

The Caudal-Related Homeodomain Proteins Upregulate *catalase* Expression in *Drosophila* Hindgut and Human Colorectal Carcinoma Cells

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Caudal-related homeodomain proteins play critical roles in intestine development and maintenance from *Drosophila* to humans. The loss or reduction of CDX1 and CDX2 are known to be associated with colon cancers. It has been well known that colorectal carcinogenesis is associated with serious oxidative stress and that catalase is decreased in colon carcinomas. However, the underlying molecular mechanisms remain elusive. Here, we report that Caudal-related homeodomain proteins positively regulate catalase expression in both *Drosophila* and humans. We found that *Drosophila caudal* heterozygotes have a decreased *catalase* expression and increased ROS generation in the hindgut, and that the overexpression of Caudal increases *catalase* promoter activity and catalase mRNA levels. We also found that CDX1 and CDX2 up-regulate *catalase* promoter activity and protein levels in HCT116 cells - human colorectal carcinoma cell lines. The level of catalase protein in several colorectal carcinoma cell lines was associated with CDX1 expression. These results suggest that CDX1 and CDX2 may be involved in intestinal homeostasis and tumorigenesis via regulation of *catalase* expression.

Key words : *Drosophila caudal*, CDX1, CDX2, catalase, colorectal carcinoma

Introduction

Homeodomain proteins are essential in the control of normal embryonic development [18]. Cdx1 and Cdx2 are members of the *caudal*-related homeobox gene family based on their sequence homology to the *caudal* gene of *Drosophila* [25]. The Caudal-related homeodomain proteins play critical roles for intestine development and maintenance from *Drosophila* to human [8,20]. *Drosophila* Caudal is required for definition of the anteroposterior axis during early embryogenesis [26] and hindgut development [41]. In vertebrates, Cdx1 is expressed in the crypts where epithelial cells are proliferating, whereas Cdx2 is mainly expressed in the villi lined by differentiated cells [33]. These homeobox genes participate in the control of intestinal homeostasis by regulating the equilibrium between proliferation and differentiation during the constant renewal of the gut epithelium. Alternated expression of homeodomain proteins is associated with the initiation and progression of cancer [23]. Cdx1 and Cdx2 expression are preferentially silenced or reduced with the progression to colon cancer in human [3,40].

Heterozygous Cdx2 knockout mice have been known to develop colonic tumors [6]. These facts indicate that the loss or reduction of Cdx1 and Cdx2 are associated with colorectal carcinogenesis. However, the underlying molecular mechanisms remain elusive.

Colorectal carcinogenesis has been well known to be associated with serious oxidative stress [4]. Recently, it was reported that gradual advancement of oxidative-antioxidative disorders is followed by progression of colorectal cancer [34]. In considering a critical function for oxidative stress in progression of colorectal cancers, regulation of the activities of H₂O₂ metabolizing enzymes, such as superoxide dimutase (SOD), catalase and glutathione peroxidase (GPx) are likely to be required [34]. Among the anti-oxidants, catalase is a heme-containing peroxisomal enzyme and protects the H₂O₂ generated in normal human tissues [27]. Catalase has been well known to be decreased in colon carcinomas [21]. Although several studies have demonstrated tissue-specific expression of the *catalase* gene in mammals [24,36] and constitutively elevated *catalase* expression at the transcriptional level by both SP1 and CCAAT-recognizing factors [28], the mechanisms of *catalase* regulation are little known. In previous study, we found that *Drosophila catalase* gene expression is regulated by DREF [29], which is also a key regulator of *Drosophila caudal* gene expression [7]. Therefore, we thought that Caudal-related transcription fac-

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tors may modulate *catalase* gene expression. We did BLAST search and found that, interestingly, several consensus DNA-binding sites for Caudal-related homeodomain protein are located in the 5'-flanking regions of *Drosophila* and human *catalase* indicating some possible links between *catalase* expression and Caudal-related homeodomain proteins. In the present study, we have investigated the role of Caudal-related homeodomain proteins in regulation of the *catalase* gene in *Drosophila* hindgut and human colorectal carcinoma cell lines. Our results suggest that Caudal-related homeodomain proteins positively regulate the expression of *catalase* in both *Drosophila* and human.

Materials and Methods

Fly stock

Fly stocks were maintained at 25°C on standard food. The *catalase-lacZ* [29] and *UAS-cad* [15] transgenic flies described previously were used. The *cad^Δ/Gla* and hs-GAL4 strain were kindly supplied by the Bloomington Stock Center. *Oregon-R* was used as wild type. The *cad^Δ/+;catalase-lacZ/+* flies were derived from a cross of *cad^Δ/Gla/+* male flies to female flies carrying homozygous *catalase-lacZ* on the third chromosome, and *+;/catalase-lacZ/+* flies were derived from a cross of the *Oregon-R* males to homozygous *catalase-lacZ* female flies. For ectopic expression of Caudal using the GAL4-UAS system, the *UAS-cad/+;hs-GAL4/+* flies were derived from a cross of the males carrying homozygous *UAS-cad* on the second chromosome to females carrying homozygous *hs-GAL4* on the third chromosome, and the *+;/hs-GAL4/+* flies were derived from a cross of the *Oregon-R* males to females carrying homozygous *hs-GAL4* on the third chromosome. The progeny third instar larvae were heat-shocked at 37°C for 45 min and then returned to 25°C.

X-Gal staining

The tissues were dissected and fixed for 15 min in PBS containing 1% glutaraldehyde, washed in PBS and immersed in 0.2% 5-bromo-4-chloro-3-indolyl-β-D-galactopyranoside (X-gal) in staining buffer containing 6.1 mM K₄Fe(CN)₆, 6.1 mM K₃Fe(CN)₆, 1 mM MgCl₂, 150 mM NaCl, 10 mM Na₂HPO₄ and 10 mM NaH₂PO₄. Incubation was in the dark at 37°C.

Plasmid construction

To construct *pCat-Luc*, the upstream region of human

Catalase gene (-906 to +18 with respect to the transcription initiation site) was amplified by PCR using human umbilical vein endothelial cell genomic DNA and inserted into the *KpnI* sites of *pGL2-basic* (Promega, Madison, USA).

DCFH-DA assay

For determination of total ROS generation, the hindgut of wild type and *cad^Δ* heterozygous adult flies were homogenized in homogenizing buffer (50 mM K⁺ phosphate buffer, pH 7.4) and centrifuged at 12,000 rpm at 4°C for 10 min. 25 mM DCFH-DA (2', 7'-dichlorofluorescein diacetate) was added to the extracts and changes in fluorescence intensity were measured on a Fluorescence Microplate Reader (FL500, Bio-Tek Instruments, Winooski, USA) with excitation wavelength of 486 nm and emission wavelength of 530 nm for 1 hr. The fluorescence intensities were normalized to protein amounts determined by the BCA protein kit (Sigma, St. Louis, MO).

Cell culture

Drosophila Kc cells were grown at 25°C in M3 (BF) medium (Sigma) supplement with 1% fetal bovine serum (FBS), 100 units/ml of penicillin and 100 μg/ml of streptomycin (Gibco BRL, Gaithersburg, MD). Human colorectal carcinoma cell lines, DLD1, HCT116, SW480 and SW620 were cultured in RPMI 1640 Medium (Gibco/BRL), supplemented with 10% FBS, 100 units/ml of penicillin and 100 μg/ml of streptomycin at 37°C incubator containing 5% CO₂. HCT-pcDNA3, HCT-CDX1, and HCT-CDX2 cell lines were obtained by stable transfecting the pcDNA3.0, pcDNA3/CDX1 [25], and pcDNA3/CDX2 [25] plasmids into HCT116 cells; and stable clones were isolated after 2-3 weeks of G418 (700 μg/ml) selection.

DNA transfection and luciferase assay

Transfection of DNA mixtures into *Drosophila* Kc cells was performed using DDAB (Dimethyldioctadecyl ammonium bromide) [13] and the cells were harvested at 48 hr after transfection. Lipofectamine Reagent (Gibco/BRL)-mediated transfection into HCT-pcDNA3, HCT-CDX1, and HCT-CDX2 cell lines was performed using 24-well tissue culture plates according to the supplier's protocol. The luciferase assay was carried out by means of a Luciferase Assay System (Promega), as described previously [16]. Normalized luciferase activities were calculated by determining the luciferase/β-galactosidase activity ratios, and reported as the

mean±S.D. from triplicate transfections.

RT-PCR

Total RNA from larvae was isolated with Trizol Reagent and cDNAs were synthesized with M-MLV-RT. The RT-PCR products were analyzed on agarose gels stained with ethidium bromide. Oligonucleotide primers for PCR were designed as follows.

caudal, 5' CGAGCAACAAGAAGGGTAGC 3' and 5' TGATGGGGCTCCATGTAGTT 3'

catalase, 5' GGTACCCT TTGAGGTGACCCACGACA 3' and 5' TACGAGCAGGCC AAGAAGTTGGTACC 3'

ribosomal protein 49 (rp49), 5' GACAACAGAGYCGG TCGC 3' and 5' GTTGTGCACCAGGAAGTT 3'

CDX1, 5' TGTGGCAGCGGTAAGACCCGAACC 3' and 5' GGACTTGCGCCGGATAGT TGAGTA 3'

CDX2, 5' CGGCTGGAGCTGGAGAAGG 3' and 5' TCAGCCTGGAATTGCTCTGC 3'

β-actin, 5' GACTACCTCATGAAGATC 3' and 5' GATCCACATCTGCTGGAA 3'

Immunoblotting

A preparation of cell extracts and immunoblotting were performed as described earlier [12]. Immunoblotting was performed using specific antibody for CDX1, Catalase (Santa-Cruz, Santa cruz, CA), α -tubulin antibody (Goma Biotech) and HRP-conjugated anti-mouse antibody (Santa Cruz). CDX1 and CDX2 antibody were kindly supplied by Dr W.J. Lee. Proteins were visualized with enhanced chemiluminescence (ECL) detection system (Amersham, Piscataway, NJ).

Results and Discussion

We established previously *catalase-lacZ* reporter transgenic flies for *Drosophila catalase* gene expression [29]. The *catalase-lacZ* expression is detected in the hindgut of adults, *Drosophila* homolog of colon [29]. Therefore, to investigate the role of Caudal in *catalase* expression, we examined whether the expression of *catalase-lacZ* is modulated by a Caudal mutant *cad^Δ*, which was generated by γ -ray-induced mitotic recombination [5]. The *catalase-lacZ* expression was significantly reduced in the hindgut of *cad^Δ* heterozygous adult flies compared to that of wild type (Fig. 1A). We also examined quantitative β -galactosidase activities in the hindgut of these flies. The β -galactosidase activity in *cad^Δ*

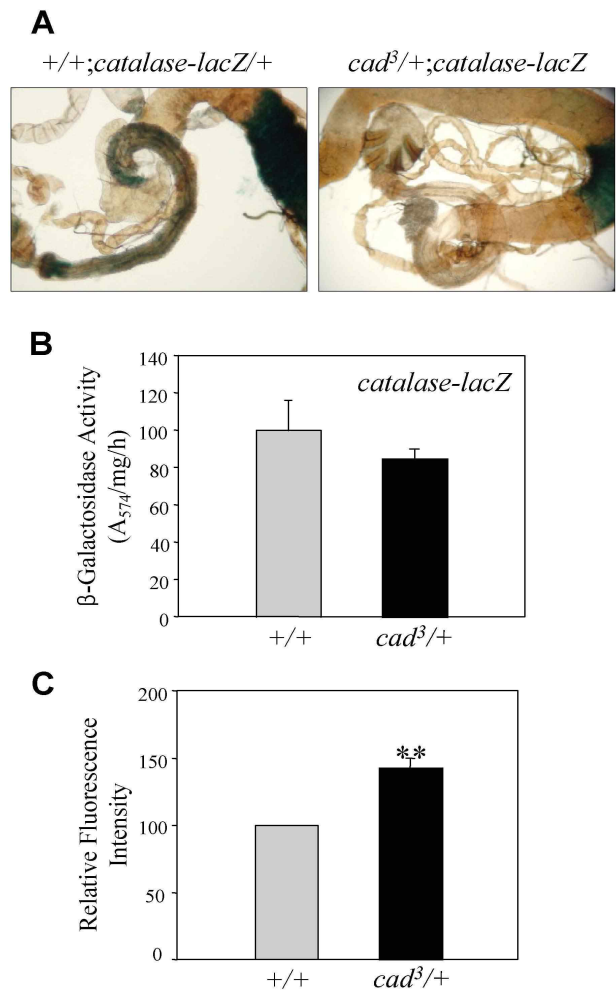


Fig. 1. A decreased expression of *catalase* gene and increased ROS generation in the hindgut of Caudal mutant heterozygous (*cad^Δ*) adult flies. (A) Expression of *catalase-lacZ* in the hindgut of wild type and *cad^Δ* heterozygous adult flies. The adult tissues were dissected and stained with 0.2% X-gal solution in the dark. Expression of β -galactosidase in the hindgut was decreased by *cad^Δ* allele. (B) Quantitative β -galactosidase activities of the hindgut of wild type and *cad^Δ* heterozygous adult flies. The adult hindgut extracts were prepared from 3-day-old adult transgenic flies as described in Materials and Methods. The β -galactosidase activities are expressed as absorbance units at 574 nm/mg of protein. Averaged values obtained from four independent experiments with \pm S.E. values are shown. (C) Comparison of ROS generation in the hindgut of wild type and *cad^Δ* heterozygous adult flies. The generation of ROS was measured by DCFH-DA assay. Changes of fluorescence intensity were measured with excitation and emission wavelength of 486 nm and 530 nm, respectively for 1 hr. Fluorescence intensities are shown as relative intensity compared to wild type. Each value is the mean \pm S.E. from four independent experiments. A statistical significance is marked as ** $p < 0.01$.

heterozygous adult hindgut was reduced to 83-85% of wild type, as shown in Fig. 1B). It has been well known that the *catalase* gene expression is critical for ROS homeostasis [32]. Therefore, we measured ROS generation in the hindgut of wild type and *cad*¹ heterozygous adults. ROS generation in the hindgut of *cad*¹ heterozygous adults was significantly higher than that of wild type (Fig. 1C). To further understand the role of Caudal on *catalase* gene expression, we examined whether Caudal overexpression stimulates *catalase* gene expression. The effect of Caudal on *catalase* promoter activity was examined by transient transfection experiment in the *Drosophila* Kc cell line using the *catalase* reporter plasmid *pcatalase-Luc* and Caudal expression plasmid *pAct-cad*. The *catalase* promoter activity was dose-dependently transactivated by Caudal (Fig. 2A). To investigate the upregulation of *catalase* expression by Caudal in living flies, Caudal overexpression in living flies was performed with the GAL4-UAS system [11]. Transgenic flies carrying *UAS-Cad* were crossed with transgenic flies carrying GAL4 cDNA placed under control of the hsp70 gene promoter (*hs-GAL4*). Caudal overexpression in third instar larvae carrying single copies of both *hs-GAL4* and *UAS-Cad* was confirmed by RT-PCR (Fig. 2B). The level of catalase mRNA was increased by heat shock-induced Caudal (Fig. 2B). Thus, these results indicate that Caudal regulates positively *catalase* gene expression at transcriptional level.

Since we found several sequences similar to the Caudal-related homeodomain binding consensus sequences in the 5'-flanking region of human *Catalase* gene, to investigate whether human *Catalase* gene expression is regulated by CDX1 and CDX2, human homologs of caudal, the reporter plasmid *phCat-Luc* was constructed (Fig. 3A). Effects of CDX1 and CDX2 on *Catalase* gene promoter activity were examined by transient transfection experiments with *phCat-Luc* reporter plasmid and *pcDNA3/CDX1* or *pcDNA3/CDX2* expression plasmid in colorectal carcinoma cell line HCT116. Human *Catalase* promoter activity was transactivated dose-dependently by CDX1 and CDX2 expression plasmids, up to nearly 7- and 4-fold, respectively (Fig. 3B). We established CDX1 and CDX2 stable transfectant clones in HCT116 cell line (HCT-CDX1 and HCT-CDX2). The overexpression of CDX1 and CDX2 in these stable transfectants was confirmed by RT-PCR, respectively (Fig. 4A). To confirm whether *Catalase* promoter activity is regulated by CDX1 and CDX2, *phCat-Luc* was transfected into CDX1 and CDX2 stable transfectants,

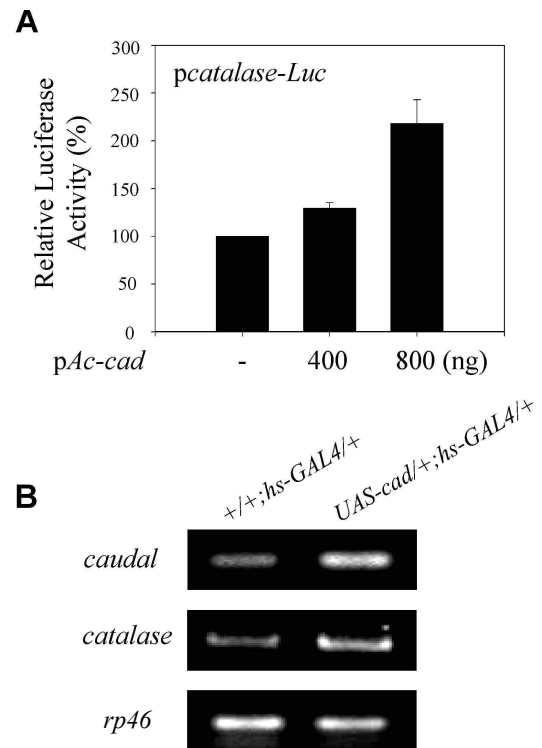


Fig. 2. Caudal over-expression induces *catalase* gene expression. (A) Luciferase reporter assay shows a strong and dose-dependent stimulation of the *catalase* reporter by Caudal. Each transfection mixture consisted of the *pcatalase-Luc* reporter plasmid and *pactin-LacZ* internal control plasmid. The 400 or 800 ng of *pAc-cad* or the plasmid *pAc* for the control (mock) of effector was co-transfected in *Drosophila* Kc cells. Normalized luciferase activities were calculated by determining the luciferase/ β -galactosidase activity ratios. (B) Total RNA was prepared from the third instar larvae incubated at 25°C for 24 hr after heat shock at 37°C for 45 min. Caudal over-expression was confirmed by RT-PCR. Level of *Drosophila* catalase mRNA was examined by RT-PCR using specific primers. control, *+/+;hs-GAL4/+*. Caudal, *UAS-cad/+;hs-GAL4/+*.

respectively. The luciferase activities in the HCT-CDX1 and in the HCT-CDX2 cell lines were 9- to 11-fold and 4-fold of the HCT-pcDNA3, respectively (Fig. 4B). To confirm the stimulatory effects of CDX1 and CDX2 on *Catalase* promoter activity, we examined levels of Catalase protein in HCT-CDX1 and HCT-CDX2 stable transfectants by Western blot. The protein levels of Catalase in HCT-CDX1 and HCT-CDX2 were higher than that in HCT-pcDNA3 (Fig. 4C). These results indicate that human *Catalase* is up-regulated by Caudal-related homeodomain proteins CDX1 and CDX2 and that transactivating activity of CDX1 on *Catalase* promoter activity is higher than that of CDX2.

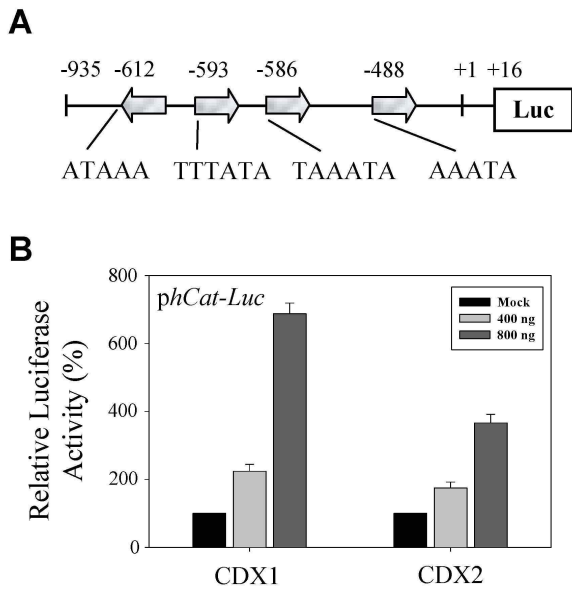


Fig. 3. CDX1 and CDX2 increase *Catalase* promoter activity in colorectal carcinoma cell line HCT116. (A) The potential Caudal-related homeodomain protein binding sites in human *Catalase* gene promoter are shown. The transcription initiation site is numbered +1. (B) Luciferase reporter assay shows dose-dependently stimulation of the *Catalase* reporter by CDX1 and CDX2. Transfection mixture was consisted of the *phCat-Luc* reporter plasmid and *SV40-LacZ* internal control plasmid. The 400 or 800 ng of *pcDNA3/CDX1* or *pcDNA3/CDX2* plasmid and *pcDNA3.0* plasmid for the control of effector were cotransfected in colorectal carcinoma cell line HCT116. Normalized luciferase activities were calculated by determining the luciferase/ β -galactosidase activity ratios.

To further confirm the effect of CDX1 on *Catalase* expression, levels of *Catalase* in four colorectal carcinoma cell lines (DLD1, HCT116, SW480 and SW620) were examined by Western blot. Recently, it was reported that CDX1 mRNA is expressed in SW480 and SW620 cell lines, but not in DLD1 and HCT116 cell lines [32]. We examined the protein expression of CDX1 in these cell lines. CDX1 expression was detected in SW480 and SW620 cell lines, but not in DLD1 and HCT116 cell lines. This result is consistent with the previous report. The protein levels of *Catalase* in SW480 and SW620 cell lines expressing CDX1 were higher than those in DLD1 and HCT116 (Fig. 5). This result indicates that the level of *Catalase* in four colorectal carcinoma cell lines is correlated with or without CDX1 expression, supporting the transactivating activity of CDX1 on *Catalase* promoter activity.

In the present study, we have shown that the Caudal-related homeodomain proteins positively regulate *catalase* expression in both *Drosophila* hindgut and human colorectal carcinoma cells. The expression of CDX1 and/or CDX2 is decreased in the majority of the colon carcinomas [3,40], although the expression of CDX1 is controversial during colorectal cancer progression [10]. Oxidative stress can induce DNA damages that lead to genomic instability and possibly stimulate cancer progression [16]. Elevated Reactive oxygen species (ROS) levels and oxidative stress are responsible for constant activation of transcription factors, such as NF κ B and AP-1, during tumor progression

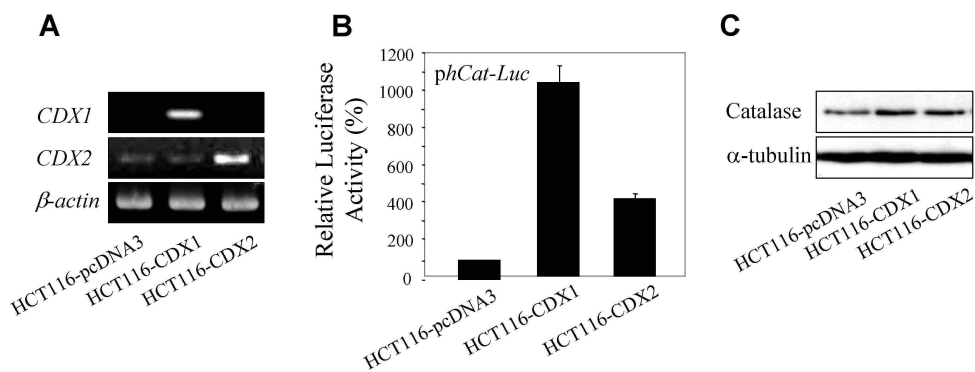


Fig. 4. *Catalase* promoter activities and protein levels in HCT-CDX1 and HCT-CDX2 stable transformants. (A) Total RNA was isolated from HCT-pcDNA3, HCT-CDX1, and HCT-CDX2 stable HCT116 cell lines. mRNA expression was monitored by RT-PCR. β -Actin was used as a positive control. (B) *Catalase* promoter activities are increased in HCT-CDX1 and HCT-CDX2 stable transformants. Transfection mixture was consisted of the *phCat-Luc* reporter plasmid and *SV40-LacZ* internal control plasmid. The mixture was transfected into HCT-pcDNA3, HCT-CDX1 and HCT-CDX2 stable HCT116 cell lines. Normalized luciferase activities were calculated by determining the luciferase/ β -galactosidase activity ratios. (C) *Catalase* protein levels are increased in HCT-CDX1 and HCT-CDX2 stable transformant. Protein extracts (50 μ g) of the HCT-pcDNA3, HCT-CDX1 and HCT-CDX2 cell lines were fractionated on a 10% SDS-PAGE and analyzed by Western blotting using the specific antibodies as noted. α -tubulin was served as a loading control.

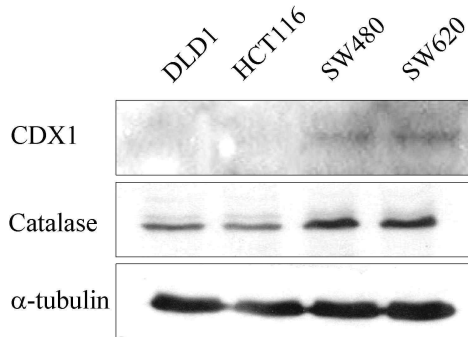


Fig. 5. The protein levels of human Catalase in different colorectal cancer cell lines. Protein extracts (50 µg) isolated from DLD1, HCT116, SW480 and SW620 cells were separated on 10% SDS-PAGE and Western blot analysis was performed with the specific antibodies as noted. α -tubulin was served as a loading control.

[12]. Thus, ROS and oxidative stress are thought to play multiple roles in tumor initiation, progression and maintenance. Colorectal carcinogenesis has been well known to be associated with serious oxidative stress [4]. Evidences have been given in some reports that H_2O_2 could promote cell growth and related gene expression in human tumors such as breast cancer and colon cancer [17,31,42]. From these facts, our results suggest that loss or reduction of CDX1 and/or CDX2 causes decreased expression of Catalase, which may increase oxidative stress in colon, accumulate genetic alterations and induce tumorigenesis.

Numerous studies also demonstrate evidences that oxidative stress occurs the pathogenesis of endotoxin-induced motility disturbances in animals and humans, and that oxygen-derived free radicals are implicated as relevant mediators in sepsis and septic shock [1,9,39]. Oxidative stress participates in the upregulation of iNOS expression, COX2 expression and NF- κ B activity, which are inhibited by Catalase [37,39]. Thus, down-regulation of Catalase expression due to loss or reduction of CDX1 and/or CDX2 may attenuate the endotoxin-related defense system, which may induce intestinal diseases such as intestinal ischemia and chronic inflammatory bowel disease as well as cancer.

Ectopic expression of CDX1 and CDX2 in adenomas of the stomach, esophagus, and liver has been reported [2,30,38]. Overexpression of CDX1 in colon cancer was also reported [10]. Since many anticancer drugs have been suggested to cause oxidative stress [22,35] and Catalase activity is associated with the acquisition of cellular resistance to anticancer drugs [14,19], increased Catalase in cancer cells

overexpressing CDX1 and CDX2 may also convey resistance to anticancer drugs.

In summary, our results indicate that upregulation of *catalase* gene by Caudal-related homeodomain proteins is conserved evolutionarily from flies to human and suggest the possibility that Caudal-related homeodomain proteins CDX1 and CDX2 are involved in intestinal homeostasis and tumorigenesis via up-regulation of *catalase* expression.

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References

1. Albuszies, G. and U. B. Bruckner. 2003. Antioxidant therapy in sepsis. *Intensive Care Med* **29**, 1632-1636.
2. Almeida, R., E. Silva, F. Santos-Silva, D. G. Silberg, J. Wang, C. De Bolos, and L. David. 2003. Expression of intestine-specific transcription factors, CDX1 and CDX2, in intestinal metaplasia and gastric carcinomas. *J. Pathol.* **199**, 36-40.
3. Baba Y., K. Nosho, K. Shima, E. Freed, N. Irahara, J. Philips, J. A. Meyerhardt, J. L. Hornick, R. A. Shivdasani, C. S. Fuchs, and S. Ogino. 2009. Relationship of CDX2 loss with molecular features and prognosis in colorectal cancer. *Clin. Cancer Res.* **15**, 4665-4673.
4. Bartsch, H., J. Nair, and R. W. Owen. 2002. Exocyclic DNA adducts as oxidative stress markers in colon carcinogenesis: potential role of lipid peroxidation, dietary fat and antioxidants. *Biol. Chem* **383**, 915-921.
5. Butler, H., S. Levine, X. Wang, S. Bonyadi, G. Fu, P. Lasko, B. Suter, and R. Doerig. 2001. Map position and expression of the genes in the 38 region of *Drosophila*. *Genetics* **158**, 1597-1614.
6. Chawengsaksophak, K., R. James, V. E. Hammond, F. Kontgen, and F. Beck. 1997. Homeosis and intestinal tumours in *Cdx2* mutant mice. *Nature* **386**, 84-87.
7. Choi, Y. J., T. Y. Choi, M. Yamaguchi, A. Matsukage, Y. S. Kim, and M. A. Yoo. 2004. Transcriptional regulation of the *Drosophila caudal* homeobox gene by DRE/DREF. *Nucleic Acids Res.* **32**, 3734-3742.
8. Crissey, M. A., R. J. Guo, S. Funakoshi, J. Kong, J. Liu, and J. P. Lynch. 2010. *Cdx2* levels modulate intestinal epithelium maturity and Paneth cell development. *Gastroenterology* **140**, 517-528.
9. Cuzzocrea, S., D. P. Riley, A. P. Caputi, and D. Salvemini. 2001. Antioxidant therapy: a new pharmacological approach in shock, inflammation, and ischemia/reperfusion

- injury. *Pharmacol. Rev.* **53**, 135-159.
10. Domon-Dell, C., A. Schneider, V. Moucadel, E. Guerin, D. Guenot, S. Aguillon, I. Duluc, E. Martin, J. Iovanna, J. F. Launay, B. Duclos, M. P. Chenard, C. Meyer, P. Oudet, M. Kedinger, M. P. Gaub, and J. N. Freund. 2003. Cdx1 homeobox gene during human colon cancer progression. *Oncogene* **22**, 7913-7921.
 11. Fischer, J. A., E. Giniger, T. Maniatis, and M. Ptashne. 1988. GAL4 activates transcription in *Drosophila*. *Nature* **332**, 853-856.
 12. Gupta, A., S. F. Rosenberger, and G. T. Bowden. 1999. Increased ROS levels contribute to elevated transcription factor and MAP kinase activities in malignantly progressed mouse keratinocyte cell lines. *Carcinogenesis* **20**, 2063-2073.
 13. Han, K. 1996. An efficient DDAB-mediated transfection of *Drosophila* S2 cells. *Nucleic Acids Res.* **24**, 4362-4363.
 14. Hunt, C. R., J. E. Sim, S. J. Sullivan, T. Featherstone, W. Golden, C. Von Kapp-Herr, R. A. Hock, R. A. Gomez, A. J. Parsian, and D. R. Spitz. 1998. Genomic instability and *catalase* gene amplification induced by chronic exposure to oxidative stress. *Cancer Res.* **58**, 3986-3992.
 15. Hwang, M. S., Y. S. Kim, N. H. Choi, J. H. Park, E. J. Oh, E. J. Kwon, M. Yamaguchi, and M. A. Yoo. 2002. The caudal homeodomain protein activates *Drosophila* *E2F* gene expression. *Nucleic Acids Res.* **30**, 5029-5035.
 16. Jackson, A. L. and L. A. Loeb. 2001. The contribution of endogenous sources of DNA damage to the multiple mutations in cancer. *Mutat. Res.* **477**, 7-21.
 17. Joseph, P., T. K. Muchnok, M. L. Klishis, J. R. Roberts, J. M. Antonini, W. Z. Whong, and T. Ong. 2001. Cadmium-induced cell transformation and tumorigenesis are associated with transcriptional activation of *c-fos*, *c-jun*, and *c-myc* proto-oncogenes: role of cellular calcium and reactive oxygen species. *Toxicol. Sci.* **61**, 295-303.
 18. Kessel, M. and P. Gruss. 1990. Murine developmental control genes. *Science* **249**, 374-379.
 19. Kim, H. S., T. B. Lee, and C. H. Choi. 2001. Down-regulation of *catalase* gene expression in the doxorubicin-resistant AML subline AML-2/DX100. *Biochem Biophys. Res. Commun.* **281**, 109-114.
 20. Kuhnlein, R. P., G. Bronner, H. Taubert, and R. Schuh. 1997. Regulation of *Drosophila* spalt gene expression. *Mech. Dev.* **66**, 107-118.
 21. Lauer, C., A. Volkl, S. Riedl, H. D. Fahimi, and K. Beier. 1999. Impairment of peroxisomal biogenesis in human colon carcinoma. *Carcinogenesis* **20**, 985-989.
 22. Lewis, A. D., I. D. Hickson, C. N. Robson, A. L. Harris, J. D. Hayes, S. A. Griffiths, M. M. Manson, A. E. Hall, J. E. Moss, and C. R. Wolf. 1988. Amplification and increased expression of alpha class glutathione S-transferase-encoding genes associated with resistance to nitrogen mustards. *Proc. Natl. Acad. Sci. USA* **85**, 8511- 8515.
 23. Lewis, M. T. 2000. Homeobox genes in mammary gland development and neoplasia. *Breast Cancer Res.* **2**, 158-169.
 24. Luo, D. and T. A. Rando. 2003. The regulation of *catalase* gene expression in mouse muscle cells is dependent on the CCAAT-binding factor NF-Y. *Biochem Biophys. Res. Commun.* **303**, 609-618.
 25. Mallo, G. V., H. Rechreche, J. M. Frigerio, D. Rocha, A. Zweibaum, M. Lacasa, B. R. Jordan, N. J. Dusetti, J. C. Dagorn, and J. L. Iovanna. 1997. Molecular cloning, sequencing and expression of the mRNA encoding human Cdx1 and Cdx2 homeobox. Down-regulation of Cdx1 and Cdx2 mRNA expression during colorectal carcinogenesis. *Int. J. Cancer* **74**, 35-44.
 26. McGinnis, W. and R. Krumlauf. 1992. Homeobox genes and axial patterning. *Cell* **68**, 283-302.
 27. Michiels, C., M. Raes, O. Toussaint, and J. Remacle. 1994. Importance of Se-glutathione peroxidase, catalase, and Cu/Zn-SOD for cell survival against oxidative stress. *Free Radic. Biol. Med.* **17**, 235-248.
 28. Neno, M., S. Ichimura, K. Mita, O. Yukawa, and I. L. Cartwright. 2001. Regulation of the *catalase* gene promoter by Sp1, CCAAT-recognizing factors, and a WT1/Egr-related factor in hydrogen peroxide-resistant HP100 cells. *Cancer Res.* **61**, 5885-5894.
 29. Park, S. Y., Y.S. Kim, D. J. Yang, and M. A. Yoo. 2004. Transcriptional regulation of the *Drosophila catalase* gene by the DRE/DREF system. *Nucleic Acids Res.* **32**, 1318-1324.
 30. Ren, P., D. G. Silberg, and A. E. Sirica. 2000. Expression of an intestine-specific transcription factor (CDX1) in intestinal metaplasia and in subsequently developed intestinal type of cholangiocarcinoma in rat liver. *Am. J. Pathol.* **156**, 621-627.
 31. Rhee, S. G. 1999. Redox signaling: hydrogen peroxide as intracellular messenger. *Exper. Mol. Med.* **35**, 53-59.
 32. Sanders, L. M., C. E. Henderson, M. Y. Hong, R. Barhoumi, R. C. Burghardt, N. Wang, C. M. Spinka, R. J. Carroll, N. D. Turner, R. S. Chapkin, and J. R. Lupton. 2004. An increase in reactive oxygen species by dietary fish oil coupled with the attenuation of antioxidant defenses by dietary pectin enhances rat colonocyte apoptosis. *J. Nutr.* **134**, 3233-3238.
 33. Silberg, D. G., G. P. Swain, E. R. Suh, and P. G. Traber. 2000. Cdx1 and Cdx2 expression during intestinal development. *Gastroenterology* **119**, 961-971.
 34. Skrzydlewska, E., S. Sulkowski, M. Koda, B. Zalewski, L. Kanczuga-Koda, and M. Sulkowska. 2005. Lipid peroxidation and antioxidant status in colorectal cancer. *World J. Gastroenterol.* **11**, 403-406.
 35. Tsutsumishita, Y., T. Onda, K. Okada, M. Takeda, H. Endou, S. Futaki, and M. Niwa. 1998. Involvement of H₂O₂ production in cisplatin-induced nephrotoxicity. *Biochem Biophys. Res. Commun.* **242**, 310-312.
 36. Van Remmen, H., M. D. Williams, H. Yang, C. A. Walter, and A. Richardson. 1998. Analysis of the transcriptional activity of the 5'-flanking region of the rat *catalase* gene in transiently transfected cells and in transgenic mice. *J. Cell Physiol.* **174**, 18-26.
 37. Wang, W. W., D. L. Smith, and S. D. Zucker. 2004. Bilirubin inhibits iNOS expression and NO production in response to endotoxin in rats. *Hepatology* **40**, 424-433.

38. Werling, R. W., H. Yaziji, C. E. Bacchi, and A. M. Gown. 2003. CDX2, a highly sensitive and specific marker of adenocarcinomas of intestinal origin: an immunohistochemical survey of 476 primary and metastatic carcinomas. *Am. J. Surg. Pathol.* **27**, 303-310.
39. de Winter, B. Y., L. van Nassauw, J. G. de Man, F. de Jonge, A. J. Bredenoord, T. C. Seerden, A. G. Herman, J. P. Timmermans, and P. A. Pelckmans. 2005. Role of oxidative stress in the pathogenesis of septic ileus in mice. *Neurogastroenterol. Motil.* **17**, 251-261.
40. Wong, N. A., M. P. Britton, G. S. Choi, T. K. Stanton, D. C. Bicknell, J. L. Wilding, and W. F. Bodmer. 2004. Loss of CDX1 expression in colorectal carcinoma: promoter methylation, mutation, and loss of heterozygosity analyses of 37 cell lines. *Proc. Natl. Acad. Sci. USA* **101**, 574-579.
41. Wu, L. H. and J. A. Lengyel. 1998. Role of caudal in hindgut specification and gastrulation suggests homology between *Drosophila* amnioproctodeal invagination and vertebrate blastopore. *Development* **125**, 2433-2442.
42. Zhu, J. W., B. M. Yu, Y. B. Ji, M. H. Zheng, and D. H. Li. 2002. Upregulation of vascular endothelial growth factor by hydrogen peroxide in human colon cancer. *World J. Gastroenterol.* **8**, 153-157.

초록 : 초파리 대장조직과 인간 대장암세포주에서의 caudal 단백질에 의한 catalase 발현 조절

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Caudal homeodomain 단백질은 초파리에서 사람에게 이르기까지 보존되어 있으며, 장조직의 발생과 유지에 필수적인 역할을 한다. CDX1과 CDX2의 발현 감소 혹은 소실이 대장암과 연관되어 있음이 잘 알려져 있다. 대장암 발생은 산화성 스트레스와 밀접한 관련이 있으며, 대장암 조직에서 항산화효소인 catalase의 발현이 감소되어 있다. 하지만 그 분자적 기전은 잘 밝혀져 있지 않다. 본 연구에서는 초파리와 사람의 caudal homeodomain 단백질들이 catalase의 발현을 조절하는 것을 보여준다. 초파리 caudal heterozygous 돌연변이체의 대장조직에서 catalase의 발현이 감소되어 있고 ROS 생성이 증가되어 있음을 관찰하였다. 그리고 caudal 유전자의 과발현에 의해 catalase promoter의 활성과 mRNA 수준이 각각 증가함을 확인하였다. 또한 사람의 대장암 세포주인 HCT116 세포에서 CDX1과 CDX2가 catalase promoter의 활성과 단백질 수준에서 catalase를 상향 조절함을 관찰하였다. 이러한 결과들은 CDX1과 CDX2가 catalase 발현 조절을 통하여 장의 항상성 유지와 암발생 과정에 관여함을 시사한다.