

Effect of *Epimedium Koreanum* on the 2-Bromopropane Induced-Reproductive Damage

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Abstract : *Epimedium koreanum* nakai (EKN) has been used for treatment of reproductive disorders. In this study, we evaluated the effects of EKN water extract on the 2-bromopropane (2-BP) induced reproductive dysfunction. The daily sperm production (DSP), sperm counts and histological changes of reproductive organs were investigated after 4 weeks of EKN administration in 2-BP induced reproductive damaged rats. Although the weights of epididymis and seminal vesicle were increased dose-dependently, they were lower than control group. DSP of EKN-dosing groups were increased compared to control group. In EKN-dosing groups, the number of degenerative seminiferous tubules and the number of epididymal tubules showing epithelial cell vacuolation and decreased spermatozoa in the lumen were significantly (p < 0.01) decreased compared to control group dose-dependently. Significant (P < 0.05 or P < 0.01) increases of the number of sertoli cells, spermatogonia, patchytene spermatocytes, round spermatids and elongated spermatids were observed in EKN-dosing groups compared control group. Consequently, based on the results, EKN water extract could treat 2-BP induced reproductive damages dramatically.

Key words: Epimedium koreanum, reproductive disorder, 2-bromopropane, rat.

Introduction

2-bromopropane (2-BP) is well known toxic material as cause of amenorrhea, oligozoospermia, azoospermia and anemia (16) as well as immunotoxic potentials (12,14). It can also decrease spermatogenesis by adversely effecting spermatogonia followed by depletion of spermatocytes, spermatids and spermatozoa with subsequent testicular atrophy (25,33).

Epimedium Koreanum Nakai (EKN) has been traditionally used as diuretic, tonic and antirheumatic and for improving of impotency in the oriental medicine (20). It also has other effects such as: anti-oxidant (19); anti-cancer in human lung cancer cells and prostate carcinoma (7); anti-osteoporosis by stimulating proliferation of bone marrow stromal cells (2); stimulation of erectile function of penis (18,20,35); anti-depressant by decreasing serum corticotropin-releasing factor and cortisol levels in rats (27).

EKN is known to contain a lot of medicinal components, such as prenylflavonol glycoside (26), icariin (31,32), magnof-

lorine (4), chlorogenic acid (30), korepimedoside A and B (34), hyperin (29) and etc. Especially, many researchers have been interested in icariin as main component of sexual function and antiosteoporosis (3,6,18,24,35,40).

Although EKN has shown medical effects for the aforementioned disorders and some of the effects were pharmaceutically proven, many of pharmacological actions and components of the EKN remain to be systemically determined.

Therefore, we performed this study to investigate the effects of EKN water extract on reproductive failure induced by 2-BP in Sprague-Dawley rats.

Materials and Methods

Preparation of materials

We obtained dried leaves of EKN from herbal market in Daegu, Korea. The extraction procedure was according to the method of *Kim et al* (15). Briefly, ground powder of dried EKN leaves were mixed with distilled water (100 g : 1000 ml) and the mixture was shaken with 140 rpm at 37°C water bath for 4 h. It was centrifuged at 3000 rpm, 4°C for 20 min, and the supernatant was filtered by No. 2 filter paper. The filtrate

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was concentrated by a rotator evaporator, lyophilized (19.5%) and stored at -20° C until use.

2-BP was obtained from Sigma (MO, USA) as liquid ((CH_{3})₂ CHBr, 99%). Olive oil was obtained from Junsei Chemical Co. Ltd (Japan). 2-BP was homogenized in olive oil (2-BP : olive oil = 1 : 1, v/v) and was administered 1,355 mg/kg subcutaneously. Homogenized 2-BP in olive oil was stored in stopper tube at room temperature.

Animals

Thirty-two 8 week-old male Sprague-Dawley rats (Crj/Bgi: CD(SD)IGS: 236 ± 21 g body weight) were obtained from Oriental Bio (Seoul, Korea) and used after acclimatization for 1 week. The rats were housed in a room with humidity of $50 \pm 10\%$ at $20 \pm 2^{\circ}$ C, under 12h-intervals of light and dark cycle, and food and water were given *ad libitum*.

Experimental design

All experimental procedures were conducted according to the Guide for the Care and Use of Laboratory Animals of the Korea Food and Drug Administration (Seoul, 2005). All rats were injected 2-BP at dose of 1,355 mg/kg subcutaneously once a day for 28 days. In the day after last injection of 2-BP, the rats were divided into 3 EKN-dosing groups by body weights: EKN-10 (10 mg/kg EKN-dosing group), EKN-100 (100 mg/kg EKN-dosing group), EKN-500 (500 mg/kg EKNdosing group) and control group in which each of 8 consisted. Then the test article was administered to the 3-EKN-dosing groups *per os* using a gastric gavage once a day for 28 days while control group was given with DW in the same manner instead of the test article.

Sample collection and histological examination

The day after last administration of the test article, all rats were anesthetized and bled via caudal vena cava. Blood samples were subjected to hematological analysis by an automatic analyzer (MS9-5, Japan). Organ samples were dissected, weighed and fixed with bouin solution or 10% neutral buffered formaldehyde. All of the samples were embedded, sectioned ($3\sim4 \mu m$) and stained with hematoxylin and eosin, and observed under light microscope. The right testis and epididymis were stored at -20° C and used for calculation of DSP and total sperm count, respectively.

DSP of the right testis and Sperm count of the right cauda epididymis

DSP was examined with the right testis according to the methods of Blazak *et al* (1). Briefly, frozen testis was thawed on ice, weighed after decapsulation, and homogenized in 50 ml of Hank's solution. The homogenate was allowed to settle down tissue debris for 1 min, and the supernatant sperm was decanted to a new tube. The sperm solution was evenly suspended and 10 ml-aliquot of the solution was collected into each glass vial and stored on ice. After complete mixing, the number of sperm heads was counted with a hemocytometer.

The number of produced sperms per gram of testicular tissue was calculated according to the following formula: the average count of sperm heads in four chambers × square factor [5] × hemocytometer factor $[10^4]$ × dilution factor [50] divided by testis weight [g] and the time of 6.1 days (28) in which the sperm was undamaged by homogenization after spermatogenesis.

To count the sperm number in the cauda epididymis, 0.5 ml of homogenate of right cauda epididymis was diluted with 4.5 ml of Hank's solution. The total number of sperm heads was counted using a hemocytometer, and the mean of three calculations per gram of cauda epididymis was calculated (28,37).

Histomorphometrical examination

The number of seminiferous tubules showing atrophic changes or containing vacuolated sertoli cells in testis, the number of epididymal tubules showing oligospermatozoa or aspermatozoa, and the number of epididymal tubules showing epithelial cell vacuolation were calculated as the number per 100 tubules, respectively. The histomorphometry was conducted using automated image analysis (DMI-300 Image Processing; DMI, Korea) under magnification 200 of microscopy (Nikon, Japan) at 5 fields (n = 5), respectively.

Spermatogenesis in seminiferous tubules

The number of sertoli cells, spermatogonia (Type A and B), preleptotene spermatocytes, leptotene spermatocytes, zygote spermatocytes, pachytene spermatocytes, round and elongated spermatids were counted at stages II-III, V, VII and XII in a total of 10 round or ovoid seminiferous tubules (number/tubules) as previously reported (21). These stages were chosen as the representative to include morphologically identifiable stages. Spermatogenic staging evaluation of testis was performed according to the previously described method (5).

Statistics

Multiple comparison tests (Mann-Whitney U-Wilcoxon Rank Sum W test) for different dosing groups were conducted in all data. Statistical analyses were conducted using SPSS for Windows (Release 10.0.7., SPSS Inc., USA). The values significantly different from control group were indicated at P < 0.05, P < 0.01

Results

Organ weight

After administration of EKN extract for 28 days in reproductive damaged rats induced by 2-BP, the absolute spleen weight in EKN-500 group was significantly (P < 0.05) decreased compared to control group but no significant change was observed in relative weight. The kidney weight in EKN-100 group was significantly (P < 0.05) increased compared to control group but no significant change was observed in other groups. The liver weight was not affected by 2-BP (data not shown). In EKN-10 and EKN-100 groups, not only the both absolute epididymis weights were significantly (P < 0.05) decreased but the both absolute and relative seminal vesicle weights were significantly (P < 0.01 or P < 0.05) decreased compared to control group. The weights of epididymis and seminal vesicle were increased dose-dependently in EKN-dosing groups. No significant change was observed in testis and prostate (Table 1).

Hematological and serum chemistry exam

The platelet and white blood cell (WBC) of EKN-dosing groups were significantly (P < 0.01 or P < 0.05) decreased, respectively, and mean corpuscular volume (MCV) of EKN-500 group was decreased significantly (P < 0.01) compared to control group. There was no significant change in red blood cell (RBC), hemoglobin (Hb), packed cell volume (PCV), mean corpuscular hemoglobin (MCH) and mean corpuscular hemoglobin concentration (MCHC) (data not shown).

DSP and sperm count changes

The testis DSPs of EKN-dosing groups were increased compared to control group. Especially, the DSP in EKN-10 and EKN-100 groups was significantly (P < 0.05) increased compared to control group. The sperm count of the right cauda epididymis in EKN-10 and EKN-500 groups was increased while EKN-100 group was decreased compared to control group. Sperm count in EKN-500 group was significantly (P < 0.01) increased compared to control group (Table 2).

Histological changes

The number of degenerative seminiferous tubules and the number of epididymal tubules showing epithelial cell vacuolation and decreased spermatozoa in the lumen in EKN-dosing groups were significantly (p < 0.01) decreased compared to control group dose-dependently (Table 3, fig 1). Significant (p < 0.01) increases of the number of sertoli cells, spermatogonia, preleptotene spermatocytes, leptotene spermatocytes, zygote spermatocytes, pachytene spermatocytes, round and elongated spermatids were observed in EKN-100 and EKN-500 at all stages and significant (P < 0.05 or P < 0.01) increase of those were observed in EKN-10 at all stages except stage XII compared to control group (Table 4, fig 2).

No abnormal histopathological change was observed in the liver, spleen, kidney, prostate and seminal vesicle in this study including control.

Table 1.	Changes	of reproductive	organ weights	after administration	of EKN extract in 2-BF	⁹ induced-reproductive	damaged rats
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Gro	oups	A dose of EKN extract administered ^{a)}				
Items control		EKN-10	EKN-100	EKN-500		
Absolute organ weight (g)						
Testis	$0.81\pm0.07^{\text{b})}$	0.81 ± 0.03	0.84 ± 0.05	0.81 ± 0.02		
Epididymis	0.49 ± 0.01	$0.45\pm0.02^{\#}$	$0.47\pm0.01^{\#}$	0.50 ± 0.05		
Prostate	0.79 ± 0.11	0.75 ± 0.22	0.69 ± 0.07	0.85 ± 0.10		
Seminal vesicle	0.76 ± 0.13	$0.33 \pm 0.99^{\text{\#}}$	$0.51 \pm 0.18^{\#\#}$	0.76 ± 0.16		
Relative organ weight (%)	c					
Testis	0.26 ± 0.02	0.26 ± 0.03	0.27 ± 0.03	0.27 ± 0.03		
Epididymis	0.16 ± 0.00	0.14 ± 0.02	0.15 ± 0.01	0.16 ± 0.03		
Prostate	0.25 ± 0.03	0.24 ± 0.08	0.22 ± 0.02	0.28 ± 0.06		
Seminal vesicle	0.24 ± 0.04	$0.11 \pm 0.03^{\#\!\#}$	$0.17\pm0.05^{\text{\#}}$	0.25 ± 0.07		

^{a)}; each of 10, 100 and 500 mg/kg of EKN extract was administered each 8 reproductive damaged rats orally for 28 days. ^{b)}; The values were expressed with mean \pm SD. ^C; Ratio of the organ weight divided by body weight.

###; each of indicated P < 0.05, P < 0.01, respectively compared to control group.

Table 2. Changes of DPS of the testis and sperm count of cauda epididymis after administration of EKN extract in 2-BP inducedreproductive damaged rats

Groups		A dose of EKN extract administered ^{a)}			
Items	control	EKN-10	EKN-100	EKN-500	
Right testis weight (g)	$0.79\pm0.13^{\text{b})}$	0.65 ± 0.10	0.69 ± 0.06	0.77 ± 0.04	
DSP (× $10^8/g$)	4.08 ± 0.72	$6.02\pm0.94^{\#}$	$5.41\pm0.67^{\text{\#}}$	4.80 ± 0.13	
Cauda epididymis weight (g)	0.20 ± 0.02	0.19 ± 0.03	0.19 ± 0.02	0.20 ± 0.01	
Sperm count (× 10 ⁸ /g)	9.31 ± 0.41	10.54 ± 1.77	8.38 ± 0.80	$12.64 \pm 1.06^{\#}$	

^{a)}; each of 10, 100 and 500 mg/kg of EKN extract was administered each 8 reproductive damaged rats orally for 28 days. ^{b)}; The values were expressed with mean \pm SD.

^{t, ##}; indicated P < 0.05, P < 0.01 compared to control group.</sup>

Table 3. Changes of the	e number of abnormal	seminiferous and	epididymal t	tubules after	administration of	of EKN extract	t in 2-BP
induced-reproductive dan	naged rats						

	No. of seminiferous tubules	No. of epididymal tubules			
Items Groups (N/100 ubules)	showing atrophic changes or contains vacuolated sertoli cells	Decreases spermatozoa of spermatozoa in the lumens	vacuolated epithelia lining		
Control	$80.80 \pm 6.65^{\text{b}}$	86.60 ± 5.41	76.00 ± 4.47		
EKN-10 ^{a)}	$64.80\pm7.63^{\#}$	$67.00 \pm 4.36^{\#}$	$51.80\pm 3.11^{\text{\#}}$		
EKN-100	$41.40\pm 3.05^{\#}$	$39.60 \pm 6.02^{\#}$	$36.20 \pm 4.87^{\text{\#}}$		
EKN-500	$30.20 \pm 4.66^{\#}$	$31.40 \pm 3.21^{\#}$	$28.80 \pm 7.85^{\#}$		

^{a)}; each of 10, 100 and 500 mg/kg of EKN extract was administered each 8 reproductive damaged rats orally for 28days. ^{b)}; The values were expressed with mean \pm SD. n=5 (5 fields of microscopy)

^{##}; indicated P < 0.01 compared to control group.



Fig 1. Histological profiles of the testis in the control (a), EKN-10 (b), EKN-100 (c) and EKN-500 (d). Note that epithelial lining cell vacuolation (arrows) and aspermatozoa (A) were observed in the lumen of epididymal tubules after administration of control. However, these epididymal histopathological changes were dramatically inhibited by administration of the EKN-dosing group. All HE stain; Scale bars = $80 \mu m$.

Discussion

EKN has been traditionally used for improving of impotency in oriental medicine (20). Recently, some researchers lively have been studying the components, flavonoids (26,32,31,4) of EKN, and its sexual or immunological effects on normal mice or rats. To investigate of effects of EKN in reproductive damage, we used 2-BP to induced reproductive damage, especially inhibition of spermatogenesis. Over 1,000 ppm exposure level of 2-BP, the spermatogenesis in the most of rats were inhibited. The 1,000 ppm exposure level is calculated to be approximately 1,355 mg/kg per day (9,22).

In the previous studies (17,36,9,12,39,13), the weights of body, thymus, spleen, testis, epididymis, prostate and seminal

vesicle were decreased in 2-BP treated rats. In this study, however, EKN did not affect the weights of liver, spleen and kidney in 2-BP treated rats. Spleen weight was slightly decreased whereas kidney weight was slightly increased compared to control group. But there was no scientific evidence about increases of liver, kidney and spleen weight after administration of EKN extract in reproductive damaged rats (Table 1). The weights of epididymis, prostate, and seminal vesicle were dose-dependently increased in the EKN-dosing groups but they were lower than control group. In EKN-10 and EKN-100 groups, they were significantly (P < 0.01 or P < 0.05) lower than control group but those of EKN-500 group were higher than control group or similar. Testis did not changed in this study (Table 2). Unexpected result of reproductive tissue weight was showed in EKN-10

Joo-Wan Kim et al.

Groups		A dose of EKN extract administered ^{a)}			
Items	control	EKN-10	EKN-100	EKN-500	
Stage I-III					
Sertoli cells	$8.30\pm2.36^{\text{b})}$	$12.40 \pm 2.59^{\#}$	$13.50 \pm 1.27^{\#}$	$14.00 \pm 2.26^{\#}$	
Spermatogonia	1.90 ± 0.57	$8.70 \pm 3.16^{\#}$	$11.00 \pm 1.15^{\#}$	$12.40 \pm 1.65^{\#}$	
Patchytene spermatocytes	10.40 ± 1.90	14.60 ± 3.50	$16.60 \pm 3.37^{\#}$	$17.90 \pm 5.22^{\#}$	
Round spermatids	80.40 ± 31.79	$125.00\pm8.65^{\#\!\#}$	$131.10\pm7.68^{\#\#}$	$138.20 \pm 12.19^{\#}$	
Elongated spermatids	93.00 ± 20.34	$119.90 \pm 13.24^{\#\#}$	$131.80\pm 8.97^{\text{\#}}$	$132.80\pm7.61^{\#}$	
Stages V					
Sertoli cells	8.30 ± 2.00	$11.10 \pm 1.10^{\#\#}$	$12.40 \pm 1.17^{\#}$	$13.00 \pm 1.63^{\#\#}$	
Spermatogonia	1.90 ± 2.56	$6.50 \pm 2.76^{\text{\#}}$	$8.80 \pm 2.10^{\#\!\!\!/}$	$10.80 \pm 1.32^{\#}$	
Patchytene spermatocytes	20.20 ± 4.78	$32.40 \pm 5.02^{\#}$	$35.50 \pm 9.59^{\#\#}$	$44.40 \pm 10.15^{\#}$	
Round spermatids	104.40 ± 14.91	$121.20\pm6.30^{\#\#}$	$125.20\pm9.98^{\#\#}$	$128.70 \pm 7.47^{\text{\#}}$	
Elongated spermatids	94.90 ± 16.64	$115.00 \pm 10.01^{\#\#}$	$126.80\pm8.84^{\text{\#}}$	$131.30 \pm 11.74^{\#}$	
Stages VII					
Sertoli cells	8.70 ± 3.37	$12.50 \pm 1.08^{\#}$	$12.60 \pm 1.71^{\#}$	$12.70 \pm 1.34^{\#}$	
Spermatogonia	0.90 ± 0.74	2.20 ± 0.79	$2.40 \pm 0.84^{\text{\#}}$	$2.40 \pm 0.84^{\text{\#}}$	
Preleptotene spermatocytes	12.40 ± 3.13	$18.60 \pm 2.88^{\#\!$	$21.60 \pm 3.63^{\#\!$	$24.60 \pm 4.14^{\#\#}$	
Patchytene spermatocytes	21.40 ± 2.28	$27.10 \pm 4.93 \#^{\!\#}$	$29.60 \pm 3.03^{\#\!\#}$	$31.70 \pm 3.53^{\#\#}$	
Round spermatids	101.80 ± 29.59	$131.10\pm 5.97^{\#}$	$133.50 \pm 10.78^{\#}$	$141.30 \pm 15.57^{\#}$	
Elongated spermatids	90.60 ± 11.53	$113.60 \pm 10.01^{\#\#}$	$120.80 \pm 9.60^{\#}$	$126.20 \pm 11.31^{\#}$	
Stages XII					
Sertoli cells	10.70 ± 1.49	12.40 ± 1.17	$13.10 \pm 2.51^{\#}$	$13.90 \pm 2.13^{\#}$	
Spermatogonia	1.00 ± 0.94	$2.20 \pm 0.63^{\#\!\#}$	$2.30\pm 0.67^{\#}$	$2.40 \pm 0.70^{\text{\#}}$	
Patchytene spermatocytes	11.60 ± 2.63	$20.20 \pm 4.52^{\#}$	$22.10 \pm 4.12^{\#}$	$24.10\pm 7.67^{\#}$	
Round spermatids	8.60 ± 2.37	$13.10 \pm 2.47^{\#}$	$13.60 \pm 2.55^{\#}$	$14.20\pm 3.22^{\#}$	
Elongated spermatids	83.90 ± 20.84	$112.60 \pm 13.38^{\#\#}$	$119.80\pm8.40^{\text{\#}}$	$124.90 \pm 10.39^{\#\!\#}$	

Table 4. Cell types and numbers of seminiferous tubules after administration of EKN extract in 2-BP induced-reproductive damaged rats

^{a)}; each of 10, 100 and 500 mg/kg of EKN extract was administered each 8 reproductive damaged rats orally for 28 days. ^b; The values were expressed with mean \pm SD. n = 10 (10 round or ovoid seminiferous tubules). ^{#, ##}; each of indicated p < 0.05, p < 0.01 compared to control group.



Fig 2. Histological profiles of the epididymis in the control (a), EKN-10 (b), EKN-100 (c) and EKN-500 (d). Note that epithelial lining cell vacuolation (arrows) and oligospermatozoa (O) were observed in the lumen of epididymal tubules after administration of control. However, these epididymal histopathological changes were dramatically inhibited by administration of EKN-dosing group. All HE stain; Scale bars = $80 \mu m$.

and EKN-100.

In this study, any significant change of reproductive organ was not observed in the low dose (EKN-10) group. Therefore further investigation about it might be needed. Over the 500 mg/kg of EKN extract has anti-impotent effect against 2-BP induced-genital damage by increasing the weight of reproductive organ.

Jeong *et al* (12) reported RBC was slightly decreased and platelet was significantly (P < 0.05) decreased and PCV, MCV and MCH were not affected in 2-BP treated rats. In this study, some parts of hematological items, MCV in EKN-500, platelet in EKN-10, were shown significant (P < 0.01) changes, but all of the hematological data were within the reference index. EKN did not affect hematological changes in 2-BP induced-reproductive damaged rats.

In the previous studies (17,36), the decreases of DSP and sperm number in 2-BP treated rats resulted from apoptosis. In this study, DSPs of EKN-dosing groups were decreased with dose-dependent pattern while this result came into conflict with histomorphological results (low damage of spermatozoa). It was supposed that DSP ability had relation to sperm count, that is, plenty of sperms stimulated negative feedback as low DSP. Sperm counts of EKN-dosing groups were increased dose-dependently. Although epididymis weights of EKN-dosing groups were decreased, but sperm count was increased compared to control group. It is considered that the sperm production could be affected by EKN and DSP could be affected by decreased sperm count. So we assume that EKN has an active function as stimulating spermatozoa and/or sertoli cell, because in control group, they did not act like in EKN-dosing groups (Table 3). So we examined the histological aspect, well corresponded to the previous studies (9,10,17,38), atrophic changes of semniferous tubules, vacuolation of sertoli cells were detected, and epithelial lining cell vacuolation and oligo- or aspermatozoa were detected in the lumen of epididymal tubules at histopathological observation after 28 days of serial injection of 1,355 mg/kg of 2-BP treatment, and these 2-BP induced reproductive damages are reconfirmed with histomorphometry. 2-BP induced-reproductive damages were dramatically decreased in EKN-dosing groups compared to control group (Figs 1 and 2). The number of degenerative seminiferous tubules and the number of epididymal tubules showing epithelial cell vacuolation and decreased spermatozoa in the lumen were significantly (p < 0.01 or p < 0.05) decreased after treatment of 10, 100 or 500 mg/kg of EKN (Table 4). These are considered as direct evidences that EKN effectively treated the 2-BP induced-reproductive damages.

DSP parameter in the histological sections of testis can be used as a valid assay of spermatogonia survival for cytotoxic agents (23) and has been used as an indicator of spermatogenesis (8,11). At dose of 1,355 mg/kg of 2-BP, the numbers of sertoli cells, spermatogonia, preleptotene spermatocytes, leptotene spermatocytes, zygote spermatocytes, pachytene spermatocytes, round and elongated spermatids were dramatically decreased on 15 days of continuous treatment (17). 2-BP primarily targets to spermatogonia by apoptosis (17,25,39) and consequently decreases of spermatocytes and spermatids were induced. In this study, significant (p < 0.01 or p < 0.05) increases of the numbers of sertoli cells, spermatogonia, preleptotene spermatocytes, leptotene spermatocytes, zygote spermatocytes, pachytene spermatocytes, round and elongated spermatids were observed in EKN-100 and EKN-500 at all stages, compared to control group. Although significant (p < 0.01 or p < 0.05) increases were restricted to some cell types of some stages, dramatic increases of all cell types at all stages were also detected in EKN-dosing groups compared to control group, respectively (Table 4). These are considered as direct evidences that EKN effectively inhibits decreases of spermatogenesis induced by 2-BP. No abnormal histopathological change was observed in liver, spleen, kidney, prostate and seminal vesicle including control group.

Based on the results, it is concluded that EKN water extract dramatically treats the 2-BP induced-reproductive damages from 10 mg/kg of dose levels, the lowest levels. Although 10 mg/kg of EKN extract also inhibited the decreases of spermatogenesis, the effective dosage of EKN inhibiting both reproductive damages and decreases of spermatogenesis by 2-BP was considered as 100 mg/kg in rats because significant inhibition of spermatogenesis decreases was restricted to some stages of some cell types in 10 mg/kg dosing group.

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2-bromopropane으로 유발된 생식기 장애에서 음양곽 물추출물의 효과

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요 약 : 음양곽(*Epimedicum Koreanum* nakai)은 예부터 생식기 장애치료를 위해 사용되어 왔다. 본 연구에서는 2-브 로모프로판(2-bromopropane; 2-BP)으로 유발된 생식기 장애에서 음양곽 물추출물의 생식기장애 치료효과를 평가하였 다. 2-BP를 체중kg당 1,355 mg씩 28일간 피하로 투여하여 생식기 장애를 유발하였으며, 음양곽 추출물은 용량별(10, 100, 500 mg/kg)로 4주간 경구투여 하였다. 그 결과 일일정자생성(daily sperm production)은 대조군에 비하여 증가하 였으나 용량의존적 감소를 나타내었으며, 정세관과 부고환관에서는 조직학적 유의한 변화를 나타내었다. 이러한 결과 는 음양곽물추출이 2-BP로 유발된 생식기장애의 치료에 도움이 될 것으로 생각된다.

주요어 : 음양곽, 생식기장애, 2-브로모프로판, 랫