.6 \mathrm{~Hz}), 58.4,57.4,38.4(\mathrm{~d}, J=16.4 \mathrm{~Hz})$.
(rel)-(1'R,2'R,3'S)-9-(3'-Vinyl-2'-fluoro-tetrahydrofuran-1'-yl) 2-fluoro-6-chloropurine\} phosphonate (20). Wittig olefination of the aldehyde $\mathbf{1 0}$ was performed using a procedure similar to that described for 10 : yield $59 \% ;{ }^{1} \mathrm{H}$ NMR ( $\left.\mathrm{CDCl}_{3}, 300 \mathrm{MHz}\right) \delta 8.46(\mathrm{~s}, 1 \mathrm{H}), 6.21(\mathrm{dd}, J=18.4$, $5.4 \mathrm{~Hz}, 1 \mathrm{H}), 5.73(\mathrm{~m}, 1 \mathrm{H}), 5.05-4.98(\mathrm{~m}, 2 \mathrm{H}), 3.73-3.68(\mathrm{~m}$, $3 \mathrm{H}), 3.50(\mathrm{~m}, 2 \mathrm{H}), 2.81-2.76(\mathrm{~m}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C} \mathrm{NMR}\left(\mathrm{CDCl}_{3}, 75\right.$ $\mathrm{MHz}) \delta 157.6(\mathrm{~d}, J=219.4 \mathrm{~Hz}), 154.5,144.3,142.5,134.6$, 123.6, 112.3, $95.6(\mathrm{~d}, J=167.8 \mathrm{~Hz}), 90.2(\mathrm{~d}, J=16.8 \mathrm{~Hz})$,
61.6, 39.1 (d, $J=16.4 \mathrm{~Hz}$ ); Anal. Calc. for $\mathrm{C}_{11} \mathrm{H}_{9} \mathrm{ClF}_{2} \mathrm{~N}_{4} \mathrm{O}$ : C, 46.09; H, 3.16; N, 19.54; Found: C, 46.12; H, 3.15; N, 19.55; MS m/z 287(M+H) ${ }^{+}$.
(rel)-(1'R,2'R,3'S)-Diethyl \{9-(3'-vinyl-2'-fluoro-tetra-hydrofuran-1'-yl) 2-fluoro-6-chloropurine\} phosphonate (21). Phosphonate nucleoside analogue 21 was prepared from 20 using a cross metathesis procedure similar to that described for 11: yield $60 \%$; ${ }^{1} \mathrm{H}$ NMR (DMSO- $d_{6}, 300 \mathrm{MHz}$ ) $\delta 8.44(\mathrm{~s}, 1 \mathrm{H}), 6.67$ (dd, $J=20.2,18.6 \mathrm{~Hz}, 1 \mathrm{H}), 6.21-6.15$ (m, 2H), 4.15-4.10 (m, 4H), 3.72-3.66 (m, 3H), 2.80-2.75 $(\mathrm{m}, 1 \mathrm{H}), 1.35-1.33(\mathrm{~m}, 6 \mathrm{H}),{ }^{13} \mathrm{C}$ NMR (DMSO- $\left.d_{6}, 75 \mathrm{MHz}\right)$ $\delta 157.3(\mathrm{~d}, J=218.8 \mathrm{~Hz}), 154.7,149.5,143.1,133.9,124.3$, 115.3, 95.7 (d, $J=166.6 \mathrm{~Hz}), 89.4(\mathrm{~d}, J=16.4 \mathrm{~Hz}), 62.2$, $61.5,60.9,40.4(\mathrm{~d}, J=16.2 \mathrm{~Hz}), 15.7$; Anal. Calc. for $\mathrm{C}_{15} \mathrm{H}_{18} \mathrm{ClF}_{2} \mathrm{~N}_{4} \mathrm{O}_{4} \mathrm{P}(+1.0 \mathrm{MeOH}): \mathrm{C}, 42.25 ; \mathrm{H}, 4.87 ; \mathrm{N}$, 12.32; Found: C, 42.21; H, 4.89; N, 12.30; MS m/z 423 $(\mathrm{M}+\mathrm{H})^{+}$.
(rel)-(1'R,2'R,3'S)-Diethyl \{9-(3'-vinyl-2'-fluoro-tetra-hydrofuran-1'-yl) 2-fluoro-6-aminopurine\} phosphonate (22a) and (rel)-(1'R,2'R,3'S)-diethyl \{9-(3'-vinyl-2'-fluoro-tetrahydrofuran-1'-yl) 2-amino-6-chloropurine\} phosphonate (22b). Dry ammonia gas was bubbled into a stirred solution of $21(180 \mathrm{mg}, 0.426 \mathrm{mmol})$ in DME $(8.0 \mathrm{~mL})$ at room temperature overnight. Salts were removed by filtration and the filtrate was concentrated under reduced pressure. The residue obtained was purified by silica gel column chromatography ( $\mathrm{MeOH} / \mathrm{CH}_{2} \mathrm{Cl}_{2}, 1: 10$ ) to give 22a ( 24 mg , $14 \%$ ) and 22b ( $80 \mathrm{mg}, 45 \%$ ). Data for 22a; UV (MeOH) $\lambda_{\text {max }} 261.0 \mathrm{~nm} ;{ }^{1} \mathrm{H}$ NMR (DMSO- $\left.d_{6}, 300 \mathrm{MHz}\right) \delta 8.21(\mathrm{~s}$, $1 \mathrm{H}), 7.74\left(\mathrm{br} \mathrm{s}, \mathrm{NH}_{2}, 2 \mathrm{H}\right), 6.65(\mathrm{dd}, J=19.8,16.4 \mathrm{~Hz}, 1 \mathrm{H})$, 6.21 (dd, $J=19.7,18.2 \mathrm{~Hz}, 1 \mathrm{H}), 6.09$ (dd, $J=12.8,5.2 \mathrm{~Hz}$, $1 \mathrm{H}), 4.13-4.09(\mathrm{~m}, 4 \mathrm{H}), 3.73-3.65(\mathrm{~m}, 3 \mathrm{H}), 2.81-2.74(\mathrm{~m}$, $1 \mathrm{H}), 1.54-1.50(\mathrm{~s}, 6 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR (DMSO- $\left.d_{6}, 75 \mathrm{MHz}\right) \delta$ 160.7 (d, $J=268.8 \mathrm{~Hz}), 155.2,152.3,148.8,142.3,119.4$, $115.4,94.8(\mathrm{~d}, J=168.4 \mathrm{~Hz}), 87.2(\mathrm{~d}, J=17.2 \mathrm{~Hz}), 63.4$, $62.8,61.5,39.6(\mathrm{~d}, J=16.6 \mathrm{~Hz}), 14.4$; Anal. Calc. for $\mathrm{C}_{15} \mathrm{H}_{20} \mathrm{~F}_{2} \mathrm{~N}_{5} \mathrm{O}_{4} \mathrm{P}(+1.0 \mathrm{MeOH}): \mathrm{C}, 44.14 ; \mathrm{H}, 5.55 ; \mathrm{N}, 16.08$; Found: C, 44.11; H, 5.57; N, 16.09; MS m/z $404(\mathrm{M}+\mathrm{H})^{+}$. Data for 22b; UV (MeOH) $\lambda_{\text {max }} 308.5 \mathrm{~nm} ;{ }^{1} \mathrm{H}$ NMR (DMSO$\left.d_{6}, 300 \mathrm{MHz}\right) \delta 8.14(\mathrm{~s}, 1 \mathrm{H}), 7.71\left(\mathrm{br} \mathrm{s}, \mathrm{NH}_{2}, 2 \mathrm{H}\right), 6.62(\mathrm{dd}$, $J=20.2,17.8 \mathrm{~Hz}, 1 \mathrm{H}), 6.24(\mathrm{dd}, J=19.6,17.7 \mathrm{~Hz}, 1 \mathrm{H}), 6.12$ (dd, $J=14.6,5.0 \mathrm{~Hz}, 1 \mathrm{H}), 4.16-4.10(\mathrm{~m}, 4 \mathrm{H}), 3.75-3.68(\mathrm{~m}$, 3 H ), 2.82-2.75 (m, 1H), 1.53-1.49 (s, 6H), ${ }^{13}$ C NMR (DMSO$\left.d_{6}, 75 \mathrm{MHz}\right) \delta 158.5,154.4,151.7,149.4,144.0,125.2$, $114.6,93.9(\mathrm{~d}, J=166.8 \mathrm{~Hz}), 88.7(\mathrm{~d}, J=16.8 \mathrm{~Hz}), 62.8$, $62.2,61.6,41.2(\mathrm{~d}, J=16.8 \mathrm{~Hz}), 14.7$; Anal. Calc. for $\mathrm{C}_{15} \mathrm{H}_{20} \mathrm{ClFN}_{5} \mathrm{O}_{4} \mathrm{P}(+1.0 \mathrm{MeOH}): \mathrm{C}, 42.53 ; \mathrm{H}, 5.35 ; \mathrm{N}, 15.50$; Found: C, 42.48; H, 5.36; N, 15.48; MS m/z $420(\mathrm{M}+\mathrm{H})^{+}$. (rel)-(1'R,2'R,3'S)-9-\{(3'-Vinyl-2'-fluoro-tetrahydrofuran-$\mathbf{1}^{\prime}$-yl) guanine\} phosphonic acid (23). To a solution of 22b ( $159 \mathrm{mg}, 0.379 \mathrm{mmol}$ ) in dry $\mathrm{CH}_{3} \mathrm{CN}(15 \mathrm{~mL})$ was added trimethylsilyl bromide ( $0.0873 \mathrm{~mL}, 6.62 \mathrm{mmol}$ ) at room temperature. The mixture was stirred for 24 h , and solvent was removed by co-evaporation with methanol three times. The residue was dissolved in $\mathrm{MeOH}(15.0 \mathrm{~mL})$ and 2mercaptoethanol ( $105.5 \mathrm{~mL}, 1.52 \mathrm{mmol}$ ), and then NaOMe ( $80.6 \mathrm{mg}, 1.52 \mathrm{mmol}$ ) was added. The mixture was refluxed
for 18 h under $\mathrm{N}_{2}$, cooled, neutralized with glacial AcOH , and evaporated to dryness in vacuo. The residue obtained was co-evaporated from conc $\mathrm{NH}_{4} \mathrm{OH}(2 \times 25 \mathrm{~mL})$ and the resultant solid was triturated with acetone $(2 \times 10 \mathrm{~mL})$. After evaporating the acetone, the residue was purified by preparative column chromatography using reverse-phase C18 silica gel and elution with water. Lyophilization of the appropriate fraction provided $23(53.52 \mathrm{mg}, 39 \%)$ as a yellowish salt (ammonium salt). mp $162-164{ }^{\circ} \mathrm{C}$; UV $\left(\mathrm{H}_{2} \mathrm{O}\right) \lambda_{\max } 254.0$ $\mathrm{nm} ;{ }^{1} \mathrm{H}$ NMR ( $\mathrm{D}_{2} \mathrm{O}, 300 \mathrm{MHz}$ ) $\delta 10.8$ (br s, NH, 1 H ), 8.12 (s, 1H), 7.03 (br s, $\left.\mathrm{NH}_{2}, 2 \mathrm{H}\right), 6.63(\mathrm{dd}, J=19.4,17.2 \mathrm{~Hz}$, $1 \mathrm{H})$, 6.17-6.08 (m, 2H), 3.72-3.65 (m, 3H), 2.80-2.75 (m, 1 H ); ${ }^{13} \mathrm{C}$ NMR ( $\left.\mathrm{D}_{2} \mathrm{O}, 75 \mathrm{MHz}\right) \delta 157.4,154.6,152.3,149.4$, 136.2, 119.1, 115.3, $96.0(\mathrm{~d}, J=168.1 \mathrm{~Hz}), 86.9(\mathrm{~d}, J=17.4$ $\mathrm{Hz}), 63.0,62.5,61.8,40.4(\mathrm{~d}, J=17.2 \mathrm{~Hz}) ; \operatorname{HPLC} t_{\mathrm{R}}=9.82$ min ; HRMS $[\mathrm{M}-\mathrm{H}]^{+}$req. 344.0678, found 344.0676.
(rel)-(1'R,2'R,3'S)-Diethyl \{9-(3'-ethyl-2'-fluoro-tetra-hydrofuran-1-yl) 2-fluoro-6-chloropurine\} phosphonate (24). Compound 24 was synthesized from 21 by transfer catalytic hydrogenation similar to that described for 14 : yield $73 \%$; ${ }^{1} \mathrm{H}$ NMR ( $\mathrm{DMSO}-d_{6}, 300 \mathrm{MHz}$ ) $\delta 8.43(\mathrm{~s}, 1 \mathrm{H})$, 6.12 (dd, $J=15.8,5.0 \mathrm{~Hz}, 1 \mathrm{H}), 4.12-4.09$ (m, 4H), 3.73-3.66 $(\mathrm{m}, 3 \mathrm{H}), 2.81-2.76(\mathrm{~m}, 1 \mathrm{H}), 2.14-2.02(\mathrm{~m}, 4 \mathrm{H}), 1.52-1.50$ $(\mathrm{m}, 6 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR (DMSO- $\left.d_{6}, 75 \mathrm{MHz}\right) \delta 157.1(\mathrm{~d}, J=$ $218.8 \mathrm{~Hz}, 1 \mathrm{H}), 153.1,145.3,136.2,121.2,96.2(\mathrm{~d}, J=166.8$ $\mathrm{Hz}), 88.5(\mathrm{~d}, J=16.7 \mathrm{~Hz}), 61.8,41.1(\mathrm{~d}, J=17.2 \mathrm{~Hz}), 28.5$, 18.4, 14.7; Anal. Calc. for $\mathrm{C}_{15} \mathrm{H}_{20} \mathrm{ClF}_{2} \mathrm{~N}_{4} \mathrm{O}_{4} \mathrm{P}(+1.0 \mathrm{MeOH})$ : C, 42.07; H, 5.29; N, 12.26; Found: C, 42.11; H, 5.31; N, 12.24; MS m/z $425(\mathrm{M}+\mathrm{H})^{+}$.
(rel)-(1'R,2'R,3'S)-Diethyl \{9-(3'-ethyl-2'-fluoro-tetra-hydrofuran-1-yl) 2-fluoro-6-aminopurine\} phosphonate (25a) and (rel)-( $\left.1^{\prime} R, 2^{\prime} R, 3 ' S\right)$-diethyl \{9-(3'-ethyl-2'-fluoro-tetrahydrofuran-1-yl) 2-amino-6-chloropurine\} phosphonate (25b). Ammonolysis of 24 using the same procedure described for 14 gave 25a and 25b. Data for 25a; yield 13\%; $\mathrm{UV}(\mathrm{MeOH}) \lambda_{\max } 261.5 \mathrm{~nm} ;{ }^{1} \mathrm{H}$ NMR (DMSO- $d_{6}, 300 \mathrm{MHz}$ ) $\delta 8.20(\mathrm{~s}, 1 \mathrm{H}), 7.74\left(\mathrm{br} \mathrm{s}, \mathrm{NH}_{2}, 2 \mathrm{H}, \mathrm{D}_{2} \mathrm{O}\right.$ exchangeable), 6.08 (d, $J=15.8,5.2 \mathrm{~Hz}, 1 \mathrm{H}), 4.14-4.10(\mathrm{~m}, 4 \mathrm{H}), 3.71-3.64(\mathrm{~m}$, $3 \mathrm{H}), 2.82-2.75(\mathrm{~m}, 1 \mathrm{H}), 2.12-1.99(\mathrm{~m}, 4 \mathrm{H}), 1.55-1.53(\mathrm{~m}$, 6 H ); ${ }^{13} \mathrm{C}$ NMR (DMSO- $\left.d_{6}, 75 \mathrm{MHz}\right) \delta 161.0(\mathrm{~d}, J=267.8$ $\mathrm{Hz}, 1 \mathrm{H}), 155.3,152.3,142.6,123.4,98.6(\mathrm{~d}, J=168.4 \mathrm{~Hz})$, 89.4 (d, $J=16.8 \mathrm{~Hz}$ ), $60.6,40.4$ (d, $J=17.4 \mathrm{~Hz}$ ), 29.4, 18.7, 15.4; Anal. Calc. for $\mathrm{C}_{15} \mathrm{H}_{22} \mathrm{~F}_{2} \mathrm{~N}_{5} \mathrm{O}_{4} \mathrm{P}(+1.0 \mathrm{MeOH})$ : C , 43.94; H, 5.99; N, 16.01; Found: C, 43.90; H, 6.01; N, 16.00; MS $m / z 406(\mathrm{M}+\mathrm{H})^{+}$. Data for 25b; yield $43 \%$; UV $(\mathrm{MeOH}) \lambda_{\text {max }} 307.0 \mathrm{~nm} ;{ }^{1} \mathrm{H}$ NMR (DMSO- $\left.d_{6}, 300 \mathrm{MHz}\right) \delta$ $8.22(\mathrm{~s}, 1 \mathrm{H}), 7.75\left(\mathrm{br} \mathrm{s}, \mathrm{NH}_{2}, 2 \mathrm{H}, \mathrm{D}_{2} \mathrm{O}\right.$ exchangeable), 6.13 (d, $J=16.0,5.0 \mathrm{~Hz}, 1 \mathrm{H}), 4.15-4.11(\mathrm{~m}, 4 \mathrm{H}), 3.72-3.67(\mathrm{~m}$, $3 \mathrm{H}), 2.81-2.74(\mathrm{~m}, 1 \mathrm{H}), 2.14-2.02(\mathrm{~m}, 4 \mathrm{H}), 1.53-1.52(\mathrm{~m}$, 6 H ); ${ }^{13} \mathrm{C}$ NMR (DMSO- $\left.d_{6}, 75 \mathrm{MHz}\right) \delta 158.6,154.8,151.0$, 143.8, 125.7, 99.2 (d, $J=167.8 \mathrm{~Hz}), 88.6(\mathrm{~d}, J=16.4 \mathrm{~Hz})$, $61.7,41.2(\mathrm{~d}, J=16.7 \mathrm{~Hz}), 28.7,19.0,14.8$; Anal. Calc. for $\mathrm{C}_{15} \mathrm{H}_{22} \mathrm{ClFN}_{5} \mathrm{O}_{4} \mathrm{P}(+1.0 \mathrm{MeOH})$ : C, $42.34 ; \mathrm{H}, 5.77$; N, 15.43; Found: C, 42.30; H, 5.75; N, 15.45; MS m/z 422.
(rel)-(1'R,2'R,3'S)-9-\{(3'-Ethyl-2'-fluoro-tetrahydrofuran-1-yl) guanine\} phosphonic acid (26). Nucleoside phosphonic acid 26 was synthesized using the hydrolysis conditions
used for 23. Yield $42 \%$; mp $159-162{ }^{\circ} \mathrm{C}$; UV $\left(\mathrm{H}_{2} \mathrm{O}\right) \lambda_{\text {max }}$ $252.5 \mathrm{~nm} ;{ }^{1} \mathrm{H}$ NMR (DMSO- $d_{6}, 300 \mathrm{MHz}$ ) $\delta 10.8$ (br s, NH, $\mathrm{H}, \mathrm{D}_{2} \mathrm{O}$ exchangeable), $8.11(\mathrm{~s}, 1 \mathrm{H}), 7.10\left(\mathrm{br} \mathrm{s}, \mathrm{NH}_{2}, 2 \mathrm{H}\right.$, $\mathrm{D}_{2} \mathrm{O}$ exchangeable), $6.11(\mathrm{~d}, J=16.2,5.2 \mathrm{~Hz}, 1 \mathrm{H}), 3.70-3.64$ $(\mathrm{m}, 3 \mathrm{H}), 2.77-2.71(\mathrm{~m}, 1 \mathrm{H}), 2.09-1.94(\mathrm{~m}, 4 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR (DMSO- $d_{6}, 75 \mathrm{MHz}$ ) d 157.8, 154.4, 152.3, 136.5, 119.2, $97.2(\mathrm{~d}, J=168.0 \mathrm{~Hz}), 90.1(\mathrm{~d}, J=16.4 \mathrm{~Hz}), 60.5,40.4(\mathrm{~d}, J$ $=16.4 \mathrm{~Hz}$ ), 28.5, 18.6, 15.3; HPLC $t_{\mathrm{R}}=10.02 \mathrm{~min} ; \mathrm{HRMS}$ [M-H] ${ }^{+}$req. 346.0676, found 346.0675.
(rel)-(1'R,2'R,3'S)- Bis(SATE) phosphoester of [9-(3'-ethylphosphonate-2'-fluoro-tetrahydrofuran-1-yl)] adenine (28). A solution of adenine phosphonic acid (ammonium salt) derivative $\mathbf{1 6}(73.80 \mathrm{mg}, 0.212 \mathrm{mmol})$ and tri- $n$-butylamine ( $117 \mathrm{mg}, 0.636 \mathrm{mmol}$ ) in methanol ( 4.5 mL ) was mixed for 30 min and concentrated under reduced pressure. The residue was thoroughly dried with anhydrous ethanol and toluene. The resulting foamy solid was dissolved in anhydrous pyridine ( 15 mL ) to which thioester $27(649 \mathrm{mg}$, 4.0 mmol ) and 1-(2-mesitylenesulfonyl)-3-nitro-1 $\mathrm{H}-1,2,4-$ triazole ( $251 \mathrm{mg}, 0.848 \mathrm{mmol}$ ) were added. The mixture was stirred overnight at room temperature and quenched with tetrabutylammonium bicarbonate buffer ( $12.0 \mathrm{~mL}, 1 \mathrm{M}$ solution, pH 8.0 ). The mixture was concentrated under reduced pressure and the residue was diluted with water (70 $\mathrm{mL})$ and extracted with $\mathrm{CHCl}_{3}(80 \mathrm{~mL})$ two times. The combined organic layer was washed with brine, dried over $\mathrm{Na}_{2} \mathrm{SO}_{4}$, filtered, and evaporated. The residue was purified by silica gel column chromatography ( $\mathrm{MeOH} / \mathrm{Hexane} / \mathrm{EtOAc}$, $0.05: 4: 1)$ to give $\mathbf{2 8}(48 \mathrm{mg}, 37 \%)$ as a white solid: $\mathrm{mp} 131-$ $133{ }^{\circ} \mathrm{C} ; \mathrm{UV}(\mathrm{MeOH}) \lambda_{\max } 262.0 \mathrm{~nm} ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}, 300\right.$ $\mathrm{MHz}) \delta 8.30(\mathrm{~s}, 1 \mathrm{H}), 8.15(\mathrm{~s}, 1 \mathrm{H}), 6.11(\mathrm{~d}, J=16.1,5.0 \mathrm{~Hz}$, $1 \mathrm{H}), 4.03-4.02(\mathrm{~m}, 4 \mathrm{H}), 3.56(\mathrm{~d}, J=9.6 \mathrm{~Hz}, 2 \mathrm{H}), 3.16(\mathrm{t}, J=$ 6.4 Hz, 4H), 2.21-2.13 (m, 2H), 1.22-1.16 (s, 18H); ${ }^{13} \mathrm{C}$ NMR ( $\left.\mathrm{CDCl}_{3}, 75 \mathrm{MHz}\right) \delta 204.2,157.1,154.7,152.8,148.2$, 145.6, 124.6, 119.4, $96.6(\mathrm{~d}, J=168.2 \mathrm{~Hz}), 88.8(\mathrm{~d}, J=18.4$ Hz ), $83.6,67.5,67.3,62.4,61.6,53.4,38.4(\mathrm{~d}, J=16.4 \mathrm{~Hz})$, 30.2, 28.7, 28.5, 23.7, 14.6; Anal. Calc. for $\mathrm{C}_{25} \mathrm{H}_{39} \mathrm{FN}_{5} \mathrm{O}_{6} \mathrm{PS}_{2}$ (+1.0 MeOH): C, 47.95; H, 6.65; N, 10.75. Found: C, 47.90; H, 6.61; N, 10.79; MS m/z $620(\mathrm{M}+\mathrm{H})^{+}$.

## References

1. (a) Votruba, I.; Bernaerts, R.; Sakuma, T.; De Clercq, E.; Merta, A.; Rosenberg, I.; Holý, A. Mol. Pharmacol. 1987, 32, 524. (b) Balzarini, J.; Hao, Z.; Herdewijn, P.; Johns, D. G.; De Clercq, E. Proc. Natl. Acad. Sci. USA 1991, 88, 1499.
2. (a) Hong, J. H.; Lee, K.; Choi, Y.; Chu, C. K. Tetrahedron Lett. 1998, 39, 3443. (b) Jeong, B. S.; Lee, J. W.; Son, H. J.; Kim, B. Y.; Ahn, S. K. Terahedron; Asymmetry 2005, 16, 3795.
3. (a) Boojamra, C. G.; Mackman, R. L.; Markevitch, D. Y.; Prasad, V.; Ray, A. S.; Douglas, J.; Grant, D.; Kim, C. U.; Cihlar, T. Bioorg. Med. Chem. Lett. 2008, 18, 1120. (b) Cihlar, T.; Ray, A. S.; Boojamra, C. G.; Zhang, L.; Hui, H.; Laflamme, G.; Vela, J. E.; Grant, D.; Chen, J.; Myrick, F.; White, K. L.; Gao, Y.; Lin, K. Y.; Douglas, J. L.; Parkin, N. T.; Carey, A.; Pakdaman, R.; Mackman, R. L. Antimicrob. Agents Chemother 2008, 52, 655.
4. Vina, D.; Wu, T.; Renders, M.; Laflamme, G.; Herdewijn, P. Tetrahedron 2007, 63, 2634.
5. Schöning, K.; Scholz, P.; Guntha, S.; Wu, X.; Krishnamurthy, R.; Eschenmoser, A. Science 2000, 5495, 1347.
6. (a) Kempeneers, V.; Vastmans, K.; Rozenski, J.; Herdewijn, P. Nucleic Acids Res. 2003, 31, 6221. (b) Chaput, J. C.; Szostak, J. W. J. Am. Chem. Soc. 2003, 125, 9274.
7. Kempeneers, V.; Froeyen, M.; Vastmans, K.; Herdewijn, P. Chem Biodivers. 2004, 1, 112.
8. Wu, T.; Froeyen, M.; Kempeneers, V.; Pannecouque, C.; Wang, J.; Busson, R.; De Clercq, E.; Herdewijn, P. J. Am. Chem. Soc. 2005, 127, 5056.
9. (a) Secrist, J. A. III; Riggs, R. M.; Comber, R. N.; Montgomery, J. A. Nucleosides, Nucleotides 1992, 11, 947. (b) Montgomery, J. A.; Thomas, H. J.; Kisliuk, R. L.; Gaumont, Y. J. Med. Chem. 1979, 22, 109.
10. Hong, J. H.; Kim, H. O.; Moon, H. R.; Jeong, L. S. Arch. Pharm. Res. 2001, 24, 95.
11. Handbook of Nucleoside Synthesis; Vorbruggen, H., Ruh-Pohlenz, C., Eds.; John Wiley \& Sons. Inc.: New York, 2001.
12. (a) Amey, R. L.; Martin. J. C. J. Am. Chem. Soc. 1978, 100, 300. (b) Amey, R. L.; Martin, J. C. J. Am. Chem. Soc. 1979, 101, 5294.
13. Maryanoff, B. E.; Reitz, A. B. Chem. Rev. 1989, 89, 863.
14. (a) Kumamoto, H.; Topalis, D.; Broggi, J.; Pradere, U.; Roy, V.; Berteina-Raboin, S.; Nolan, S. P.; Deville-Bonne, D.; Andrei, G.; Snoeck, R.; Garin, D.; Crance, J.-M.; Agrofoglio, L. A. Tetrahedron 2008, 64, 3517. (b) Montagu, A.; Pradere, U.; Roy, V.; Nolan, S. P.; Agrofoglio, L. A. Tetrahedron 2011, 67, 5317.
15. Scholl, M.; Ding, S.; Lee, C. W.; Grubbs, R. H. Org. Lett. 1999, 1,

## 953.

16. Hocková, D.; Holý, A.; Masojídková, M.; Keough, D. T.; De Jersey, J.; Guddat, L. W. Bioorg. Med. Chem. 2009, 17, 6218.
17. Huang, Q.; Herdewijn, P. J. Org. Chem. 2011, 76, 3742.
18. Robins M. J.; Uznanski, B. Can. J. Chem. 1981, 59, 2608.
19. Montgomery, J.; Hewson, K. J. Med. Chem. 1969, 12, 498.
20. Tong, G. L.; Ryan, K. J.; Lee, W. W.; Acton, E. M.; Goodman, L. J. Org. Chem. 1967, 32, 859.
21. Lefebvre, I.; Périgaud, C.; Pompon, A.; Aubertin, A. M.; Girardet, J. L.; Kirn, A.; Gosselin, G.; Imbach, J. L. J. Med. Chem. 1995, 38, 3941.
22. Périgaud, C.; Gosselin, G.; Lefebvre, I.; Girardet, J. L.; Benzaria, S.; Barber, I.; Imbach, J. L. Bioorg. Med. Chem. Lett. 1993, 3, 2521.
23. Holy, A.; Votruba, I.; Merta, A.; Cerny, J.; Vesely, J.; Vlach, J.; Sediva, K.; Rosenberg, I.; Otmar, M.; Hrebabecky, H.; Travniekb, M.; Vonkac, V.; Snoeck, R.; De Clercq, E. Antiviral Res. 1990, 13, 295.
24. Pauwels, R.; Balzarini, J.; Baba, M.; Snoeck, R.; Schols, D.; Herdewijn, P.; Desmyter, J.; De Clercq, E. J. Virol. Methods 1988, 20, 309.
25. All geometries were optimized with the framework of the density functional theory (DFT), with Spartan modeling software. The B3LYP functional with 6-31G* basis set was employed.
