

## Characterization of Crop Residue-Derived Biochars Produced by Field Scale Biomass Pyrolyzer

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Application of biochar to soils is proposed as a significant, long-term, sink for atmospheric carbon dioxide in terrestrial ecosystems. In addition to reducing emissions and increasing the sequestration of carbon, production of biochar and its application to soils will contribute improve soil quality and crop productivity. Objectives were i) to evaluate biochar productivity from crop residues using a low-cost field scale mobile pyrolyzer and ii) to evaluate characteristics of feedstocks and biochars from locally collected crop residues. Pyrolysis experiments were performed in a reactor operated at 400-500°C for 3-4 hours using biomass samples of post-harvest residues of corn (*Zea mays* L.), cotton (*Gossypium* spp.), rice (*Oryza sativa* L.), sorghum (*Sorghum bicolor* L.) and wheat (*Triticum aestivum* L.). Feedstocks differed, but average conversion to biochar was 23%. Carbon content of biomass feedstock and biochar samples were 445 g kg<sup>-1</sup> and 597 g kg<sup>-1</sup>, respectively. Total carbon content of biochar samples was 34% higher than its feedstock samples. Significant increases were found in P, K, Ca, Mg, and micro-nutrients contents between feedstock and biochar samples. Biochar from corn stems and rice hulls can sequester by 60% and 49% of the initial carbon input into biochar respectively when biochar is incorporated into the soils. Pyrolysis conversion of corn and rice residues sequestered significant amounts of carbon as biochar which has further environmental and production benefits when applied to soils. Field experiment with crop residue biochar will be investigated the stability of biochars to show long-term carbon sequestration and environmental influences to the cropping systems.

**Key words:** Biochar, Pyrolysis, Carbon sequestration, Crop residue

### Introduction

Judicious management of crop residue is important not only to sustain crop production but also to reduce carbon emissions from crop production systems. Burning of crop residue leads to emission of CO<sub>2</sub> and other greenhouse gases (GHGs) (Zhang et al., 2008). Conversely, crop residue can be composted and recycled as a major nutrient source. And direct return of crop residue to the soil can reduce chemical fertilizer use through improving nutrient use efficiency (Malhi et al., 2006; Teasdale et al., 2008). Use of compost and manure also improves the soil organic carbon (SOC) pool and enhances crop yield. Carbon sequestration in soil is a technique for long-term storage of carbon and has been proposed to mitigate global warming (Baldock and Smernik, 2002; Lal, 2004). Carbon sequestration

can be taken a step further by pyrolysis, the burning of plant biomass in a limited O<sub>2</sub> environment to form biochar (Baldock and Smernik, 2002; Lehmann, 2007a).

Biochar resists microbial decomposition in the soil for a much longer time than regular biomass (Lehmann, 2009) to provide environmental benefits. Large amounts of biochar carbon may be sequestered in the soil for hundreds to thousands of years, but few studies have quantified or estimated the carbon sequestration as a result of biochar application (McHenry, 2009). Biochar application into soils lowered greenhouse gas emission by 12-84% compared with its direct combustion for energy purposes (Hansen et al., 2008).

Studies also show crop yields can increase with application of biochar as a soil amendment (DeLuca et al., 2009). Biochar improves soil fertility due to its strong exchange capacity to retain nutrients for plant uptake. This capacity also acts as an environmental buffer in which the biochar retains large quantities of nutrients and pesticides to reduce their losses by infiltration into ground water and runoff into surface

waters (Van Zwieten et al., 2010). Crop residue biochar can be produced by pyrolysis, a thermal decomposition process used widely for converting solid fossil fuels into liquid and gaseous hydrocarbons and a solid char residue. Coal pyrolysis has been widely studied (Howard, 1981; Liang et al., 2007; Teng et al., 1997), but few investigations of crop biomass pyrolysis have been reported. Biomass carbon from crop residues is composed of cellulose, hemicellulose and lignin (Demirba, 2001; Goldstein and Company, 1981; McKendry, 2002).

Biomass pyrolysis to biochar by thermal decomposition of organic matter results in the production of char, liquid and gaseous products (Demirba, 2001). The process can sequester up to 50% of the initial carbon input that can be returned to the soil (Lehmann et al., 2006). Pyrolysis of biomass was evaluated for recovering a bio-oil (Mohan et al., 2006; Oasmaa et al., 2009). Lignin decomposes over a wider temperature range compared to cellulose and hemicelluloses which rapidly degrade over narrower temperature ranges. Lignin gives higher yields of charcoal and tar from biomass (Alves et al., 2006).

Historically, evidence for soil benefits from highly reduced carbon compounds have been known since the discovery of a black colored soil in the Amazon basin of Brazil termed *Terra Preta*. The charcoal in these soils was added by native Amazonians to create arable farmland (Glaser et al., 2000; Lehmann et al., 2006). *Terra Preta* soils contain higher levels of P and Ca (Glaser, 2007). They also have high soil organic matter (SOM) content and cation exchange capacity (Cunha et al., 2009), which can contribute to improve soil fertility. Carbon enhanced SOM offers other direct values through improved water infiltration, water holding capacity, structural stability, cation exchange capacity and soil biological activity (Lehmann, 2007b).

Several laboratory scale pyrolysis studies have been reported. However there are few practical field-scale studies involving biomass composition, small-scale pyrolysis and evaluation of land application of biochar in a somewhat closed system. Such a system may be easily adaptable to local producers for their field scale application. Therefore, objectives of this research were i) to evaluate biochar productivity from crop residues using a low-cost mobile pyrolyzer and ii) to evaluate characteristics of feedstocks and biochars from locally grown crop residues. In subsequent studies field testing of the biochar will be conducted to evaluate the long-

term carbon sequestration and soil fertilization.

## Methods and Materials

A low-cost field scale biomass pyrolyzer was constructed at University of Missouri-Delta Research Center and consisted of a steel cylinder with an inside diameter of 0.65 m, height of 1 m and capacity of 0.33 m<sup>3</sup> (Figure 1). Air-flow inlet and exhaust pipes were installed on the bottom and top of the pyrolyzer cylinder through which air flow is controlled manually during pyrolysis. A fine-mesh screen was installed 15 cm above the bottom of the reactor cylinder. Temperature during the pyrolysis process was monitored with a thermometer probe in the middle of the reactor. Crop residues were added to occupy about 80 to 85% of reactor capacity. Charcoal lighter fluid, a volatile mixture of phenol distillates, was sprayed onto the biomass surface to accelerate ignition and to nurture uniform burning of crop residue. Soon after ignition, the steel lid was closed with bolts to provide an oxygen limited reaction. Temperature was maintained at 400-500°C for 3-4 hours. When finished, the lid was removed and water was sprayed on the biochar to prevent burn down. Three separate batches of each feedstock (replications) were pyrolyzed, after which the biochar samples were air-dried.

Residue from corn (*Zea mays* L.), cotton (*Gossypium* spp.), rice (*Oryza sativa* L.), grain sorghum (*Sorghum bicolor* L.) and wheat (*Triticum aestivum* L.) were used as biomass samples. Samples were obtained during 2010 at the University of Missouri-Delta Research Center research farm near Portageville, MO. Corn, cotton, grain sorghum were grown with ground water irrigation and conventional management practices recommended by the University of Missouri. The Sharkey soils of the research farm are on flood plains and low terraces of the Mississippi River; MLRA 131A. They are classified as very-fine, smectitic, thermic Chromic Epiaquerts (Soil Survey Staff, 1997). Crop residue samples were obtained after grain or cotton harvest and consisted of the whole plant except roots and grain/or bolls. Crop residues were air dried, chopped and sieved manually to give a particle size of 0.4-10 mm. Wheat straw was collected at the local farm after grain harvest in May 2010. Rice residue (i.e., hulls) was collected from local rice mill (i.e., Riceland food, Inc., New Madrid, MO), and used

**Table 1. Moisture content of feedstock and pyrolysis results from biomass samples. Standard deviations are in parentheses (n=3).**

Biomass sample	Moisture content of feedstock	Feedstock input	Biochar output	Ash content of biochar	Biochar yield
	----- % -----	----- kg -----	----- % -----	----- % -----	----- % -----
Corn stems	26.2 (0.46)	11.4 (2.7)	4.1 (1.0)	22.9 (0.7)	36.9 (0.89)
Cotton stems	13.7 (1.19)	13.1 (1.6)	2.2 (0.5)	13.6 (2.0)	17.0 (0.32)
Rice hulls	12.0 (0.42)	26.9 (1.5)	9.9 (0.5)	37.2 (2.5)	36.8 (0.36)
Sorghum stems	37.8 (0.89)	30.9 (5.1)	4.4 (1.2)	16.4 (1.8)	14.5 (0.48)
Wheat straw	4.2 (0.72)	8.2 (0.1)	0.7 (0.2)	24.1 (1.8)	8.1 (0.25)

directly without drying and chopping. Moisture contents of crop residue feedstock are given in Table 1.

Total carbon concentration of the biomass was determined by dry combustion in a Leco C-144 carbon analyzer (Merry and Spouncer, 1988). Total N was determined using a block digester method (Wendt, 1997) and total P by an acid persulfate digestion method (Liao and Marten, 2001). Exchangeable cations (i.e., K, Ca and Mg) were determined by atomic absorption spectrometry using ammonium acetate extraction procedures (Thomas, 1982). Micronutrient (i.e., Zn, Fe, Cu and Mn) contents were determined by atomic absorption spectrophotometer extracted by using the DTPA extraction procedure (Jones Jr and Case, 1990). Ash content of biochar was determined by combustion at 900°C in the muffle furnace for an hour. SAS GLM procedure and Duncan's multiple range tests were employed to test for statistical significance between and among treatments.

## Results and Discussion

Moisture content of biomass samples ranged from 4.2% for wheat straw to 37.8% for sorghum stems (Table 1). Average input amount of biomass sample as a feedstock to fill the pyrolysis reactor (i.e., 0.33 m<sup>3</sup>) ranged from 8.2 kg for wheat straw to 30.9 kg for sorghum stems. Average output amount of biochar ranged from 0.7 kg for wheat straw to 9.9 kg for rice hull. Ash content of biochar was observed from 16.6% for sorghum to 37.2% for rice hulls. Such a high ash content of biochar can be explained by manual operation of pyrolyzer, which has limitation of precise oxygen control in the reactor. Highest biochar yield was about 36.8% from corn stems and rice hulls, while the lowest



**Fig. 1. Field scale biomass pyrolyzer developed by University of Missouri Delta Research Center. Pyrolysis converts crop biomasses in the reactor (i.e., 0.33 m<sup>3</sup>) that is run at 400-500°C for 3-4 hours produce each batch of biochar. Note the air intake on the lower left of the reactor and the exhaust chimney above. The container on the right is catching the bio-oil.**

was about 8.1% from wheat straw. Contents of C, N, P, K, Ca, Mg, Fe, Zn, Mn, and Cu in the feedstock and biochar are given in Table 2, Fig. 2 and Fig. 3. Total carbon content was significantly highest in cotton stem biomass samples and lowest in rice hulls. Average carbon content of biomass (445 g kg<sup>-1</sup>) was increased in biochar (597 g kg<sup>-1</sup>) by 34%. The total carbon content of biochar from cotton stems and sorghum stems was increased by 50% and 40%, respectively, whereas that of rice hulls was increased by only 18%.

**Table 2. Elemental content of crop biomass samples based on dry weight (n=3). Means among feedstock samples or biochar samples followed by the same letter do not differ at the < 0.05 level.**

Biomass sample	C	N	P	K	Ca	Mg	Fe	Zn	Mn	Cu
	----- g kg <sup>-1</sup> -----					----- mg kg <sup>-1</sup> -----				
	<u>Feedstock</u>									
Corn stems	448 <sub>c</sub>	10.9 <sub>a</sub>	2.63 <sub>a</sub>	8.42 <sub>b</sub>	3.58 <sub>b</sub>	1.92 <sub>a</sub>	30.0 <sub>a</sub>	380 <sub>a</sub>	101.0 <sub>b</sub>	6.7 <sub>a</sub>
Cotton stems	466 <sub>a</sub>	8.4 <sub>b</sub>	1.04 <sub>b</sub>	7.07 <sub>bc</sub>	6.60 <sub>a</sub>	1.29 <sub>b</sub>	15.6 <sub>b</sub>	146 <sub>b</sub>	25.0 <sub>d</sub>	5.0 <sub>b</sub>
Rice hulls	409 <sub>d</sub>	5.1 <sub>d</sub>	0.52 <sub>c</sub>	1.48 <sub>d</sub>	1.28 <sub>e</sub>	0.46 <sub>c</sub>	16.0 <sub>b</sub>	69 <sub>c</sub>	256.3 <sub>a</sub>	4.3 <sub>b</sub>
Sorghum stems	449 <sub>c</sub>	6.6 <sub>c</sub>	1.01 <sub>b</sub>	6.50 <sub>c</sub>	2.28 <sub>d</sub>	1.61 <sub>ab</sub>	24.4 <sub>a</sub>	55 <sub>c</sub>	43.0 <sub>c</sub>	5.0 <sub>b</sub>
Wheat straws	454 <sub>b</sub>	5.3 <sub>d</sub>	0.43 <sub>c</sub>	15.34 <sub>a</sub>	3.02 <sub>c</sub>	1.45 <sub>b</sub>	8.1 <sub>c</sub>	27 <sub>c</sub>	28.0 <sub>d</sub>	4.6 <sub>b</sub>
F-value	200.6	102.0	67.3	93.6	167.4	25.9	18.3	43.5	1466.4	3.9
Pr > F	<.0001	<.0001	<.0001	<.0001	<.0001	<.0001	<.0001	<.0001	<.0001	0.04
	<u>Biochar</u>									
Corn stems	549 <sub>b</sub>	16.2 <sub>a</sub>	6.56 <sub>a</sub>	9.31 <sub>c</sub>	8.70 <sub>c</sub>	4.04 <sub>c</sub>	89.3 <sub>a</sub>	1844 <sub>b</sub>	273.6 <sub>b</sub>	9.2 <sub>b</sub>
Cotton stems	697 <sub>a</sub>	12.9 <sub>b</sub>	5.86 <sub>b</sub>	15.38 <sub>b</sub>	25.89 <sub>a</sub>	5.10 <sub>a</sub>	49.3 <sub>b</sub>	3462 <sub>a</sub>	141.6 <sub>c</sub>	16.6 <sub>a</sub>
Rice hulls	481 <sub>b</sub>	6.1 <sub>d</sub>	0.90 <sub>e</sub>	2.91 <sub>d</sub>	1.97 <sub>d</sub>	0.72 <sub>d</sub>	22.8 <sub>c</sub>	93 <sub>d</sub>	521.3 <sub>a</sub>	6.6 <sub>b</sub>
Sorghum stems	627 <sub>a</sub>	7.3 <sub>c</sub>	3.87 <sub>c</sub>	13.68 <sub>b</sub>	7.41 <sub>c</sub>	4.54 <sub>b</sub>	82.2 <sub>a</sub>	2899 <sub>a</sub>	157.6 <sub>c</sub>	8.6 <sub>b</sub>
Wheat straws	630 <sub>a</sub>	5.8 <sub>d</sub>	1.86 <sub>d</sub>	20.29 <sub>a</sub>	11.79 <sub>b</sub>	4.73 <sub>ab</sub>	24.4 <sub>c</sub>	897 <sub>c</sub>	127.3 <sub>c</sub>	6.5 <sub>b</sub>
F-value	13.8	232.4	240.6	85.3	314.4	182.7	72.8	47.6	21.6	9.2
Pr > F	0.004	<.0001	<.0001	<.0001	<.0001	<.0001	<.0001	<.0001	<.0001	0.002

**Table 3. Estimation of biochar returns for carbon and nutrients from pyrolysis of 1 Mg biomass. Standard deviations are in parentheses (n=3).**

Biomass sample	C	N	P	K
	----- kg Mg <sup>-1</sup> -----			
Corn stems	203 (50)	6.0 (1.4)	2.4 (0.7)	3.4 (0.6)
Cotton stems	119 (28)	2.2 (0.5)	1.0 (0.1)	2.6 (0.5)
Rice hulls	177 (18)	2.3 (0.3)	0.3 (0.1)	1.1 (0.2)
Sorghum stems	90 (30)	1.1 (0.4)	0.6 (0.2)	2.0 (0.6)
Wheat straws	52 (18)	0.5 (0.1)	0.2 (0.1)	1.7 (0.6)

The total N content differed among biomass samples, being highest in corn stems and cotton stems. Following pyrolysis, biochar from corn stems had 48% higher total N content and that from cotton stems had 54% higher total N content than the feedstock. Rice hulls had the lowest total N content in the biomass and the total N content of the biochar was not significantly higher. Contents of P, K, Ca, and Mg were different among feedstock and biochar samples which generally were consistent with their contents in the biochar formed. Average P content was >3-fold higher in biochar samples than feedstock samples. Significant high P content was found in biochar samples of cotton stems and wheat straws samples when compared to feedstock

samples.

Content of K differed among feedstocks and the biochar formed from them. Biochar K from cotton and sorghum stems was double that of their respective feedstock samples. Average Ca and Mg content were >3-fold higher in biochar samples than in feedstock samples. Micronutrient (i.e., Fe, Zn, Mn, and Cu) contents were significantly different among the biomass samples in feedstock and biochar. But, no significant difference between feedstock and biochar was found in Zn, Fe, and Cu content for rice hulls sample. Zn, Fe, Mn and Cu content of biochar samples were >3-fold, >13-fold, 2.5-fold, and 1.5-fold higher than feedstock samples, respectively.

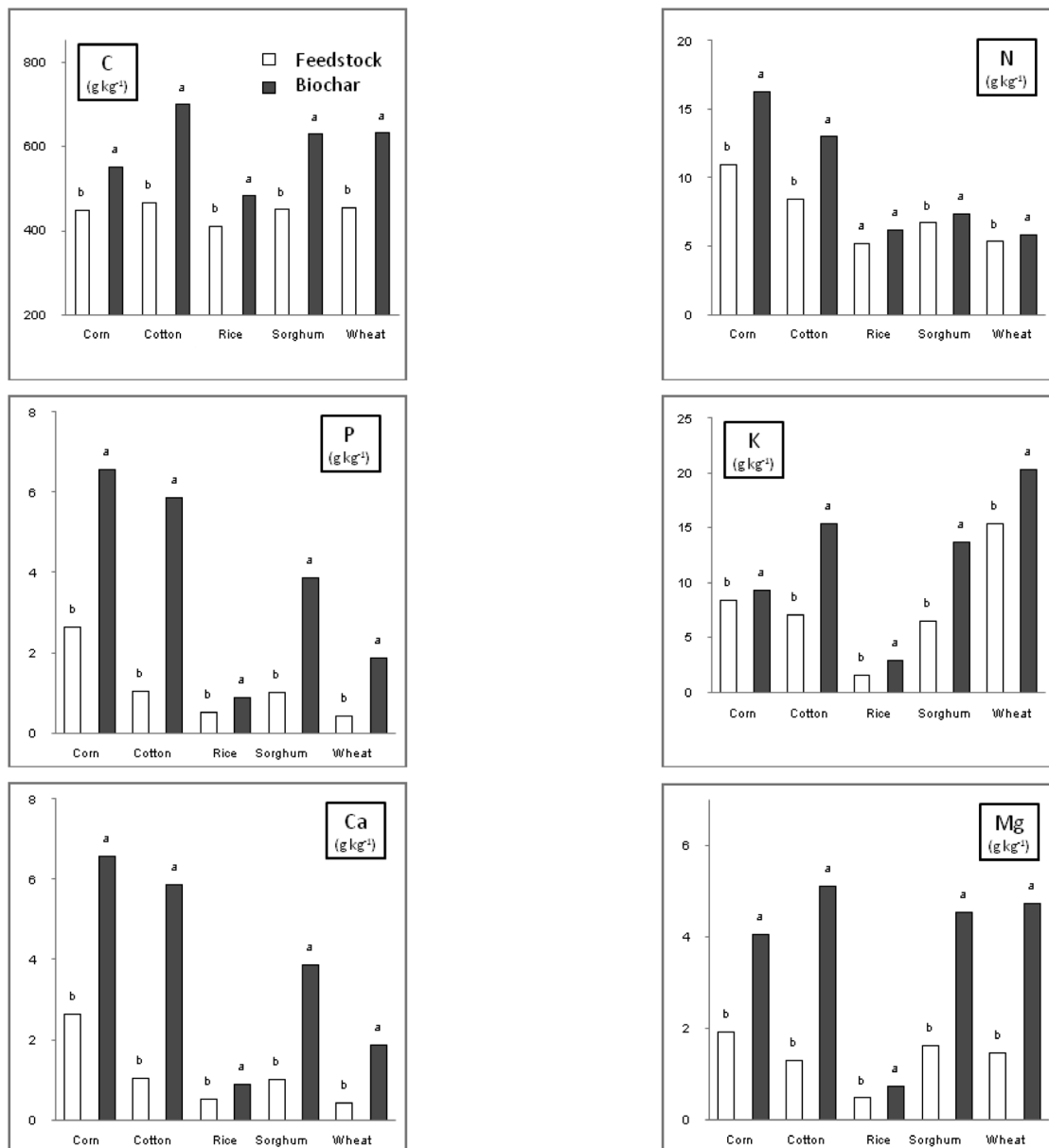


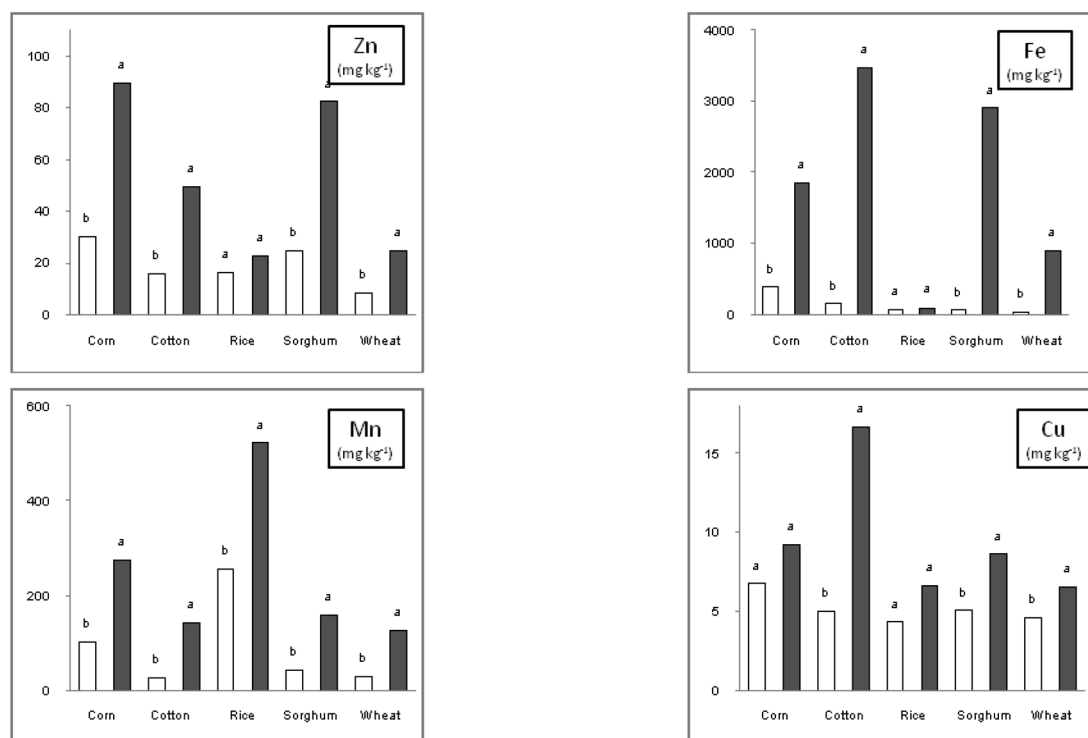
Fig. 2. Comparison of nutrient contents of the crop biomass feedstock and the biochar it produced (n=3). Means within a feedstock with the same letter are not different at p < 0.05.

Biochar production from corn stems and rice hulls was relatively high in carbon retention (i.e., 203 kg and 177 kg from 1 Mg of biomass) while wheat straw was relatively low in carbon retention (i.e., 52 kg from 1 Mg of biomass) (Table 3). Cotton stems biochar retained the highest amount of N (i.e., 6.0 kg), P (i.e., 2.4 kg), and K (i.e., 3.4 kg) while wheat straw biochar retained the lowest amount of N (i.e., 0.5 kg) and P (i.e., 0.2 kg). Rice hulls biochar retained the lowest amount of K (i.e., 1.1 kg).

The biomass feedstock samples differed in dry moisture content (Table 1). Those of corn stems, cotton stems, and sorghum stems decreased in moisture content

by 30% for a week of air drying, but there was no change in moisture content of rice hulls and wheat straw (data not shown). Because rice hull biomass already has a uniform particle size (i.e., < 5 mm in diameter), the chopping process is not necessary. Therefore, rice hulls biomass was already dried and required the least use of energy to prepare for pyrolysis. Despite a range in moisture contents of biomass samples, the pyrolysis system runs very well under natural outdoor conditions.

Previous research has reported that organic carbon in biochar is very resistant against microbial decomposition and works in the soil much like soil organic matter (DeLuca et al., 2009; Lehmann, 2009). When



**Fig. 3.** Comparison of micro-nutrients contents between crop biomass feedstock and the biochar produced (n=3). Means within a feedstock with the same letter are not different at  $p < 0.05$ .

biochar is returned to soil, we expect that on average 13.3% of the biomass weight could be sequestered in the soil for several decades or centuries. Specifically, biochar from corn stems and rice hulls, respectively, can sequester 20% and 17.7% the biomass weight. Lehmann et al. (2006) reported that biochar can sequester up to 50% of the initial carbon input. However, we found that biochar from corn stems and rice hulls can sequester by 60% and 49% of the initial carbon input from the biomass respectively when biochar is incorporated into the soils. This result implies that pyrolysis conversion of biomass to biochar contributes significantly to sequester carbon that can be applied to soils. However, few research data have been reported especially for practical application of pyrolysis and biochar use at the local farm scale. Therefore, these results provide fundamental data for future research on small scale biochar production. Furthermore, agronomic and environmental impacts and conditions for biochar application should be investigated with various biomass sources and crops.

## Conclusions

We developed low-cost on farm biomass pyrolysis system, which can be used to produce biochar at the

farm level from locally collected feedstocks. This pyrolysis system is designed for operating without continuous non-feedstock energy use. Bio-oil, a by-product with economic value, was collected during pyrolysis and is being evaluated. Foremost advantage of this biomass pyrolyzer is ease to adopt this manual system for producing biochar at the local farm. Furthermore, pyrolysis conversion of crop residues can sequester significant amounts of carbon as biochar which has further environmental and production benefits when applied to soils.

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## Disclaimer

Mention of trade names or commercial products in this publication is solely for the purpose of providing specific information and does not imply recommendation or endorsement by the University of Missouri.

## References

- Alves A., M. Schwanninger, H. Pereira, and J. Rodrigues. 2006. Analytical pyrolysis as a direct method to determine the lignin content in wood: Part 1: Comparison of pyrolysis lignin with Klason lignin. *Journal of Analytical and Applied Pyrolysis* 76:209-213.
- Baldock, J. and R. Smernik. 2002. Chemical composition and bioavailability of thermally altered *Pinus resinosa* (Red pine) wood. *Organic Geochemistry* 33:1093-1109.
- Cunha, T., B. Madari, L. Canellas, L. Ribeiro, V. Benites, and G. Santos. 2009. Soil organic matter and fertility of anthropogenic dark earths (Terra Preta de Índio) in the Brazilian Amazon basin. *Revista Brasileira de Ciência do Solo* 33:85-93.
- DeLuca, T., M. MacKenzie, and M. Gundale. 2009. Biochar Effects on Soil Nutrient Transformations. *In: Lehmann J and Joseph S. (eds.) Biochar for environmental management: science and technology:pp., 251-285. Eatthscan Publishing: London, Sterlin, VA.*
- Demirba, A. 2001. Biomass resource facilities and biomass conversion processing for fuels and chemicals. *Energy Conversion and Management* 42:1357-1378.
- Glaser, B. 2007. Prehistorically modified soils of central Amazonia: a model for sustainable agriculture in the twenty-first century. *Philosophical Transactions of the Royal Society B: Biological Sciences* 362:187.
- Glaser, B., E. Balashov, L. Haumaier, G. Guggenberger, and W. Zech. 2000. Black carbon in density fractions of anthropogenic soils of the Brazilian Amazon region. *Organic Geochemistry* 31:669-678.
- Goldstein, I. and C.R. Company. 1981. *Organic chemicals from biomass.* CRC press Boca Raton, FL.
- Hansen, J., M. Sato, P. Kharecha, D. Beerling, R. Berner, V. Masson-Delmotte, M. Pagani, M. Raymo, D. Royer, and J. Zachos 2008. Target atmospheric CO<sub>2</sub>: Where should humanity aim? *Open Atmospheric Science Journal* 2:217-231.
- Howard, J. 1981. Fundamentals of coal pyrolysis and hydrolysis. *Chemistry of Coal Utilization* 2:665-784.
- Jones Jr., J. and V. Case. 1990. Sampling, handling, and analyzing plant tissue samples. *Soil Testing and Plant Analysis* 3:389-428.
- Lal, R. 2004. Soil carbon sequestration impacts on global climate change and food security. *Science* 304:1623.
- Lehmann, J. 2007a. Bio-energy in the black. *Frontiers in Ecology and the Environment* 5:381-387.
- Lehmann, J. 2007b. A handful of carbon. *Nature* 447:143-144.
- Lehmann, J. 2009. Biological carbon sequestration must and can be a win-win approach. *Climatic Change* 97:459-463.
- Lehmann, J., J. Gaunt, and M. Rondon. 2006. Bio-char sequestration in terrestrial ecosystems-a review. *Mitigation and Adaptation Strategies for Global Change* 11:395-419.
- Liang, P., Z. Wang, and J. Bi. 2007. Process characteristics investigation of simulated circulating fluidized bed combustion combined with coal pyrolysis. *Fuel Processing Technology* 88:23-28.
- Liao, N. and S. Marten. 2001. Determination of total phosphorus by flow injection analysis colorimetry (acid persulfate digestion method). *QuikChem® Method:10-115.*
- Malhi, S., R. Lemke, Z. Wang, and B. Chhabra. 2006. Tillage, nitrogen and crop residue effects on crop yield, nutrient uptake, soil quality, and greenhouse gas emissions. *Soil and Tillage Research* 90:171-183.
- McHenry, M. 2009. Agricultural bio-char production, renewable energy generation and farm carbon sequestration in Western Australia: Certainty, uncertainty and risk. *Agriculture, Ecosystems & Environment* 129:1-7.
- McKendry, P. 2002. Energy production from biomass (part 1): overview of biomass. *Bioresource Technology* 83:37-46.
- Merry, R. and L. Spouncer. 1988. The measurement of carbon in soils using a microprocessor-controlled resistance furnace. *Communications in Soil Science and Plant Analysis* 19: 707-720.
- Mohan, D., C. Pittman Jr., and P. Steele. 2006. Pyrolysis of wood/biomass for bio-oil: a critical review. *Energy Fuels* 20:848-889.
- Oasmaa, A., D. Elliott, and S. Müller. 2009. Quality control in fast pyrolysis bio-oil production and use. *Environmental Progress & Sustainable Energy* 28:404-409.
- Soil Survey Staff. 1997. *Keys to soil taxonomy.* Pocahontas Press, Blacksburg, VA.
- Teasdale, J., A. Abdul-Baki, and Y. Bong Park. 2008. Sweet corn production and efficiency of nitrogen use in high cover crop residue. *Agronomy for Sustainable Development* 28: 559-565.
- Teng, H., H. Lin, and J. Ho. 1997. Thermogravimetric analysis on global mass loss kinetics of rice hull pyrolysis. *Industrial & Engineering Chemistry Research* 36:3974-3977.
- Thomas, G. 1982. Exchangeable cations. *In: A.L Page (ed.) Methods of Soil Analysis, Part 2 Chemical and Microbiological Properties, 2<sup>nd</sup> edition.* Agronomy 9:159-165.
- Van Zwieten, L., S. Kimber, S. Morris, K. Chan, A. Downie, J. Rust, S. Joseph, and A. Cowie. 2010. Effects of biochar from slow pyrolysis of papermill waste on agronomic performance and soil fertility. *Plant and Soil* 327:235-246.
- Wendt, K. 1997. Determination of total Kjeldahl nitrogen by flow injection analysis colorimetry (block digester method). *QuikChem Method:10-107.*
- Zhang, X., S. Kondragunta, C. Schmidt, and F. Kogan. 2008. Near real time monitoring of biomass burning particulate emissions (PM<sub>2.5</sub>) across contiguous United States using multiple satellite instruments. *Atmospheric Environment* 42: 6959-6972.