

Antioxidant Activity of *Curcuma Longa* L., Novel Foodstuff

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Abstract

The potential antioxidant activities of different fractions from methanolic extract of *Curcuma longa* L. were assayed *in vitro*. All of the fractions exception of *n*-hexane and H₂O showed a strong antioxidant activity, especially the ethylacetate (EtOAc) fraction, which showed the highest 1,1-diphenyl-2-picrylhydrazyl (DPPH) free radical scavenging activity (IC₅₀ = 9.86 µg/mL). The results of 2,2'-azinobis (3-ethylbenzothiazoline-6-sulfonic acid) diammonium salt (ABTS) radical scavenging activity and ferric reducing antioxidant power (FRAP) assay showed concentration dependency, the EtOAc fraction demonstrating a better result than the other fractions at the same concentration in this studies. Additionally, when the total phenolic contents of fractions were measured, the EtOAc fraction contained the highest level. Meanwhile, correlation analysis indicated a high correlation between the antiradical activity and the total phenolic contents, suggesting that fractions obtained from the methanolic extract of *Curcuma longa* L. have wide potential for use as sources of antioxidant material.

Keywords: *Curcuma longa* L., Antioxidant, DPPH, ABTS, FRAP, Phenolic contents

Natural antioxidants in fruits and vegetables have been interested among consumers and the scientific community. Through epidemiological studies, it has been reported that frequent consumption of natural antioxidants is associated with prevention of cardiovascular disease and cancer^{1,2}. Oxygen Consumption involved in the respiration process generates harmful intermediates called reactive oxygen species (ROS). Excess ROS in the body are capable of cumulative

damaging in proteins, lipids, and DNA, resulting in so-called oxidative stress. Oxidative stress, defined as the imbalance between oxidants and antioxidants in favor of the oxidants³, has been suggested to be the cause of aging and various diseases in humans⁴. It could be partly attributed to the presence of antioxidant compounds, especially phenolic compounds, which are the most abundant hydrophilic antioxidants in the food and are involved in many biological activities^{5,6}. Dietary antioxidants can stimulate cellular defenses and help to prevent cellular components against oxidative damage⁷. For these usages, the most commonly used synthetic antioxidants are butylated hydroxyanisole (BHA), butylated hydroxytoluene (BHT), propylgallate, and *tert*-butyl hydroquinone. However, BHA and BHT have been suspected of inducing liver damage and carcinogenesis^{8,9}. Therefore, a need for identifying alternative natural and safe sources of dietary antioxidants has been created, and related studies have increased in recent years.

In East Asia, the rhizome of *Curcuma longa* L. (turmeric) is widely used as a spice, coloring, flavoring, and traditional medicinal preparations. It has been also considered as an analgesic in the treatment of menstrual disorders, rheumatism, and traumatic diseases due to a number of components, such as monoterpenoids, sesquiterpenoids, and curcuminoids¹⁰. Furthermore, it has been noted that the materials of rhizome of *Curcuma longa* L. have antiplatelet¹¹, fungicidal¹², and repellent¹³ properties. The constituents having antiplatelet and insect repellent properties in rhizome of *Curcuma longa* L. are curcuminoids¹¹ and turmerones¹⁴, respectively. Curcuminoids (curcumin, demethoxycurcumin, and bisdemethoxycurcumin), the phenolic yellowish pigments of *Curcuma longa* L., have been suggested to have antioxidative, anticarcinogenic, antiinflammatory, and hypocholesterolemic activities¹⁵.

There are several studies that these beneficial properties of *Curcuma longa* L. have been associated to the antioxidant activity¹⁶⁻¹⁸. So, this study examined the antioxidative activity of various fractions from methanolic extract of *Curcuma longa* L. against DPPH, ABTS, and FRAP and to estimate total phenolic contents. Additionally, the correlation between the total phenolic contents in the tested fractions and their antioxidant activities was investigated.

Table 1. DPPH free radical scavenging activity of different fractions from MeOH extracts of *Curcuma longa* L..

Fractions	IC ₅₀ ¹⁾ (µg/mL)
EtOAc	9.86
CHCl ₃	16.70
MeOH crude extract	58.17
<i>n</i> -BuOH	81.09
<i>n</i> -Hexane	280.42
H ₂ O	759.28
Trolox	17.19
Vit. C	2.44

¹⁾The effective concentration at which DPPH radical was scavenged by 50%.

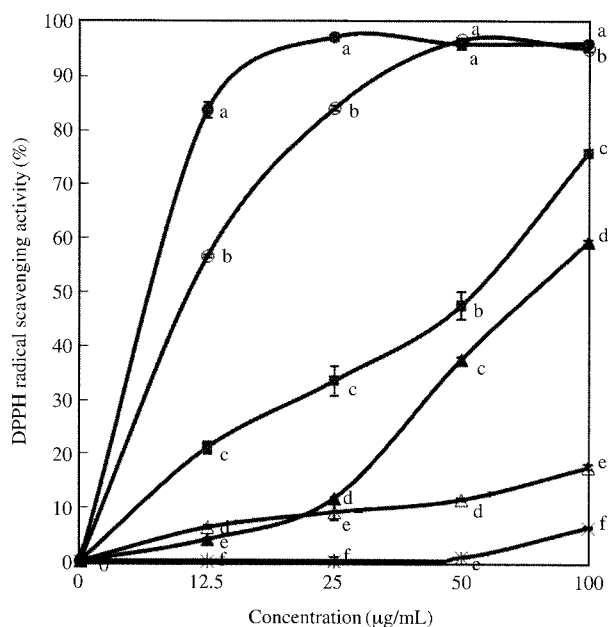


Figure 1. Radical scavenging activity of various fractions from methanolic extracts of *Curcuma longa* L. determined by the DPPH assay: (■) MeOH crude extract, (Δ) *n*-Hexane, (○) CHCl₃, (●) EtOAc, (▲) BtOH, (*) H₂O fraction. Results are mean ± SD (n=3).^{a,b,c,d,e,f} Different superscripts indicate significant differences at *P* < 0.05 in the each concentration.

DPPH Radical Scavenging Activity of Fractions of MeOH-Extracted *Curcuma Longa* L.

The free radical scavenging activity of its derived fraction of *Curcuma longa* L. was assessed by DPPH assay. The IC₅₀ values (the concentration required to inhibit radical formation by 50%) is a parameter widely used to measure that activity¹⁹. As shown in Table 1 and Figure 1, except for the *n*-hexane and H₂O fraction, all the test samples showed a significant antiradical activity against the DPPH radical (IC₅₀ range: 9.86-81.09 µg/mL). Among them, EtOAc fraction showed the highest scavenging activity. In addition, the IC₅₀

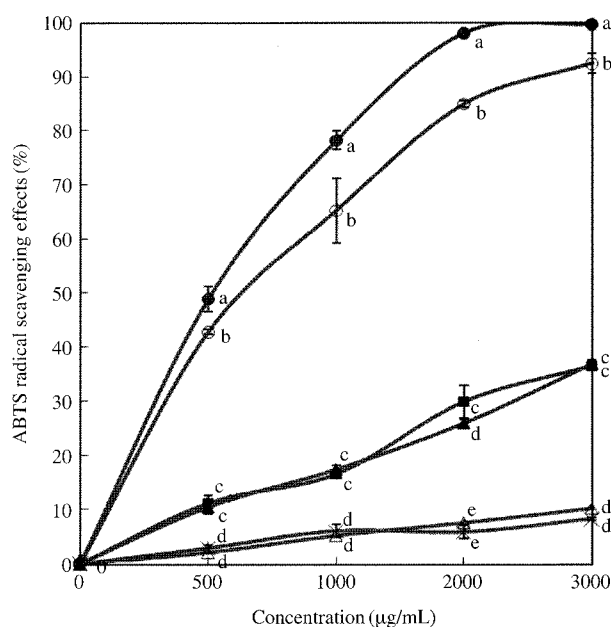


Figure 2. Radical scavenging activity of various fractions from methanolic extracts of *Curcuma longa* L. determined by the ABTS assay: (■) MeOH crude extract, (Δ) *n*-Hexane, (○) CHCl₃, (●) EtOAc, (▲) BtOH, (*) H₂O fraction. Results are mean ± SD (n=3).^{a,b,c,d,e} Different superscripts indicate significant differences at *P* < 0.05 in the each concentration.

values of the EtOAc, CHCl₃, MeOH crude extract, BuOH, *n*-Hexane, and H₂O fraction were 9.86, 16.70, 58.17, 81.09, 280.42, and 759.28 µg/mL, respectively. In this study, trolox and vitamin C were measured as the positive controls, showing the IC₅₀ values of 17.19 and 2.44 µg/mL, respectively.

ABTS Radical Scavenging Activity of Fractions of MeOH-Extracted *Curcuma Longa* L.

ABTS assay is based on the reaction between ABTS and potassium persulfate giving blue/green ABTS radical (ABTS^{•+}). With the addition of the antioxidants, decolorization is attained and measured spectrophotometrically at 734 nm. The ABTS radical scavenging activities of different fractions were increased with the sample concentrations (Figure 2). EtOAc and CHCl₃ fractions showed a high and dose-dependent ABTS radical scavenging activity. EtOAc fraction demonstrated the highest scavenging activity for the same concentration. The 5 fractions in descending order of strength of radical scavenging activity, were EtOAc > CHCl₃ > MeOH crude extract ≥ BuOH > *n*-Hexane ≥ H₂O fraction (Figure 2). The ABTS radical scavenging activity of the tested fractions was in the same descending order as the DPPH radical scavenging activity, and this fact might be associated with the relationship

between the antioxidant activity and the free radical scavenging activity of fractions.

Antioxidative Capacity of Fractions of MeOH-Extracted *Curcuma Longa* L. by FRAP Assay

The FRAP assay measures the reduction of ferric iron (Fe^{3+}) to ferrous iron (Fe^{2+}) in the presence of antioxidants, which are reductants with half-reaction reduction potentials above $\text{Fe}^{3+}/\text{Fe}^{2+}$. The ferric complexes reducing ability of different fractions was presented in Figure 3. The trend for the ferric ion reducing

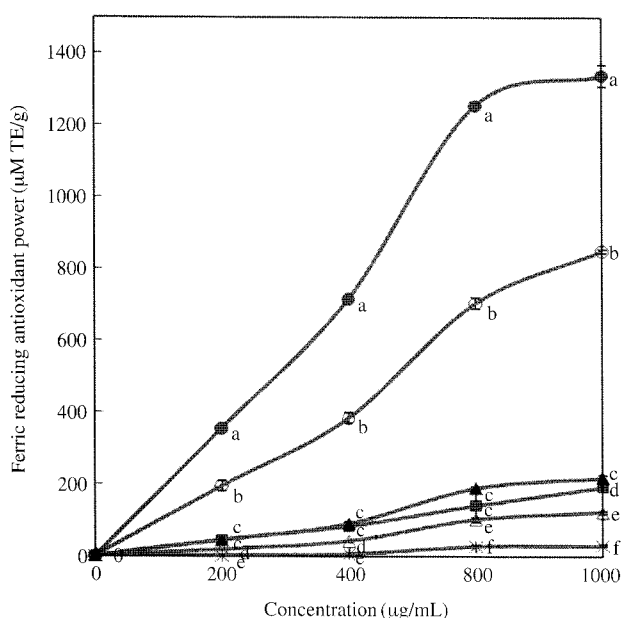


Figure 3. Antioxidant activity of various fractions from methanolic extracts of *Curcuma longa* L. determined by the FRAP assay. Data expressed as μM equivalents of Trolox (TE) released from 1 g of sample. FRAP assay: (■) MeOH crude extract, (Δ) *n*-Hexane, (\circ) CHCl_3 , (\bullet) EtOAc, (\blacktriangle) *n*-BuOH, ($*$) H_2O fraction. Results are mean \pm SD ($n=3$).^{a,b,c,d,e,f} Different superscripts indicate significant differences at $P < 0.05$ in the each concentration.

activities of 5 fractions tested was not markedly different from their DPPH and ABTS scavenging activities. Similar to the results obtained for radical scavenging assay, EtOAc and CHCl_3 fractions showed very strong ferric ion reducing activities for the same concentration. The 5 fractions in descending order of strength of ferric ion reducing activity, were EtOAc $>$ CHCl_3 $>$ BuOH $>$ MeOH crude extract $>$ *n*-Hexane $>$ H_2O fraction.

Total Phenolic Contents of Fractions of MeOH-Extracted *Curcuma Longa* L.

Plant phenolics, in general, are highly effective free radical scavengers and antioxidants. Therefore, the content of total phenolics in each fraction determined spectrophotometrically according to the Folin-Ciaccateu method was expressed as gallic acid equivalents (GAE) (Table 2). The total phenolic content of the EtOAc fraction was higher than that of the other fractions. These results indicated that the components having free radical scavenging activity of methanolic extracts of *Curcuma longa* L. could be effectively enriched in the EtOAc fraction. Thus, the EtOAc fraction will be further investigated for its phytochemical characteristics and *in vitro* antioxidant activity. The phenolic contents of EtOAc, CHCl_3 , MeOH crude extract, BuOH, *n*-Hexane, and H_2O fractions were 228.7, 140.7, 38.7, 30.7, 26.9, and 6.0 mg GAE/g. A strong positive corre-

Table 2. Total phenolic contents of different fractions from MeOH extract of *Curcuma longa* L..

Fraction	Total phenolic contents ¹⁾ (mg GAE/g)
EtOAc	228.7 \pm 2.3
CHCl_3	140.7 \pm 10.6
MeOH crude extract	38.7 \pm 6.7
<i>n</i> -BuOH	30.7 \pm 3.0
<i>n</i> -Hexane	26.9 \pm 7.4
H_2O	6.0 \pm 1.3

¹⁾Each value is expressed as mean \pm SD ($n=3$).

Table 3. Correlation coefficients (*R*) between the antioxidant properties and content of total phenol in *Curcuma longa* L. fractions.

	Total phenol content	Antioxidant activity ^a		
		DPPH radical scavenging activity	ABTS radical scavenging activity	Ferric reducing antioxidant power (FRAP)
Total phenol content	1.000			
DPPH radical scavenging activity	0.979***	1.000		
ABTS radical scavenging activity	0.914***	0.954***	1.000	
Ferric reducing antioxidant power (FRAP)	0.960***	0.969***	0.984***	1.000

^aThe correlations of *Curcuma Longa* L. fractions in the scavenging activity of DPPH radical, ABTS radical and ferric reducing antioxidant power used for correlation analysis were 25, 1,000, and 400 $\mu\text{g}/\text{mL}$, respectively. *** $P < 0.001$

lation has been reported between total polyphenolic contents and DPPH or ABTS radical scavenging activity^{20,21}, which had been strongly demonstrated in this study. DPPH and ABTS radical scavenging activities and the phenolic contents of 5 fractions were, in the same descending order: EtOAc > CHCl₃ > MeOH crude extract > BuOH > *n*-Hexane > H₂O fraction.

Correlations Analysis

The correlations between antioxidant contents and antioxidant activities were summarized in Table 3. There were positively high correlation between polyphenolic contents in the tested fractions and their antioxidant activities as determined by DPPH, ABTS, and FRAP assays. Significant correlations were found between the various methods used to determine the antioxidant potential, especially between ABTS and FRAP assays ($R=0.984$, $P<0.001$), DPPH and ABTS assays ($R=0.954$, $P<0.001$), and DPPH and FRAP assays ($R=0.969$, $P<0.001$). Results obtained from DPPH, ABTS, and FRAP assays were also correlated to the total phenolic compound concentration ($R=0.979$, $R=0.914$, and $R=0.960$, $P<0.001$).

Discussion

In this study, antioxidative capacities, total phenolic contents, and their correlations of the various fractions from methanolic extract of *Curcuma longa* L. were examined with useful methods.

DPPH, ABTS, and FRAP assays used for testing have been widely used to examine the antioxidant activities of plant extracts. It reported that these assays require relatively standard equipment and yield the fast and most reproducible results²². From the results, it was demonstrated that the EtOAc fraction from methanolic extract of *Curcuma longa* L. possessed an excellent antioxidant activity based on the DPPH, ABTS, and FRAP assay.

And significant correlations were also found between DPPH, ABTS, and FRAP assays and total phenolic contents. Phenolic compounds are found in both eatable and uneatable plants, which have various biological effects, especially, including antioxidant activity. Phenolic compounds having one or more aromatic rings bearing one or more hydroxyl groups can potentially quench free radicals by forming resonance-stabilized phenoxyl radicals and therefore have redox properties^{23,24}. In our results, strong positive correlations were found between DPPH, ABTS, and FRAP assays and total phenolic contents. These results indicated a relationship between phenolic compound concentration in various fractions from methanolic extract

of *Curcuma longa* L. and their free radical scavenging and ferric reducing capacities. Therefore, the presence of phenolic compounds in fractions contributes significantly to their antioxidant potential. This result is in agreement with previous studies that a highly positive relationship existed between total phenol contents and antioxidativity in many plants^{20,21}.

It was reported that the methanol extract of *Curcuma longa* L. exhibited scavenging activity against DPPH radicals²⁵ and its ethanol and water extracts exhibited antioxidant activity. However, there are few studies on antioxidant effects of various fractions of its extracts. From this study, we could know that antioxidant activity of *Curcuma longa* L. was due to constituents including in EtOAc and CHCl₃ fraction. These constituents may be curcuminoids, of phenolic constituents of *Curcuma longa* L., and its antioxidant activity was correlated to phenolic contents. Among these curcuminoids, curcumin with its proven anti-inflammatory and antioxidant properties had the strongest activity²⁵. It was shown to be a potent scavenger of a variety of ROS including hydroxyl radicals²⁶ and nitrogen dioxide radicals²⁷. It was also shown to inhibit lipid peroxidation in different animal models²⁶. And our results was similar that curcumin has an effective DPPH· scavenging, ABTS·⁺ scavenging, DMPD·⁺ scavenging, superoxide anion radical scavenging, hydrogen peroxide scavenging, ferric ions (Fe³⁺) reducing power, and ferrous ions (Fe²⁺) chelating activities²⁸.

In this study, it was demonstrated for the first time examined an excellent antioxidant activity testing assay and total phenolic contents of various fractions of the methanolic extracts from *Curcuma longa* L.. Some components including curcumin, of EtOAc and CHCl₃ fraction exhibited the strongest radical scavenging activity. Future studies should focus on the identification of these components and purification of this plant ingredient into better agents with high efficacy and activity.

Materials & Methods

Sample Preparation

The *Curcuma longa* L. was obtained from local medicine stores in Seoul, Korea. The dried powder (500 g) was extracted with solvent of MeOH for 3 hr (1 L × 3). This MeOH crude extract was collected by filtering and evaporating using a rotary evaporator at 45°C. The MeOH crude extract was subsequently fractionated with *n*-hexane, CHCl₃, EtOAc, *n*-BuOH, and H₂O with a separatory funnel and then each fraction was evaporated at 50°C to give 12.5 g of *n*-hexane, 22.65 g of CHCl₃, 3.95 g of EtOAc, 6.75 g of *n*-BuOH,

and 7 g of H₂O partitioned fractions, respectively.

DPPH Radical Scavenging Capacity Determinations

The free radical scavenging capacity of various fractions of *Curcuma longa* L. was evaluated by the method described by Gue and Wang²⁹ with some modifications. First, 4 mL of test sample in MeOH (final concentrations were 12.5, 25, 50, and 100 µg/mL, respectively) were mixed with 1 mL of 1.5×10^{-4} M DPPH solution. After 30 min of incubation at room temperature, the reduction of the DPPH free radical was measured by reading the absorbance at 517 nm. The DPPH radical scavenging activity was calculated according to the following equation:

$$\text{Scavenging rate} = [1 - (A_1 - A_2)/A_0] \times 100\%$$

where A_0 was the absorbance of the control (blank, without extract) and A_1 was the absorbance in the presence of the extract, A_2 was the absorbance without DPPH.

ABTS Radical Scavenging Activity Determinations

The scavenging activity of fractions on ABTS radical cation was measured according to the method of Re *et al.*³⁰ with slight modification. ABTS radical cation was generated by adding 7 mM ABTS to 2.45 mM potassium persulfate solution and the mixture was left to stand for overnight in the dark at room temperature. The ABTS radical cation solution was diluted with MeOH to obtain an absorbance of 0.7 ± 0.02 at 734 nm. 990 µL of ABTS radical cation solution and 10 µL of each sample were mixed and measured after 6 min at 734 nm. The ABTS radical scavenging activity was calculated according to the following equation:

$$\text{Scavenging rate} = [1 - (A_1 - A_2)/A_0] \times 100\%$$

where A_0 was the absorbance of the control (blank, without extract) and A_1 was the absorbance in the presence of the extract, A_2 was the absorbance without ABTS.

Antioxidative Capacity Determinations by FRAP Assay

The FRAP assay was performed according to the modified Benzie and Strain method³¹. The stock solutions included 300 mM acetate buffer (3.1 g C₂H₃NaO₂ · 3H₂O and 16 mL C₂H₄O₂), pH 3.6, 10 mM 2,4,6-tripyridyl-s-triazine (TPTZ) solution in 40 mM HCl and 20 mM FeCl₃ · 6H₂O solution. The fresh working solution was prepared by mixing 25 mL acetate buffer, 2.5 mL TPTZ solution and 2.5 mL FeCl₃ · 6H₂O solu-

tion and then warmed at 37°C before using. Different concentrations of various fractions (150 µL) were allowed to react with 2,850 µL of FRAP solutions for 30 min in the dark condition. Readings of the colored product (ferrous tripyridyltriazine complex) were then taken at 593 nm. The standard curve was linear between 25 and 800 µM Trolox. Results are expressed in µM Trolox equivalents (TE)/g used dried weight. Additional dilution was needed of the FRAP value measured was over the linear range of the standard curve.

Determination of Total Phenolic Contents

The phenolic compounds were determined by the Folin-Ciocalteu method, which was adapted from Swain and Hillis³². The 150 µL of various concentrations of different fractions, 2,400 µL of nanopure water, and 150 µL of 0.25 N Folin-Ciocalteu reagent were combined in a vial and then mixed well using a vortex. The mixture was allowed to react for 3 min then 300 µL of 1 N Na₂CO₃ solution was added and mixed well. The solution was incubated at room temperature (23°C) in the dark for 2 hr. The absorbance was measured at 725 nm using a UV/VIS spectrophotometer (Jasco V-530, Tokyo, Japan) and the results were expressed in mg gallic acid equivalents (GAE)/g sample using a gallic acid (0-0.3 mg/mL) standard curve. Additional dilution was done if the absorbance value measured was over the linear range of the standard curve.

Statistical Analysis

Each antioxidant activity assay was done three times from the same extract in order to determine their reproducibility. Statistical significance was evaluated by one-way analysis (ANOVA) of variance using SAS version 9.1. Duncan's new multiple range test was used to determine significant differences. Correlations among data obtained were calculated using Pearson's correlation coefficient (*R*).

References

1. Liu, S. *et al.* Intake of vegetables rich in carotenoids and risk of coronary heart disease in men: the physicians' health study. *Int J Epidemiol* **30**:130-135 (2001).
2. Bazzano, L. A. *et al.* Fruit and vegetable intake and risk of cardiovascular disease in US adults: the first national health and nutrition examination survey epidemiologic follow-up study. *Am J Clin Nutr* **76**:93-99 (2002).
3. Sies, H. Oxidative stress: oxidants and antioxidants. *Exp Physiol* **82**:291-295 (1997).
4. Halliwell, B. & Gutteridge, J. M. Role of free radicals and catalytic metal ions in human disease: an overview.

- Method Enzymol* **186**:1-85 (1990).
5. Kahkonen, M. P. *et al.* Antioxidant activity of plant extracts containing phenolic compounds. *J Agric Food Chem* **47**:3954-3962 (1999).
 6. Sugihara, N., Arakawa, T., Ohnishi, M. & Furuno, K. Anti- and prooxidative effects of flavonoids on metal induced lipid hydroperoxide dependent lipid peroxidation in cultured hepatocytes located with α -linolenic acid. *Free Radic Biol Med* **27**:1313-1323 (1999).
 7. Lorenzo, Y. *et al.* The carotenoid beta-cryptoxanthin stimulates the repair of DNA oxidation damage in addition to acting as an antioxidant in human cells. *Carcinogenesis* **30**:308-314 (2009).
 8. Grice, H. C. Safety evaluation of butylated hydroxyanisole from the perspective of effects on forestomach and oesophageal squamous epithelium. *Food Chem Toxicol* **26**:717-723 (1988).
 9. Sherwin, E. R. *Antioxidants*. In: Branen AL, Davidson PM, Salminen S (eds) Food additives. Marvel Dekker inc, New York, p. 139-193 (1990).
 10. Tang, W. & Eisenbrand, G. Chinese Drugs of Plant Origin. SpringerVerlag, Berlin Heidelberg, Germany (1992).
 11. Lee, H. S. Antiplatelet property of *Curcuma longa* L. rhizome-driven ar-turmerone. *Bioresource Technol* **97**:1372-1376 (2006).
 12. Kim, M. K., Choi, G. J. & Lee, H. S. Fungicidal property of *Curcuma longa* L. rhizome-driven curcumin against phytopathogenic fungi in greenhouse. *J Agr Food Chem* **51**:1587-1581 (2003).
 13. Jilani, G. & Saxena. R. C. Repellent and feeding deterrent effects of turmeric oil, sweetflag oil, neem oil, and neem-based insecticide against lesser grain borer (Coleoptera: Bostrychidae). *J Econ Entomol* **83**:629-634 (1990).
 14. Lee, H. S. *et al.* Insecticidal activities of ar-turmerone identified *Curcuma longa* L. rhizome against *Nilaparvata lugens* (Homoptera: Delphacidae) and *Plutella xyostella* (Lepidoptera: Yponomeutidae). *J Asia-Pacific Entomol* **4**:181-185 (2001).
 15. Ammon, H. P. T. & Wahl, M. A. Pharmacology of *Curcuma longa*. *Planta Med* **57**:1-7 (1991).
 16. Ramírez-Tortosa, M. C. *et al.* Oral administration of a turmeric extract inhibits LDL oxidation and has hypocholesterolemic effects in rabbits with experimental atherosclerosis. *Atherosclerosis* **147**:371-378 (1999).
 17. Devasagayam, T. P. A., Kamat, J. P. & Sreejayan, N. *Antioxidant action of curcumin*. In Micronutrients and Health: Molecular Biological Mechanisms. Nesaret-nam K, Packer L (eds). AOCs Press: Champaign, IL, USA, p. 42-59 (2001).
 18. Devasagayam, T. P. A. & Sainis, K. B. Immune system and antioxidants, especially those derived from Indian medicinal plants. *Indian J Exp Biol* **40**:639-655 (2002).
 19. Cuvelier, M. E., Richard, H. & Berset, C. Comparison of the antioxidative activity of some acid-phenols: Structure-activity relationships. *Biosci Biotech Bioch* **56**:324-325 (1992).
 20. Katalinic, V., Milos, M., Kulisic, T. & Jukic, M. Screening of 70 medicinal plant extracts for antioxidant capacity and total phenols. *Food Chem* **94**:550-557 (2006).
 21. Wong, C. C., Li, H. B., Cheng, K. W. & Chen, F. A. Systematic survey of antioxidant activity of 30 chinese medicinal plants using the ferric reducing antioxidant power assays. *Food Chem* **97**:705-711 (2006).
 22. Buenger, J. *et al.* An interlaboratory comparison of methods used to assess antioxidant potentials. *Int J Cosmet Sci* **28**:135-146 (2006).
 23. Rice-Evans, C. A., Miller, N. J. & Paganga, G. Structure-antioxidant activity relationships of flavonoids and phenolic acid. *Free Radical Biol Med* **20**:933-956 (1996).
 24. Bors, W. & Michel, C. Chemistry of the antioxidant effect of polyphenols. *Ann NY Acad Sci* **957**:57-69 (2002).
 25. Pozharitskaya, O. N., Ivanova, S. A., Shikov, A. N. & Makarov, V. G. Separation and free radical-scavenging activity of major curcuminoids of *Curcuma longa* using HPTLC-DPPH method. *Phytochem Anal* **19**:236-243 (2008).
 26. Reddy, A. C. & Lokesh, B. R. Studies on the inhibitory effects of curcumin and eugenol on the formation of reactive oxygen species and the oxidation of ferrous iron. *Mol Cell Biochem* **137**:1-8 (1994).
 27. Sreejayan, N. & Rao, M. N. Curcuminoids as potent inhibitors of lipid peroxidation. *J Pharm Pharmacol* **46**:1013-1016 (1994).
 28. Ak, T. & Gülçin, I. Antioxidant and radical scavenging properties of curcumin. *Chem Biol Interact* **174**:27-37 (2008).
 29. Gue, J. & Wang, M. H. Antioxidant and antidiabetic activities of *Ulmus davidiana* extracts. *Food Sci Biotechnol* **16**:55-66 (2007).
 30. Re, R. *et al.* Antioxidant activity applying an improved ABTS radical cation decolorization assay. *Free Radical Bio Med* **26**:1231-1237 (1999).
 31. Benzie, I. F. F. & Strain, J. J. The ferric reducing ability of plasma (Frap) as a measure of antioxidant power the Frap assay. *Anal Biochem* **239**:70-76 (1996).
 32. Swain, Y. & Hillis, W. E. The phenolic constituents of *Prunus Domestica* I-the quantitative analysis of phenolic constituents. *J Sci Food Agr* **10**:63-68 (1959).