

# Early Morphological Development of the Brown Croaker, *Miichthys miiuy* (Basilewsky): Fin Differentiation, Head Dimensions, and Squamation

By In-Seok Park\*, Young Ja Kim<sup>1</sup>, In Bon Goo and Dong Soo Kim<sup>2</sup>

Division of Marine Environment and Bioscience, College of Ocean Science and Technology, Korea Maritime University, 727 Taejong-ro, Yeong do-gu, Busan 606-791, Korea

<sup>1</sup>Korea Environmental Industry & Technology Institute (KEITI), 215 Jinheung-ro, Eunpyeong-gu, Seoul 122-706, Korea

<sup>2</sup>Institute of Marine Living Modified Organisms (iMLMO), Pukyong National University, 45 Yongso-ro, Nam-gu, Busan 608-737, Korea

**ABSTRACT** We describe early morphological development in laboratory-reared specimens of the brown croaker, *Miichthys miiuy*, in relation to fin differentiation, head dimensions, and squamation. From the yolk sac stage to the flexion larval stage (a period of 12 days following hatching, at which time the larvae were <4.2 mm in total length; TL) we observed the presence of a fin-fold around the body, while the caudal fin appeared rounded and lacked scales. Rays developed in the dorsal, anal, and pectoral fins in a process that was almost complete in larvae 12 days, while ray segmentation occurred between 26 and 29 days of age. Elongation of the middle rays of the caudal fin was initiated at 32 days, and the rays were remarkably elongated by 37 days. By 68 days the caudal fin was lanceolated (50.7 mm TL). Scales began to develop from the midlateral lines of the caudal peduncle at 9.1 mm TL (28 days), eventually encompassing the entire operculum (22.1 mm TL; 44 days). The head dimensions were largely stabilized at >12 mm TL (30 day).

**Key words** : Fin differentiation, head dimensions, *Miichthys miiuy*, squamation

## INTRODUCTION

The brown croaker, *Miichthys miiuy* Basilewsky has adapted well to the western coastal waters of Korea, which are characterized by high turbidity resulting from strong tidal currents and low water temperatures in winter (Park *et al.*, 2007). Given the commercial importance of brown croaker, especially to the aquaculture industry (Seo, 2004), information on aspects of its biology and early morphological development are of great interest (Park *et al.*, 2007).

Detailed morphological information is important for the early detection of both morphological and physiological abnormalities in reared fish (Mana and Kawamura, 2002). Fish reared in hatcheries for release as juveniles into the wild experience conditions during the maturation process that differ markedly from those encountered by

fish raised in the wild (Hard *et al.*, 2000). As hatchery-reared (hereafter, reared) fish are not well equipped to survive in the natural environment, most die during early stages of development (Hughes *et al.*, 1992; Mana and Kawamura, 2002; Seo, 2004), or often harbor a variety of morphological abnormalities if they survive (Kanazawa, 1993; Dedi *et al.*, 1997).

Although more information about the developmental stages of this fish is required because of the high levels of hatchery mortality that commonly occur during the early life stages of reared larvae, there have been no detailed reports of the anatomy of this species other than those on egg development and morphological changes in brown croaker larvae (Han *et al.*, 2002). The early life stages in ichthyoplankton have been characterized in surveys of developmental series of specimens (Russell, 1976; Dunn, 1984). To construct a morphological database of early life stages, we studied fin development, head dimensions, and squamation in a series of laboratory-reared specimens.

\*Corresponding author: In-Seok Park Tel: 82-51-410-4321,  
Fax: 82-51-405-4322, E-mail: ispark@hhu.ac.kr

## MATERIALS AND METHODS

The brown croaker, *Miichthys miiuy* were produced from naturally fertilized eggs of wild adults. The eggs were stocked in 20-tonne concrete tanks and reared according to established commercial procedures. Briefly, hatched larvae were fed enriched rotifers from 4 days, and then fed *Artemia nauplii* with artificial diets from 15 days. The water temperature during the fish production period was not controlled and ranged from 25.1 to 27.3°C. All observations and measurements were made on pre-larvae (n=72) of 3.0 mm total length (TL), and juveniles (n=48; TL=60.0 mm). The two size groups were preserved in 5% and 10% formalin solution, respectively.

As shown in Fig. 1, for specimens of >40.0 mm TL, four head dimensions were measured to the nearest 0.1 mm using a digital vernier caliper (CD-20CP, Mitutoyo, Japan). These included: head length (HL; the most anterior extension of the head to the most posterior point of the operculum); postorbital length (PL; the most posterior point of the eye to the most posterior point of the operculum); snout length (SNL; the most anterior extension of the head to the most anterior point of the eye); and eye diameter (ED). Larvae and juveniles of <40.0 mm TL were observed using a microscope (Axioskop 40 FL, Zeiss, Germany) with a mounted video camera (AxioCam MRm, Zeiss, Germany) connected to a computer; images were interpreted using image analysis software (Axiovision4, Zeiss, Germany). Specimens were periodically sampled for assessment of fin differentiation and squamation, following staining with alizarin red S

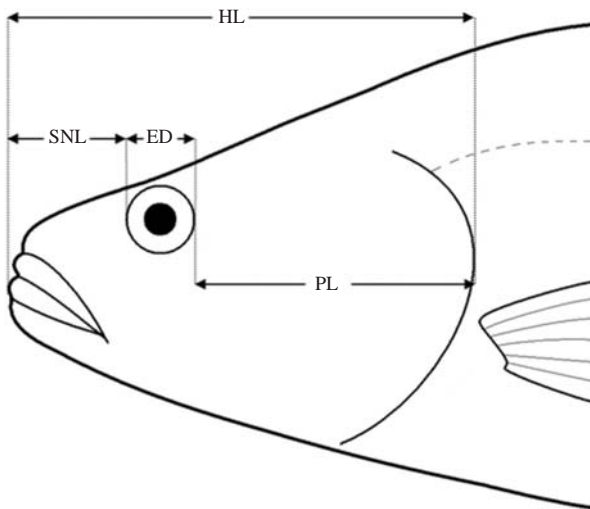


Fig. 1. Head dimensions used in this study. HL, head length (from the most anterior extension of the head to the most posterior point of the operculum); PL, postorbital length (from the most posterior point of the eye to the most posterior point of the operculum); SNL, snout length (most anterior extension of the head to the most anterior point of the eye); ED, eye diameter.

(Sigma, USA). The developmental stages were identified according to the criteria described by Russell (1976) and Han *et al.* (2002).

## RESULTS AND DISCUSSION

The dorsal fin anlage in brown croaker, *Miichthys miiuy* appeared when the larvae were at 3.7 mm TL (8 days); ray formation occurred at 4.2 mm TL, 12 days after hatching; ray segmentation was initiated at 11.9 mm TL (29 days); and the formation of ray branching was initiated at 38.5 mm TL (>51 days). Anal fin anlage appeared at 3.7 mm TL (8 days); ray formation was initiated at 4.2 mm TL (12 days); and ray segmentation was initiated at 7.4 mm TL (26 days). The pectoral fin bud appeared at 3.5 mm TL (2 days) and was initially fanlike. Ray formation was initiated at 4.2 mm TL (12 days), and elongated with elongation of the middle ray of the caudal fin ray at 13.2 mm TL (35 days). The caudal fin became rounded immediately following hatching, and at 5.9 mm TL, during the postlarval stage (16 days), it was particularly long and rounded at the end margin. The middle fin ray was prominently elongated at >32 days, but then became lanceolated at 50.7 mm TL in juveniles 68 days. The full complement of ray counts was initially observed in the caudal ray, followed by the anal, dorsal, and pectoral rays.

The use of reared larvae in studying fish ontogeny has been previously proposed (Hunter, 1984; Myung *et al.*, 2004). However, some critical considerations must be taken account of because the rearing method has significant effects on development (Hunter, 1984; Koumoundouros *et al.*, 2001). In this study we found slightly different values for several parameters relative to those reported by Han *et al.* (2002). These include length of fish, day of hatching, and initiation and completion of ray formation, segmentation, and branching. The differences may be a consequence of differences in the sampling and rearing methods used. In the present study all fin rays were observed to have been completely developed between 30 and 37 days of age, and the size corresponding to these ages largely encompassed the juvenile stage, as reported by Han *et al.* (2002). Specifically, the length of the middle rays of the caudal fin began to increase at 32 days, and then became prominently elongated after 37 days.

Fin elongation and spination have roles in maintaining buoyancy, but may also be central to the species' predator avoidance strategy (Moser, 1981). Fin development is an important process in the early life stages of fish, and has been intimately correlated with both swimming speed, and feeding techniques and preferences (Fukuhara, 1992). Consequently, specific fin ray elongation is assumed to be key to distinguishing development stages among

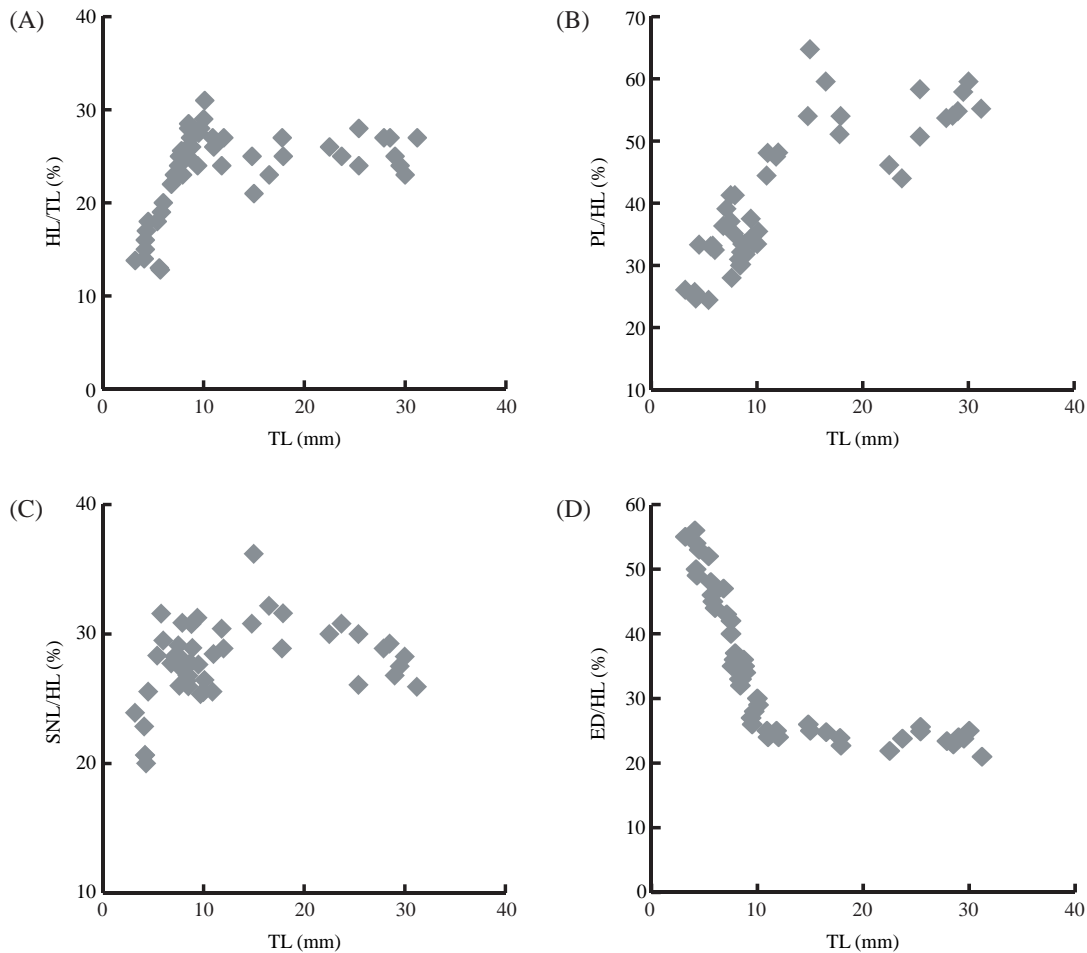


Fig. 2. Scatter diagrams showing the proportion of various dimensions to total length (TL) in fingerlings of reared brown croaker, *Miichthys miuy*. (A) head length (HL/TL), (B) postorbital length (PL/HL), (C) snout length (SNL/HL), and (D) eye diameter (ED/HL).

related species.

The initial head length to total length (HL/TL) proportion was 13.0~17.4% in specimens of 3.2~4.9 mm TL, and this increased gradually with increasing size, eventually reaching 23.8~37.4% at 5.0~6.3 mm TL (Fig. 2A). In specimens of 6.6~7.8 mm TL, the relative degree to which growth in head length had occurred was quite variable (23.8~32.0%), but it stabilized gradually with further growth to an average of 25.0%.

The postorbital length to head length (PL/HL) proportion was found to increase with size, reaching 33.1% in specimens of <8.0 mm TL, but then stabilized with further growth, eventually ranging from 50.0~60.3% (Fig. 2B). The snout length to head length (SNL/HL) proportion increased markedly in specimens of >6.2 mm TL (from 20.0% to 32.1%), decreased to 25.3% at 8.0 mm TL, and stabilizing at 25.9~31.9% with further growth (Fig. 2C). The eye diameter to head length (ED/HL) proportion was approximately 50.0~55.6% in specimens of 8.0 mm HL (Fig. 2D). The growth of all head dimen-

sions stabilized at approximately 30 days of age and a HL > 12 mm, coinciding with extension of the middle ray of the caudal fin.

The origin of scales and squamation in the brown croaker is illustrated in Fig. 2. The initial site of scale formation was parallel to the longitudinal principal axis of the caudal peduncle, and occurred at 9.1 mm TL (28 days); no scales were observed on any fish younger than this age. Scale development progressed in an anterior direction along the midlateral line, and reached the midlateral line area posterior to the head at 11.7 mm TL (29 days).

New scales then developed on the dorsal and ventral regions, with scale formation around the circumference of the caudal peduncle being completed at 13.2 mm TL (35 days). Scales subsequently emerged around the gill cover and ventral region of the eye at 15.8 mm TL (37 days), and the entire operculum was covered with scales at 22.1 mm TL (44 days). Squamation in juveniles was complete at 25.0 mm TL (57 days). The single patch place

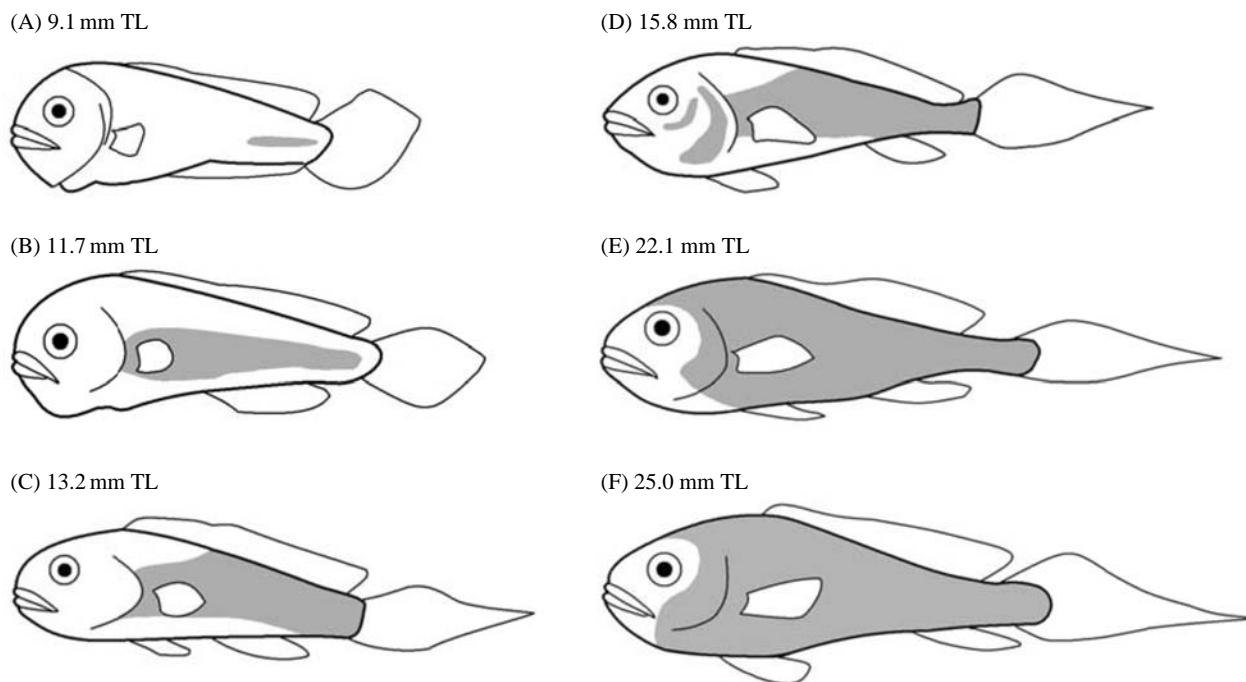


Fig. 3. Development of squamation in the brown croaker, *Miichthys miiuy*. The area of the body covered by scales is shaded in the diagrams. TL, total length.

on the caudal peduncle is a common pattern (White, 1977). However, scale development varies among fish, especially with regard to the number and location of patches, as does the correlation of scale formation with the length and age of the fish (Fukuhara and Fushimi, 1988; Park and Lee, 1988; Fukuhara, 1992). In this study, the specimens tended to develop patches in three places on the body. Squamation typically progressed along the lateral line, and then moved dorsally and ventrally. Scale development initially occurred laterally on the central portion of the caudal peduncle, progressed along the midlateral line, then extended dorsally and ventrally. A similar pattern was observed in *Rivulus marmoratus* (Cyprinodontidae), but without the caudal peduncle being involved initially (Park and Lee, 1988).

Larger and longer fish develop more squamation than smaller fish, including the zebra fish *Brachydanio rerio* (Armstrong, 1973) and *R. marmoratus* (Park and Lee, 1988). We noted some differences among fish with respect to the scale patch region. Yellow sail red bass, *Callanthis japonicus* (Kim and Okiyama, 1989), red sea bream, *Pagrus major*, and black sea bream, *Acanthopagrus schlegeli* (Fukuhara, 1992) initially develop scales on the middle of the body. However, in the Japanese amberjack, *Seriola quinqueradiata*, scale patches first emerge around the caudal peduncle and the anal fin (Fukuhara, 1992). Table 1 was shown sequence of fin differentiation, and head and scale development in relation to TL in the brown croaker. The identification of early life stages in

Table 1. Sequence of fin differentiation, and head and scale development in relation to total length in the brown croaker, *Miichthys miiuy*

Stage	Range (mm)	Age
Stage A (Yolksac larva to flexion)		
Finfold exist	2.3 ~ 3.8	< 10-d
No scales	2.3 ~ 8.8	< 26-d
Stage B (Post flexion larva)		
Fin ray formation begins		
Dorsal fin	4.2 ~ 4.8	12-d
Anal fin	4.0 ~ 4.5	12-d
Pectoral fin	4.0 ~ 5.7	12-d
Caudal fin is getting longer	5.9 ~ 6.5	16-d
Stage C (Post flexion larva)		
Fin segmentation initiated		
Dorsal fin	11.9 ~ 12.0	29-d
Anal fin	7.4 ~ 8.0	26-d
Elongation of middle ray of caudal fin		
Initiated	12.3 ~ 13.9	30-d
Remarkably elongated	15.9 ~ 16.8	< 32-d
Scale developed along midlateral lines	9.1 ~ 13.2	28-d to 35-d
Gill cover and ventral region of eye	15.8 ~ 22.1	37-d to 44-d
Entirely covering the operculum	25.0 ~ 27.0	57-d
Growth of head dimension stabilized	12.3 ~ 30.8	> 30-d
Stage D (Juvenile)		
Caudal fin initially lanceolated	56.1 ~ 70.0	68-d
Squamation completed	25.0	57-d

ichthyoplankton was previously surveyed by examination of developmental stages of specimens (Russell, 1976).

These developmental data are of critical importance to the early detection and elimination of morphological deformities in reared fish (Koumoundouros *et al.*, 2001). These result may prove to be useful indicators in the successful rearing of brown croaker fingerlings.

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## REFERENCES

- Armstrong, J.G. 1973. Squamation chronology of the zebra-fish (Cyprinidae), *Brachydnio rerio*. *Copeia*, 4: 823-824.
- Dedi, J., T. Takeuchi, T. Seikai, T. Watanabe and K. Hosoya. 1997. Hypervitaminosis A during vertebral morphogenesis in larval Japanese flounder. *Fish. Sci.*, 63: 466-473.
- Dunn, J.R. 1984. Developmental osteology. In: Moser, H.G., W.J. Richards, D.M. Cohen, M.P. Fahay, A.W. Kendal and S.L. Richardson (eds.), *Ontogeny and systematics of fishes*. American Society of Ichthyologists and Herpetologists, Special publication. 1, Allen, Lawrence, KS, pp. 48-50.
- Fukuhara, O. 1992. Study on the development of functional morphology and behaviour of the larvae and juvenile *Limanda yokohamae* (Pisces: Pleuronectidae) reared in the laboratory. *Mar. Biol.*, 99: 271-281.
- Fukuhara, O. and T. Fushimi. 1988. Fin differentiation and squamation of artificially reared grouper, *Epinephelus akaara*. *Aquaculture*, 69: 379-386.
- Han, K.H., S.H. Oh, D.S. Hwang, Y.H. Cho and D.C. Seo. 2002. Egg development and morphological change of larvae of the brown croaker, *Miichthys miiuy*. *Korean J. Ichthyol.*, 14: 93-99.
- Hard, J.J., B.A. Berejikian, E.P. Tezak, S.L. Schroder, C.M. Knudsen and L.T. Parker. 2000. Evidence for morphometric differentiation of wild and captively reared adult coho salmon: a genometric analysis. *Environ. Biol. Fishes.*, 58: 61-73.
- Hughes, R.N., M.J. Kaiser, P.A. Mackney and K. Warburton. 1992. Optimizing foraging behavior through learning. *J. Fish Biol.*, 41: 77-91.
- Hunter, J.R. 1984. Synopsis of culture methods for marine fish larvae. In: Moser, H.G., W.J. Richards, D.M. Cohen, M.P. Fahay, A.W. Kendal and S.L. Richardson (eds.), *Ontogeny and systematics of fishes*, American Society of Ichthyologists and Herpetologists, Special pub. 1, Allen, Lawrence, KS, pp. 24-27.
- Kanazawa, A. 1993. Nutritional mechanism involved in the occurrence of abnormal pigmentation in hatchery-reared flatfish. *J. World Aquacul. Soc.*, 24: 162-166.
- Kim, J.-M. and M. Okiyama. 1989. Larval morphology and distribution of *Callanthias japonicus* (Franz) (Serranidae). *Ocean Res.*, 11: 1-7.
- Koumoundouros, G., P. Divanach and M. Kentouri. 2001. Osteological development of *Dentex dentex* (Osteichthyes: Sparidae): dorsal, anal, paired fins and squamation. *Mar. Biol.*, 138: 399-406.
- Mana, R.R. and G. Kawamura. 2002. A comparative study on morphological differences in the olfactory system of red sea bream (*Pagrus major*) and black sea bream (*Acanthopagrus schlegeli*) from wild and cultured stocks. *Aquaculture*, 209: 285-306.
- Moser, H.G. 1981. Morphological and functional aspects of marine fish larvae. In: Lasker, R. (ed.), *Marine Fish Larvae*, Washington Sea Grant Program, Washington University Press, Seattle, USA, pp. 99-131.
- Myung, J.G., Y.U. Kim, Y.J. Park, P.K. Kim, J.M. Kim and H.T. Huh. 2004. Embryonic development, larvae and juveniles of the small yellow croaker (*Larimichthys polyactis*) reared in aquarium. *J. Korean Fish. Soc.*, 37: 478-484.
- Park, E.H. and S.H. Lee. 1988. Scale growth and squamation chronology for the laboratory-reared hermaphroditic fish *Rivulus marmoratus* (Cyprinodontidae). *Japanese J. Ichthyol.* 34: 476-482.
- Park, I.-S., Y.J. Kim, H.J. Choi, S.Y. Oh, C.H. Noh and S.H. Lee. 2007. Total length estimation from head dimensions of artificially propagated brown croaker *Miichthys miiuy*. *Korean J. Ichthyol.*, 19: 128-131.
- Russell, F.S. 1976. *The eggs and planktonic stages of British marine fishes*. Academic Press, London, 524pp.
- Seo, D.C. 2004. Developmental ecology and early life growth of brown croaker *Miichthys miiuy*. Doctoral dissertation, Yosun National University, Yosun, Korea, pp. 1-49.
- White, D.S. 1977. Early development and pattern of scale formation in the spotted sucker, *Minytrema melanops* (Catostomidae). *Copeia*, 19: 400-403.

## 민어, *Miichthys miiuy*의 초기 형태 발달: 지느러미 분화, 두부 계측 및 비늘 도포

박인석 · 김영자<sup>1</sup> · 구인본 · 김동수<sup>2</sup>

한국해양대학교 해양환경·생명과학부, <sup>1</sup>한국환경산업기술원, <sup>2</sup>부경대학교 해양수산형질전환생물연구소

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**요 약** : 민어, *Miichthys miiuy* (Basilewsky)에서의 지느러미 분화, 두부 계측 및 비늘 도포 양상의 초기 형태학적 발달을 조사하였다. 부화 후 12일(전장 4.2 mm 미만)에, 어체 주위로 fin-fold의 존재가 관찰되었다. 등지느러미, 뒷지느러미 및 가슴지느러미의 기조 형성은 부화 후 12일에 거의 완전히 이루어진 반면, 지느러미 분절은 부화 후 26일과 29일 사이에 이루어졌다. 꼬리지느러미 중간 기조의 신장은 부화 후 32일에 시작되었으며 부화 후 37일에 현저하였다. 부화 후 68일에 꼬리지느러미가 뽀족해지기 시작하였다(전장 70.7 mm). 비늘의 발달은 전장이 9.1 mm(부화 후 28일)일 때 미병의 측선부위로부터 시작되어 결국 아가미덮개 전체를 도포하였다(전장 22.1 mm, 부화 후 44일). 두부 계측치들은 전장이 12 mm 이상(부화 후 30일)에서 거의 안정화되었다.

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**찾아보기 낱말** : 민어, 두부 계측, 비늘 도포, 지느러미 분화