Climate Effects on Greenhouse Gas Emissions and Microbial Communities in Wetlands

Seon-Young Kim1 and Hojeong Kang2*

¹Department of Environmental Science and Engineering, Ewha Womans University, Seoul, 120-750, Korea ²School of Civil and Environmental Engineering, Yonsei University, Seoul, 120-749, Korea (Received March 6, 2007; Accepted September 3, 2007)

기후변화가 습지 내 온실기체 발생과 미생물 군집구조에 미치는 영향

김선영1 · 강호정2*

이화여자대학교 환경공학과, ² 연세대학교 토목환경공학부 (2007년 3월 6일 접수; 2007년 9월 3일 수락)

ABSTRACT

Global climate changes including elevated CO₂, drought, and global warming may influence greenhouse gas emissions in wetlands. A variety of microbial communities including denitrifiers and methanogens play a key role in determining such processes. In this paper we summarize current knowledge on the effects of climate changes on CO₂, CH₄, and N₂O production and microbial communities mediating those processes in wetlands. Elevated atmospheric CO₂ and warming generally increase gas emissions, but effects of droughts differ with gas type and drying level. The responses of microbial community to climate changes in terms of composition, diversity and abundance are still in question due to lack of studies in wetlands. Based on the present review, it is suggested that future studies on microbial processes should consider microbial community and relationships between microbial function and structure with diverse environmental factors including climate changes. Such knowledge would be crucial to better understand and predict accurately any shifts in ecological functions of wetlands.

Key words: Elevated CO₂, Drought, Greenhouse gas emissions, Microbial community, Warming, Wetland

I. INTRODUCTION

Wetland ecosystems including peat-forming wetland cover only 2-6% of global land surface (Gorham, 1991). However, they play a pivotal role in global biogeochemical cycles. Firstly, peat accumulation over thousands of years has resulted in a vast store of carbon of 455 Pg-C in peatland ecosystems (Gorham, 1991; Roulet, 2000). This represents 20-30% of the world's pool of soil organic carbon and is comparable to the total carbon in the atmosphere as CO₂ (IPCC, 2001). Secondly, wetlands are substantial sources of the radi-

atively active trace gases such as CH_4 and N_2O (Freeman *et al.*, 1994). For example, natural wetlands and rice paddies release about 40-50% of global emissions of CH_4 which is 25 times more radiatively active than CO_2 on a molar basis (Cicerone and Oremland, 1988). As such, even small changes in gas emissions from wetland soils could significantly influence the balance of carbon flux between the atmosphere and biosphere. This would be of great importance in future trajectory of global warming scenario.

Anthropogenic activities have increased the concentration of atmospheric CO₂ continuously from about

Corresponding Author: Hojeong Kang(hj kang@yonsei.ac.kr)

280 parts per million (ppm) at the beginning of the industrial revolution to 382 ppm at the present time (IPCC, 2001). The linear warming trend over the last 50 years (0.13 per decade) is nearly twice that for the last 100 years. Extremely hot and dry summers have also occurred throughout much of the earth. Substantial carbon (C) losses have already been reported in boreal and sub-arctic bogs and fens due to lower water levels during warm and dry summers (Schreader *et al.*, 1998; Alm *et al.*, 1999; Moore *et al.*, 2002). As such, concerns have been raised that persistent warming and drought could reduce the capacity of wetlands to act as a sink for global C, and may eventually turn it into a source of C to the atmosphere.

Wetland soils undergo intermittent flooding, draining, and dewatering, potentially supporting both aerobic and anaerobic microbial communities. Moreover, chemical characteristics differ substantially among different types of wetlands, ranging from mineral to organic, eutrophic to oligotrophic and saline to freshwater, allowing each wetland to support distinct microbial communities. Those diverse microbial communities may exploit a wide range of electron acceptors during organic matter decomposition. Critical processes controlling CO₂, N₂O, and CH₄ production from wetlands are carried out by diverse microbial communities such as denitrifiers, methanogens, and so on. Those microbes potentially influence the balance among anaerobic gas production in soils (Yavitt and Seidman-Zager, 2006).

Climate changes are possible to influence soil microbial processes mediating organic matter decomposition and gas emissions through altering abundance and/or structure of microbial communities inhabiting wetlands. The knowledge on the links between microbial structure and soil functioning is also essential by considering that 80-90% of the processes in soil are reactions mediated by microbes (Nannipieri *et al.*, 2003). However, the effects of climate changes on soil microbial community and activity, and their relationship remain largely unknown in wetlands. The aim of this paper is to summarize current knowledge on the effects of elevated CO₂, drought, and warming on biogenic gas emissions (CO₂, CH₄, and N₂O) and microbial communities in wetland ecosystems.

II. EFFECTS OF ELEVATED CO₂ CONCENTRATIONS

Atmospheric CO₂ concentrations have increased

from pre-industrial level of 280 ppm to 382 ppm at present. Considering the long-term average CO₂ increase of 1.5 ppm per year (http://www.cmdl.noaa. gov/gmd/ccgg/trends), it is likely to reach approximately 530 ppm by 2100. Previous studies have mainly focused on the effects of elevated CO₂ on wetland vegetation in terms of growth, leaf photosynthesis, evaporation, nutrients and community structure (Jauhiainen et al., 1997; Mitchell et al., 2002). Some studies have drawn attention on the response of below-ground processes to elevated CO₂ in terms of dissolved organic carbon (DOC) release, soil enzyme activity, and greenhouse gas emissions (Saarnio et al., 2000; Freeman et al., 2004; Kang et al., 2005).

2.1. Gas emissions

Elevated atmospheric CO2 concentrations increased CO2 emissions from wetlands via increased root biomass and DOC (Schrope et al., 1999; Kang et al., 2001). CH₄ emissions differently responded to elevated CO₂. Although many studies showed an increase in CH₄ emissions under elevated CO2 (Megonigal and Schlesinger, 1997; Wang and Adachi, 1999), no changes or even decrease in CH₄ emissions were also observed in wetlands (Kang et al., 2001; Saarnio and Silvola, 1999; Schrope et al., 1999; Saarnio et al., 2000). It is possible that changes in litter chemistry (Hirschel et al., 1997), nutrient deficiency (Niklaus and Körner, 1996), or heights of water table (Roulet et al., 1992) might change the magnitude and direction of methane emissions in wetland ecosystem under elevated CO2 conditions.

2.2. Microbial communities

In many studies conducted in terrestrial ecosystems, there is an uncertainty in the responses of microbial communities to elevated CO₂; no change (Griffiths *et al.*, 1998; Deiglmayr *et al.*, 2004; Grüter *et al.*, 2006; Jossi *et al.*, 2006; Chung *et al.*, 2006) and change (Marilley *et al.*, 1999; Montealegre *et al.*, 2000; Janus *et al.*, 2005; Jossi *et al.*, 2006; Lipson *et al.*, 2006) (Table 1). Unfortunately, understanding of microbial communities under elevated CO₂ is still limited in wetland ecosystems. Lee *et al.* (2004) reported a shift of denitrifiers in the marsh incubated at elevated CO₂.

The lack of consistent results is likely related with analytical tools applied. Most molecular tools (e.g., PLFA, T-RFLP, and DGGE) detect both active and inactive ones in microbial community. More recently, it

Plant type	Molecular methods applied	Responses	Reference	
	PLFA ^a	No	Wiemken et al., 2001	
	PLFA	No/changes	Phillips et al., 2002	
Forest	PLFA / DGGE ^b	No	Chung et al., 2006	
	PLFA	No	Zak et al., 2000	
	PLFA / T-RFLP ^c	Change ^d	Janus et al., 2005	
,	TTGE ^e / T-RFLP	No	Grüter et al., 2006	
Grassland	PCR-DGGE	No	Jossi <i>et al.</i> , 2006	
	Reverse transcriptase-PCR-DGGE	Change		
	PLFA	Change	Montealegre et al., 2002	
	ARDRA ^f	Change	Marilley et al., 1999	
	BOX-PCR	Change	Montealegre et al., 2000	
	PCR-RFLP	No	Deiglmayr et al., 2004	
	DNA hybrization	No	Griffiths et al., 1998	
	%G + C base profiling	No	Niklaus et al., 2003	
	Sequencing	Change	Lipson et al., 2006	
Wetlands	T-RFLP	Change	Lee et al., 2004	
	T-RFLP	Nog	Kim (unpublished data)	

Table 1. The list of studies reporting the effects of elevated CO₂ on microbial community

(a) PLFAs (phospholipids fatty acid); (b) DGGE (denaturing gradient gel electrophoresis); (c) T-RFLP (terminal restriction fragments length profile); (d) bacterial and fungal communities; (e) TTGE (temporal temperature gradient gel electrophoresis); (f) ARDRA (Amplified ribosomal DNA restriction analysis); (g) bacterial, denitrifier, and sulphate reducing bacterial communities

has been reported that only "active" species presenting in the rhizosphere responded to elevated CO₂ (Jossi *et al.*, 2006). Additionally descriptive explanation without statistical analysis might attribute to controversial results. It is necessary to apply adequate statistical methods to community composition and structure such as nonmetric multi-dimensional scaling, diversity indices, multi-response permutation procedure and indicator species analysis (McCune and Grace, 2002).

III. EFFECTS OF DROUGHTS

In the future climate, droughts and floods are expected to be more frequent, severe, and longer-lasting (IPCC, 2001). Persistent droughts are likely to reduce the capacity of wetlands to act as a sink for global carbon, and may eventually turn it into a source of carbon to the atmosphere. Substantial carbon losses have already been reported in various wetlands including temperate, boreal, and sub-arctic peatlands (Shurpali *et al.*, 1995; Alm *et al.*, 1999; Moore *et al.*, 2002) and marsh (Heinsch *et al.*, 2004) during warm and dry summers. After drought wetlands lost their own characteristics in relation with anoxic environments they behaved more like a terrestrial ecosystem (Heilman *et al.*, 2000).

3.1. Gas emissions

The main control on a wetland's ability to retain carbon and to generate trace gases is determined by its ability to remain wet. Partial drying of wetlands significantly increased CO₂ emissions into the atmosphere (Shurpali *et al.*, 1995; Oechel *et al.*, 1998; Heinsch *et al.*, 2004), which is related to the stimulation of hydrolytic enzyme activities (Freeman *et al.*, 1996; Corstanje, 2003). Denitrification and nitrous oxide emissions also increased with drying (Robertson and Tiedje, 1987; Davidsson *et al.*, 2002), because nitrification in the topsoil layers supplied substrates for denitrification (Baldwin and Mitchell, 2000).

Many studies have focused on methane emission in relation with drought (Moore and Knowles, 1989; Freeman *et al.*, 1993; Moore and Roulet, 1993; Ratering and Conrad, 1998). It is generally accepted that droughts reduce the amount of methane emissions from wetlands. Enhanced methane oxidation has been considered as a major reason for the decline in CH₄ emissions. However, recently it has been reported that methane oxidation is unable to completely account for the decreased methane emissions during droughts (Freeman *et al.*, 2002). During drought, suppressed primary production may reduce CH₄ emissions via

	Genes	Riparian	Fen	Bog
Abundance	Bacterial 16S rDNAs	-	Decrease*	Increase**
	mcrA	-		Decrease***
	nirS	-	Decrease***	Decrease**
Diversity	Bacterial 16S rDNAs	Decrease*	-	Increase(*)
	mcrA	-	-	-
	nirS	Increase(*)	-	_

Table 2. Changes in abundance and diversity of bacteria, methanogens, and denitrifiers amplified bacterial 16S rRNA, mcrA and mirS genes, respectively, following the drought treatment in riparian, fen, and bog soils. Data were analyzed by t-test at p < 0.001***, p < 0.01**, and p < 0.05*, n = 6 (2-tails).

decreased labile organic substrates which normally drive methanogenesis (Whiting and Chanton, 1993). Additionally, an increased sulphate and ferric iron may allow sulphate and ferric iron reducing bacteria to outcompete methanogenic bacteria on H₂ (Ratering and Conrad, 1998). Although wetlands are restored with rewetting, methane production rates do not recover like before. It is because that rewetting may result in subsequent sulphate releases into surface water, suppressing the recovery of methane emissions via methanogens inhibition (Freeman *et al.*, 1994; Ratering and Conrad, 1998; Dowrick *et al.*, 2006).

3.2. Microbial communities

Microbial communities have been assumed to be affected by drought. However, such reports were just based on changes in microbial processes such as enzyme activity, organic matter decomposition, and gas emissions without specific measure of community structure. For example, no release of nutrients following the adding of sulphate into rewetted sediments speculated a loss of sulphate reducing bacteria during droughts (Bayley *et al.*, 1986). Changes in CH₄ emissions may imply a shift in methanogens following droughts (e.g., Boon *et al.*, 1997).

Recently, DNA-based approaches have been used to measure microbial communities with droughts. We observed the response of eubacteria, denitrifiers, and methanogens to droughts based on abundance, composition or diversity in a bog, a fen, and a riparian wetland (Table 2). The abundance and diversity of microbial communities differently responded to droughts according to the type of wetland and microbial community (Table 2). It has been reported that drying of sediments significantly changed the bacterial community structure in a semi-permanent stream (Rees *et al.*, 2006). However, following rewetting they did not return to their original

structures at pre-drought conditions. As for methanogens, regardless of water regimes, the structure and biomass were relatively stable in rice paddy soil (Mayer and Conrad, 1990; Lueders and Friedrich, 2000). We also found that droughts did not affect the diversity and composition of methanogens in wetlands examined (Table 2).

IV. EFFECTS OF GLOBAL WARMING

Climate change models predicted that the mean global surface air temperatures will increase by between 1.4 and 5.8°C until the end of the 21st century (IPCC, 2001). Higher temperatures may transform permafrost soils to wetlands and accelerate mineralization in wetlands. Previous studies have demonstrated that warming potentially influenced DOC release (Freeman *et al.*, 2001; Cole *et al.*, 2002), soil enzyme activity (Waldrop and Firestone, 2004; Fenner *et al.*, 2005), and nutrient availability (Fenner *et al.*, 2006) in wetlands. However, the impacts of this projected warming on carbon balance in wetlands are still uncertain.

4.1. Gas emissions

Terrestrial studies demonstrates that CO₂ production is positively related with temperature (Eliasson *et al.*, 2005), which is primarily regulated by enhancement of plant growth (Zhang *et al.*, 2005), dominant soil fauna (Briones *et al.*, 2004) and carbon quality (Fierer *et al.*, 2006). There is also a positive correlation between temperature and N₂O emission rates (Carnol and Ineson, 1999). As for wetlands, ecosystem respiration (ER) as indicated by CO₂ emission showed a positive relationship with temperature in peatlands (Kim and Verma, 1992; Updegraff *et al.*, 2001; Lafleur *et al.*, 2005). However, little attention has been given to possible

effects of warming on N2O emissions from wetlands.

CH₄ efflux is positively correlated with soil temperature (Hutchin et al., 1995; Kim et al., 1999; Updegraff et al., 2001). CH₄ production rates in anaerobically incubated rice field soil increased with temperature ranging from 10 to 37°C (Fey and Conrad, 2000). The optimum temperature and the lower limit for methanogenesis were 25 and -5°C, respectively, in acidic peats (Metie and Frenzel, 2005). It is consistent with a study showing that a marked increase in CH₄ emissions from peat soils between 6 and 15 (Fenner et al., 2006). Temperature sensitivity of methanogenesis is potentially influenced by substrate quality, wetland type, and soil nutrient dynamics (Updegraff et al., 1995, 2001). Global warming would also influence a shift in growth stages of wetland vegetation, resulting in altered plant mediated CH₄ and CO₂ transport (Kim et al., 1998, 1999). Additionally, warming may increase CH₄ emissions possibly at the global scale via increases in newly formed peatlands as a result of melting of the permafrost (Gorham, 1991).

4.2. Microbial communities

Warming changes substrate quality and quantity in belowground (Hooper *et al.* 2000; Biasi *et al.*, 2005), which is likely to affect microbial community. We observed changes in bacterial community following warming treatment (+3°C) over three years (Kim,

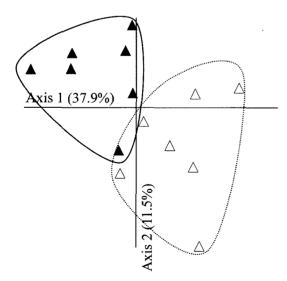


Fig. 1. Non-metric multidimensional scaling (NMS) ordination of bacterial 16S rRNA genes in control (\triangle) and warming (\blacktriangle) treatments. The percent variation explained by each axis is shown in parentheses.

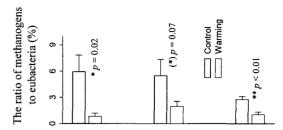


Fig. 2. The ratio of methanogens to eubacteria based on abundance with depth at warming and control treatments (2-tails).

2007) (Fig. 1). The diversity and composition of nitrogen-fixing microbial communities changed with warming (1-3°C) in an arctic ecosystem (Deslippe *et al.*, 2005). However, increased temperatures did not change methanogens in a rice field soil (Chin *et al.*, 1999) and a bog (Metje and Frezel, 2005). Warming relatively increased the relative contribution of fungi to soil microbial community (Thormann *et al.*, 2004), whereas warming decreased that of methanogens to eubacterial community in peatlands (Kim, 2007) (Fig. 2).

V. SUMMARY AND CONCLUDING REMARKS

In order to better understand and predict more accurately future C dynamics in wetlands, we suggest several future directions of research. First, the combined effects of climate changes should be considered. Interactions of elevated CO₂, drought, and warming may be different from separate effect. For example, elevated CO₂ treatment increased the temperature sensitivity of soil CO₂ emissions through an increase in substrate availability (Tingey *et al.*, 2006). In contrast, rising temperature down-regulates CH₄ emissions via an increased O₂ availability caused by droughts.

Secondly, effects of climatic changes on wetlands should be considered in various geographical regions, because wetlands located in different latitudes and locations exhibit substantial differences in geology, litter chemistry, carbon pool size, and future scenario of climate change. In fact, the effects of elevated CO₂ on CH₄ emissions have been reported to be slighter in northern peatlands (Saarnio and Silvola, 1999; Saarnio *et al.*, 2000; Kang *et al.*, 2001) compared to those in temperate or subtropical wetlands. Furthermore, sensitivity to rainfall patterns differed in northern and southern Europe (Sowerby *et al.*, 2005), depending on

previous exposure to moisture stress (Fierer *et al.*, 2003). Another example is differences in litter chemistry (Thormann *et al.*, 2004) and labile carbon pools (Eliasson *et al.*, 2005), which affect sensitivity of SOM to warming effects (Knorr *et al.*, 2005).

Lastly, the relationship between microbial community structure and their ecological functions should be interpreted with caution, because differences in microbial communities may lead to no changes in microbial functions. Measuring whole microbial community does not represent microbial groups involved in critical processes measured. Therefore, understanding of functional groups like denitrifiers, methanogens, and sulphate reducing bacteria subjected by climate changes would be essential to predict wetland C cycling.

ACKNOWLEDGEMENTS

This work was supported by the Korea Science and Engineering Foundation through the Advanced Environmental Biotechnology Research Center (R11-2003-006) at Pohang University of Science and Technology. S-Y. Kim is grateful to Korea Research Foundation Grant funded by the Korean Government (MOEHRD, Basic Research Promotion Fund) (KRF-2006-C00587) and the International Research Internship Program of KOSEF for financial support. H. Kang thanks Ewha Womans University for the financial support.

적 요

대기 중 이산화탄소 농도 및 온도 증가와 강수 패 턴 변화에 따른 가뭄 정도 및 횟수의 변화는 습지에 서 발생하는 온실가스의 양에 영향을 미칠 수 있다. 습지에 존재하는 다양한 미생물 군집(탈질세균 및 메 탄생성세균) 이 온실가스 생성에 있어 중요한 역할을 감당한다. 본 논문은 지금까지 전지구적 기후변화가 습 지에서의 온실가스 발생과 관련 미생물 군집에 미치는 영향에 관한 다양한 연구를 정리하는 데 그 목적이 있다. 대기 중 이산화탄소 농도와 기온 증가는 일반적 으로 온실가스 생성을 증가시켰다. 반면, 가뭄의 영향 은 기체 종류와 가뭄 정도에 따라 다양한 결과가 보 고되었다. 기후변화에 따른 미생물 군집의 변화는 습지 시스템에서 보고된 연구의 부족으로 인해 특정한 결론 을 도출할 수 없었다. 본 총설은 습지에서 미생물을 매개로 한 반응을 연구함에 있어 관련 미생물 군집구 조의 특성을 파악하고, 다양한 환경인자에 대한 그들의 반응을 알아내는 것과 미생물 반응과 군집구조간의 상 관 관계를 도출하는 것의 중요성을 제안한다. 이는 향후 전지구적 기후 변화가 습지의 생태학적 기능에 미칠 영향을 더 잘 이해하고 예측하는데 있어 매우 중요할 것이라 사료된다.

REFERENCES

- Alm, J., L. Schulman, J. Walden, H. Nykänen, P. J. Martikainen, and J. Silvola, 1999: Carbon balance of a boreal bog during a year with an exceptionally dry summer. *Ecology* **80**, 161-174.
- Baldwin, D. S., and A. M. Mitchell, 2000: The effects of drying and re-flooding on the sediment and soil nutrient dynamics of lowland river-flood plain systems: a synthesis. Regulated Rivers Research and Management 16(5), 457-467.
- Bayley, S. E., R. S. Behr, and C. A. Elly, 1986: Retention and release of S from A freshwater wetland. Water, Air, and Soil Pollution 31(1-2), 101-114.
- Biasi, C., O. Rusalimova, H. Meyer, C. Kaiser, W. Wanek, P. Barsukov, H. Junger, and A. Richter, 2005: Temperature-dependent shift from labile to recalcitrant carbon sources of arctic heterotrophs. *Rapid Communications in Mass Spectrometry* 19, 1401-1408.
- Boon, P. I., A. Mitchell, and K. Lee, 1997: Effects of wetting and drying on methane emissions from ephemeral floodplain wetlands in south-eastern Australia. *Hydrobiologia* 357, 73-87.
- Briones, M. J. I., J. Poskitt, and N. Ostle, 2004: Influence of warming and enchytraeid activities on soil CO₂ and CH₄ fluxes. Soil Biology and Biochemistry 36, 1851-1859.
- Carnol, M., and P. Ineson, 1999: Environmental factors controlling NO₃⁻ leaching, N₂O emissions and numbers of NH₄⁺ oxidisers in a coniferous forest soil. *Soil Biology and Biochemistry* **31**(7), 979-990.
- Chin, K. J., T. Lukow, and R. Conrad, 1999: Effect of temperature on structure and function of the methanogenic archaeal community in an anoxic rice field soil. *Applied and Environmental Microbiology* **65**(6), 2341-2349.
- Chung, H., D. R. Zak, and E. A. Lilleskov, 2006: Fungal community composition and metabolism under elevated CO₂ and O₃. *Oecologia* **147**(1), 143-154.
- Cicerone, R. J., and R. S. Oremland, 1988: Biogeochemical aspects of atmospheric methane. *Global Biogeochemical Cycles* **2**, 299-327.
- Cole, L., R. D. Bardgett, D. P. Ineson, and P. J. Hobbs, 2002: Enchytraeid worm (Oligochaeta) influences on microbial community structure, nutrient dynamics and plant growth in blanket peat subjected to warming. Soil Biology and Biochemistry 34(1), 83-92.
- Corstanje, R., 2003: Experimental and multivariate analysis of biogeo-indicators of change in wetland ecosystems.

- Ph.D. dissertation. University of Florida, Gainesville.
- Davidsson, T. E., M. Trepel, and J. Schrautzer, 2002: Denitrification in drained and rewetted minerotrophic peat soils in Northern Germany (*Pohnsdorfer Stauung*). *Jour*nal of Plant Nutrition and Soil Science 165(2), 199-204.
- Deiglmayr, K., L. Philippot, U. A. Hartwig, and E. Kandeler, 2004: Structure and activity of the nitrate-reducing community in the rhizosphere of *Lolium perenne* and *Trifolium repens* under long-term elevated atmospheric pCO₂. FEMS Microbiology Ecology 49, 445-454.
- Deslippe, J. R., K. N. Egger, and G. H. R. Henry, 2005: Impacts of warming and fertilization on nitrogen-fixing microbial communities in the Canadian High Arctic. FEMS Microbiology Ecology 53(1), 41-50.
- Dowrick, D. J., C. Freeman, M. A. Lock, and B. Reynolds, 2006: Sulphate reduction and the suppression of peatland methane emissions following summer drought. *Geoderma* 132, 384-390.
- Eliasson, P. E., R. E. McMurtrie, D. A. Pepper, M. Strömgren, S. Linder, and G. I. Ågren, 2005: The response of heterotrophic CO₂ flux to soil warming. *Global Change Biology* 11(1), 167-181.
- Fenner, N., C. Freeman, and B. Reynolds, 2005: Observations of a seasonally shifting thermal optimum in peatland carbon-cycling processes; implications for the global carbon cycle and soil enzyme methodologies. *Soil Biology and Biochemistry* **37**(10), 1814-1821.
- Fenner, N., D. J. Dowrick, M. A. Lock, C. R. Rafarel, and C. Freeman, 2006: A novel approach to studying the effects of temperature on soil biogeochemistry using a thermal gradient bar. Soil Use and Management 22, 267-273.
- Fey, A., and R. Conrad, 2000: Effect of temperature on carbon and electron flow and on the archaeal community in methanogenic rice field soil. *Applied and Envi*ronmental Microbiology 66(11), 4790-4797.
- Fierer, N., B. P. Colman, J. P. Schimel, and R. B. Jackson, 2006: Predicting the temperature dependence of microbial respiration in soil: A continental-scale analysis. *Glo*bal Biogeochemical Cycles 20, GB3026.
- Fierer, N., J. P. Schimel, and P. A. Holden, 2003: Influence of drying-rewetting frequency on soil bacterial community structure. *Microbial Ecology* 45, 63-71.
- Freeman, C, J. Hudson, M. A. Lock, B. Reynolds, and C. Swanson, 1994: A possible role for sulphate in the suppression of methane fluxes following drought. Soil Biology and Biochemistry 26, 1439-1442.
- Freeman, C, N. Fenner, N. J. Ostle, H. Kang, D. J. Dowrick, B. Reynolds, M. A. Lock, D. Sleep, S. Hughes, and J. Hudson, 2004. Dissolved organic carbon export from peatlands under elevated carbon dioxide levels. *Nature* 430, 195-198.
- Freeman, C., G. B. Nevison, H. Kang, S. Hughes, B. Reynolds, and J. A. Hudson, 2002: Contrasted effects of

- simulated drought on the production and oxidation of methane in a mid-Wales wetland. *Soil Biology and Biochemistry* **34**, 61-67.
- Freeman, C., G. Liska, N. J. Ostle, J. A. Hudson., M. A. Lock, and B. Reynolds, 1996: Microbial activity and enzymic decomposition processes following peatland water table drawdown. *Plant and Soil* 180, 121-127.
- Freeman, C., N. J. Ostle, and H. Kang, 2001: An enzymic latch on a global carbon store. *Nature* **409**: 149.
- Freeman, C., M. A. Lock, and B. Reynolds, 1993: Fluxes of carbon dioxide, methane and nitrous oxide from a Welsh peatland following simulation of water table draw-down: Potential feedback to climatic change. *Biogeochemistry* **19**, 51-60.
- Gorham, E., 1991: Northern peatlands: role in the carbon cycle and probable to climatic warming. *Ecological Applications* 1, 182-195.
- Griffiths, B. S., K. Ritz, N. Ebblewhite, E. Paterson, and K. Killham, 1998: Ryegrass rhizosphere microbial community structure under elevated carbon dioxide concentrations with observations on wheat rhizosphere. *Soil Biology and Biochemistry* 30(3), 315-321.
- Grüter, D., B. Schmid, and H. Brandl, 2006: Influence of plant diversity and elevated atmospheric carbon dioxide levels on belowground bacterial diversity. *BMC Microbiology* 6, 68-76.
- Heilman, J. L., F. A. Heinsch, D. R. Cobos, and K. J. McInnes, 2000: Energy balance of a high marsh on the Texas Gulf Coast: effect of water availability. *Journal of Geophysical Research* 105, 22371-22377.
- Heinsch, F. A., J. L. Heilman, K. J. McInnes, D. R. Cobos, D. A. Zuberer, and D. L. Roelke, 2004: Carbon dioxide exchange in a high marsh on the Texas Gulf Coast: effects of freshwater availability. *Agricultural and For*est Meteorology 125, 159-172.
- Hirschel, G, C. Körner, and J. A. Arnone III, 1997: Will rising atmospheric CO₂ affect leaf litter quality and in situ decomposition rates in native plant communities? *Oecologia* 110, 387-392.
- Hooper, D. U., D. E. Bignell, V. K., Brown, L. Brussaard,
 J. M, Dangerfield, D. H. Wall, and A. D. Wardle, D. C.
 Coleman, K. E. Giller, P. Lavelle, W. H. Van Del Putten,
 P. C. de Ruiter, J. Rusek, W. L. Silver, J. M. Tiedje, and
 V. Wolters, 2000: Interactions between aboveground and belowground biodiversity in terrestrial ecosystems: Patterns, mechanisms, and feedbacks. *Bioscience* 50, 1049-1061.
- Hutchin, P. R., M. C. Press, J. A. Lee, and T. W. Ashenden, 1995: Elevated concentrations of CO₂ may double methane emissions from mires. *Global Change Biology* 1, 125-128.
- IPCC, 2001: Climate Change 2001: the scientific basis. Cambridge University Press.
- Janus, L. R., N. L. Angeloni, J. McCormack, S. T. Rier, N.

- C. Tuchman, and J. J. Kelly, 2005: Elevated atmospheric CO₂ alters soil microbial communities associated with Trembling Aspen (*Populus tremuloides*) roots. *Microbial Ecology* **50**(1), 102-109.
- Jauhiainen, J., J. Silvola, K. Tolonen, and H. Vasander, 1997: Response of Sphagnum fuscum to water levels and CO₂ concentration. Journal of Bryology 19, 391-400.
- Jossi, M., N. Fromin, S. Tarnawski, F. Kohler, F. Gillet, M. Aragno, and J. Hamelin, 2006: How elevated pCO₂ modifies total and metabolically active bacterial communities in the rhizosphere of two perennial grasses grown under field conditions. FEMS Microbiology Ecology 55, 339- 350.
- Kang, H. J., C. Freeman, and T. W. Ashendon, 2001: Effects of elevated CO₂ on fen peat biogeochemistry. The Science of the Total Environment 279, 45-50.
- Kang, H. J., S. Y. Kim, N. Fenner, and C. Freeman, 2005: Shifts of soil enzyme activities in wetlands exposed to elevated CO₂. Science of the Total Environment 337, 207-212.
- Kim, J., S. B. Verma, and D. P. Billesbach, 1999: Seasonal variation in methane emission from a temperate Phragmites-dominated marsh: effect of growth stage and plant-mediated transport. *Global Change Biology* **5**(4), 433-440.
- Kim, J., and S. B. Verma, 1992: Soil surface CO₂ flux in a Minnesota peatland. *Biogeochemistry* 18, 37-51.
- Kim, J., S. B. Verma, D. P. Billesbach, and R. J. Clement, 1998: Diel variation in methane emission from a midlatitude prairie wetland: significance of convective throughflow in phragmites australis. *Journal of Geophysical* research 103, 28029-28039.
- Kim, S. Y., 2007: A study of wetland vegetation and microbial communities under elevated CO₂, warming, and drought. PhD dissertation, Ewha Womans University.
- Knorr, W., I. C. Prentice, J. I. House, and E. A. Holland, 2005: On the available evidence for the temperature dependence of soil organic carbon. *Biogeosciences Dis*cussions 2, 749-755.
- Lafleur, P. M., T. R. Moore, N. T. Roulet, and S. Frolking, 2005: Ecosystem respiration in a cool temperate bog depends on peat temperature but not water table. *Ecosys*tems 8, 619-629.
- Lee, S. H., S. Y. Kim, and H. J. Kang, 2004: Influence of elevated CO₂ on denitrifying bacterial community in a wetland soil. *Korean Journal of Microbiology* **40**(3), 244-247. (in Korean with English abstract)
- Lipson, D. A., M. Blair, G. Barron-Gafford, K. Grieve, and R. Murthy, 2006: Relationships between microbial community structure and soil processes under elevated atmospheric carbon dioxide. *Microbial Ecology* 51(3), 302-314.
- Lueders, T., and M. Friedrich, 2000: Archaeal population

- dynamics during sequential reduction processes in rice field soil. *Applied and Environmental Microbiology* **66**(7), 2732-2742.
- Marilley, L., U. A. Hartwig, and M. Aragno, 1999: Influence of an elevated atmospheric CO₂ content on soil and rhizosphere bacterial communities beneath *Lolium perenne* and *Trifolium repens* under field conditions. *Microbial Ecology* **38**, 39-49.
- Mayer, H. P., and R. Conrad, 1990: Factors influencing the population of methanogenic bacteria and the initiation of methane production upon flooding of paddy soil. *FEMS Microbiology Ecology* 73, 103-112.
- McCune, B., and J. B. Grace, 2002: *Analysis of Ecological Communities*. Gleneden Beach. MjM Software Design, 300pp.
- Megonigal, J. P., and W. H. Schlesinger, 1997: Enhanced CH₄ emission from a wetland soil exposed to elevated CO₂. *Biogeochemistry* **37**(1), 77-88.
- Metje, M., and P. Frenzel, 2005: Effect of temperature on anaerobic ethanol oxidation and methanogenesis in acidic peat from a northern wetland. Applied and Environmental Microbiology 71(12), 8191-8200.
- Mitchell, E. A. D., A. Buttler, P. Grosvernier, H. Rydin, A. Siegenthaler, and J. M. Gobat, 2002: Contrasted effects of increased N and CO₂ supply on two keystone species in peatland restoration and implications for global change. *Ecology* **90**, 529-533.
- Montealegre, C. M., C. van Kessel, J. M. Blumenthal, H. Hur, U. A. Hartwig, and M. J. Sadowsky, 2000: Elevated Atmospheric CO₂ alters microbial population structure in a pasture ecosystem. *Global Change Biology* 6, 475-482.
- Montealegre, C. M., C. van Kessel, M. P. Russelle, and M. J. Sadowsky, 2002: Changes in microbial activity and composition in a pasture ecosystem exposed to elevated atmospheric carbon dioxide. *Plant and Soil* 243(2), 197-207.
- Moore, T. R., and N. T. Roulet, 1993: Methane flux: Water table relations in northern wetlands. *Geophysical Research Letters* 20(7), 587-590.
- Moore, T. R., and R. Knowles, 1989: Influence of water table levels on methane and carbon dioxide emissions from peatland soils. *Canadian Journal of Soil Science* 69(1), 33-38.
- Moore, T. R., J. L. Bubier, S. E. Frolking, P. M. Lafleur, and N. T. Roulet, 2002: Plant biomass and production and CO₂ exchange in an ombrotrophic bog. *Journal of Ecology* 90(1), 25-36.
- Nannipieri, P., J. Ascher, M. T. Ceccherini, L. Landi, G. Pietramellara, and G. Renella, 2003: Microbial diversity and soil functions. *European Journal of Soil Science* **54**(4), 655-670.
- Niklaus, P. A., D. Alphei, D. Ebersberger, C. Kampichler, E. Kandeler, and D. Tscherko, 2003: Six years of in situ

- CO₂ enrichment evoke changes in soil structure and soil biota of nutrient-poor grassland. *Global Change Biology* **9**, 585-600.
- Niklaus, P. A., and C. Körner, 1996: Responses of soil microbiota of a late successional alpine grassland to long term CO₂ enrichment. *Plant and Soil* 184(2), 219-229.
- Oechel, W. C., G. L. Vourlitis, S. J. Hastings, R. P. Ault, and P. Bryant, 1998: The effects of water table manipulation and elevated temperature on the net CO₂ flux of wet sedge tundra ecosystems. *Global Change Biology* 4, 77-90.
- Phillips, R. L., D. R. Zak, W. E. Holmes, and D. C. White, 2002: Microbial community composition and function beneath temperate trees exposed to elevated atmospheric carbon dioxide and ozone. *Oecologia* 131(2), 236-244.
- Ratering, S., and R. Conrad, 1998: Effects of short-term drainage and aeration on the production of methane in submerged rice soil. Global Change Biology 4(4), 397-407.
- Rees, G. N., G. O. Watson, D. S. Baldwin, and A. M. Mitchell, 2006: Variability in sediment microbial communities in a semipermanent stream: impact of drought. *Journal of the North American Benthological Society* 25(2), 370-378.
- Robertson, G. P., and J. M. Tiedje, 1987: Nitrous oxide sources in aerobic soils: Nitrification, denitrification and other biological processes. *Soil Biology and Biochemis*try 19(2), 187-193.
- Roulet, N. T, R. Ash, and T. R. Moore, 1992: Low boreal wetlands as a source of atmospheric methane. *Journal of Geophysical Research* **97**, 3739-3749.
- Roulet, N. T., 2000: Peatlands, carbon storage, greenhouse gases, and the kyoto protocol: prospects and significance for Canada. *Wetlands* **20**(4), 605-615.
- Saarnio, S., and J. Silvola, 1999: Effects of increased CO₂ and N on CH₄ efflux from a boreal mire: a growth chamber experiment. *Oecologia* 119(3), 349-356.
- Saarnio, S., T. Saarinen, H. Vasander, and J. Silvola, 2000: A moderate increase in the annual CH₄ efflux by raised CO₂ or NH₄NO₃ supply in a boreal oligotrophic mire. *Global Change Biology* **6**(2), 137-144.
- Schreader, C. P., W. R. Rouse, T. J. Griffis, L. D. Boudreau, and P. D. Blanken, 1998: Carbon dioxide fluxes in a northern fen during a hot-dry summer. *Global Biogeochemical Cycles* 12, 729-740.
- Schrope, M. K., J. P. Chanton, L. H. Allen, and J. T. Baker, 1999: Effect of CO₂ enrichment and elevated temperature on methane emissions from rice *Oryza sativa*. Global Change Biology 5, 587-599.
- Shurpali, N. J., S. B. Verma, J. Kim, and T. J. Arkebauer, 1995: Carbon dioxide exchange in a peatland ecosystem. *Journal of Geophysical Research* 100, 14319-

- 14326.
- Sowerby, A., B. Emmett, C. Beier, A. Tietema, J. Penuelas, M. Estiarte, M. J. M.Van Meeteren, S. Hughes, and C. Freeman, 2005: Microbial community changes in heathland soil communities along a geographical gradient: interaction with climate change manipulations. Soil Biology and Biochemistry 37, 1805-1813.
- Thormann, M. N., S. E. Bayley, and R. S. Currah, 2004: Microcosm tests of the effects of temperature and microbial species number on the decomposition of *Carex aquatilis* and *Sphagnum fuscum* litter from southern boreal peatlands. *Canadian Journal of Microbiology* **50**, 793-802.
- Tingey, D. T., E. H. Lee, R. Waschmann, M. G. Johnson, and P. T. Rygiewicz, 2006: Does soil CO₂ efflux acclimatize to elevated temperature and CO₂ during ong-term treatment of Douglas-fir seedlings? *New Phytologist* 170, 107-118.
- Updegraff, K., S. D. Bridgham, J. Pastor, P. Weishampel, and C. Harth, 2001: Response of CO₂ and CH₄ emissions from peatlands to warming and water table manipulation. *Ecological Applications* 11(2), 311-326.
- Updegraff, K., J. Pastor, S. D. Bridgham, and C. A. Johnston, 1995: Environmental and substrate controls over carbon and nitrogen mineralization in northern wetlands. *Ecological Applications* 5(1), 151-163.
- Waldrop, M. P., and M. K. Firestone, 2004: Altered utilization patterns of young and old soil C by microorganisms caused by temperature shifts and N additions. *Biogeochemistry* **67**, 235-248.
- Wang, B., and K. Adachi, 1999: Methane production in a flooded soil in response to elevated atmospheric carbon dioxide concentrations. *Biology of Fertile Soils* 29, 218-220.
- Whiting, G. J., and J. P. Chanton, 1993: Primary production control of methane emission from wetlands. *Nature* 364, 794-795.
- Wiemken, V., E. Laczko, K. Ineichen, and T. Boller, 2001: Effects of elevated carbon dioxide and nitrogen fertilization on mycorrhizal fine roots and the soil microbial community in Beech-Spruce ecosystems on siliceous and calcareous soil. *Microbial Ecology* 42(2), 126-135.
- Yavitt, J. B., and M. Seidman-Zager, 2006: Methanogenic conditions in northern peat soils. *Geomicrobiology Jour*nal 23, 119-127.
- Zak, D. R., K. S. Pregitzer, J. S. King, and W. E. Holmes, 2000: Elevated atmospheric CO₂, fine roots and the response of soil microorganisms: a review and hypothesis. *New Phytologist* 147(1), 201-222.
- Zhang, W., K. M. Parker, Y. Luo, S. Wan, L. L. Wallace, and S. Hu, 2005: Soil microbial responses to experimental warming and clipping in a tallgrass prairie. *Global Change Biology* 11, 266-277.