# Cytotoxic Sesquilignans from the Roots of Saururus chinensis 

Young-Won Chin, Xing-Fu Cai, Kyung-Seop Ahn, Hyeong-Kyu Lee, and Sei-Ryang Oh ${ }^{*}$<br>Bio-Therapeutics Research Institute, Korea Research Institute of Bioscience and Biotechnology, Ochangeup, Cheongwongun, ChungBuk 363-883, Korea. *E-mail: seiryang@kribb.re.kr Received April 26, 2010, Accepted May 25, 2010

Key Words: Saururus chinensis, Sesquilignans, Cytotoxicity, Saururaceae

Saururus chinensis Hort. ex Loudon (Saururaceae) is a perennial herb distributed in China and Korea, and has been used as a folk medicine for the treatment of edema, gonorrhea, jaundice, pneumonia, and several inflammatory diseases in Korea. ${ }^{1}$ Previous studies of $S$. chinensis reported the occurrence of lignans, ${ }^{2-9}$ aristolactams, ${ }^{10}$ flavonoids, ${ }^{11}$ and furanoditerpenes, ${ }^{12}$ and a wide range of biological activities including antioxidant activity, ${ }^{5}$ hepatoprotective activity, ${ }^{6}$ cytotoxic activity, ${ }^{13-18}$ antiinflammatory activity, ${ }^{19-22}$ anti-atherogenic activity, ${ }^{21,23}$ and immunosuppressive activity. ${ }^{24}$ This paper reports the structures elucidation of the two new lignans and six known compounds, along with their cytotoxicity.

Compound $\mathbf{1}$ was obtained as a colorless powder and its molecular formula was determined to be $\mathrm{C}_{31} \mathrm{H}_{38} \mathrm{O}_{8}$, based on the [M-H] peak at $m / z 537.2471$ (calcd 537.2488) in the HRESIMS. The ${ }^{1} \mathrm{H}-\mathrm{NMR}$ spectroscopic data of compound $\mathbf{1}$ indicated the presence of a tetrahydrofuran-type lignan unit, as judged from the signals for two methines at $\delta_{\mathrm{H}} 2.34$ ( $\mathrm{H}-8$ and $\mathrm{H}-8^{\prime}$ ), for oxymethines at $\delta_{\mathrm{H}} 5.65$ (H-7 and $\mathrm{H}-7^{\prime}$ ), and for methyl groups at $\delta_{\mathrm{H}} 0.78(\mathrm{H}-9)$ and $0.74\left(\mathrm{H}-9{ }^{\prime}\right)$, as well as the signals at $\delta_{\mathrm{H}}$ 7.20 (d, $J=1.7 \mathrm{~Hz}, \mathrm{H}-2$ ), $7.30(J=8.2 \mathrm{~Hz}, \mathrm{H}-5), 7.36$ (br d, $J=$ $8.2 \mathrm{~Hz}, \mathrm{H}-6), 7.19$ (d, $\left.J=1.7 \mathrm{~Hz}, \mathrm{H}-2^{\prime}\right), 7.34(\mathrm{~d}, J=8.2 \mathrm{~Hz}$, H-5'), and 7.09 (dd, $\left.J=8.2,1.7 \mathrm{~Hz}, \mathrm{H}^{\prime} 6^{\prime}\right)$ corresponding to two 1,3,4-trisubstitued benzene rings. ${ }^{4,25}$ An additional phenylpropanoid unit was observed in compound $\mathbf{1}$ from the proton signals for two oxymethines at $\delta_{\mathrm{H}} 4.95\left(\mathrm{H}-8^{\prime \prime}\right)$, and 5.42 (H-7"),

7",8"
1 erythro

Figure 1. Structures of compounds 1-2.



Figure 2. Key ${ }^{1} \mathrm{H}-{ }^{1} \mathrm{H} \operatorname{COSY}(\boldsymbol{\sim})$ and $\mathrm{HMBC}(\mathrm{H} \longrightarrow \mathrm{C})$ and $\operatorname{NOESY}(\longleftrightarrow)$ correlations of compound $\mathbf{1}$.
for a methyl at $\delta_{\mathrm{H}} 1.60\left(\mathrm{H}-9{ }^{\prime \prime}\right)$ and for a 1,3,4-trisubstituted benzene ring at $\delta_{\mathrm{H}} 7.58\left(\mathrm{~d}, J=1.6 \mathrm{~Hz}, \mathrm{H}-2^{\prime \prime}\right), 7.01(\mathrm{~d}, J=8.2$ $\left.\mathrm{Hz}, \mathrm{H}-55^{\prime \prime}\right)$, and 7.07 (dd, $J=8.2,1.6 \mathrm{~Hz}, \mathrm{H}-6$ "). ${ }^{4}$ The location of the phenylpropanoid group was confirmed by the HMBC correlation between $\mathrm{H}-8^{\prime \prime}\left(\delta_{\mathrm{H}} 4.95\right)$ and C-4' ( $\delta_{\mathrm{C}} 147.5$ ), suggesting this phenylpropanoid is attached to $\mathrm{C}-4$ ' on the tetrahydro-furan-type lignan moiety through an ether linkage. The relative stereochemistry of the tetrahydrofuran ring in compound $\mathbf{1}$ was established by the observed NOE correlations of H-9 with H-8', $\mathrm{H}-2$, and $\mathrm{H}-6$ as well as $\mathrm{H}-9$ ' with $\mathrm{H}-8, \mathrm{H}-2^{\prime}$, and $\mathrm{H}-6$ ', indicating the 7,8 -cis- $8,8^{\prime}$-trans- $7^{\prime}, 8^{\prime}$-cis configuration. ${ }^{4,26}$ In addition, the chemical shifts of C-9" ( $\delta_{\mathrm{C}} 15.3$ ), C-7" ( $\delta_{\mathrm{C}} 75.7$ ), and C-8" ( $\delta_{\mathrm{C}}$ 81.1 ), and the coupling constant ( $J=3.4 \mathrm{~Hz}$ ) of $\mathrm{H}-7$ " was supportive of the relative configuration of $\mathrm{C}-7{ }^{\prime \prime}$ and $\mathrm{C}-8$ " as being erythro. ${ }^{4,26}$ Futhermore, the positive Cotton effect at 231 nm enabled to assign the configuration of C-7" and $8^{\prime \prime}$ as $R$ and $S$, respectively. ${ }^{27}$ Thus, the structure of compound $\mathbf{1}$ was determined as 7 " $R, 8$ " $S$-saucerneol, a diastereomer of (-)-saucerneol (3).

Saucerneol J (2) had a molecular formula $\left(\mathrm{C}_{30} \mathrm{H}_{36} \mathrm{O}_{8}\right)$ and exhibited a close resemblance to compound $\mathbf{1}$ in their ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR spectroscopic data except for the presence of three methoxy groups. There were differences in the chemical shifts and coupling constants of $\mathrm{H}-7$ ' in compounds 2 ( $\delta_{\mathrm{H}} 4.54, J=9.3$ Hz ) and $\mathbf{1}\left(\delta_{\mathrm{H}} 5.65, J=6.4 \mathrm{~Hz}\right)$, indicating the opposite configuration at $\mathrm{C}-\mathbf{7}^{\prime}$ position in compound $\mathbf{2}$ compared to compound $\mathbf{1}$.


2 erythro



Table 1. ${ }^{1} \mathrm{H}$-and ${ }^{13} \mathrm{C}$-NMR data ( $\delta$ ) of 1-2 $\left(\text { Pyridine- } d_{5}\right)^{a}$

| position | 1 |  | 2 |  |
| :---: | :---: | :---: | :---: | :---: |
|  | $\delta_{\mathrm{H}}$ | $\delta_{C}$ | $\delta_{\mathrm{H}}$ | $\delta_{\text {c }}$ |
| 1 | - | 136.5 (s) | - | 135.6 (s) |
| 2 | 7.20 d, 1.7 | 111.6 (d) | 7.25 d, 1.7 | 112.1 (d) |
| 3 | d | 151.5 (s) | - | 151.5 (s) |
| 4 |  | 147.6 (s) | - | 147.8 (s) |
| 5 | 7.30 d, 8.2 | 117.5 (d) | 7.28 d, 8.1 | 116.5 (d) |
| 6 | 7.36 br d, 8.2 | 120.1 (d) | 7.31 br d, 8.1 | 120.6 (d) |
| 7 | 5.65 d, 6.4 | 84.6 (d) | 5.30 d, 8.6 | 82.7 (d) |
| 8 | 2.34 ddq, 12.8, 6.4, 6.4 | 44.5 (d) | 2.24 ddq, 8.6, 7.0, 6.5 | 45.7 (d) |
| 9 | 0.78 d, 6.4 | 15.3 (q) | 0.73 d, 7.0 | 14.9 (q) |
| $1^{\prime}$ | - | 133.8 (s) | - | 133.2 (s) |
| $2^{\prime}$ | 7.19 d, 1.7 | 112.2 (d) | $7.42 \mathrm{~d}, 1.7$ | 111.2 (d) |
| $3^{\prime}$ | - | 148.9 (s) | - | 149.2 (s) |
| $4^{\prime}$ | - | 147.5 (s) | - | 148.5 (s) |
| $5 '$ | 7.34 d, 8.2 | 116.6 (d) | 7.30 d, 8.1 | 116.1 (d) |
| $6{ }^{\prime}$ | $7.09 \mathrm{dd}, 8.2,1.7$ | 119.8 (d) | $7.32 \mathrm{dd}, 8.1,1.7$ | 120.5 (d) |
| 71 | 5.65 d, 6.4 | 84.3 (d) | 4.54 d, 9.3 | 87.4 (d) |
| 8' | 2.34 ddq, 12.8, 6.4, 6.4 | 44.5 (d) | $1.93 \mathrm{ddq}, 9.3,6.5,6.5$ | 47.9 (d) |
| $9{ }^{\prime}$ | 0.74 d, 6.4 | 15.3 (q) | 1.05 d, 6.5 | 14.5 (q) |
| $1{ }^{\prime \prime}$ | - | 136.8 (s) | - | 135.1 (s) |
| 2 " | 7.58 d, 1.6 | 112.2 (d) | 7.59 d, 1.6 | 111.3 (d) |
| $3 "$ | - | 150.2 (s) | - | 148.9 (s) |
| $4 "$ | - | 149.5 (s) | - | 147.7 (s) |
| 5" | 7.01 d, 8.2 | 112.6 (d) | 7.31 d, 8.1 | 115.8 (d) |
| $6 "$ | $7.07 \mathrm{dd}, 8.2,1.6$ | 120.3 (d) | $7.29 \mathrm{dd}, 8.1,1.6$ | 120.7 (d) |
| $7{ }^{\prime \prime}$ | 5.42 d, 3.4 | 75.7 (d) | $5.42 \mathrm{~d}, 3.9$ | 74.8 (d) |
| 8" | $4.95 \mathrm{dq}, 6.2,3.4$ | 81.1 (d) | 4.90 dq, 6.2, 3.9 | 80.3 (d) |
| $9 "$ | 1.60 d, 6.2 | 15.3 (q) | $1.60 \mathrm{~d}, 6.2$ | 14.3 (q) |
| $3-\mathrm{OCH}_{3}$ | 3.84 s | 56.5 (q) | 3.82 s | 56.4 (q) |
| $3{ }^{\prime}-\mathrm{OCH}_{3}$ | 3.85 s | 56.5 (q) | 3.86 s | 56.4 (q) |
| $3 \mathrm{H}-\mathrm{OCH}_{3}$ | 3.82 s | 56.3 (q) | 3.80 s | 56.3 (q) |
| $4 \mathrm{H}-\mathrm{OCH}_{3}$ | 3.77 s | 56.4 (q) | - | - |

${ }^{a}$ Assignments were based on ${ }^{1} \mathrm{H}-{ }^{1} \mathrm{H}$ COSY, DEPT, HMQC and HMBC spectra.

The CD spectroscopic data exhibited the positive Cotton effect at 232 nm in the same manner. Therefore, the structure of compound $\mathbf{2}$ was confirmed to be 7 '-epi-7" $R, 8$ "S-4"-demethylsaucerneol.

The known compounds were in good agreement with previously reported NMR data and were consequently identified as $(-)$-saucerneol (3) with a $[\alpha]_{\mathrm{D}}$ value of $-71.1(c 0.1 \mathrm{MeOH})$ $\left[\right.$ Lit. $\left.[\alpha]_{\mathrm{D}}-91.8\right],{ }^{25}$ threo,erythro-manassantin A (4), ${ }^{16} 4-\mathrm{O}$ demehylmanassantin $\mathrm{B}(\mathbf{5})$, ${ }^{18}$ erythro, erythro-manassantin A (6), ${ }^{16}$ manassantin B (7) ${ }^{28}$ and manassantin A (8). ${ }^{17}$

All the compounds isolated were evaluated against HL-60 (human promyelocytic leukemia) cells. Compounds 1-8 exhibited cytotoxicity ( $\mathrm{IC}_{50}, 0.5,7.1,3.3,5.2,3.6,2.3,8.5$ and 0.8 $\mu \mathrm{M}$, respectively) against HL-60 cell lines (camptothecin, $\mathrm{IC}_{50}$ $0.8 \mu \mathrm{M})$.

## Experimental Section

General experimental procedures. Melting points were determined on a Kofler micro-hotstage (uncorrected). Optical rota-
tions were measured on a JASCO P-1020 polarimeter. UV spectra were measured on a Shimadzu UV-1601 UV-visible. CD spectroscopic data were obtained from JASCO-720 CD spectrometer. The NMR spectra were recorded on a Varian Unity 400 FT-NMR spectrometers with the tetramethylsilane as an internal standard. Chemical shifts are presented in ppm. HRESIMS were measured on a Waters Q-Tof Premier mass spectrometer. Column chromatography (CC) was performed on silica gel (70-230 and 230-400 mesh, Merck), reverse-phase C18 gel ( $40 \mu \mathrm{~m}$, Nacalai Inc., Japan). Thin layer chromatography (TLC) was performed on Kieselgel $60 \mathrm{~F}_{254}$ (Merck) or RP-18 F 254 (Merck) plates. Spots were visualized by spraying $10 \%$ aqueous $\mathrm{H}_{2} \mathrm{SO}_{4}$ solution on the plates and heating them for 5 min .

Plant material. The roots of Saururus chinensis was collected at Jeju (Korea) in July 2008 and dried at room temperature. A voucher specimen (00250) is deposited at the Plant Extract Bank, Korea Research Institute of Bioscience and Biotechnology, Daejeon, Korea.

Extraction and isolation. The dried roots of S. chinensis
( 6.5 kg ) was extracted with MeOH at room temperature ( $3 \times$ 20 L ) to obtain 0.65 kg of the solid extract. The MeOH extract was suspended in $\mathrm{H}_{2} \mathrm{O}$ and extracted with EtOAc $(3 \times 3 \mathrm{~L})$ to give the EtOAc-soluble fractions ( 85 g ). The EtOAc-soluble fraction ( 84 g ) was chromatographed on a silica gel column eluted with a stepwise gradient of hexane and EtOAc to yield 14 fractions (fr. SC1-SC14). Fr. SC12 (1.2 g) was chromatographed on a RP C-18 column ( $\mathrm{MeOH} / \mathrm{H}_{2} \mathrm{O}, 7: 3$ ) to yield 19 sub-fractions (fr. SC12-1-SC12-19). Fr. SC12-15 (0.14 g) was chromatographed on a RP C-18 column $\left(\mathrm{MeOH} / \mathrm{H}_{2} \mathrm{O}, 2: 1\right)$ to give compounds $\mathbf{1}(13.2 \mathrm{mg})$ and $\mathbf{3}(98.1 \mathrm{mg})$. Fr. SC12-19 $(0.103 \mathrm{~g})$ was chromatographed on a RP C-18 column $(\mathrm{MeOH} /$ $\left.\mathrm{H}_{2} \mathrm{O}, 2: 1\right)$ to give compound $4(17.8 \mathrm{mg})$. Fr. SC14 ( 0.88 g ) was chromatographed on a RPC-18 column ( $\mathrm{MeOH} / \mathrm{H}_{2} \mathrm{O}, 8: 2$ ) to yield seven sub-fractions (Fr. SC14-1-SC14-7). Fr. SC14-3 $(0.12 \mathrm{~g})$ was chromatographed on a RP C-18 column ( $\mathrm{MeOH} /$ $\mathrm{H}_{2} \mathrm{O}, 3: 2$ ) to give compound $2(5.3 \mathrm{mg})$. Fr. SC14-6 ( 0.36 g ) was chromatographed on a RP C-18 column ( $\mathrm{MeOH} / \mathrm{H}_{2} \mathrm{O}, 3: 2$ ) to give compounds $\mathbf{5}(76.7 \mathrm{mg}), \mathbf{6}(5.3 \mathrm{mg}), 7(18.3 \mathrm{mg})$, and 8 ( 43.3 mg ).
erythro-Saucerneol (1): Colorless powder, $\mathrm{mp} 85-86^{\circ} \mathrm{C} .[\alpha]_{\mathrm{D}}^{25}$ -18 (c 0.1, MeOH). UV $\lambda_{\text {max }}(\mathrm{MeOH}) \mathrm{nm}(\log \varepsilon): 206$ (3.74), 284 (2.94). ${ }^{1} \mathrm{H}$ - and ${ }^{13} \mathrm{C}$-NMR data see Tables 1. HRESIMS $m / z 537.2471$ [M-H] (Calcd for $\mathrm{C}_{31} \mathrm{H}_{37} \mathrm{O}_{8}: 537.2488$ ). CD (c $0.0004 \mathrm{MeOH}):[\theta]_{231}+12,075$.
Saucerneol J(2): Colorless powder, mp $75-76^{\circ} \mathrm{C} .[\alpha]_{\mathrm{D}}^{25}-10$ ( $c 0.1, \mathrm{MeOH}) . \mathrm{UV} \lambda_{\max }(\mathrm{MeOH}) \mathrm{nm}(\log \varepsilon): 206$ (3.81), 282 (2.67). ${ }^{1} \mathrm{H}$ - and ${ }^{13} \mathrm{C}-$ NMR data see Tables 1 and 2. HRESIMS $m / z 523.2310[\mathrm{M}-\mathrm{H}]$ (Calcd for $\mathrm{C}_{30} \mathrm{H}_{35} \mathrm{O}_{8}: 523.2332$ ). CD (c $0.0006 \mathrm{MeOH}):[\theta]_{232}+18,121$.
Cytotoxicity evaluation. All the isolates were assessed with the HL-60 (human promyelocytic leukemia) cells according to the established protocol. ${ }^{29}$

Acknowledgments. This research was supported by a grant of KRIBB Research Initiative Program (KGS2241012).

## References and Notes

1. Chung, B. S.; Shin, M. G. Dictionary of Korean Folk Medicine; Young Lim Publishing: Seoul, 1990; pp 813-814.
2. Ahn, B. T.; Lee, S.; Lee, S. B.; Lee, E. S.; Kim, J. G.; Bok, S. H.; Jeong, T. S. J. Nat. Prod. 2001, 64, 1562.
3. Hwang, B. Y.; Lee, J. H.; Jung, H. S.; Kim, K. S.; Nam, J. B.; Hong, Y. S.; Paik, S. G.; Lee, J. J. Planta Med. 2003, 69, 1096.
4. Hwang, B. Y.; Lee, J. H.; Nam, J. B.; Hong, Y. S.; Lee, J. J. Phytochemistry 2003, 64, 765.
5. Lee, W. S.; Baek, Y. I.; Kim, J. R.; Cho, K. H.; Sok, D. E.; Jeong, T. S. Bioorg. Med. Chem. Lett. 2004, 14, 5623.
6. Sung, S. H.; Kim, Y. C. J. Nat. Prod. 2000, 63, 1019.
7. Sung, S. H.; Lee, E. J.; Cho, J. H.; Kim, H. S.; Kim, Y. C. Biol. Pharm. Bull. 2000, 23, 666.
8. Seo, C. S.; Zheng, M. S.; Woo, M. H.; Lee, C. S.; Lee, S. H.; Jeong, B. S.; Chang, H. W.; Jahng, Y.; Lee, E. S.; Son, J. K. J. Nat. Prod. 2008, 71, 1771.
9. Sung, S. H. Fitoterapia 2006, 77, 487.
10. Kim, S. R.; Sung, S. H.; Kang, S. Y.; Koo, K. A.; Kim, S. H.; Ma, C. J.; Lee, H. S.; Park, M. J.; Kim, Y. C. Planta Med. 2004, 70, 391.
11. Kang, T. H.; Cho, H.; Oh, H.; Sohn, D. H.; Kim, Y. C. Fitoterapia 2005, 76, 115.
12. Hwang, B. Y.; Lee, J. H.; Nam, J. B.; Kim, H. S.; Hong, Y. S.; Lee, J. J. J. Nat. Prod. 2002, 65, 616.
13. Seo, B. R.; Lee, K. W.; Ha, J.; Park, H. J.; Choi, J. W.; Lee, K. T. Carcinogenesis 2004, 25, 1387.
14. Seo, B. R.; Yoo, C. B.; Park, H. J.; Choi, J. W.; Seo, K.; Choi, S. K.; Lee, K. T. Biol. Pharm. Bull. 2004, 27, 1594.
15. Song, S. Y.; Lee, I.; Park, C.; Lee, H.; Hahm, J. C.; Kang, W. K. Int. J. Mol. Med. 2005, 16, 517.
16. Hahm, J. C.; Lee, I. K.; Kang, W. K.; Kim, S. U.; Ahn, Y. J. Planta Med. 2005, 71, 464.
17. Hossain, C. F.; Kim, Y. P.; Baerson, S. R.; Zhang, L.; Bruick, R. K.; Mohammed, K. A.; Agarwal, A. K.; Nagle, D. G.; Zhou, Y. D. Biochem. Biophys. Res. Commun. 2005, 333, 1026.
18. Hodges, T. W.; Hossain, C. F.; Kim, Y. P.; Zhou, Y. D.; Nagle, D. G. J. Nat. Prod. 2004, 67, 767.
19. Hwang, B. Y.; Lee, J. H.; Koo, T. H.; Kim, H. S.; Hong, Y. S.; Ro, J. S.; Lee, K. S.; Lee, J. J. Planta Med. 2002, 68, 101.
20. Lee, A. K.; Sung, S. H.; Kim, Y. C.; Kim, S. G. Br. J. Pharmacol. 2003, 139, 11.
21. Rho, M. C.; Kwon, O. E.; Kim, K.; Lee, S. W.; Chung, M. Y.; Kim, Y. H.; Hayashi, M.; Lee, H. S.; Kim, Y. K. Planta Med. 2003, 69, 1147.
22. Moon, T. C.; Kim, J. C.; Song, S. E.; Suh, S. J.; Seo, C. S.; Kim, Y. K.; Jin, M.; Yang, J. H.; Son, J. K.; Jahng, Y.; Kim, C. H.; Chang, H. W. Int. Immunopharmacol. 2008, 8, 1395.
23. Lee, W. S.; Lee, D. W.; Baek, Y. I.; An, S. J.; Cho, K. H.; Choi, Y. K.; Kim, H. C.; Park, H. Y.; Bae, K. H.; Jeong, T. S. Bioorg. Med. Chem. Lett. 2004, 14, 3109.
24. Park, S. Y.; Lee, S. H.; Choi, W. H.; Koh, E. M.; Seo, J. H.; Ryu, S. Y.; Kim, Y. S.; Kwon, D. Y.; Koh, W. S. Planta Med. 2007, 73, 674.
25. Hanessian, S.; Reddy, G. J.; Chahal, N. Org. Lett. 2006, 8, 5477.
26. Tofern, B.; Jenett-Siems, K.; Siems, K.; Jakupovic, J.; Eich, E.; Phytochemistry 2000, 53, 119.
27. Huo, C.; Liang, H.; Zhao, Y.; Wang, B.; Zhang, Q. Phytochemistry 2008, 69, 788.
28. Park, H. J.; Kim, R. G.; Seo, B. R.; Ha, J.; Ahn, B. T.; Bok, S. H.; Lee, Y. S.; Kim, H. J.; Lee, K. T. Planta Med. 2003, 69, 947.
29. Park, B. Y.; Oh, S. R.; Ahn, K. S.; Kwon, O. K.; Lee, H. K. Int. Imтипорharmcol. 2008, 8, 967.
