# Synthesis of Facial Amphiphile 3,7-Diamino-5α-cholestane Derivatives as a Molecular Receptor

Md. Wasi Ahmad, Young Mee Jung, Sharaf Nawaz Khan, and Hong-Seok Kim

Department of Applied Chemistry, Kyungpook National University, Daegu 702-701. Korea. 'E-mail: kimhs@knu.ac.kr Received July 21, 2009, Accepted August 4, 2009

A series of facial amphiphiles 3,7-diaminocholestane were synthesized from 3,7-diketocholestane via 2 sequential reductive aminations and anion recognition was evaluated with acetate, chloride, bromide, fluoride and phosphate anions. The stereo-selective reductive amination protocol was utilized to synthesized facial amphiphiles afforded receptors in high yields. The molecular receptor 2 showed the highest binding constant with acetate in a 1:1 ratio.

Key Words: Facial amphiphile. Anion receptor. Reductive amination. Aminosteroids. Stereoselectivity

#### Introduction

Design of preorganized molecular receptor provides the advantage of rigid scaffolds that could be functionalized further with ligands for anion recognition. Steroid molecules are notorious for their preorganized structures provide a platform to constitute a molecular receptor and offer ease of functionalization. The ligands for the H-bonding e.g. amine group in the steroid scaffold of cholic acid were introduced through ether, ester or amide linkages.<sup>1</sup> The hydroxyl group could be transformed to ether linkages or ester linkages conveniently but were found to be unstable in higher pH values.<sup>2</sup> Hence introduction of the amine group directly to steroids were cynosure in steroid based molecular receptor syntheses.<sup>3</sup>

Cholic acid scaffolds having a *cis* AB ring or 5 $\beta$  configuration supports functionalization at C3. C7 and C12 while cholesterol-based receptors having an AB *trans* ring or 5 $\alpha$  configuration supports modification at C3 and C7. The advantage of cholesterol-based receptors is that they offer exactly the same bond length attachment of ligands at C3 and C7 in an axial manner, while in the same position with 5 $\beta$ -configuration it is not possible. The introduction of an axial amino group at C3 and C7 could be derivatized rapidly in a quantitative yield. The preferred method by Davis *et al.* to introduce the NH<sub>2</sub> group was through inversion at stereogenic centers and azide formation or by oximation and metal-assisted reduction in a multi-step synthesis afforded overall low yield.<sup>4</sup> The amino group becomes highly hydrophilic, which gives an edge in intramolecular hydrogen bonding and solubility in non-polar



Figure 1. Perspective drawing of cholesterol-based molecular receptor.

solvents. Figure 1 showed cholesterol based facial amphiphilic anionic receptors. which was derivatized with urea at C3 and C7 in an axial fashion. A highly stereoselective synthesis of 3  $\alpha$ .7 $\alpha$ -diaminocholestane (1) from 3 $\beta$ -acetoxy-5 $\alpha$ -cholest-7-one (3) by reductive amination methodology has been investigated, and was further elaborated on to synthesize anionic receptors.<sup>5</sup>

To get the facial amphiphile 2. one-step direct reductive amination of diketone 4 with NH<sub>4</sub>OAc in the presence of NaBH<sub>3</sub>CN was carried out which resulted in the formation of 6a (34%) along with a mixture of  $3\alpha/3\beta$ -isomers of 3.7-dihydroxycholestane. To improve the yield and stereo-selectivity of 6a, the sequential procedure was investigated.

### **Experimental Section**

Melting points were determined using a Thomas-Hoover capillary melting point apparatus and are uncorrected. The NMR spectra were recorded on a Bruker AM-400 spectrometer in CDCl<sub>3</sub> using Me<sub>4</sub>Si as the internal standard. Elemental analyses were performed on a Calro Erba 1106 at the Center for Scientific Instruments, Kyungpook National University. HR-FAB Mass spectra were taken at KBSI Daegu branch. TLC analyses were carried out on a plate precoated with 0.2 mm of HPTLC silica gel 60; substances were visualized by spraying with 5% ammonium molybdate in 10% H<sub>2</sub>SO<sub>4</sub> followed by heating. Flash column chromatography was performed with Merck silica gel 60 (70 - 230 mesh). Reactions were washed with brine and dried over anhydrous sodium sulfate. Compound **4** was obtained by literature method.<sup>5</sup>

**3a-(tert-Butyloxycarbonyl)amino-5a-cholestane (5a).** NaBH  $(OEh)_3^8$  (4 mL. 2 eq) was added to a solution of **4** (200 mg, 0.50 mmol) and NH<sub>4</sub>OTf (250 mg, 1.50 mmol) in a dry THF (10 mL) and stirred at room temperature for 1 h. After the solvent was removed, the residue was extracted with ethyl acetate. The organic layer was washed, dried, and concentrated. Without further purification, the residue was treated with (Boc)<sub>2</sub>O (164 mg, 0.75 mmol) in methanol (20 mL) for 3 h. After the solvent was removed, the residue was extracted with ethyl acetate. The organic layer was washed, dried, and concentrated.

centrated. The residue was purified by a column chromatography (elution with 5% EtOAc-hexane) to give 190 mg of **5a** (76%) and 23 mg of 3 $\beta$ -isomer (9%). **5a**: TLC R<sub>f</sub> 0.52 (EtOAchexane 1:4): mp 169 - 172 °C (CH<sub>2</sub>Cl<sub>2</sub>-hexane): <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  0.59 (s. 3H. 18-CH<sub>3</sub>), 0.80 (d. *J* = 6.5 Hz, 3H. 26-CH<sub>3</sub>), 0.84 (d. *J* = 6.5 Hz, 3H. 27-CH<sub>3</sub>), 0.86 (d. *J* = 6.5 Hz, 3H. 21-CH<sub>3</sub>), 1.00 (s. 3H, 19-CH<sub>3</sub>), 1.40 (s. 9H. -COC(CH<sub>3</sub>)<sub>3</sub>), 3.83 (s. 1H. 3 $\beta$ -H), 4.88 (s. 1H. 3 $\alpha$ -NH); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  11.0, 12.2, 14.2, 18.9, 21.3, 22.6, 22.9, 23.8, 25.0, 26.2, 28.1, 28.5, 29.2, 30.2, 32.9, 33.5, 35.7, 36.2, 36.5, 38.8, 39.6, 42.6, 43.1, 46.0, 49.1, 50.3, 55.1, 55.9, 65.3, 79.2, 155.3, 212.1; FAB-MS Calcd for C<sub>32</sub>H<sub>35</sub>NO<sub>3</sub>Na: 524.4080, Found: *m/z* 524.4080 [M+Na]<sup>-</sup>.

**3α-(***n***-Butyl)amino-5α-cholestan-7-one (5b).** yield: 77%. TLC R<sub>f</sub> 0.75 (CH<sub>2</sub>Cl<sub>2</sub>-MeOH-NH<sub>4</sub>OH 20:1:0.2); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 0.63 (s. 3H. 18-CH<sub>3</sub>). 0.78 (s. 3H. 19- CH<sub>3</sub>). 0.84 (d. 3H, J = 6.6 Hz. 26-CH<sub>3</sub>), 0.85 (d. 3H, J = 6.6 Hz. 27-CH<sub>3</sub>), 0.87 (d. 3H. J = 6.3 Hz. 21-CH<sub>3</sub>). 1.74-1.94 (m. 3H). 2.34 (dt. J = 11.2, 7.1 Hz. 1H), 2.53 (t, J = 7.3 Hz. 2H). 2.69 (m, 1H), 2.82 (bm. 1H. 3β-H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 11.2, 12.2, 14.1, 18.9, 20.6, 21.3, 22.7, 22.9, 23.9, 25.1, 25.7, 28.1, 28.6, 32.3, 32.8, 35.8, 36.2, 36.7, 38.8, 39.6, 41.8, 42.6, 46.2, 47.0, 49.0, 50.3, 52.4, 55.1, 55.5, 212.3; FAB-MS Calcd for C<sub>31</sub>H<sub>56</sub>NO: 458.4362, Found: *m/z* 458.4342 (M+H)<sup>-</sup>.

**3***a*-*N*-**[(3**-*tert*-butyloxycarbonyl)propylamino]-5*α*-cholestan-7-one (5c), yield: 72%, TLC R<sub>f</sub> 0.61 (CH<sub>2</sub>Cl<sub>2</sub>-MeOH-NH<sub>4</sub>OH 20:1.5:0.5); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 0.60 (s. 3H, 18-CH<sub>3</sub>), 0.81 (d. 3H, J = 6.6 Hz, 26-CH<sub>3</sub>), 0.82 (d. 3H, J = 6.6 Hz, 27-CH<sub>3</sub>), 0.86 (d, 3H, J = 6.3 Hz, 21-CH<sub>3</sub>), 1.00 (s, 3H, 19-CH<sub>3</sub>), 1.39 (s. 9H, -COC(CH<sub>3</sub>)<sub>3</sub>), 2.12 (bs. 1H, 3*α*-NH), 2.12 (t. J = 13.4Hz, 1H), 2.39 (t. J = 11.4 Hz, 1H), 2.59 (bt. 2H, HNCH<sub>2</sub>), 2.84 (bs. 1H, 3β-H), 3.16 (bm. 1H, CH<sub>2</sub>NH-Boc), 5.41 (bs. 1H, NH-Boc); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 11.4, 12.4, 19.1, 21.5, 22.9, 23.1, 24.1, 25.3, 28.3, 28.5, 28.7, 28.8, 29.3, 32.3, 33.1, 36.0, 36.5, 36.9, 39.0, 39.5, 39.9, 42.0, 42.9, 45.7, 46.3, 49.2, 50.5, 53.2, 55.4, 55.5, 79.3, 156.7, 212.4; FAB-MS Calcd for C<sub>35</sub>H<sub>63</sub>N<sub>2</sub>O<sub>3</sub>: 559.4839, Found: *m/z* 559.4842 (M+H)<sup>+</sup>.

**3α-N-[(4-***tert***-butyloxycarbonyl)butylamino]-5α-chole stan-7-one (5d),** yield 76%, TLC R<sub>f</sub> 0.62 (CH<sub>2</sub>Cl<sub>2</sub>-MeOH-NH<sub>4</sub>OH 15:2:0.5); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 0.59 (s, 3H, 18-CH<sub>3</sub>), 0.80 (d, 3H, J = 7.0 Hz, 26-CH<sub>3</sub>), 0.81 (d, 3H, J = 7.0 Hz, 27-CH<sub>3</sub>), 0.85 (d, 3H, J = 7.5 Hz, 21-CH<sub>3</sub>), 1.00 (s, 3H, 19-CH<sub>3</sub>), 1.38 (s, 9H, -COC(CH<sub>3</sub>)<sub>3</sub>), 2.57 (m, 2H, HN(Boc)CH<sub>2</sub>), 2.93 (bs, 1H, 3β-H), 3.06 (bs, 1H, NH-Boc), 4.93 (bs, 1H, 3α-NH); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 11.4, 12.4, 12.7, 14.5, 19.1, 21.5, 22.9, 23.1, 23.2, 24.1, 25.3, 26.2, 26.5, 28.3, 28.8, 30.4, 32.3, 32.8, 36.0, 36.4, 36.8, 39.0, 39.8, 40.5, 41.9, 42.8, 46.3, 46.7, 49.2, 50.5, 52.7, 55.3, 55.6, 79.2, 156.4, 212.3; FAB-MS Calcd for C<sub>36</sub>H<sub>65</sub>N<sub>2</sub>O<sub>3</sub>: 573.4995, Found: *m/z* 573.4998 (M+H)<sup>+</sup>.

**3a**-*N*-[(**4**,**9**-**Di**-*tert*-**butyloxycarbonyl)spemidinyl**]-**5a**-**cho**lestan-7-one (5e). yield 78%. TLC R<sub>f</sub> 0.69 (CH<sub>2</sub>Cl<sub>2</sub>- MeOH-NH<sub>4</sub>OH 15:2:0.5); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  0.59 (s. 3H. 18-CH<sub>3</sub>). 0.80 (d. 3H. *J* = 6.5 Hz. 26-CH<sub>3</sub>), 0.81 (d. 3H. *J* = 6.5 Hz. 27-CH<sub>3</sub>). 0.85 (d. 3H. *J* = 6.5 Hz. 21-CH<sub>3</sub>), 0.99 (s. 3H. 19-CH<sub>3</sub>) 1.38 (s, 18H. -COC(CH<sub>3</sub>)<sub>3</sub>), 2.51 (bm, 2H, CH<sub>2</sub>NH), 2.83 (s. 1H. 3α-H). 2.90 (bs. 1H. 3β-H). 3.00-3.15 (bm. 6H. CH<sub>2</sub>). 4.62 (bs. 1H, NHBoc): <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  11.2, 12.2, 14.3, 18.9, 21.3, 22.7, 22.9, 23.9, 25.1, 25.4, 25.7, 25.9, 27.5, 28.1, 28.5, 29.8, 32.2, 32.4, 32.6, 35.8, 36.2, 36.7, 38.8, 39.6, 40.3. 41.7, 42.0, 42.6, 44.5, 45.4, 46.1, 46.7, 47.0, 49.1, 50.3, 53.0, 55.1, 55.4, 55.8, 79.4, 79.7, 155.7, 156.1, 212.1; FAB Mass Calcd for  $C_{44}H_{80}N_3O_5$ : 730.6098, Found: *m/z* 730.6086 (M+H)<sup>-</sup>.

**3α-[3-(1-Imidazolyl)propyl]amino-5α-cholestan-7-one (5f).** yield: 81%, TLC R<sub>f</sub> 0.64 (CH<sub>2</sub>Cl<sub>2</sub>-MeOH-NH<sub>4</sub>OH 15:2:0.5); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 0.60 (s. 3H. 18-CH<sub>3</sub>). 0.81 (d. J = 6.6 Hz. 3H. 26-CH<sub>3</sub>), 0.82 (d, J = 6.6 Hz. 3H. 27-CH<sub>3</sub>), 0.86 (d. J = 6.6 Hz. 3H, 21-CH<sub>3</sub>), 1.00 (s. 3H, 19-CH<sub>3</sub>), 2.26 (m. 2H, HNCH<sub>2</sub> CH<sub>2</sub>CH<sub>2</sub>Im). 2.58 (bs. 2H. HNCH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>Imi). 2.94 (bs. 1H. 3β-H), 4.03 (bm, 2H, CH<sub>2</sub>-Im). 5.69 (bs.1H, NH), 6.91 (s. 1H. Im H-5). 6.99 (s. 1H. Im H-4). 7.52 (s. 1H. Im H-2); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 10.1, 11.0, 17.8, 20.2, 21.5, 21.8, 22.8, 23.9, 26.9, 27.4, 28.7, 28.7, 30.9, 34.7, 35.1, 35.5, 37.6, 38.4, 40.7, 41.5, 42.7, 43.6, 44.9, 45.9, 47.9, 48.9, 49.2, 54.1, 54.4, 60.7, 71.3, 117.9, 128.2, 136.2, 211.4; FAB-MS Calcd for C<sub>33</sub>H<sub>56</sub>N<sub>3</sub>O: 510.4424, Found: *m*/*z* 510.4421 (M+H)<sup>+</sup>.

**3a-[2-(2-Pyridyl)ethyl]amino-5a-cholestan-7-one (5g).** yield: 84%. TLC R<sub>f</sub> 0.47 (CH<sub>2</sub>Cl<sub>2</sub>-MeOH-NH<sub>4</sub>OH 20:1.5:0.5); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  0.62 (s. 3H, 18-CH<sub>3</sub>), 0.82 (d. *J* = 6.6 Hz, 3H, 26-CH<sub>3</sub>), 0.83 (d. *J* = 6.6 Hz, 3H, 27-CH<sub>3</sub>), 0.88 (d. *J* = 6.3 Hz. 3H. 21-CH<sub>3</sub>), 1.04 (s. 3H. 19-CH<sub>3</sub>), 3.15 (bs. 1H. 3β-H), 3.57 (m. 2H, NHCH<sub>2</sub>CH<sub>2</sub>Py). 3.71 (m, 2H. NHCH<sub>2</sub>CH<sub>2</sub>Py) 4.39 (bs. 1H, NH). 7.16 (d. 1H. *J* = 7.6 Hz, 1H), 7.17 (d. 1H. *J* = 7.8 Hz. 1H). 7.61 (ddd, 1H, *J* = 7.8, 7.6, 1.5 Hz. 1H). 8.37 (d, 1H, *J* = 4.8, Hz. 1H); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  11.2, 12.1, 18.8, 21.2, 22.6, 22.8, 23.8, 24.8, 25.0, 28.0, 28.4, 35.7, 36.2, 36.5, 38.7, 39.5, 42.2, 42.5, 45.8, 46.0, 49.0, 50.3, 53.1, 55.0, 55.9, 61.6, 72.4, 122.1, 123.7, 137.1, 148.7, 159.4, 211.9; FAB-MS Calcd for C<sub>3</sub><sub>3</sub>H<sub>55</sub>N<sub>2</sub>O: 507.4314, Found: *m/z* 507.4317 (M+H)<sup>+</sup>.

**3a-(2-Pyiidylmethyl)amino-5a-cholestan-7-one (5h).** yield: 78%. TLC R<sub>f</sub> 0.60 (CH<sub>2</sub>Cl<sub>2</sub>-MeOH-NH<sub>4</sub>OH 20:1.5:0.5): <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  0.61 (s. 3H, 18-CH<sub>3</sub>), 0.82 (d. *J* = 6.6 Hz, 3H, 26-CH<sub>3</sub>), 0.83 (d. *J* = 6.6 Hz, 3H, 27-CH<sub>3</sub>), 0.87 (d. *J* = 6.3 Hz, 3H, 21-CH<sub>3</sub>), 1.02 (s. 3H, 19-CH<sub>3</sub>), 3.00 (bs, 1H, 3β-H), 3.89 (d. 2H, *J* = 10.6 Hz, NH*CH*<sub>2</sub>Py), 3.94 (bs, 1H, NH), 7.14 (dd, 1H, *J* = 7.8, 7.8 Hz, 1H), 7.31 (d. 1H, *J* = 7.8, Hz, 1H), 7.61 (ddd, 1H, *J* = 7.8, 7.5, 1.8 Hz, 1H), 8.50 (d. 1H, *J* = 4.5 Hz, 1H); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  11.1, 12.1, 18.8, 21.2, 22.6, 22.9, 23.8, 25.1, 25.7, 28.0, 28.5, 32.3, 32.9, 35.7, 36.2, 36.7, 38.8, 39.5, 41.8, 42.6, 46.1, 49.0, 50.3, 52.4, 52.5, 55.0, 55.6, 122.2, 122.7, 136.7, 149.1, 158.8, 212.2; FAB-MS Calcd for C<sub>33</sub>H<sub>53</sub>N<sub>2</sub>O: 493.4158, Found: *m/z* 493.4158 (M+H)<sup>+</sup>.

**3α-[Di-(2-pyridylmethyl)amino]-5α-cholestan-7-one (5i).** yield: 56%. TLC R<sub>f</sub> 0.50 (CH<sub>2</sub>Cl<sub>2</sub>-MeOH-NH<sub>4</sub>OH 20:1.5:0.5); H NMR (CDCl<sub>3</sub>) δ 0.55 (s. 3H. 18-CH<sub>3</sub>). 0.77 (d. J = 6.6 Hz. 3H. 26-CH<sub>3</sub>), 0.78 (d, J = 6.6 Hz. 3H. 27-CH<sub>3</sub>), 0.82 (d. J = 6.6 Hz. 3H. 21-CH<sub>3</sub>), 0.95 (s. 3H. 19-CH<sub>3</sub>), 2.99. (bs. 1H. 3β-H). 3.82 (d. J = 15.2 Hz, 2H. CH<sub>2</sub>), 3.98 (d, J = 15.2 Hz. 2H, CH<sub>2</sub>). 7.05 (dd. J = 7.3, 7.3 Hz, 2H). 7.29 (d. J = 7.8 Hz, 2H). 7.54 (ddd. J = 7.8, 7.6, 1.0 Hz, 2H). 8.40 (dd. J = 5.0, 1.0 Hz, 2H). <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 11.8, 12.1, 18.8, 21.2, 22.6, 22.8, 23.8, 24.8, 25.0, 28.0, 28.4, 31.8, 33.2, 35.7, 36.1, 36.5, 38.8, 39.5, 42.4, 42.5, 45.8, 48.9, 50.2, 55.1, 56.2, 57.9, 58.0, 61.6, 72.4, 122.0, 123.4, 136.6, 148.6, 159.5, 212.3.

3a,7a-Bis(*tert*-butyloxycarbonylamino)-5a-cholestane (6a). NaBH<sub>3</sub>CN (79 mg. 1.20 mmol) was added to a mixture of 5a (200 mg, 0.40 mmol), NH<sub>4</sub>OAc (925 mg. 12.00 mmol), and bromocresol green in THF-MeOH ( $\nu\nu$ ; 1:1, 20 mL) and stirred at room temperature for 2 h. At the same time, pH was adjusted to 6 by acetic acid. After the solvent was removed, the residue was neutralized with aqueous NaOH solution, and extracted with CH<sub>2</sub>Cl<sub>2</sub>. The organic layer was washed, dried and concentrated. The residue was treated with (Boc)<sub>2</sub>O (131 mg, 0.60 mmol) in methanol (10 mL). After 1 h, a few drops of 1N NaOH solution were added to the reaction mixture and stirred for 5 h. The solvent was removed and residue was extracted with CH<sub>2</sub>Cl<sub>2</sub>. The organic layer was washed, dried, and concentrated. The residue was purified by column chromatography (10% EtOAc-hexane) to give 175 mg of **6a** (71%). TLC R<sub>f</sub> 0.33 (EtOAc-hexane).<sup>8</sup>

**3a**,7a-**Bis**[*(n*-butyl)amino]-5α-cholestane (6b). yield: 80%. TLC R<sub>f</sub> 0.56 (CH<sub>2</sub>Cl<sub>2</sub>-MeOH-NH<sub>4</sub>OH 20:1.5:0.5); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 0.58 (s. 3H. 18-CH<sub>3</sub>), 0.74 (s. 3H. 19-CH<sub>3</sub>), 0.80 (d. 3H. J = 7.0 Hz. 26-CH<sub>3</sub>), 0.81 (d. 3H. J = 7.0 Hz, 27-CH<sub>3</sub>), 0.84 (d. 3H. J = 7.5 Hz. 21-CH<sub>3</sub>), 0.85 (t. J = 7.5 Hz. CH<sub>3</sub>), 1.70 - 1.88 (m. 2H. CH<sub>2</sub>), 2.48 (bt. J = 7.5 Hz, 2H. CH<sub>2</sub>), 2.54 (bm. 1H. 7β-H), 2.76 (bs, 1H, 3β-H); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 11.0, 11.9, 14.2, 14.2, 18.7, 20.8, 22.7, 23.0, 23.7, 23.9, 26.0, 28.1, 31.7, 31.8, 32.6, 32.7, 32.7, 32.9, 35.9, 36.3, 36.7, 39.2, 39.6, 39.6, 42.7, 46.4, 47.1, 47.6, 50.8, 52.3, 54.6, 56.2; Anal. Calcd for C<sub>35</sub>H<sub>68</sub>Cl<sub>2</sub>N<sub>2</sub>; C, 71.51, H. 11.66, N 4.77, Found: C, 71.20, H, 12.47, N, 4.86; FAB-MS Calcd for C<sub>35</sub>H<sub>67</sub>N<sub>2</sub>; 515.5304, Found: *m/z* 515.5300 (M+H)<sup>+</sup>.

**3a**,7**a**-**Bis**[*N*-(3-*tert*-butyloxycarbonyl)propylamino]-5**a**cholestane (6c). yield: 71%. TLC R<sub>f</sub> 0.33 (CH<sub>2</sub>Cl<sub>2</sub>-MeOH-NH<sub>4</sub>OH 20:1.5:0.5): <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  0.67 (s, 3H. 18-CH<sub>3</sub>), 0.84 (d, 6H. *J* = 6.6 Hz. 26.27-CH<sub>3</sub>). 0.86 (s, 3H. 19-CH<sub>3</sub>). 0.89 (d, 3H, *J* = 6.6 Hz. 21-CH<sub>3</sub>). 1.41 (s. 9H. -COC(CH<sub>3</sub>)<sub>3</sub>), 1.43 (s, 9H. -COC(CH<sub>3</sub>)<sub>3</sub>). 1.99 (bm, 2H. CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>). 2.02 (bm. 2H. CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>). 2.16 (bm. 2H. NH). 2.96 (bm. 2H. 3β. 7β-H), 3.59 (t. *J* = 4.5 Hz, 4H, NHCH<sub>2</sub>), 3.74 (t. *J* = 4.5 Hz. 4H. CH<sub>2</sub>NHBoc). 4.86 (bs. 1H. NH-Boc). 5.41(bs. 1H. NH-Boc): <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  11.0, 11.9, 18.7, 20.8, 22.6, 22.9, 23.7, 24.1. 26.2, 28.1, 28.2, 28.6, 28.7, 29.4, 29.7, 29.9, 31.4, 31.7, 32.3, 32.7, 35.9, 36.2, 36.7, 39.2, 39.8, 40.6, 42.7, 45.6, 46.3, 46.6, 50.8, 52.7, 54.9, 56.3, 78.8, 78.9, 156.2, 156.3; FAB-MS Calcd for C<sub>43</sub>H<sub>81</sub>N<sub>4</sub>Q<sub>4</sub>; 715.6083, Found: *m/z* 715.6092 (M+H)<sup>-</sup>.

**3α**,7*α*-**Bis**[*N*-(4-*tert*-butyloxycarbonyl)butylamino]-5*α*cholestane (6d). yield: 80%. TLC R<sub>f</sub> 0.50 (CH<sub>2</sub>Cl<sub>2</sub>-MeOH-NH<sub>4</sub>OH 15:2:0.5): <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 0.57 (s. 3H, 18-CH<sub>3</sub>), 0.74 (s. 3H, 19-CH<sub>3</sub>). 0.78 (d. 3H. *J* = 6.5 Hz, 26-CH<sub>3</sub>), 0.79 (d. 3H. *J* = 6.5 Hz, 27-CH<sub>3</sub>). 0.82 (d. 3H. *J* = 7.0 Hz, 21-CH<sub>3</sub>). 1.36 (s. 18H, COC(CH<sub>3</sub>)<sub>3</sub>). 2.02 (m, 4H). 2.55 (bs. 1H), 2.73 (bm. 4H). 3.06 (bs. 4H). 3.11 (m. 2H. 3β-H. 7β-H). 4.80 (bs. 1H). 5.24 (bs. 2H. N*H*-Boc); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 11.1, 11.9, 12.6. 14.3, 14.4, 18.8, 20.8, 22.8, 23.1, 23.1, 24.2, 26.3, 26.3, 27.6, 28.2, 28.3, 28.7, 30.3, 31.6, 31.9, 32.8, 32.9, 36.0, 36.3, 36.4, 38.9, 39.3, 39.7, 40.0, 40.2, 43.0, 45.7, 45.9, 49.9, 50.2, 53.5, 55.7, 56.2, 79.0, 156.4, 182.4; Anal. Calcd for C<sub>35</sub>H<sub>72</sub> Cl<sub>4</sub>N<sub>4</sub>·5H<sub>2</sub>O: C, 53.58. H, 10.58. N. 7.18; Found: C. 54.23, H, 10.74. N, 7.74; FAB-MS Calcd for C<sub>45</sub>H<sub>85</sub>N<sub>4</sub>O<sub>4</sub>: 745.6571. Found: *m*/z 745.6577 (M+H)<sup>+</sup>.

 $3\alpha$ , $7\alpha$ -Bis[*N*-(4,9-Di-*tert*-butyloxycarbonyl)spermidinyl]- $5\alpha$ -cholestane (6e), yield: 81%. TLC R<sub>f</sub> 0.4 (CH<sub>2</sub>Cl<sub>2</sub>-MeOH- NH<sub>4</sub>OH 15:2:0.5): <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 0.58 (s. 3H. 18-CH<sub>3</sub>), 0.72 (s. 3H. 19-CH<sub>3</sub>), 0.79 (d. J = 6.5 Hz, 3H. 26-CH<sub>3</sub>), 0.80 (d. J = 6.5 Hz, 3H. 27-CH<sub>3</sub>), 0.83 (d. J = 6.0 Hz, 3H. 21-CH<sub>3</sub>), 1.34 (s. 18H, -COC(CH<sub>3</sub>)<sub>3</sub>), 1.38 (s. 18H. -COC(CH<sub>3</sub>)<sub>3</sub>), 3.20 (bs. 1H. 7β-H), 3.73 (bs. 1H. 3β-H), 4.63 (bs. 2H. 3α-NH. 7α-NH); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 10.6, 11.1, 11.9, 12.0, 18.8, 20.7, 22.7, 23.0, 23.7, 23.8, 23.9, 25.7, 26.0, 27.5, 28.2, 28.3, 28.4, 28.5, 28.6, 30.9, 31.9, 32.0, 32.1, 32.4, 35.9, 36.3, 36.4, 36.6, 39.6, 39.7, 39.9, 40.4, 42.7, 42.8, 44.7, 45.4, 45.9, 46.3, 46.4, 46.9, 47.3, 49.3, 50.8, 55.7, 56.2, 68.1, 79.1, 79.3, 79.4, 155.7, 155.9, 156.2, 156.2; FAB-MS Calcd for C<sub>61</sub>H<sub>115</sub>N<sub>6</sub>O<sub>8</sub>: 1059.8838, Found: *m/z* 1059.8840 (M+H)<sup>-</sup>.

**3a**,7**a**-**Bis**{**3**-(**1**-imidazolyl)propyl]amino}-**5***a*-cholestane (6f), yield: 78%, TLC R<sub>f</sub> 0.55 (CH<sub>2</sub>Cl<sub>2</sub>-MeOH-NH<sub>4</sub>OH 15:2:0.5); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  0.57 (s. 3H, 18-CH<sub>3</sub>), 0.72 (s. 3H, 19-CH<sub>3</sub>), 0.79 (d. *J* = 6.6 Hz, 3H, 26-CH<sub>3</sub>), 0.793 (d, *J* = 6.6 Hz, 3H, 27-CH<sub>3</sub>), 0.83 (d. *J* = 6.6 Hz, 3H, 21-CH<sub>3</sub>), 1.95 (m. 4H, HNCH<sub>2</sub> CH<sub>2</sub>CH<sub>2</sub>Imi), 2.30 (dd, *J* = 11.6, 5.8 Hz, 7β-H), 2.51 (m. 4H, HNCH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>Imi), 2.61 (bs, 1H, 3β-H), 3.97 (t, 2H, *J* = 6.8 Hz, CH<sub>2</sub>-Imi), 4.03 (dd, 2H, *J* = 13.6, 6.8 Hz, CH<sub>2</sub>-Imi), 4.80 (bs, 1H, N-H)), 6.83 and 6.86 (s, 1H, Im H-5), 6.95 and 6.96 (s, 1H, Im H-4), 7.40 and 7.55 (s. 1H, Im H-2); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  10.5, 11.5, 18.3, 20.4, 22.2, 22.5, 23.3, 23.5, 24.7, 27.7, 27.8, 31.3, 31.6, 32.1, 32.3, 35.4, 35.8, 36.2, 38.6, 39.1, 42.3, 43.6, 44.1, 44.4, 44.6, 45.9, 50.5, 52.9, 54.6, 55.9, 118.6, 128.7, 129.0, 136.9, 137.4; FAB-MS Calcd for C<sub>39</sub>H<sub>6</sub><sup>+</sup>N<sub>6</sub>: 619.5427, Found: *m/z* 619.5431 (M+H)<sup>+</sup>.

**3a**,7**a**-**Bis**[**2**-(**2**-**pyridy**])ethylamino]-**5a**-cholestane (6g). yield: 80%, TLC R<sub>f</sub> 0.53 (CH<sub>2</sub>Cl<sub>2</sub>-MeOH-NH<sub>4</sub>OH 15:2:0.5); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  0.55 (s. 3H. 18-CH<sub>3</sub>), 0.73 (s. 3H, 19-CH<sub>3</sub>), 0.81 (d. *J* = 6.8 Hz, 3H, 26-CH<sub>3</sub>), 0.82 (d. *J* = 6.3 Hz, 3H, 27-CH<sub>3</sub>), 0.83 (d. *J* = 7.1 Hz, 3H, 21-CH<sub>3</sub>), 1.84 (m. 1H.), 1.96 (m. 1H.), 2.58 (bs, 1H,-NH ). 3.05 (m, 4H. HNCH<sub>2</sub>CH<sub>2</sub>Py), 3.08 (m, 4H, HNCH<sub>2</sub>CH<sub>2</sub>Py), 3.69 (t. 1H. 3β-H), 3.54 (t. 1H. 7β-H), 7.04 (t. *J* = 7.4 Hz, 2H, PyH-2), 7.13 (t. *J* = 7.5 Hz, 2H, Py H-4), 7.53 (m. 2H. Py H-3), 8.42 and 8.50 (s. 2H. *J* = 4.8 Hz, Py H-5); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  11.5, 12.2, 191, 21.1, 22.9, 23.2, 23.7, 24.1, 28.4, 28.6, 31.6, 32.2, 32.8, 36.1, 36.5, 36.8, 39.4, 39.8, 39.9, 43.1, 46.4, 46.6, 48.1, 50.9, 53.2, 55.8, 56.4, 62.0, 72.9, 75.2, 121.7, 121.9, 123.8, 123.9, 136.8, 137.1, 149.3, 149.5, 160.2, 161.1; FAB-MS Calcd for C<sub>41</sub>H<sub>65</sub>N<sub>4</sub>: 613.5209, Found: *m*/z 613.5212 (M+H)<sup>+</sup>.

**3a**,7**a**-**Bis**[**2**-(**2**-**pyridylmethyl)amino**]-**5***a*-**cholestane** (**6h**). yield: 84%, TLC R<sub>f</sub> 0.70 (CH<sub>2</sub>Cl<sub>2</sub>-MeOH-NH<sub>4</sub>OH 15:2:0.5); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  0.58 (s, 3H, 18-CH<sub>3</sub>), 0.75 (s, 3H, 19-CH<sub>3</sub>). 0.80 (d, 3H, *J* = 6.5 Hz, 26-CH<sub>3</sub>), 0.81 (d, 3H, *J* = 7.0 Hz, 27-CH<sub>3</sub>), 3.86 (d, *J* = 2.5 Hz, N-CH<sub>2</sub>), 7.04-7.09 (m, 2H), 7.50-7.58 (m, 2H). 8.44-8.47 (m, 2H): <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  11.1, 11.9, 18.7, 20.8, 22.7, 23.0, 23.5, 23.9, 25.9, 28.1, 30.2, 31.8, 32.6, 32.9, 35.9, 36.2, 36.7, 39.3, 39.6, 42.0, 42.8, 46.2, 50.7, 52.4, 52.8, 53.3, 54.6, 56.1, 121.8, 122.7, 136.4, 149.1, 160.0, 160.9; FAB-MS Calcd for C<sub>39</sub>H<sub>61</sub>N<sub>4</sub>: 585.4896. Found: *m/z* 585.4897 (M+H)<sup>-</sup>.

 $3\alpha$ ,7*a*-Di(phenylureido)-5a-cholestane (2). Compound 6a (200 mg, 0.33 mmol) was dissolved in THF (10 mL) and trifluoroacetic acid (0.26 mL, 3.30 mmol) was stirred at room temperature for 3 h. After the completion of reaction. THF was removed under *vacuo*, and the mixture was neutralized with

# 2104 Bull. Korean Chem. Soc. 2009. Vol. 30, No. 9

aqueous NaOH solution (25 mL), and extracted with ethyl acetate. The organic layer was washed, dried and concentrated to give crude product 1. Without further purification crude product was dissolved in dry chloroform (10 mL) and phenyl isocyanate (0.09 mL, 0.86 mmol, 2.5 eq) was added at room temperature. After the completion of reaction, chloroform (15 mL) was added and the mixture was washed with NaHCO3 solution (20 mL) followed by water (20 mL  $\times$  2) and brine (25 mL). The organic layer was dried and concentrated. The crude product was subjected to column chromatography (10% EtOAchexane) to give 158 mg of 2 (75%). TLC R<sub>f</sub> 0.50 (EtOAc-hexane 1:1); mp 150 - 152 °C (MeOH-hexane); <sup>1</sup>H NMR (DMSO- $d_6$ )  $\delta 0.53$  (s. 3H. 18-CH<sub>3</sub>), 0.84 (d. 3H. J = 6.5 Hz, 26-CH<sub>3</sub>), 0.85 (d. 3H, J = 6.5 Hz, 27-CH<sub>3</sub>), 3.95 (bs, 2H, 3 $\alpha$ -H, 7 $\alpha$ -H), 6.97 (bs. 2H, NH), 7.52 (bs. 2H, NH), <sup>13</sup>C NMR (DMSO-d<sub>6</sub>) δ 10.5, 12.0, 14.3, 18.8, 20.5, 21.3, 22.6, 22.7, 22.8, 23.0, 23.6, 24.0, 26.5, 28.1, 29.5, 29.8, 32.0, 33.7, 34.5, 35.8, 35.9, 36.2, 37.4. 38.9. 39.2, 39.6, 42.6. 46.9. 51.2, 56.1, 119.4, 122.7. 129.1, 139.3, 156.1; Anal. Calcd for C41H60N4O22H2O: C. 72.74, H. 9.53, N, 8.28, Found: C, 72.43, H, 9.68, N, 8.08.

### **Results and Discussion**

The requisite starting material 4 was prepared from commercially available 3B-acetoxy-cholest-5-ene.5 The reductive amination of 4, with NaBH(OAc)<sub>3</sub> and NH<sub>4</sub>OTf, followed by protection with  $(Boc)_2O$ , gave 5a as a mixture of a  $3\alpha/3\beta$ -isomer in a 69% yield. High diastereo-selectivity has been achieved by modifying NaBH<sub>4</sub> with different carboxylic acids, in the synthesis of squalamine analogues.<sup>6</sup> Thus, in order to improve selectivity toward the  $3\alpha$  isomer, various acyloxyborohydrides were prepared from NaBH<sub>4</sub> and the bulky carboxylic acids. NaBH(OEh)<sub>3</sub> prepared from 2-ethylhexanoic acid (Eh) improved the stereo-selectivity of 5a, and reaction time was shortened to 1 h: it provided  $3\alpha/3\beta$  with a ratio of 9:1 in an 85% vield. The stereochemistry of 5a was determined based on the  $R_f$  value and the chemical shift of the  $3\alpha$ -NH proton in the <sup>1</sup>H, and those of C-3 and C-7 in the <sup>13</sup>C NMR spectrum. The <sup>1</sup>H NMR of **5a** showed a  $3\alpha$ -NH proton of an  $\alpha$  isomer at  $\delta$  4.88. In the <sup>13</sup>C NMR spectrum of 5a C-3 carbamate and C-7 carbonyl carbons appeared at & 155.3 and 212.1, and tert-butyloxy carbon at 8 79.2. Further relative stereochemistry

was confirmed by data obtained from COSY. HETCO. DEPT, and comparisons were made with the similar published structure.<sup>8</sup>

The reductive amination of 5a at C-7 was accomplished by NaBH<sub>3</sub>CN and NH<sub>4</sub>OAc in a co-solvent of THF and methanol in high yield. The <sup>1</sup>H NMR of 6a showed the 7 $\alpha$ -NH and 7 $\beta$ -H protons of the 7 $\alpha$  isomer at  $\delta$  4.73 and 3.67, respectively. In the <sup>13</sup>C NMR, C-7 and C-3 of the  $\alpha$  isomer appeared at  $\delta$  46.1 and 43.0, respectively. These values are in accordance with published values.<sup>5</sup> It is interesting to note that a bulky reducing reagent such as NaBH(OEh)<sub>3</sub> did not give any amino product. Hence it was concluded from the pattern of reducing reagent that bulkier reagents gave 3 $\alpha$  product and smaller reagents furnish 7 $\alpha$  products preferentially.

Compound 4 was further derivatized with *n*-BuNH<sub>2</sub>, BocNH (CH<sub>2</sub>)<sub>3</sub>NH<sub>2</sub>, BocNH(CH<sub>2</sub>)<sub>4</sub>NH<sub>2</sub> and BocNH(CH<sub>2</sub>)<sub>3</sub>N(Boc) (CH<sub>2</sub>)<sub>4</sub> N(Boc)(CH<sub>2</sub>)<sub>3</sub>NH<sub>2</sub> (Boc-spermidine) having various alkyl chain lengths in consistently high yield and stereoselectivity. 1-(3-Aminopropyl)imidazole and 2-(2-pyridyl)ethylamine were also used to prepare receptors. They reacted with 4 smoothly to give 5f and 5g in 92% and 93% yields, respectively. The latter could be converted to facial amphiphiles 6f and 6g in a high yield by reductive amination at C7 with NaBH<sub>3</sub>CN. When 2-picolylamine reacted with 4 which gave 5h in a 78%. and it undergo reductive amination at C7 with NaBH<sub>3</sub>CN to yield 6h in an 84% yield (entry 8, Table 1). The reductive amination of 4 with secondary amine, di-(2-picolyl)amine, proceeded smoothly at C3 but failed to aminate at C7. It was evident from Table 1 that steric hindrance played an important role while synthesizing  $3\alpha$ -amines 5 by reductive animation from 4.

Boc deprotection of **6a** with TFA gave diamine **1**. and subsequent reaction with phenyl isocyanate in a dry chloroform provided receptor **2** in good yield. The anion binding properties of **2** were initially investigated by <sup>1</sup>H NMR titrations in CDCl<sub>3</sub> with tetrabuty lammonium salts of F<sup>-</sup>. Cl<sup>-</sup>. CH<sub>3</sub>CO<sub>2</sub><sup>-</sup>. and H<sub>2</sub>PO<sub>4</sub><sup>-</sup>. The highest association constant ( $K_a = 42.000 \text{ M}^{-1}$ ) was obtained for CH<sub>3</sub>CO<sub>2</sub><sup>-</sup> anion as shown in Table 2. The complexation of Y-shaped CH<sub>3</sub>CO<sub>2</sub><sup>-</sup> anion with **5** easily formed H-bonding with urea protons.<sup>3a</sup> The molecular calculations of complex between **2** and acetate ion showed acetate ion bind with two 3 $\alpha$ -urea N-H and two 7 $\alpha$ -urea N-H protons through hydrogen bonds



Scheme 1. Reagents and conditions: (i) KOH/EtOH, r.t.; (ii) PCC,  $CH_2Cl_2$ , r.t.; (iii) NH<sub>4</sub>OTf, NaBH(OEh)<sub>3</sub>, THF, r.t.; (iv) (Boc)<sub>2</sub>O, MeOH; (v) NH<sub>4</sub>OAc, NaBH<sub>3</sub>CN, MeOH/THF = 1:1; (vi) TFA/THF = 3:1; (vii) PhNCO, CHCl<sub>3</sub>, reflux.

Synthesis of Facial Amphiphile as a Molecular Receptor



Scheme 2. (i) Amine, NaBH(OEh)<sub>3</sub>, THF, r.t.; (ii) Amine, NaBH<sub>3</sub>CN, MeOH/THF = 1:1.

Table 1.	. The	reductive	amination	of 4	with	various	amines
----------	-------	-----------	-----------	------	------	---------	--------

Entry	Amine <sup>a</sup>	$R^1, R^2$	Time (h) 1 <sup>st</sup> RA	Yield <sup>(</sup> (%) 3α/3β	Product 3α	Time (h) 2 <sup>nd</sup> RA	Yield <sup>e</sup> (%)	$\frac{\text{Product}^{d}}{7\alpha}$
1	NH4OTf	H, Boc	I	76/9	5a <sup>b</sup>	3	71	<b>6</b> a <sup>b</sup>
2	n-BuNH <sub>2</sub>	Н, <b><i>n-</i>Ви</b>	I	77/9	5b	3	80	6b
3	BocNH(CH <sub>2</sub> ) <sub>3</sub> NH <sub>2</sub>	H, BocNH(CH <sub>2</sub> ) <sub>3</sub>	2	72/8	5c	3	71	6c
4	BocNH(CH <sub>2</sub> ) <sub>4</sub> NH <sub>2</sub>	H, BocNH(CH <sub>2</sub> ) <sub>4</sub>	1	76/9	5d	4	80	6d
5	Boc-spermidine	H, Boc-spermidine	1	78/9	5e	4	81	6e
6	3-Im(CH <sub>2</sub> ) <sub>3</sub> NH <sub>2</sub>	H, 3-Im(CH <sub>2</sub> ) <sub>3</sub>	2	81/9	5f	3	78	6f
7	$2-Py(CH_2)_2NH_2$	$H, 2-Py(CH_2)_2$	2	84/8	5g	3	80	6g
8	2-PvCH <sub>2</sub> NH <sub>2</sub>	$H, 2-PyCH_2$	I	78/9	5h	3	84	6h
9	$(2-PyCH_2)_2NH$	$(2-PyCH_2)_2$	3	56/33	5i	72	No Ran	6i

 $^{\circ}$ See experimental section.  $^{\circ}$ Both **5a** and **6a** are Boc-protected amines, see reagents and conditions in the Scheme 1. Isolated yields.  $^{\circ}$ The 7 $\beta$  isomer was obtained in negligible amount.

Table 2. Association constants  $(K_a)$  of receptor 2 with various anions<sup> $\sigma$ </sup>

OAc	$H_2PO_4$	F	Cl
<b>42</b> ,100 <sup><i>b</i></sup>	$2,700^{b}$	1,300	1,500

<sup>a</sup>TBA salt of the anions were used in CDCl<sub>3</sub> at 298 K. [host] =  $4.5 \times 10^{-3}$  M. <sup>b</sup>Errors estimated to be  $\leq 10^{\circ}$  b.



Figure 2. The H-bonding of 2 with acetate anion was shown by simulated molecular modeling.

(distances of N-H $\cdots$  O<sub>2</sub>CCH<sub>3</sub>: a = 1.79, b = 1.66, c = 1.78, d = 1.68 Å) as shown in Figure 2.<sup>9</sup>

In conclusion,  $3\alpha$ . $7\alpha$ -aminocholestane-based anionic receptors have been synthesized by reductive amination protocol with modified sodium acyloxyborohydride reagent in high yield and stereoselectivity. This procedure will be used to prepare various molecular receptors with compounds **6a-6g**. The anion-binding studies of the remainder of the receptors are currently under investigation.

Acknowledgments. This work was supported by the Korea Research Foundation Grant funded by the Korean Government(MOEHRD, KRF-2007-C00155) and the Kyungpook National University Research Fund, 2007.

## **References and Notes**

- (a) Li, C.; Peters, A. S.; Meredith, E. L.; Allman, G. W.; Savage, P. B. J. Am. Chem. Soc. **1998**, 120, 2962. (b) Bhattacharjya, S.; David, S. A.: Mathan, V. I.; Balaram, P. Biopolymers **1997**, 41, 251. (c) Bruch, M. D.; Cajal, Y.; Koh, J. T.; Jain, M. K. J. Am. Chem. Soc. **1999**, 121, 11993. (d) Hancock, R. E. W. Annu. Rev. Microbiol. **1984**, 38, 237. (e) Li, C.; Budge, L. P.; Driscoll, C. D.; Willardson, B. M.; Allman, G. W.; Savage, P. B. J. Am. Chem. Soc. **1999**, 121, 931. (f) Ding, B.; Guan, Q.; Walsh, J. P.; Boswell, J. S.; Winter, T. W.; Winter, E. S.; Boyd, S. S.; Li, C.; Savage, P. B. J. Med. Chem. **2002**, 45, 663.
- (a) Guan, Q.; Li, C.; Schmidt, E. J.; Boswell, J. S.; Walsh, J. P.; Allman, G. W.; Savage, P. B. Org. Lett. 2000, 2, 2837. (b) Rehman, A.; Li, C.; Budge, L. P.; Street, S. E.; Savage, P. B. Tetrahedron Lett. 1999, 40, 1865. (c) Broderick, S.; Davis, A. P.; Williams, R. P. Tetrahedron Lett. 1998, 39, 6083 (d) Li, C.; Rehman, A.; Dalley, N. K.; Savage, P. B. Tetrahedron Lett. 1999, 40, 1861. (e) Zhou, X.-T.; Rehman, A.; Li, C.; Savage, P. B. Org. Lett. 2000, 2, 3015.
- (a) Clare, J. P.; Ayling, A. J.; Joos, J.-B.; Sission, G. M.; Perez-Payan, M. N.; Lambert, T. N.; Shukla, R.; Smith, B. D.; Davis, A. P. J. Am. Chem. Soc. 2005, 127, 10739. (b) Bhattarai, K. M.; del Amo, V.; Magro, G.; Sission, A. L.; Joos, J.-B.; Charmant, J. P. H.; Kantacha, A.; Davis, A. P. Chem. Commun. 2006, 2335. (c) Rivera, D. G.; Concepcion, O.; Perez-Labrada, K.; Coll, F. Tetrahedron 2008, 64, 5298.
- (a) McNally, B. A.; Koulov, A. V.; Smith, B. D.; Joos, J. B.; Davis, A. P. Chem. Commun. 2005, 1087. (b) del Amo, V.; Siracusa, L.; Markidis, T.; Baragaña, B.; Bhattarai, K. M.; Galobardes, M.; Naredo, G.; Perez-Payan, M. N.; Davis, A. P. Org. Biomol. Chem. 2004, 2, 3320. (c) Koulov, A. V.; Lambert, T. N.; Jain, M.; Boon, J. M.; Smith, B. D.; Li, H. Y.; Sheppard, D. N.; Joos, J. B.;

2106 Bull. Korean Chem. Soc. 2009, Vol. 30, No. 9

Clare, J. P.; Davis, A. P. Angew. Chem. Int. Ed. 2003, 42, 4931.
(d) Lambert, T. N.; Boon, J. M.; Smith, B. D.; Pérez-Payán, M. N.; Davis, A. P. J. Am. Chem. Soc. 2002, 124, 5276. (e) Ayling, A. J.; Broderick, S.; Clare, J. P.; Davis, A. P.; Pérez-Payán, M. N.; Lahtinen, M.; Nissinen, J. J.; Rissanen, K. Chem. Eur. J. 2002, 8, 2197. (f) Siracusa, L.; Hurley, F. M.; Dresen, S.; Lawless, L. J.; Pérez-Payán, M. N.; Davis, A. P. Org. Lett. 2002, 4, 4639.

 Khan, S. N.; Cho, N. J.; Kim, H.-S. Tetrahedron Lett. 2007, 48, 5189.

6. Khan, S. N.; Jung, Y. M.; Kim, B. J.; Cho, H.; Lee, J.; Kim, H.-S.

Md. Wasi Ahmad et al.

Bioorg. Med. Chem. Lett. 2008, 18, 2558. And references cited therein.

- Khan, S. N.; Bae, S. Y.; Kim, H.-S. Tetrahedron Lett. 2005, 46, 7675.
- Khan, S. N.; Cho, N. J.; Kim, H.-S. In *Catalyst for the Fine Chemical Synthesis*; Robert, S. M.; Whittall, J., Eds.; Regio- and Stereo-Controlled Oxidations and Reductions; John Wiley & Sons: Chichester, 2007; Vol. 5, p 175.
- 9. MP2 calculation at the AM1 mode level was performed SPARTAN'04 for Windows (Wavefunction, Inc.: Irvine, CA).