

The Role of Nitric Oxide in Mycobacterial Infections

Chul-Su Yang, Jae-Min Yuk and Eun-Kyeong Jo*

Department of Microbiology and Infection Signaling Network Research Center, College of Medicine, Chungnam National University, Daejeon 301-747, Korea

Although tuberculosis poses a significant health threat to the global population, it is a challenge to develop new and effective therapeutic strategies. Nitric oxide (NO) and inducible NO synthase (iNOS) are important in innate immune responses to various intracellular bacterial infections, including mycobacterial infections. It is generally recognized that reactive nitrogen intermediates play an effective role in host defense mechanisms against tuberculosis. In a murine model of tuberculosis, NO plays a crucial role in antimycobacterial activity; however, it is controversial whether NO is critically involved in host defense against *Mycobacterium tuberculosis* in humans. Here, we review the roles of NO in host defense against murine and human tuberculosis. We also discuss the specific roles of NO in the central nervous system and lung epithelial cells during mycobacterial infection. A greater understanding of these defense mechanisms in human tuberculosis will aid in the development of new strategies for the treatment of disease.

[Immune Network 2009;9(2):46-52]

INTRODUCTION

Tuberculosis (TB) is a bacterial infectious disease caused by the obligate human pathogen *Mycobacterium tuberculosis* (MTB). TB remains an urgent global health problem, with a third of the global population latently infected and eight million new cases each year. Although only 5~10% of infected individuals develop active TB, the fatality rate is nearly two million people annually (1-3). Following exposure to MTB, a series of immune responses are triggered that ultimately define the course of the infection (4,5). The pathogenesis of infection is complicated; however, recent discoveries have at-

tracted great attention due to their association with host-derived and microbial factors. Advances in free radical research have revealed that the production of reactive oxygen and nitrogen oxide species such as superoxide (O_2^-) and nitric oxide (NO) by innate immune cells is a relatively effective host defense mechanism against bacterial, viral, parasitic, and fungal infections (5,6).

The host cells that are protective against TB include macrophages, dendritic cells, T lymphocytes, and alveolar epithelial cells (2,3,7). Macrophages are believed to play a pivotal role in the immune response against mycobacteria through the production of cytokines such as tumor necrosis factor (TNF)- α and interleukin (IL)-1 β . TNF- α and IL-1 β , along with interferon (IFN)- γ , which is produced by T lymphocytes, can induce NO production in macrophages via the action of inducible forms of the enzyme NO synthase (iNOS) (8-10). NO and related reactive nitrogen intermediates (RNI)s can kill and/or inhibit intracellular pathogens such as mycobacteria (11-14). The actions of iNOS and the production of NO correlate well with antimycobacterial defense in murine models of TB infection (10,12,15). Although it has been demonstrated that iNOS expression is up-regulated in macrophages from human TB lesions (16), few reports have examined the antimycobacterial effects of cytokines and NO released by human macrophages (17,18). These data suggest that human macrophages possess a NO-independent antimicrobial mechanism, although a role for NO in human host defense cannot be excluded.

MTB infects the airways and stimulates alveolar macrophages, epithelial cells, and macrophages. As a result, NO is produced in response to the stimulation of cytokines and chemokines (19). By producing NO, alveolar epithelial cells can actively participate in alveolar inflammatory processes and

Received on March 3, 2009. Accepted on March 10, 2009.

*Corresponding Author. Tel: 82-42-580-8243; Fax: 82-42-585-3686; E-mail: hayoungj@cnu.ac.kr

Keywords: nitric oxide, mycobacteria, macrophages, host defense

defense mechanisms against MTB. In this review, we discuss the role of NO in defense mechanisms against MTB and the mechanisms regulating the production of NO in macrophages, including microglia and alveolar epithelial cells.

OVERVIEW OF NO PRODUCTION AND FUNCTION

NO is a gaseous free radical molecule with pleiotropic functions in pathophysiology that is synthesized by a two-step enzymatic reaction involving a monooxygenase (12,13). One molecule of L-arginine is oxidized at the terminal nitrogen in guanidine to produce N^{ω} -OH-L-arginine as an intermediate. This intermediate is then further oxidized to form one molecule each of NO and L-citrulline (13,14). L-arginine (a conditionally essential amino acid) is obtained from exogenous (food) and endogenous sources, including whole-body protein degradation and, to a lesser extent, *de novo* synthesis from citrulline by renal arginosuccinate synthase (20,21). Two sequential reactions are catalyzed by NOSs, resulting in the constitutive expression of enzymes primarily in endothelial cells (eNOS) and neuronal cells (nNOS), and as an inducible

isoform (iNOS). Constitutively produced NOSs contribute to several physiological processes including vasorelaxation and neurotransmission. In contrast, iNOS is expressed in various cells including macrophages, neutrophils, epithelial cells, and hepatocytes, and it produces excessive NO during infection, inflammation, and states of physiological stimulation (22-24).

Th1 cytokines such as IFN- γ , IL-1 β , and TNF- α stimulate the expression of macrophage iNOS, leading to NO production. In contrast, under the influence of Th2 cytokines such as IL-4, IL-10, and IL-13, arginine is depleted by arginases (8-10). NO is one of several RNIs with antimicrobial activity (18,25). The increase in RNIs is mediated through reactive nitrogen oxides (e.g., peroxynitrite (ONOO $^-$)) generated by the reaction of NO with O $_2^-$ (13,24) (Fig. 1). NO and RNIs can modify bacterial DNA, proteins, and lipids in both the microbe and host. NO can also deaminate and directly damage bacterial DNA by generating abasic sites and strand breaks (7). Other potential killing mechanisms by NO include interactions with accessory protein targets such as iron-sulfur groups, heme groups, thiols, aromatic or phenolic residues, tyrosyl radicals, and amines. These reactions result in enzy-

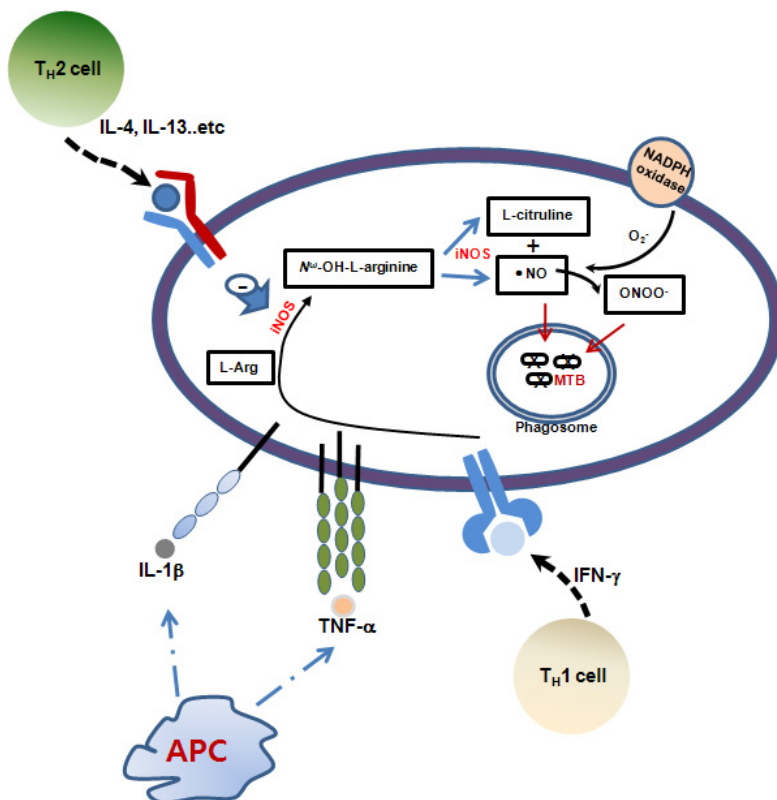


Figure 1. Yang et al. Synthesis, regulation, and antimycobacterial function of NO in mycobacterial infection. Activated inducible nitric oxide synthase (iNOS) produces N^{ω} -OH-L-arginine from L-arginine, and then N^{ω} -OH-L-arginine is transduced to form NO and L-citrulline. Synthesis of NO and reactive nitrogen oxides (RNI) are positively regulated by Th1 cytokines, whereas they are negatively regulated by Th2 cytokines. Produced NO and RNIs, which combined with NO and O $_2^-$, can directly kill intracellular MTB in the infected cells (including macrophages, epithelial and glial cells), although the action of NO is dependent on the species and specific cell types.

matic inactivation and/or other protein malfunctions (26).

THE ROLE OF NO IN HOST DEFENSE AGAINST MICROBIAL INFECTIONS

During infection with *Mycobacterium*, *Salmonella*, *Streptococcus*, *Leishmania*, or *Bordetella*, excessive NO is produced after the induction of iNOS. In many cases, excessive NO production results in innate resistance to bacterial infection. In a study of *Bordetella pertussis* infection in wild-type (WT) and iNOS-knockout (iNOS KO) mice, the iNOS KO mice displayed increased bacterial growth and susceptibility to infection as compared with the WT mice (27). In a study of murine salmonellosis (*Salmonella typhimurium*), the use of a NO inhibitor, *N*^ω-monomethyl-L-arginine (L-NMMA), or iNOS KO mice led to similar antimicrobial effects (12,28). In these studies, the lack of NO production was associated with extensive damage, including increased bacterial growth, increased apoptosis, and the exacerbation of histopathological characteristics in mouse livers infected with *Salmonella enterica* serovar Typhimurium (12). Although NO shows antimicrobial activity against bacteria, fungi, and parasites, some studies suggest dual functions during viral infection. NO produced by macrophages and phagocytic cells can act as an effector molecule during innate host defense mechanisms. For example, NO shows antiviral activity in response to certain viruses such as coxsackievirus (29-31), Epstein-Barr virus (32), and herpes simplex virus (HSV)-1 (33-35). In contrast to the antibacterial activity observed with NO, this antiviral activity is associated with nonspecific damage to host cells and tissues, leading to an exacerbation of viral pathogenesis in many infections such as influenza (36), tick-born virus (37), sendai virus (38), HSV-1 (39,40), and cytomegalovirus (41,42). Therefore, despite the antiviral activity of NO, excessive NO production may facilitate viral pathogenesis. These dual functions of NO may lead to differential outcomes during viral infection.

THE ROLE OF NO IN MYCOBACTERIAL INFECTION: MURINE STUDIES

NO plays a key role in innate immunity and host defense against mycobacteria (1,2,7,43). For example, iNOS KO and immunodeficient mice infected with MTB are at a significantly higher risk of dissemination and mortality as compared with control mice (1,43). In addition, macrophages from mice with

the *Bcg/natural resistance associated macrophage protein-1* resistance phenotype show inhibition of MTB survival through NO production (3,44). Mycobacterial species exhibit variations in susceptibility to NO and its RNIs. For example, murine macrophages have been shown to inhibit the intracellular growth of *M. leprae*, *M. bovis*, and MTB H37Rv (7,11,17,45). When IFN- γ -treated rat alveolar macrophages were infected with *M. avium*, the growth of the bacterium was significantly inhibited by NO synthesized from L-arginine (46).

Contrasting data have been reported in murine and human macrophages infected with *M. avium* (6); neither competitive inhibition by L-NMMA nor depletion of L-arginine by arginase had any effect on *M. avium* growth in murine peritoneal macrophages or human monocyte-derived macrophages (6). In addition, no significant inhibitory effects of NO produced by rat macrophages were observed on the growth of *M. intracellulare* (45). In murine models of latent infection, both NO-dependent (iNOS- and IFN- γ -dependent antimycobacterial mechanisms) and -independent (CD4⁺ T cells required for preventing reactivation of the disease) mechanisms maintain latent TB (4,47); however, the applicability of these reports to humans is uncertain.

THE ROLE OF NO IN MYCOBACTERIAL INFECTION: HUMAN STUDIES

In contrast to the murine model of TB, there is controversy surrounding the role of NO in the killing and inhibition of MTB in humans (5). The early inhibition of mycobacterial growth by human alveolar macrophages has been shown to be NO-independent (48). Specifically, exogenous IFN- γ failed to produce mycobactericidal effects in human alveolar macrophages (48). Nevertheless, a growing body of evidence suggests that NO production by MTB-infected human monocytes/macrophages, macrophage-like cell lines, and epithelial cells induces mycobacteriostatic activity against MTB (16, 49-52). For example, alveolar macrophages from healthy control subjects infected with MTB produce NO, and this production is correlated with the intracellular inhibition of MTB growth (51). One study demonstrated increased NO production in TB patients as compared with healthy controls following the infection of peripheral blood mononuclear cells (PBMC)s with MTB (50). Other studies have also demonstrated that alveolar macrophages are able to kill mycobacteria and that these antimycobacterial activities are dependent

on iNOS expression (49,53). These results suggest a significant role for NO in host defense against mycobacterial infection. Moreover, increased iNOS expression and pulmonary NO production have been reported in alveolar macrophages and PBMCs from TB patients as compared with healthy controls (8,10,16,54). In those studies, NO played a role in the enhancement of TNF- α and IL-1 β secretion, which subsequently affected NO production via feedback loops (8). These data indicate an autoregulatory role for NO. It has also been shown that iNOS and nitrotyrosine (a tissue marker of NO metabolism) are expressed in macrophages within granulomas and areas of TB pneumonitis (15,55). Human PBMCs and bronchial epithelial cells may produce NO when stimulated with MTB-produced NO (50). In addition, the avirulent strain H37Ra was shown to induce significantly higher levels of NO production as compared with the virulent strain H37Rv (50). Recent studies have shown that L-arginine depletion induces the down-regulation of CD3 ζ , thereby impairing T cell signaling, whereas the addition of L-arginine leads to CD3 ζ re-expression and the recovery of T cell proliferation (18,56). In addition, T cells from TB patients show reduced CD3 ζ expression, which is correlated with arginase-induced L-arginine deficiency. These expression levels were normalized with successful TB treatment (21). Taken together, these data suggest that NO plays a contributory role in human host defense against MTB infection.

THE ROLE OF NO IN MYCOBACTERIAL INFECTION OF THE CENTRAL NERVOUS SYSTEM (CNS)

The roles of NO and iNOS in host defense against infection of the CNS by intracellular pathogens have been reported in previous studies of several intracellular pathogens (e.g., *Toxoplasma gondii* and Sindbis virus) (57,58). The role of microglial cells in neuropathogenesis following CNS infection has been a topic of growing research interest (59). It was previously reported that in contrast to astrocytes, iNOS was not expressed in human microglia following stimulation by IL-1 β or IFN- γ (9,60). These findings suggest that NO and iNOS expression may be dependent on cell type and species. Indeed, previous reports demonstrated NO production in activated murine microglia, but not in human microglia (61). Recently, significant effort has been devoted to developing appropriate models of TB infection in the CNS (CNS-TB) using rabbits or mice. Intracerebral inoculation with MTB or *M. bovis* BCG resulted in mononuclear cell infiltration, microglial

cell activation, and an increase in the number of bacterial cells within the CNS in a mouse model (62,63). In addition, inoculation with MTB or *M. bovis* BCG led to the up-regulation of IL-1 β , TNF- α , IL-6, and IFN- γ within the CNS (63). Recently, Michael et al. (64) reported that iNOS KO mice infected intracerebrally with MTB developed clinical manifestations of CNS-TB, including high mortality rates and histopathological abnormalities resembling human tuberculous meningitis throughout the meninges. The above clinical manifestations were absent in WT mice. These studies underscore the importance of NO in defense against CNS-TB.

THE ROLE OF NO IN THE MYCOBACTERIAL INFECTION OF EPITHELIAL CELLS

Alveolar epithelial cells are able to actively participate in the pathogenesis of pulmonary inflammatory diseases by producing several cytokines and chemokines (65-67). Alveolar epithelial cells produce NO and various innate immune effectors including chemokines (IL-8), which regulate immune activation. In addition, normal T cells express and secrete RANTES in response to MTB infection (19,67). Strong NO production via iNOS also occurs in human lung epithelial cells (19,66); however, the amount of NO released in response to MTB is not mycobactericidal (65,66). Various cytokines (IFN- γ , TNF- α , and IL-1 β ; alone or in combination) and mycobacterial components stimulate MTB-infected epithelial cells, inducing NO production and mycobactericidal effects (65,66). These factors may contribute to innate immune control in epithelial cells against intracellular pathogens such as MTB.

CONCLUDING REMARKS

NO is a nonspecific, chemically reactive molecule that is important in host defense against a wide variety of microbial pathogens. However, it is becoming increasingly clear that specific killing mechanisms and cell types are not sufficient to kill mycobacteria *in vivo*. Although NO is not required for mycobactericidal activity in mouse models, the lack of a role for NO or its products (e.g., ONOO⁻) has not been definitively proven in humans. Nevertheless, a substantial body of evidence indicates a role for NO in human host defenses against MTB. Additional studies are necessary to define the role of NO in relevant human cells including alveolar macrophages, microglia, and epithelial cells. Additionally, it would be useful to generate conditions that mimic *in vivo* environ-

ments, such as the co-culture of relevant cells. Such studies, which will refine our understanding of the importance and specific role of NO in TB defense, may lead to innovative strategies for TB treatment.

ACKNOWLEDGEMENTS

This research was supported by the Korea Science & Engineering Foundation through the Infection Signaling Network Research Center (R13-2007-020-01000-0) at Chungnam National University. The authors declare that they have no competing financial interests.

CONFLICTS OF INTEREST

The authors have no financial conflict of interest.

REFERENCES

1. Adams LB, Dinanuer MC, Morgenstern DE, Krahenbuhl JL: Comparison of the roles of reactive oxygen and nitrogen intermediates in the host response to *Mycobacterium tuberculosis* using transgenic mice. *Tuber Lung Dis* 78:237-246, 1997
2. Chan J, Xing Y, Magliozzo RS, Bloom BR: Killing of virulent *Mycobacterium tuberculosis* by reactive nitrogen intermediates produced by activated murine macrophages. *J Exp Med* 175:1111-1122, 1992
3. Rojas M, Barrera LF, Puzo G, Garcia LF: Differential induction of apoptosis by virulent *Mycobacterium tuberculosis* in resistant and susceptible murine macrophages: role of nitric oxide and mycobacterial products. *J Immunol* 159:1352-1361, 1997
4. Flynn JL, Scanga CA, Tanaka KE, Chan J: Effects of aminoguanidine on latent murine tuberculosis. *J Immunol* 160:1796-1803, 1998
5. Nathan C, Shiloh MU: Reactive oxygen and nitrogen intermediates in the relationship between mammalian hosts and microbial pathogens. *Proc Natl Acad Sci U S A* 97:8841-8848, 2000
6. Bermudez LE: Differential mechanisms of intracellular killing of *Mycobacterium avium* and *Listeria monocytogenes* by activated human and murine macrophages. The role of nitric oxide. *Clin Exp Immunol* 91:277-281, 1993
7. Chan J, Tanaka K, Carroll D, Flynn J, Bloom BR: Effects of nitric oxide synthase inhibitors on murine infection with *Mycobacterium tuberculosis*. *Infect Immun* 63:736-740, 1995
8. Kuo HP, Wang CH, Huang KS, Lin HC, Yu CT, Liu CY, Lu LC: Nitric oxide modulates interleukin-1 β and tumor necrosis factor- α synthesis by alveolar macrophages in pulmonary tuberculosis. *Am J Respir Crit Care Med* 161:192-199, 2000
9. Lee SC, Dickson DW, Liu W, Brosnan CF: Induction of nitric oxide synthase activity in human astrocytes by interleukin-1 β and interferon- γ . *J Neuroimmunol* 46:19-24, 1993
10. Wang CH, Lin HC, Liu CY, Huang KH, Huang TT, Yu CT, Kuo HP: Upregulation of inducible nitric oxide synthase and cytokine secretion in peripheral blood monocytes from pulmonary tuberculosis patients. *Int J Tuberc Lung Dis* 5:283-291, 2001
11. Adams LB, Franzblau SG, Vavrin Z, Hibbs JB Jr, Krahenbuhl JL: L-arginine-dependent macrophage effector functions inhibit metabolic activity of *Mycobacterium leprae*. *J Immunol* 147:1642-1646, 1991
12. Alam MS, Akaike T, Okamoto S, Kubota T, Yoshitake J, Sawa T, Miyamoto Y, Tamura F, Maeda H: Role of nitric oxide in host defense in murine salmonellosis as a function of its antibacterial and antiapoptotic activities. *Infect Immun* 70:3130-3142, 2002
13. MacMicking J, Xie QW, Nathan C: Nitric oxide and macrophage function. *Annu Rev Immunol* 15:323-350, 1997
14. Mayer B, Hemmens B: Biosynthesis and action of nitric oxide in mammalian cells. *Trends Biochem Sci* 22:477-481, 1997
15. Choi HS, Rai PR, Chu HW, Cool C, Chan ED: Analysis of nitric oxide synthase and nitrotyrosine expression in human pulmonary tuberculosis. *Am J Respir Crit Care Med* 166:178-186, 2002
16. Nicholson S, Bonecini-Almeida Mda G, Lapa e Silva JR, Nathan C, Xie QW, Mumford R, Weidner JR, Calaycay J, Geng J, Boechat N, Linhares C, Rom W, Ho JL: Inducible nitric oxide synthase in pulmonary alveolar macrophages from patients with tuberculosis. *J Exp Med* 183:2293-2302, 1996
17. Denis M: *In vivo* modulation of atypical mycobacterial infection: adjuvant therapy increases resistance to *Mycobacterium avium* by enhancing macrophage effector functions. *Cell Immunol* 134:42-53, 1991
18. Rodriguez PC, Zea AH, DeSalvo J, Culotta KS, Zabaleta J, Quiceno DG, Ochoa JB, Ochoa AC: L-arginine consumption by macrophages modulates the expression of CD3 ζ chain in T lymphocytes. *J Immunol* 171:1232-1239, 2003
19. Robbins RA, Barnes PJ, Springall DR, Warren JB, Kwon OJ, Buttery LD, Wilson AJ, Geller DA, Polak JM: Expression of inducible nitric oxide in human lung epithelial cells. *Biochem Biophys Res Commun* 203:209-218, 1994
20. Paton NI, Chua YK, Earnest A, Chee CB: Randomized controlled trial of nutritional supplementation in patients with newly diagnosed tuberculosis and wasting. *Am J Clin Nutr* 80:460-465, 2004
21. Wu G, Morris SM Jr: Arginine metabolism: nitric oxide and beyond. *Biochem J* 336:1-17, 1998
22. Moncada S, Palmer RM, Higgs EA: Nitric oxide: physiology, pathophysiology, and pharmacology. *Pharmacol Rev* 43:109-142, 1991
23. Nathan C, Xie QW: Regulation of biosynthesis of nitric oxide. *J Biol Chem* 269:13725-13728, 1994
24. Zaki MH, Akuta T, Akaike T: Nitric oxide-induced nitrative stress involved in microbial pathogenesis. *J Pharmacol Sci*

- 98;117-129, 2005
25. Ralph AP, Kelly PM, Anstey NM: L-arginine and vitamin D: novel adjunctive immunotherapies in tuberculosis. *Trends Microbiol* 16;336-344, 2008
 26. Gow AJ, Thom SR, Ischiropoulos H: Nitric oxide and peroxynitrite-mediated pulmonary cell death. *Am J Physiol* 274;L112-L118, 1998
 27. Canthaboo C, Xing D, Wei XQ, Corbel MJ: Investigation of role of nitric oxide in protection from *Bordetella pertussis* respiratory challenge. *Infect Immun* 70;679-684, 2002
 28. Umezawa K, Akaike T, Fujii S, Suga M, Setoguchi K, Ozawa A, Maeda H: Induction of nitric oxide synthesis and xanthine oxidase and their roles in the antimicrobial mechanism against *Salmonella typhimurium* infection in mice. *Infect Immun* 65;2932-2940, 1997
 29. Zaragoza C, Ocampo C, Saura M, Leppo M, Wei XQ, Quick R, Moncada S, Liew FY, Lowenstein CJ: The role of inducible nitric oxide synthase in the host response to coxsackievirus myocarditis. *Proc Natl Acad Sci U S A* 95;2469-2474, 1998
 30. Zaragoza C, Ocampo CJ, Saura M, Bao C, Leppo M, Lafond-Walker A, Thiemann DR, Hruban R, Lowenstein CJ: Inducible nitric oxide synthase protection against coxsackievirus pancreatitis. *J Immunol* 163;5497-5504, 1999
 31. Zaragoza C, Ocampo CJ, Saura M, McMillan A, Lowenstein CJ: Nitric oxide inhibition of coxsackievirus replication in vitro. *J Clin Invest* 100;1760-1767, 1997
 32. Mannick JB, Asano K, Izumi K, Kieff E, Stampler JS: Nitric oxide produced by human B lymphocytes inhibits apoptosis and Epstein-Barr virus reactivation. *Cell* 79;1137-1146, 1994
 33. Croen KD: Evidence for antiviral effect of nitric oxide. Inhibition of herpes simplex virus type 1 replication. *J Clin Invest* 91;2446-2452, 1993
 34. Gamba G, Cavalieri H, Courreges MC, Massouh EJ, Benencia F: Early inhibition of nitric oxide production increases HSV-1 intranasal infection. *J Med Virol* 73;313-322, 2004
 35. MacLean A, Wei XQ, Huang FP, Al-Alem UA, Chan WL, Liew FY: Mice lacking inducible nitric-oxide synthase are more susceptible to herpes simplex virus infection despite enhanced Th1 cell responses. *J Gen Virol* 79;825-830, 1998
 36. Akaike T, Okamoto S, Sawa T, Yoshitake J, Tamura F, Ichimori K, Miyazaki K, Sasamoto K, Maeda H: 8-nitroguanosine formation in viral pneumonia and its implication for pathogenesis. *Proc Natl Acad Sci U S A* 100;685-690, 2003
 37. Kreil TR, Eibl MM: Nitric oxide and viral infection: NO antiviral activity against a flavivirus *in vitro*, and evidence for contribution to pathogenesis in experimental infection in vivo. *Virology* 219;304-306, 1996
 38. Yoshitake J, Akaike T, Akuta T, Tamura F, Ogura T, Esumi H, Maeda H: Nitric oxide as an endogenous mutagen for Sendai virus without antiviral activity. *J Virol* 78;8709-8719, 2004
 39. Adler H, Beland JL, Del-Pan NC, Kobzik L, Brewer JP, Martin TR, Rimm IJ: Suppression of herpes simplex virus type 1 (HSV-1)-induced pneumonia in mice by inhibition of inducible nitric oxide synthase (iNOS, NOS2). *J Exp Med* 185;1533-1540, 1997
 40. Fujii S, Akaike T, Maeda H: Role of nitric oxide in pathogenesis of herpes simplex virus encephalitis in rats. *Virology* 256;203-212, 1999
 41. Bolovan-Fritts CA, Spector SA: Endothelial damage from cytomegalovirus-specific host immune response can be prevented by targeted disruption of fractalkine-CX3CR1 interaction. *Blood* 111;175-182, 2008
 42. Zhang M, Xin H, Atherton SS: Murine cytomegalovirus (MCMV) spreads to and replicates in the retina after endotoxin-induced disruption of the blood-retinal barrier of immunosuppressed BALB/c mice. *J Neurovirol* 11;365-375, 2005
 43. MacMicking JD, North RJ, LaCourse R, Mudgett JS, Shah SK, Nathan CF: Identification of nitric oxide synthase as a protective locus against tuberculosis. *Proc Natl Acad Sci U S A* 94;5243-5248, 1997
 44. Arias M, Rojas M, Zabaleta J, Rodríguez JL, París SC, Barrera LF, García LF: Inhibition of virulent *Mycobacterium tuberculosis* by Bcg(r) and Bcg(s) macrophages correlates with nitric oxide production. *J Infect Dis* 176;1552-1558, 1997
 45. Flesch IE, Hess JH, Kaufmann SH: NADPH diaphorase staining suggests a transient and localized contribution of nitric oxide to host defence against an intracellular pathogen in situ. *Int Immunol* 6;1751-1757, 1994.
 46. Doi T, Ando M, Akaike T, Suga M, Sato K, Maeda H: Resistance to nitric oxide in *Mycobacterium avium* complex and its implication in pathogenesis. *Infect Immun* 61; 1980-1989, 1993
 47. Scanga CA, Mohan VP, Yu K, Joseph H, Tanaka K, Chan J, Flynn JL: Depletion of CD4(+) T cells causes reactivation of murine persistent tuberculosis despite continued expression of interferon gamma and nitric oxide synthase 2. *J Exp Med* 192;347-358, 2000
 48. Aston C, Rom WN, Talbot AT, Reibman J: Early inhibition of mycobacterial growth by human alveolar macrophages is not due to nitric oxide. *Am J Respir Crit Care Med* 157;1943-1950, 1998
 49. Jagannath C, Actor JK, Hunter RL Jr: Induction of nitric oxide in human monocytes and monocyte cell lines by *Mycobacterium tuberculosis*. *Nitric Oxide* 2;174-186, 1998
 50. Kwon OJ: The role of nitric oxide in the immune response of tuberculosis. *J Korean Med Sci* 12;481-487, 1997
 51. Rich EA, Torres M, Sada E, Finegan CK, Hamilton BD, Toossi Z: *Mycobacterium tuberculosis* (MTB)-stimulated production of nitric oxide by human alveolar macrophages and relationship of nitric oxide production to growth inhibition of MTB. *Tuber Lung Dis* 78;247-255, 1997
 52. Rockett KA, Brookes R, Udalova I, Vidal V, Hill AV, Kwiatkowski D: 1,25-Dihydroxyvitamin D₃ induces nitric oxide synthase and suppresses growth of *Mycobacterium tuberculosis* in a human macrophage-like cell line. *Infect Immun* 66;5314-5321, 1998
 53. Nozaki Y, Hasegawa Y, Ichiyama S, Nakashima I, Shimokata K: Mechanism of nitric oxide-dependent killing of *Mycobacterium bovis* BCG in human alveolar macrophages. *Infect Immun* 65;3644-3647, 1997

54. Wang CH, Liu CY, Lin HC, Yu CT, Chung KF, Kuo HP: Increased exhaled nitric oxide in active pulmonary tuberculosis due to inducible NO synthase upregulation in alveolar macrophages. *Eur Respir J* 11;809-815, 1998
55. Facchetti F, Vermi W, Fiorentini S, Chilosi M, Caruso A, Duse M, Notarangelo LD, Badolato R: Expression of inducible nitric oxide synthase in human granulomas and histiocytic reactions. *Am J Pathol* 154;145-152, 1999
56. Kropf P, Baud D, Marshall SE, Munder M, Mosley A, Fuentes JM, Bangham CR, Taylor GP, Herath S, Choi BS, Soler G, Teoh T, Modolell M, Müller I: Arginase activity mediates reversible T cell hyporesponsiveness in human pregnancy. *Eur J Immunol* 37;935-945, 2007
57. Gazzinelli RT, Eltoun I, Wynn TA, Sher A: Acute cerebral toxoplasmosis is induced by *in vivo* neutralization of TNF- α and correlates with the down-regulated expression of inducible nitric oxide synthase and other markers of macrophage activation. *J Immunol* 151;3672-3681, 1993
58. Tucker PC, Griffin DE, Choi S, Bui N, Wesselingh S: Inhibition of nitric oxide synthesis increases mortality in Sindbis virus encephalitis. *J Virol* 70;3972-3977, 1996
59. Rock RB, Gekker G, Hu S, Sheng WS, Cheeran M, Lokensgard JR, Peterson PK: Role of microglia in central nervous system infections. *Clin Microbiol Rev* 17;942-964, 2004
60. Rock RB, Hu S, Deshpande A, Munir S, May BJ, Baker CA, Peterson PK, Kapur V: Transcriptional response of human microglial cells to interferon- γ . *Genes Immun* 6;712-719, 2005
61. Peterson PK, Hu S, Anderson WR, Chao CC: Nitric oxide production and neurotoxicity mediated by activated microglia from human versus mouse brain. *J Infect Dis* 170; 457-460, 1994
62. Mazzolla R, Puliti M, Barluzzi R, Neglia R, Bistoni F, Barbolini G, Blasi E: Differential microbial clearance and immunoresponse of Balb/c (Nramp1 susceptible) and DBA2 (Nramp1 resistant) mice intracerebrally infected with *Mycobacterium bovis* BCG (BCG). *FEMS Immunol Med Microbiol* 32;149-158, 2002
63. van Well GT, Wieland CW, Florquin S, Roord JJ, van der Poll T, van Furth AM: A new murine model to study the pathogenesis of tuberculous meningitis. *J Infect Dis* 195; 694-697, 2007
64. Olin MR, Armien AG, Cheeran MC, Rock RB, Molitor TW, Peterson PK: Role of nitric oxide in defense of the central nervous system against *Mycobacterium tuberculosis*. *J Infect Dis* 198;886-889, 2008
65. Kwon OJ, Kim JH, Kim HC, Suh GY, Park JW, Chung MP, Kim H, Rhee CH: Nitric oxide expression in airway epithelial cells in response to tubercle bacilli stimulation. *Respirology* 3;119-124, 1998
66. Roy S, Sharma S, Sharma M, Aggarwal R, Bose M: Induction of nitric oxide release from the human alveolar epithelial cell line A549: an *in vitro* correlate of innate immune response to *Mycobacterium tuberculosis*. *Immunology* 112; 471-480, 2004
67. Sharma M, Sharma S, Roy S, Varma S, Bose M: Pulmonary epithelial cells are a source of interferon-gamma in response to *Mycobacterium tuberculosis* infection. *Immunol Cell Biol* 85;229-237, 2007