# Physiological responses of *Fucus serratus* (Phaeophyceae) to high doses of cadmium exposure

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Growth, oxidative stress and antioxidant capacity of  $Fucus\ serratus$  exposed to high doses of Cd were examined. Two sites in Southwest England (Restronguet Point and Bantham Quay) were selected since they had different histories of metal contamination.  $1 \sim 10$  mg Cd L<sup>-1</sup> were treated to Aquil medium for up to 14 days. Similar levels of lipid peroxidation but different values of relative growth rates, cupric ion reducing antioxidant capacity (CUPRAC) and 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical scavenging capacity indicated that F. serratus has population-dependent antioxidant strategies. F. serratus demonstrated cadmium resistance with no visual symptoms for 14 days and the population from the polluted area seemed to have more powerful antioxidant strategies. However Fucus from the conserved area also showed protective antioxidative mechanism.

Key words: Fucus serratus, Cadmium, Antioxidant response, Restronguet Point, Bantham Quay

Metals are known to encourage various physiological responses in plants and algae, of which chlorosis, cell lysis, necrosis, discoloration, encystment and death can be counted as some of visible signs (Hu *et al.*, 1996, Okamoto *et al.*, 1999, Küpper *et al.*, 2002, Collén *et al.*, 2003). Besides, as intracellular symptoms of metal stress, oxidative damage of proteins, lipids and DNA was reported (Asada and Takahashi, 1987, Collén and Davison, 1999b, Halliwell and Gutteridge, 2007). Metals are also able to induce reactive oxygen species (ROS) (Collén and Davison, 1999a, Dummermuth *et al.*, 2003, Güçlü *et al.*, 2006). ROS are obligatory by-products of common aerobic metabolism, such as photosynthesis and respiration (Noctor and Foyer, 1998, Collén and

Davison, 1999a), however over-production of ROS and imbalance of cellular oxidative status can be fatal to organisms (Rijstenbil *et al.*, 1998, Okamoto *et al.*, 2001a, Okamoto *et al.*, 2001b, Collén *et al.*, 2003).

To reduce the harmful effects of metal exposure, plants and algae induces antioxidants and antioxidant enzymes (Van Assche and Clijsters, 1990, Pinto *et al.*, 2003). Contreras et al. (2005) reported the increased ROS levels in *Scytosiphon lomentaria* from mine wastes and the high levels of antioxidative enzymes against oxidative stress. However, our knowledge on antioxidant mechanism of macroalgae is relatively limited (Collén and Davison, 1999a, b, c) and most studies were focused on terrestrial higher plants to date (Ratkevicius et al., 2003).

Various macroalgae have been used to investigate their antioxidant capacity and their absorption/adsorption ability. Hashim and Chu (2004) reported that brown algae

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showed the highest Langmuir maximum adsorption ability among seven different species of brown (Sargassum siliquosum, S. baccularia and Padina tetrastomatica). green (Chaetomorpha linum) and red algae (Gracilaria changii, G. edulis and G. salicornia). This enhanced performance of brown algae is based on their basic structure and biochemical constitution, which is closely linked to the external/internal constituents of cell wall and antioxidant capacity (Hu et al., 1996, Davis et al., 2003). Although Fucus and Sargassum are known as bioindicators, F. serratus is not often used compared to other Fucus species (Förstner and Wittmann, 1983. Bryan et al., 1985). Since F. serratus thrives in both of polluted and conserved areas in the UK, the investigation of antioxidant capacity of F. serrutus can reveal a relationship between metal contamination and abundant species.

Therefore, the aims of this study were to examine the oxidative stress and antioxidant capacities of F. serratus collected from different populations exposed at different levels of cadmium in seawater.

## Material and methods

Fucus serratus was collected from two sites with different history of metal contamination (Fig. 1). Bantham Quay (BQ; 2667100E, 43795N), at the mouth of the

river Avon, has been known as one of the least polluted coastal areas in the UK in which 18 µg g<sup>-1</sup> of total Cu was measured in sediment (Table 1). On the contrary, Restronguet Point (RP; 181500E, 37435N), the entrance to Restronguet Creek, is a part of the most metal-contaminated area in the Fal Estuary (Pirrie et al., 2003). In this estuary, hard rock mining had prevailed from the 13<sup>th</sup> century (Gerrard, 2000), and drainage from the old mine adits and erosion of the spoil heaps by water continues (Bryan and Gibbs, 1983, Pirrie *et al.*, 2003).

Culture conditions were maintained for 14 days at 15 °C, 250 µmol photons m<sup>-2</sup> s<sup>-1</sup> photosynthetically active radiation (PAR) and 12 : 12 h light : dark cycle. Light was controlled by cool white fluorescent lamps with a time controller. Culture tanks containing artificial culture medium, Aquil, were mixed and aerated by shakers (60 rpm) for 24 h.

Relative growth rates (RGRs) of fronds were calculated using the following equation based on fresh weight (FW, g). Pieces of *Fucus* thalli were washed with filtered seawater to remove mud/sand and epiphytes were blotted dry and weighted. RGRs were expressed as percentage (Hunt, 1982).

$$RGR(\%d^{-1}) = \frac{(\ln(m_f) - \ln(m_i))}{t} \times 100$$

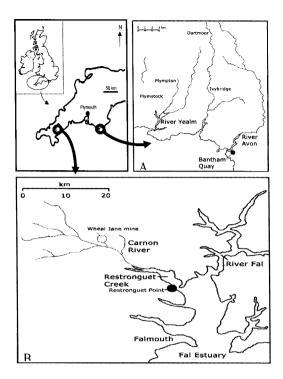


Fig. 1. Two target sites in which *Fucus serratus* was collected. A, Bantham Quay; B, Restronguet Point, South West England.

where ' $m_i$ ' and ' $m_i$ ' are masses of the fronds (g FW) on the final/initial day of measurement and 't' is time in days between measurements.

Lipid peroxidaton using 10% trichloroacetic acid (TCA) and 0.6% thiobarbituric acid (TBA) was followed by Lee (2009). The concentration of malondialdehyde (MDA) was determined spectrophotometrically (Unicam He $\lambda$ ios  $\beta$  UV-VIS spectrophotometer, Spectronic Unicam, Cambridge, U.K.) and calculated by the following equation:

MDA-TBA (mmol 
$$mL^{-1}$$
 of extract) =  $\Delta A / \varepsilon$ 

Here,  $\Delta A$  is the difference between absorbance at 532 nm and absorbance at 600 nm (A532 - A600)

Table 1. Concentrations of metals in sediments and water column of sampling area

	Water (µg L-1)	Sediment (µg g-1 D.W.)	
Metals	Restronguet Creek	Restronguet Creek	Avon Estuary
Ag		3.76	0.06
As		1740	13.0
Cd	<0.1-38	1.53	0.08
Co		21	10
Cr		32	28
Cu	2-176	2398	18
Fe		49071	18361
Hg		0.46	0.12
Mn	3 - 1513	485	326
Ni	1 - 18	58	23
Pb	<2 - 4	341	68
Sn		55.9	3.9
Zn	7 - 22460	2821	82
Reference	Bryan and Gibbs (1983)	Bryan and Langston (1992)	

and  $\varepsilon$  is the extinction coefficient, 159,200 mM<sup>-1</sup> cm<sup>-1</sup>. The value of mmol mL<sup>-1</sup> was then expressed as  $\mu$ mol g<sup>-1</sup> D.W. Three replicates were used for each treatment.

For cupric ion reducing antioxidant capacity (CUPRAC) and 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical scavenging capacity analysis, extraction of polyphenols was preceded. Harvested thalli were thoroughly rinsed with running tap water to remove salt and remaining Cd on the surface of thalli and were

ground to a powder using liquid nitrogen. Ground F. serratus (approximately 0.1 g) was extracted with 10 mL 70 : 30 MtOH :  $H_2O$  at 25  $^{\circ}C$ , in the dark, in a shaking incubator (60 rpm) for 24 hr, followed by centrifugal at 6,500 g for 10 min (25  $^{\circ}C$ ). These polyphenol extracts were stored for the following analyses for antioxidant capacity.

CUPRAC of *F. serratus* was examined according to Apak et al. (2007). Total antioxidant capacity of *F. serratus* was expressed as trolox equivalent antioxidant capacity (TEAC).

Capacity (in mmol TE 
$$g^{-1}$$
) =  $(A_f / \mathcal{E}_{TR})$  ( $V_f / V_s$ )  $r$  ( $V_{cup} / m$ )

where  $A_f$  is the absorbance,  $\varepsilon_{TR}$  is the molar absorptivity of Trolox (1.67 x 10<sup>4</sup> L mol<sup>-1</sup> cm<sup>-1</sup>),  $V_f$  is the final volume of mixture,  $V_s$  is the sample volume taken for analysis from the diluted extract, r is the dilution rates,  $V_{cup}$  is the initial volume of *Fucus* extract and m is the fresh weight (g) of algal material.

Free radical scavenging activity of Cd-exposed *F. serratus* was assessed by the DPPH free-radical method (Connan et al., 2006). Lower absorbance of the mixture indicates a higher free radical scavenging activity (Oztürka et al., 2007). The DPPH free radical scavenging capacity was calculated by the following equation (Gülçin *et al.*, 2003, Oztürka *et al.*, 2007):

DPPH Scavenging Effect (%) = 
$$\frac{A_{control} - A_{sample}}{A_{control}} \times 100$$
 ,

where  $A_{control}$  is the absorbance of control and  $A_{sample}$  is the absorbance in the presence of the sample

of F. serratus.

Data were analysed using the statistical package SPSS version 16.0 for Windows (SPSS Inc.). Before all parametric tests, the data were tested for homogeneity of variance and normality (Sokal and Rohlf, 1995). Multivariate test of General Linear Model (GLM) was used for analyzing the effects of locality, metal concentration, exposure time and their interactions, and Tukey HSD was used for the *Post Hoc* multiple comparisons.

#### Results

The effect of high concentrations of Cd ( $1 \sim 10 \text{ mg}$  L<sup>-1</sup>) on RGRs of *Fucus serratus* was measured on the basis of weight change for  $7 \sim 14$  d exposure (Fig. 2). In the RP population, RGRs of *F. serratus* decreased for the first 7 d in all treatments except 0  $\mu$ g Cd L<sup>-1</sup>. After 14 d of Cd exposure, recovery of weight was shown as increased RGRs in RP. 5 and 10 mg Cd L<sup>-1</sup> exposure increased RGRs of *F. serratus* after 14 d even though they were lower than RGRs of 0 mg L<sup>-1</sup>. 1 mg L<sup>-1</sup> showed the last recovery of growth in RP.

The BQ population also showed drastic decrease of RGRs for the first 7 d. RGRs of BQ have also decreased with increasing Cd concentrations however 1 mg  $L^{-1}$  showed the similar decrease to 10 mg  $L^{-1}$  (p > 0.05). After 14 d, RGRs were still lower than 0 except for 0 mg  $L^{-1}$  All Cd treatments above 1 mg  $L^{-1}$  demonstrated similar RGRs in BQ (p > 0.05). For 7 and 14 d, all RGR values from Cd-exposed BQ materials were significantly lower than RGRs of the RP population (p < 0.0001).

Lipid peroxidation of *Fucus* thalli from two natural sites was evaluated without additional metal exposure (Fig. 3). *F. serratus* from RP showed lower thiobarbituric acid-reactive substance (TBARS) values than *F. serratus* from BQ, but statistical differences were not significant (p > 0.05).

When high doses of Cd ( $1 \sim 10$  mg Cd L<sup>-1</sup>) were treated, lipid peroxidation of cell membrane occurred by time of exposure and Cd concentrations (p < 0.0001, respectively) but not by locality (p > 0.05, Fig. 4). The longer exposure of Cd showed the higher TBARS values (7 d < 14 d). The higher Cd concentrations (5 and 10 mg Cd L<sup>-1</sup>) induced the higher TBARS values in both populations at 14 d.

Total antioxidant capacity of Cd treated Fucus was studied by reducing a cupric ion (Fig. 5). When F. serratus was exposed to Cd, the BQ population showed significantly higher levels of trolox equivalent capacity (mmol TE DW-1) than the RP population at each day (p < 0.0001). Time of Cd exposure and Cd concentration significantly affected antioxidant capacity of F. serratus from RP (p < 0.0001 and p = 0.001, respectively). However any clear pattern of trolox equivalent capacity was not discovered according to Cd concentrations at 1 and 7 d. Meanwhile, at 14 d, Cd treated materials revealed higher values than 0 mg Cd  $L^{-1}$  (p = 0.002). On the other hand, time of exposure and Cd concentration revealed a significant effect on CUPRAC values of the BQ population (p < 0.0001, respectively). The capacity was not significant between Cd concentrations (1~10 mg Cd L-1) at 1 d and the control materials showed high values at each day (Fig. 5).

Free radical scavenging capacity of F. serratus was

evaluated using DPPH (Fig. 6). Locality and time of exposure revealed a significant effect on DPPH free radical scavenging capacity (p < 0.0001, respectively), however Cd concentration did not have a noticeable effect (p > 0.05). Same as CUPRAC, the BQ population showed much higher values than the RP population.

Effects of exposure time and Cd concentration on DPPH free radical scavenging ability were different at each population. In RP, free radical scavenging ability did not show a regular trend with Cd concentrations however Cd treated materials had significantly higher capacities than the control at 14 d (p = 0.001). In BQ, the longer time of Cd exposure showed the lower scavenging capacity for free radical (p < 0.0001). However, concentration of treated Cd did not have an apparent tendency for free radical scavenging capacity even though there were significant differences (p = 0.006).

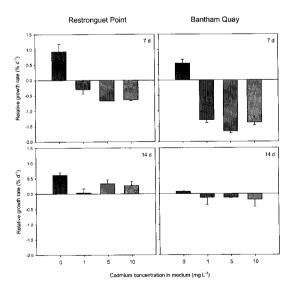


Fig. 2. Relative growth rates of *Fucus serratus* from Restronguet Point and Bantham Quay exposed to cadmium for 7 and 14 days. Values were expressed by means and standard deviations (n = 3).

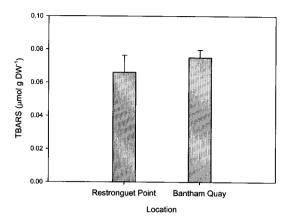


Fig. 3. Thiobarbituric acid-reactive substance (TBARS) levels, mainly malondialdehyde, in *Fucus serratus* harvested from Restronguet Point and Bantham Quay. Values represent mean values of three independent replicates  $\pm$  standard deviations.

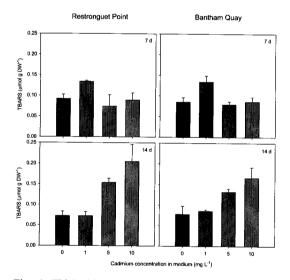


Fig. 4. Thiobarbituric acid-reactive substance (TBARS) levels, mainly malondialdehyde, in *Fucus serratus* from Restronguet Point and Bantham Quay which were exposed to cadmium for 7 and 14 days. Values represent mean values of three to six independent replicates ± standard deviations.

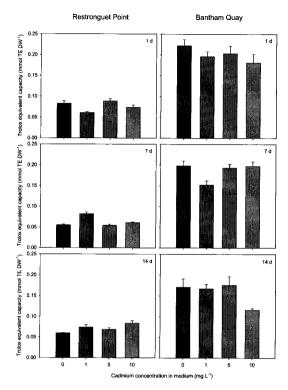


Fig. 5. Cupric ion reducing antioxidant capacity (CUPRAC) in Fucus serratus from Restronguet Point and Bantham Quay which were exposed to cadmium for 1, 7 and 14 days. Values represent mean values of three to six independent replicates  $\pm$  standard deviations.

## Discussion

Brown macroalga *F. serratus* was cultured with high doses of Cd in Aquil medium for up to 14 d. *Fucus* species has been known as bioindicators since they accumulated metals with a positive linear correlation with concentrations (Förstner and Wittmann, 1983, Bryan *et al.*, 1985). Although no visible symptoms of metal stress (e.g. discolouration, dark spot or death) were recognized for 14 d, growth of *F. serratus* was significantly affected by Cd treatment. Both populations showed drastic decreases of RGRs at 7 d, however the decrease of RGRs in the BQ population was more

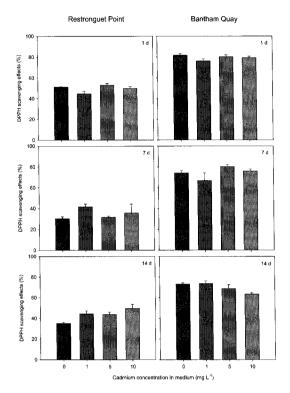


Fig. 6. 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical scavenging activity in *Fucus serratus* from Restronguet Point and Bantham Quay which were exposed to cadmium for 7 and 14 days. Values represent mean values of three to six independent replicates  $\pm$  standard deviations.

significant. RGRs of both populations were partially recovered after 14 d, but RGRs of BQ still showed negative growth compared with the initial states. It is not the first report of shrunken size and weight loss (or stable weight) of macroalgae after metal treatment (Bryan and Gibbs, 1983, Brown and Newman, 2003, Han *et al.*, 2008). Metal-induced stress, such as inhibition of cell division and/or expansion, may be connected to a decrease of cell turgor, and alteration of cell wall elasticity or acclimatization (Stauber and Florence, 1987, Brown and Newman, 2003).

More significant decrease and less recovery of RGRs in the BQ population showed more serious effects on

weight growth by Cd, which might be resulted from the different resistance ability due to the population or adaptation to their habitats. Some researchers have reported the inheritance of metal tolerance of marine macroalgae (Bryan and Gibbs, 1983, Correa *et al.*, 1996, Nielsen, 2002, Nielsen *et al.*, 2003). *F. serratus* from polluted areas revealed higher RGRs and metal resistance than *F. serratus* from unpolluted areas (Bryan and Gibbs, 1983, Nielsen *et al.*, 2003).

Polyunsaturated lipids are essential components of cell membranes, endoplasmic reticula and mitochondria (Muriel, 1997). Consequently lipid peroxidation by metal stress can be crucial to cellular function and survival. Although the TBARS test is non-specific and HPLC techniques may be more accurate, it has widely used for determination of oxidative stress in terrestrial plants, micro- and macroalgae as one of the most frequently operated tests (Halliwell and Gutteridge, 2007, Lee, 2009). Before measuring oxidative stress in Fucus from Cd-treated medium, contents of TBARS, mostly malondialdehyde, of two natural populations from RP and BO were measured. Since the RP area is significantly polluted by metals, including Cd (Table 1), two populations showed similar values in lipid peroxidation. It implies the RP population has the stronger detoxifying ability, less stress from metal exposure and/or adaptation. This indicates they may have more efficient antioxidant protective systems than the BO population.

Increases of TBARS levels with Cd treatment at 14 d suggested Cd-induced oxidative damage in the present study. Even though time of exposure and Cd concentration had clear relations with TBARS values, especially at

14 d, both populations did not reveal a difference by locality. Similar values of TBARS from the two populations may include some causes. 1) Similar lipid peroxidation due to over-burden of Cd stress, regardless of their history of metal exposure. Indeed 1~10 mg Cd L-1 for 7 and 14 d is a toxic situation that the alga may not experience at all in the natural environment. 2) Population-dependent antioxidant response. Sensitive populations revealed higher lipid peroxidation than resistant populations (Randhawa et al., 2001), which were attributed to their different antioxidative defence systems. In this research, the RP thalli still contained significantly higher concentrations of metals that were absorbed from the natural seawater before the experiment (data not shown). Nevertheless the RP population represented similar TBARS values to the BQ population, and this implies the higher and powerful antioxidant mechanism of the polluted population. 3) Species-specific stress-resistant mechanism. Similar TBARS values may not describe the higher antioxidant capacity of the RP population or the over-burden of Cd stress. Fucus spp. are well-known as a stress-tolerant species (Jiménez-Escrig et al., 2001). Therefore, F. serratus from BO may also have strong potential against oxidative stress and showed sustained high levels of antioxidant activities (Lee, 2009).

Antioxidant activities can be evaluated by many different ways. Determination of antioxidant potential with different assays would give more informative and reliable results (Oztürka et al., 2007). Consequently CUPRAC assay was processed with DPPH free radical scavenging capacity assay in this study.

The BQ population showed significantly higher antioxidant capacity in CUPRAC assay, however the values

were not likely related to the Cd exposure in the research. Highly sustained values and no clear dose-dependent pattern of the BQ population (including controls) represent a loose relationship between CUPRAC and Cd treatment. However, differences between the two populations were evident that the two populations were in significantly different status.

DPPH free radical scavenging activity represented similar patterns with CUPRAC assay in the current study. The BQ population showed stable antioxidant activities regardless of Cd doses and time of exposure than the RP population. This indicates the BQ population has more stable activities to defense oxidative stress. Since *F. serratus* exposed to lower and shorter Cd showed dose-related activities in our previous study (manuscript in preparation), less relationship with high dose of Cd may be due to the over-limit of the algal tolerance on Cd or other antioxidant activities, such as antioxidative enzymes or antioxidants.

Brown algae were reported to have excellent radical scavenging capacities compared with other algal groups (Matsukawa et al., 1997, Yan et al., 1998). The free radical scavenging activity is known to be closely related to phenolic compounds which are abundant in brown algae (Jiménez-Escrig et al., 2001, Connan et al., 2006, Oztürka et al., 2007). F. vesiculosus was also reported to have a strong radical scavenging activity due to the high contents of polyphenol (Jiménez-Escrig et al., 2001). Therefore F. serratus in this study may also contain high antioxidant potentials due to the high polyphenol contents, which was represented by high and stable levels of free radical scavenging activities. In the mean while, the lower values of the RP population represent population-specific strategy for oxidative stress.

The RP population showed the lower free radical scavenging ability and higher RGRs with the same Cd exposure and the much higher other metal contents from the natural habitat. It can be regarded that the RP population may possess other efficient antioxidant strategies (manuscript in preparation).

In conclusion, *F. serratus* had strong Cd resistance and represented the population-specific antioxidant capacities. The BQ population from the reference area also possessed high antioxidant ability and the RP population from the contaminated area seems to have higher and stronger ability against metal stress.

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#### References

- Apak, R., Güçlü, K., Demirata, B., Özyürek, M., Çelik, S.E., Bektaşoğlu, B., Berker, K.I. and Özyurt, D.: Comparative evaluation of various total antioxidant capacity assays applied to phenolic compounds with the CUPRAC assay. Molecules, 12: 1496-1547, 2007.
- Asada, K. and Takahashi, M.: Production and scavenging of active oxygen species in photosynthesis. In Photoinhibition, Kyle, D.J., Osmond, C. and Amtzen, C. (eds.), Elsevier Science Publishers, New York, pp. 227-287, 1987.
- Brown, M.T. and Newman, J.E.: Physiological responses

- of *Gracilariopsis longissima* (S.G. Gmelin) Steentoft, L.M. Irvine and Farnham (Rhodophyceae) to sub-lethal copper concentrations. Aquatic Toxicology, 64: 201-213, 2003.
- Bryan, G.W. and Gibbs, P.E.: Heavy metals in the Fal estuary,

  Cornwall: a study of long-term contamination by

  mining waste and its effects on estuarine organisms.

  Occasional Publication of Marine Biological

  Association of the United Kingdom, 2: 1-112, 1983.
- Bryan, G.W. and Langston, W.J.: Bioavailability, accumulation and effects of heavy metals in sediments with special reference to United Kingdom estuaries: a review. Environmental Pollution, 76: 89-131, 1992.
- Bryan, G.W., Langston, W.J., Hummerstone, L.G. and Burt, G.R.: A guide to the assessment of heavy metal contamination in estuaries using biological indicators. Marine Biological Association of the United Kingdom, 4: 92, 1985.
- Collén, J. and Davison, I.R.: Reactive oxygen metabolism in intertidal *Fucus* spp. (Phaeophyceae). Journal of Phycology, 35: 62-69, 1999a.
- Collén, J. and Davison, I.R.: Reactive oxygen production and damage in intertidal *Fucus* spp. (Phaeophyceae).

  Journal of Phycology, 35: 54-61, 1999b.
- Collén, J. and Davison, I.R.: Stress tolerance and reactive oxygen metabolism in the intertidal red seaweeds *Mastocarpus stellatus* and *Chondrus crispus*.

  Plant, Cell & Environment, 22: 1143-1151, 1999c.
- Collén, J., Pinto, E., Pedersen, M. and Colepicolo, P.: Induction of oxidative stress in the red macroalga *Gracilaria tenuistipitata* by pollutant metals. Archives of Environmental Contamination and Toxicology, 45: 337-342, 2003.

- Connan, S., Delisle, F., Deslandes, E. and Gall, E.A.:
  Intra-thallus phlorotannin content and antioxidant
  activity in Phaeophyceae of temperate waters.
  Botanica Marina, 49: 39-46, 2006.
- Contreras, L., Moenne, A. and Correa, J.A.: Antioxidant responses in *Scytosiphon lomentaria* (Phaeophyceae) inhabiting copper-enriched coastal environments.

  Journal of Phycology, 41: 1184-1195, 2005.
- Correa, J.A., González, P., Sánchez, P., Muñoz, J. and Orellana, M.C.: Copper-algae interactions: inheritance or adaptation? Environmental Monitoring and Assessment, 40: 41-54, 1996.
- Davis, T.A., Volesky, B. and Mucci, A.: A review of the biochemistry of heavy metal biosorption by brown algae. Water Research, 37: 4311-4330, 2003.
- Dummermuth, A.L., Karsten, U., Fisch, K.M., Konig, G.M. and Wiencke, C.: Responses of marine macroalgae to hydrogen-peroxide stress. Journal of Experimental Marine Biology and Ecology, 289: 103-121, 2003.
- Förstner, U. and Wittmann, G.T.W.: Metal Pollution in the Aquatic Environment, Springer-Verlag, Berlin, Germany, 1983.
- Güçlü, K., Altun, M., Özyürek, M., Karademir, S.E. and Apak, R.: Antioxidant capacity of fresh, sun- and sulphited-dried Malatya apricot (*Prunus armeniaca*) assayed by CUPRAC, ABTS/TEAC and folin methods. International Journal of Food Science & Technology, 41: 76-85, 2006.
- Gülçin, İ., Oktay, M., Kireçci, E. and Küfrevioğlu, Ö.İ.:

  Screening of antioxidant and antimicrobial activities of anise (*Pimpinella anisum* L.) seed extracts. Food Chemistry, 83: 371-382, 2003.
- Gerrard, S.: The Early British Tin Industry, Tempus

- Publishing Ltd, Stroud, 2000.
- Halliwell, B. and Gutteridge, J.M.C.: Free radicals in biology and medicine, Oxford University Press, Oxford, 2007.
- Han, T., Kang, S.-H., Park, J.-S., Lee, H.-K. and Brown, M.T.: Physiological responses of *Ulva pertusa* and *U. armoricana* to copper exposure. Aquatic Toxicology, 86: 176-184, 2008.
- Hashim, M.A. and Chu, K.H.: Biosorption of cadmium by brown, green, and red seaweeds. Chemical Engineering Journal, 97: 249-255, 2004.
- Hu, S., Tang, C.H. and Wu, M.: Cadmium accumulation by several seaweeds. Science of The Total Environment, 187: 65-71, 1996.
- Hunt, R.: Plant Growth Curves, Edward Arnold Publisher Ltd., London, U.K., 1982.
- Jiménez-Escrig, A., Jiménez-Jiménez, I., Pulido, R. and Saura-Calixto, F.: Antioxidant activity of fresh and processed edible seaweeds. Journal of the Science of Food and Agriculture, 81: 530-534(5), 2001.
- Küpper, H., Šetlík, I., Spiller, M., Küpper, F.C. and Prášil, O.: Heavy metal-induced inhibition of photosynthesis: targets of *in vivo* heavy metal chlorophyll formation. Journal of Phycology, 38: 429-441, 2002.
- Lee, S.J.: Physiological and Biochemical Responses to Cadmium Exposure in *Fucus serratus* (Phaeophyceae). School of Biological Sciences. University of Plymouth, Plymouth, UK, 2009.
- Matsukawa, R., Dubinsky, Z., Kishimoto, E., Masaki, K., Masuda, Y., Takeuchi, T., Chihara, M., Yamamoto, Y., Niki, E. and Karube, I.: A comparison of screening methods for antioxidant activity in seaweeds. Journal of Applied Phycology, 9: 29-35, 1997.
- Muriel, P.: Peroxidation of lipids and liver damage. In

- Oxidants, Antioxidants, and Free Radicals, 237-, Baskin, S.I.andSalem, H. (eds.), Taylor & Francis Ltd., London, UK, 1997.
- Nielsen, H.D.: Copper toxicity in the physiology and early development of *Fucus serratus*. Department of Biological Sciences. University of Plymouth, Plymouth, UK, 2002.
- Nielsen, H.D., Brownlee, C., Coelho, S.M. and Brown, M.T.: Inter-population differences in inherited copper tolerance involve photosynthetic adaptation and exclusion mechanisms in *Fucus serratus*. New Phytologist, 160: 157-165, 2003.
- Noctor, G. and Foyer, C.H.: Ascorbate and glutathione: keeping active oxygen under control. Annual Review of Plant Physiology and Plant Molecular Biology, 49: 249-279, 1998.
- Okamoto, O.K., Pinto, E., Latorre, L.R., Bechara, E.J.H. and Colepicolo, P.: Antioxidant modulation in response to metal-induced oxidative stress in algal chloroplasts. Archives of Environmental Contamination and Toxicology, 40: 18-24, 2001a.
- Okamoto, O.K., Robertson, D.L., Fagan, T.F., Hastings, J.W. and Colepicolo, P.: Different regulatory mechanisms modulate the expression of a dinoflagellate iron-superoxide dismutase. Journal of Biologica Chemistry, 276: 19989-19993, 2001b.
- Okamoto, O.K., Shao, L., Hastings, J.W. and Colepicolo, P.: Acute and chronic effects of toxic metals on viability, encystment and bioluminescence in the dinoflagellate *Gonyaulax polyedra*. Comparative Biochemistry and Physiology Part C: Pharmacology, Toxicology and Endocrinology, 123: 75-83, 1999.

- Oztürka, M., Aydoğmuş-Öztürkb, F., Duru, M.E. and Topçud, G.: Antioxidant activity of stem and root extracts of Rhubarb (*Rheum ribes*): An edible medicinal plant. Food Chemistry, 103: 623-630, 2007.
- Pinto, E., Sigaud-kutner, T.C.S., Leitão, M.A.S., Okamoto, O.K., Morse, D. and Colepicolo, P.: Heavy metal-induced oxidative stress in algae. Journal of Phycology, 39: 1008-1018, 2003.
- Pirrie, D., Power, M.R., Rollinson, G., Camm, G.S., Hughes, S.H., Butcher, A.R. and Hughes, P.: The spatial distribution and source of arsenic, copper, tin and zinc within the surface sediments of the Fal Estuary, Cornwall, UK. Sedimentology, 50: 579-595, 2003.
- Randhawa, V.K., Zhou, F., Jin, X., Nalewajko, C. and Kushner, D.J.: Role of oxidative stress and thiol antioxidant enzymes in nickel toxicity and resistance in strains of the green alga Scenedesmus acutus f. alternans. Canadian Journal of Microbiology, 47: 987-993, 2001.
- Ratkevicius, N., Correa, J.A. and Moenne, A.: Copper accumulation, synthesis of ascorbate and activation of ascorbate peroxidase in *Enteromorpha compressa* (L.) Grev. (Chlorophyta) from heavy metal-enriched environments in northern Chile. Plant, Cell & Environment, 26: 1599-1608, 2003.
- Rijstenbil, J.W., Haritonidis, S., Malea, P., Seferlis, M. and Wijnholds, J.A.: Thiol pools and glutathione redox ratios as possible indicators of copper toxicity in the green macroalgae *Enteromorpha* spp. from the Scheldt Estuary (SW Netherlands, Belgium) and Thermaikos Gulf (Greece, N Aegean Sea). Hydrobiologia, 385: 171-181, 1998.
- Sokal, R.R. and Rohlf, F.J.: Biometry: the principles and practice of statistics in biological research, Freeman,

New York, Oxford, 1995.

Stauber, J.L. and Florence, T.M.: Mechanism of toxicity of ionic copper and copper complexes to algae.

Marine Biology, 94: 511-519, 1987.

Van Assche, F. and Clijsters, H.: Effects of metals on enzyme activity in plants. Plant, Cell and Environment, 13: 195-206, 1990.

Yan, X., Nagata, T. and Fan, X.: Antioxidative activities

in some common seaweeds. Plant Foods for Human Nutrition (Formerly Qualitas Plantarum), 52: 253-262, 1998.

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