

Increase of Amyloid-Beta Peptide Generation in High Cholesterol Diet Rabbit Brain

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Abstract – Alzheimer's disease (AD) is an abnormal accumulation of the β -amyloid protein (A β) in specific brain region. It has been speculated that disturbance in cholesterol homeostasis may contribute to the etiology of AD by increasing A β generation. However, conclusive evidence and possible mechanism has not been reported. In the present study, we demonstrated that rabbits treated with 0.5% cholesterol for 16 weeks increased serum total cholesterol, triacylglycerol, and low-density lipoprotein levels. A β levels is higher in the hippocampus of brain in cholesterol dieted rabbits than that of normal diet rabbits. Expression and activities of β - and γ - secretases, the enzymes that cleave β -amyloid precursor protein to generate A β , were also increased in hippocampus of high cholesterol dieted rabbit than those of normal dieted rabbits. Our results suggest that high cholesterol diet may be associated with increased A β accumulation in the brain of rabbits, and suggest that high cholesterol diet may be causal factor in the development or progression of AD.

Key words □ Alzheimer's disease, high cholesterol diet, beta-amyloid, β - and γ - secretase, rabbit

INTRODUCTION

Alzheimer's disease (AD) is a progressive neurodegenerative process characterized by loss of memory, cognition, and behavioral stability. AD is defined pathologically by accumulation of extracellular neuritic plaque comprised of fibrillar deposits of amyloid β -peptide (A β) and neurofibrillary tangles comprised of paired helical filaments of hyperphosphorylated tau (Hardy *et al.*, 2002).

Epidemiological and animal studies suggest that disturbances in cholesterol homeostasis may contribute to the etiology of AD (Fassbender *et al.*, 2001; Galbete *et al.*, 2000; Jick *et al.*, 2000; Racchi *et al.*, 1997; Refole *et al.*, 2000). Epidemiological studies have shown that people in the UK who are prescribed statins, cholesterol lowering agent (lipid-lowering agents: LLAs), have a 37%~70% high risk of dementia than those who do not have hyperlipidaemia or who are not treated with LLAs (Jick *et al.*, 2000). In animal studies, dietary cholesterol accelerates A β deposition in the brain of transgenic mice

expressing familial AD mutant, human APP_{K671N,M671I} (Refole *et al.*, 2000). BM15.766, a cholesterol-lowering drug, reduces A β depositon in the brain of AD and in the brain of APP_{K671N,M671I} mutant mice (Refole *et al.*, 2001). Although high cholesterol can influence on A β generation, it is note worthy that a change in membrane properties, including stiffness and fluidity, has been suggested to influence activities of membrane-bond proteins and enzymes, including secretase. Cellular cholesterol content is increased in neuron by cholesterol enriched diet (Othman *et al.*, 2006). Thus, the high cholesterol content in lipid rafts, membrane regions where these enzymes are located, facilitates the clustering of the secretases with their substrates into an optimum configuration, promoting the undesirable pathogenetic cleavage of amyloid precursor protein (Bodovitz *et al.*, 1996; Witter *et al.*, 1991). However, direct relationship between high cholesterol diet and A β depositon, and the mechanism by which cholesterol affects A β production and metabolism is not fully understood.

Because 2% cholesterol diets cause severe hypercholesterolemic side effects, requiring sacrifice of the cholesterol-fed rabbits at 8 weeks (Sparks *et al.* 1994; Sparks *et al.* 2000), we have anticipated that a long-term diet supplemented with a low level of cholesterol would allow the animals to live longer, thus

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facilitating the accumulation of A β_{1-42} and the activation of enzyme involved in A β generation. In present study, we have examined the effect of 0.5% cholesterol-enriched diet on levels of serum cholesterol, beta-amyloid, and activation of enzyme involved in A β generation in rabbit.

MATERIALS AND METHODS

Hypercholesterolemia animal model

New Zealand white female rabbits were used for the proposed studies. Animals were randomly assigned to 2 groups as follows: group 1, normal chow (n=3) and group 2, chow supplemented with 0.5% cholesterol (n=5). The animals were dieted for 16 weeks. Diets were kept frozen at -10°C to reduce the risk of oxidation. Cholesterol-dieted animals and their matched controls were euthanized 16 weeks later. At necropsy, each brain was immediately frozen in liquid nitrogen, placed into a zipper-closure plastic bag, and buried in dry ice pellets until transferring to -80°C for storage before sectioning, western blot, and A β assay.

Measurement of serum cholesterol level

Total serum cholesterol and triglycerides were determined enzymatically as previously described (Haubenwalner *et al.*, 1995). Serum lipoprotein cholesterol profiles and distribution among lipoproteins were determined by on-line post column analysis on Superose 6HR high performance gel filtration chromatography (HPGC). Lipoprotein cholesterol was determined by multiplying the independently determined total serum cholesterol by the percent area for each lipoprotein distinctly separated by the HPGC method.

Measurement of brain A β level

The A β level in brain was determined using available A β_{1-42} assay kit (Immuno-Biological Laboratories Co., Aramachi, Japan) according to the manufacture's protocols. The whole brain, cerebral cortex and hippocampus were homogenized in protein extraction solution (PRO-PREP™, Intron Biotechnology, Korea). To determine A β , 100 μ l lysate was put in 96 well plate which was coated with anti-Human A β (38-42) rabbit IgG and the plate was incubated for 12 hr at 4°C. 100 μ l labeled antibody solution was added into the wells and followed by incubation 1 hr at 4°C in the dark. Tetra Methyl Benzidine (TMB) was added each well then the plate was incubated for 30 minutes for room temperature in the dark. TMB is used as coloring agent (Chromogen). The reaction was stopped by addi-

tion of 100 ml stop solution (1N H₂SO₄). The A β amount was quantified by measured light absorbance at 450 nm within 30 minutes after application of stop solution. The A β amount was linearly related to the increase in light absorbance. The A β amount was expressed as pg produced substrate which was determined by the formation of pg per mg protein.

Measurement of β - and γ -secretase activity in brain

The total activities of β - and γ -secretase present in cortical and hippocampal region which were determined using commercially available β -secretase fluorescence resonance energy transfer (BACE 1 FRET) assay kit (PANVERA, Madison, USA) and γ -secretase activity Kit (R&D systems, Wiesbaden, Germany) according to the manufacture's protocols, respectively. The cerebral cortex and hippocampus were homogenized in cold 1x cell extraction buffer (ready for use in the kit) to yield a final protein concentration of 1 mg/ml.

To determine β -secretase, 10 μ l lysate was mixed with 10 μ l BACE1 substrate (Rh-EVNLDAEFK-Quencher), and then the reaction mixture was incubated for 1 hr at room temperature in the 96 well flat bottom microtitre plate. The reaction was stopped by addition of 10 μ l BACE1 stop buffer (2.5 M sodium acetate). The formation of fluorescence was read with Fluostar galaxy fluorometer (excitation at 545 nm and emission at 590 nm) with Felix software (BMG Labetechnologies, Offenburg, Germany). The enzyme activity was linearly related to the increase in fluorescence. The enzyme activity was expressed as nM produced substrate which was determined by the formation of fluorescence per mg protein per min.

To determine γ -secretase, 50 μ l lysate was mixed with 50 μ l reaction buffer. The reaction mixture was then incubated for 1 hr in the dark at 37°C. 5 ml substrate was added after the periods of incubation time to stop the reaction. Cleaved substrate by γ -secretase was conjugated to the reporter molecules EDANS and DABCYL, and released fluorescent signal. This formation of fluorescence was read with Fluostar galaxy fluorometer (excitation at 355 nm and emission at 510 nm) with Felix software (BMG Labetechnologies, Offenburg, Germany). The level of γ -secretase enzymatic activity is proportional to the fluorometric reaction, and the γ -secretase activity was expressed the produced fluoresce unit. All controls, blanks and samples were run in triplicate.

Western blotting

Brains were homogenized with lysis buffer and centrifuged at 23,000 g for 1hr. The protein concentration was measured by

the Bradford method (Bio-Rad Protein Assay, Bio-Rad Laboratories Inc, Hercules, CA), and equal amount of proteins (20 μ g) were separated on a SDS/10%, 15% and 20% -polyacrylamid gel, and then transferred to a nitrocellulose membrane (Hybond ECL, Amersham Pharmacia Biotech Inc., Piscataway, NJ, USA). Blots were blocked for 1 hr at room temperature with 5 % (w/v) non-fat dried milk in Tris-buffered saline. The membrane was incubated for 5 hr at room temperature with specific antibodies: APP (1:1000), BACE (1:1000), C99 (1:1000) and β -actin (1:10000) as internal standard. Immunoreactive proteins were detected with the ECL western blotting detection system.

RESULTS

Serum cholesterol level in high cholesterol dieted rabbit

Examination of the lipid profiles of high cholesterol diet versus control rabbits reveals a significant elevation in the amount of total cholesterol, triacylglycerol and low density lipoprotein in the high cholesterol diet rabbits (Table I). Serum total cholesterol concentrations 43.00 ± 9.29 mg/dl in controls and this level was elevated to 1118.57 ± 126.90 mg/dl in high cholesterol dieted rabbits. Triacylglycerol level was 15.33 ± 2.33 mg/dl in normal whereas 37.17 ± 7.11 mg/dl in high cholesterol dieted rabbits. However, HDL level did not show significant changed (15.67 ± 2.03 mg/dl vs 21.21 ± 10.21 mg/dl). Especially, serum LDL level in cholesterol diet rabbits (381.43 ± 167.23 mg/dl) was increased over 10 times than controls (36.33 ± 19.14 mg/dl).

$A\beta_{1-42}$ content and secretase activity in the brain of high cholesterol dieted rabbits

We used $A\beta$ assay kit to measure $A\beta_{1-42}$ content in whole brain, cortex and hippocampus. $A\beta_{1-42}$ level was increased in the whole and hippocampus of cholesterol-treated rabbit. However, $A\beta_{1-42}$ levels was not changed in cortex of cholesterol dieted rabbits (Fig. 1). Since $A\beta$ is produced with serial cleav-

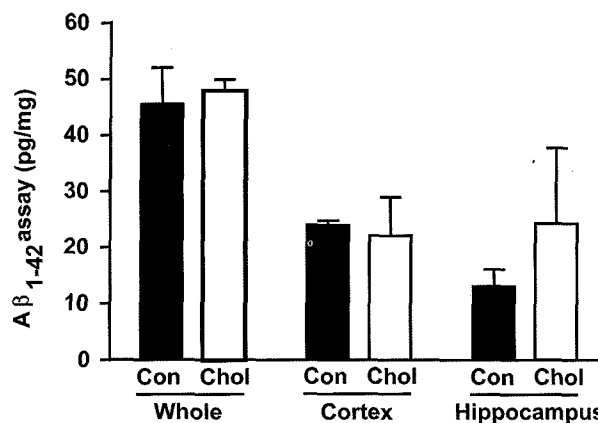


Fig. 1. $A\beta_{1-42}$ level in the brain of control and high cholesterol diet rabbit. High cholesterol diet rabbit were served 0.5% cholesterol contained food for 16 weeks. $A\beta_{1-42}$ level was measured by commercial $A\beta$ assay kit in the whole brain, hippocampus and cortex of control (con) and high cholesterol diet (chol) rabbit.

age of amyloid precursor protein (APP) by β - and γ - secretase. We determined secretase activity in brain. Consistent with the elevated levels of $A\beta$, β - and γ - secretase activities were increased in whole, and cortex and hippocampus of high cholesterol diet rabbits (Fig. 2).

APP processing change in the brain of cholesterol dieted rabbits

Western blotting was next done to determine whether APP processing could be altered in cholesterol-dieted rabbit brain. Consistent with the increase of β - and γ - secretase activity, western blot analysis showed that β -APP, BACE and its C-terminal fragment, C99, which is the product of cleavage of β -APP by β -secretase (BACE) levels were increased in whole brain and hippocampus region of high cholesterol diet rabbits. However, β -APP, BACE and C99 protein levels were not significantly changed in cortex of high cholesterol diet rabbits (Fig 3).

DISCUSSION

In the present study, we have examined the relationship between hypercholesterolemia and $A\beta$ generation using high cholesterol dieted rabbit model. We served food contained 0.5 % cholesterol to rabbit for 16 weeks. In results, serum low density lipoprotein (LDL) and triacylglyceride levels were significantly increased in high cholesterol dieted rabbits. $A\beta$ level was

Table I. Cholesterol concentrations in serum of control and cholesterol diet rabbits

	Control (n=3)	Cholesterol (n=14)
Total cholesterol (mg/dl)	43.00 ± 9.29	1118.57 ± 126.90
Triacylglycerol (mg/dl)	15.33 ± 2.33	37.14 ± 7.11
High-density lipoprotein (mg/dl)	15.67 ± 2.03	21.21 ± 10.21
Low-density lipoprotein (mg/dl)	19.67 ± 7.36	381 ± 44.70

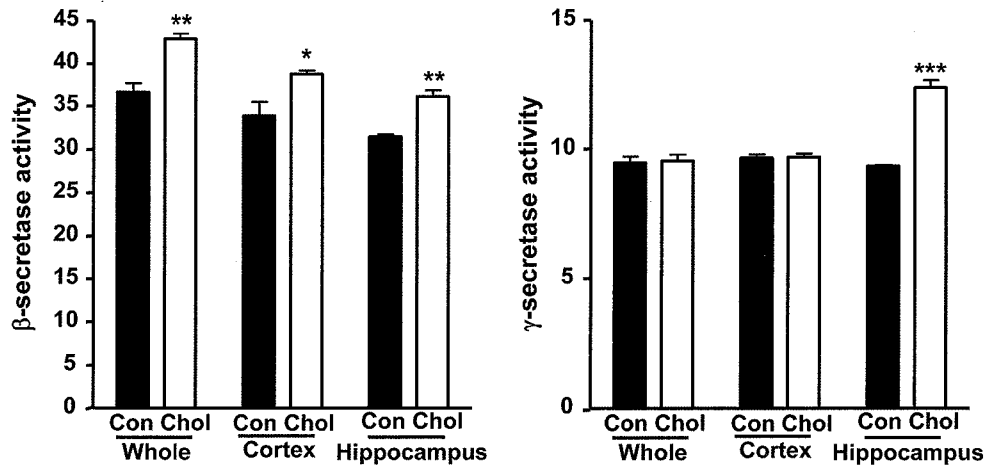


Fig. 2. β - and γ - secretase activities in the brain of control and high cholesterol diet rabbit. β - and γ - secretase activities were measured by secretase assay kit in the whole brain, cortex and hippocampus of control (con) and high cholesterol diet (chol) rabbit. High cholesterol diet rabbit was made by serving 0.5% cholesterol contained food for 16 weeks. * P <0.05 indicates statistically significant differences between control and high cholesterol diet rabbit.

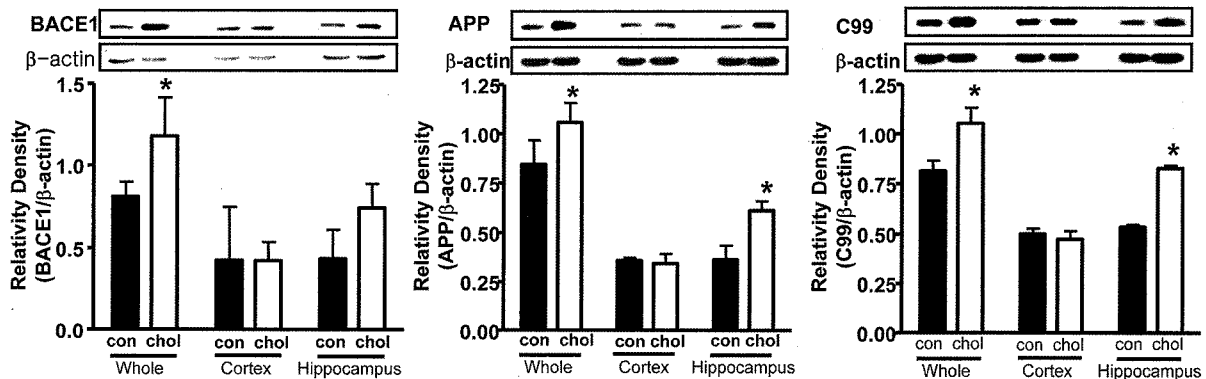


Fig. 3. Expression of BACE1, APP and C99 in the brain of control and high cholesterol diet rabbit. High cholesterol diet rabbit was served 0.5% cholesterol contained food for 16 weeks. The brains of control (con) and high cholesterol diet (chol) rabbit were used for western blot. Equal amounts of total proteins (40 mg/lane) were subjected to 6% (APP), 10% (BACE1) and 15% (C99) SDS-PAGE. Expression of BACE1, APP, C99 and β -actin were detected by western blotting using specific antibody. β -actin protein was used as an internal control. * P <0.05 indicates statistically significant differences between control and high cholesterol diet rabbit.

increased in high cholesterol dieted rabbit brain. In addition, we also found that β - and γ - secretase activities were increased in brain of high cholesterol diet rabbits accompanied by the increased expression of BACE, APP and C99 which these proteins were involved in A β generation.

A number of epidemiological studies suggest that high levels of dietary cholesterol may contribute to the pathogenesis of AD. AD patients have increased levels of total serum and low-density lipoprotein (LDL) cholesterol along with reduced levels of high-density lipoprotein (HDL) in their plasma, as compared to age-matched controls (Fernandes., 1999; Kuo *et al.*, 1998). Cholesterol abnormally accumulates in the dense cores of amy-

loid plaques in the brain of AD patients (Mori., 2001). Similar accumulation of cholesterol has also been found in amyloid plaques of transgenic mice expressing a mutant form APP₆₉₅ (Swedish mutation) associated with FAD (Mori., 2001). Studies of using transgenic animal models of AD show a strong connection between plasma cholesterol levels and A β generation (Fassbender *et al.*, 2001; Holsinger., 2002; Refolo *et al.*, 2000). A recent report shows that high-cholesterol diet containing 5% cholesterol, 10% fat, and 5.2 kcal/g for 7 weeks raise cholesterol levels in plasma and CNS of transgenic mice expressing the FAD mutant APP_{K670,M671L} and PS1_{M146V}. Both β -APP C-terminal fragment (CTF) and A β levels were

increased in the brain of these animals (Refolo *et al.*, 2000). The result suggested that high cholesterol diet may be implicated in the development of AD. Other *in vitro* studies also have shown that a high cholesterol environment results in reduced production of soluble amyloid precursor protein (Kojro *et al.*, 2001; Racchi *et al.*, 1997; Simons *et al.*, 1998; Wahrle *et al.*, 2002), a protein protecting neurons against cell stimuli including A β . Neuropathological analysis showed that a high cholesterol diet also increased the deposition of amyloid plaque in presenilin 1 transgenic mice, AD model mice (Holcomb *et al.*, 1998; McGowan *et al.*, 1999). Additionally, cholesterol-lowering agents reversed the effect of high fat/high cholesterol diet on A β accumulation and cholesterol levels in the plasma and CNS (Refolo *et al.*, 2001). Taken together these data suggest that high cholesterol diet may increase of A β level in the brain, thereby affect AD development.

The mechanism by which dietary cholesterol promotes A β peptide accumulation is not fully understood. However, cholesterol influences the activities of the enzymes involved in the metabolism of the amyloid precursor protein to generate A β . The post-translational cleavage of amyloid precursor protein by β - and γ -secretase, membrane associated proteins, results in amyloidogenic products that aggregate as extracellular plaques (Leila *et al.*, 2005). β -secretase activity has been demonstrated to be increased in lipid microdomains (Cordy *et al.*, 2003; Marlow *et al.*, 2003), and is upregulated in sporadic cases of Alzheimer's disease (Holsinger *et al.*, 2002; Yang *et al.*, 2003). Lipid microdomains that are enriched in cholesterol are critically involved in the initial cleavage of APP by β -secretase to generate A β (Ehehalt *et al.*, 2003). The cholesterol-induced increase in APP and BACE is associated with an increase in C99 levels. Therefore these results suggest that β -secretase and APP processing are upregulated by the cholesterol diet, resulting in an increase in the cleavage of APP to C99 and A β peptide.

It is interested to know that A β content was significantly accumulated in hippocampus compared with cortex region by 0.5% cholesterol diet in rabbit brain. A variety of memory task has been known that hippocampus is specifically involved in memory (Mishkin., 1978; Squire *et al.*, 1991). In addition, more recent reports have indicated that subjects with hippocampal damage can exhibit impaired memory (Simons *et al.*, 1998). Even though the hippocampal dentate gyrus receives a direct excitatory input from the ipsilateral entorhinal cortex (Witter *et al.*, 1991), an area known to be affected early in course of Alzheimer disease (Gomez-Isla *et al.*, 1996; Van Hoesen *et al.*, 1991), one of the major neuropathological find-

ings in the brains of individuals with AD is loss of synaptic contact in hippocampus (Bertoni-Freddari *et al.*, 1990; Bodovitz *et al.*, 1996; Goto *et al.*, 1990; Hamos *et al.*, 1989; Masliah *et al.*, 1989). The accumulation of the loss of synapses and a failure to replace the loss synaptic contacts in hippocampus may lead to the decline in cognitive function that is clinically assessed as a decline in memory. Thus, high cholesterol diet could affect on the memorial function of hippocampus via accumulation of A β .

The present study showing high cholesterol diet increase A β generation, and the elevated A β is associated with increase β - and γ -secretase activities and with the increase of the express of BACE, APP and C99 suggest that high cholesterol diet could contribute to the development or progression of AD.

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REFERENCES

- Bertoni-Freddari C, Fattoretti P, Casoli T, Meier-Ruge W, Ulrich J. (1990). Morphological adaptive response of the synaptic junctional zones in the human dentate gyrus during aging and Alzheimer's disease. *Brain Res.* **517**, pp. 69-75.
- Bodovitz S, Klein WL. (1996). Cholesterol modulates alpha-secretase cleavage of amyloid precursor protein. *J Biol Chem.* **271**, pp. 4436-4440.
- Cabalka LM, Hyman BT, Goodlett, CR, Ritchie TC, Van-Hoesen GW. (1992). Alteration in the pattern of nerve terminal protein immunoreactivity in the perforant pathway in Alzheimer's disease and in rats after entorhinal lesions. *Neurobiol Aging.* **13**, pp. 283-291.
- Cordy JM, Hussain I, Dingwall C, Hooper NM, Turner AJ. (2003). Exclusively targeting beta-secretase to lipid rafts by GPI-anchor addition up-regulates beta-site processing of the amyloid precursor protein. *Proc. Natl. Acad. Sci.* **100**, pp. 11735-11740.
- Ehehalt R, Keller P, Haass C, Thiele C, Simons K. (2003). Amyloidogenic processing of the Alzheimer beta-amyloid precursor protein depends on lipid raft. *J. Cell Biol.* **160**, pp. 113-123.
- Fassbender K, Simons M, Bergmann C, Stroick M, Lutjohann D, Keller P, Runz H, Kuhl S, Bertsch T, von-Bergmann K, Hennerici M, Beyreuther K, Hartmann T. (2001) Simvastatin strongly reduces levels of Alzheimer's disease beta-amyloid peptides Abeta 42 and Abeta 40 *in vitro* and *in vivo*. *Proc. Natl. Acad. Sci. USA.* **98**, pp. 5856-5861.
- Fernandes MA. (1999) Effects of apolipoprotein E genotype on blood lipid composition and membrane platelet fluidity in Alzheimer's disease. *Biochem. Biophys. Acta.* **1454**, pp. 89-96.
- Galbete JT, Martin TR, Peressini E, Modena P, Bianchi R, Forloni G. (2000). Cholesterol decreases secretion of the secreted

- form of amyloid precursor protein by interfering with glycosylation in the protein secretory pathway. *J. Biochem.* **348**, pp. 307-313.
- Gomez-Isla T, Price JL, McKeel DW, Morris JC, Growdon JH, Hyman BT. (1996). Profound loss of layer II entorhinal cortex neurons occurs in very mild Alzheimer's disease. *J. Neurosci.* **16**, pp. 4491-4500.
- Goto S, Hirano A. (1990). Neuronal inputs to hippocampal formation in Alzheimer's disease and in parkinsonism-dementia complex on Guam. *Acta Neuropathol.* **79**, pp. 545-550.
- Hamos J, DeGennaro L, Drachman D. (1989). Synaptic loss in Alzheimer's disease and other dementias. *Neurology.* **39**, pp. 355-361.
- Hardy J, Selkoe DJ. (2002). The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science.* **297**, pp. 253-256.
- Haubenwallner S, Essenburg AD, Barnett BC, Pape ME, DeMatos RS, Leff T, Bisgaier CB. (1995) Hypolipidemic activity of select fibrates correlates to changes in hepatic apolipoprotein C- expression: a potential physiologic basis for their mode of action. *J. Lipid Res.* **36**, pp. 2541-2551.
- Holcomb L, Gordon MN, McGowan E, Yu X, Benkovic S, Jantzen P, Wright K, Saad I, Mueller R, Morgan D, Sanders S, Zehr C, O'Campo K, Hardy J, Prada C, Eckman M, Younkin CS, Hsiao K, Duff K. (1998). Accelerated Alzheimer's-type phenotype in transgenic mice carrying both mutant amyloid precursor and presenilin 1 transgenes. *Nature Med.* **4**, pp. 97-100.
- Holsinger RM, McLean CA, Beyreuther K, Masters CL, Evin G. (2002). Increased expression of the amyloid precursor beta-secretase in Alzheimer's disease. *Ann. Neurol.* **51**, pp. 783-786.
- Howland DS. (1998) Modulation of secreted β -amyloid precursor protein and amyloid β -peptide in brain by cholesterol. *J. Biol. Chem.* **273**, pp. 16576-16582.
- Jick H, Zomberg GL, Jick SS, Seshadri S, Drachman DA. (2000). Statins and the risk of dementia. *Lancet.* **356**, pp. 1627-1631.
- Kojro E, Gimpl G, Lammich S, Fahrenholz F. (2001). Low cholesterol stimulates the nonamyloidogenic pathway by its effect on the alpha-secretase ADAM 10. *Proc Natl Acad Sci USA.* **98**, pp 5815-5820.
- Kuo YM. (1998). Elevated low-density lipoprotein in Alzheimer disease correlates with brain A β 1-42 levels. *Biochem. Biophys. Res. Commun.* **252**, pp. 711-715.
- Leila AS, Ging-Yuek RS, Howard HF. (2005). Cholesterol in Alzheimer's disease. *Lancet Neurol.* **4**, pp. 841-852.
- Marlow L, Cain M, Pappolla MA, Sambamurti K. (2003). Beta-secretase processing of the Alzheimer's amyloid protein precursor (APP). *J. Mol. Neurosci.* **20**, pp. 233-239.
- Masliah E, Terry R, DeTeresa R, Hansen L. (1989). Immunohistochemical quantification of the synapse-related protein synaptophysin in Alzheimer disease. *Neurosci Lett.* **103**, pp. 234-239.
- McGowan E, Sanders S, Iwatsubo T, Takeuchi A, Saido T, Zehr C, Yu X, Ulijon S, Wang R, Mann D, Dickson D, Duff K. (1999). Amyloid phenotype characterization of transgenic mice overexpressing both mutant amyloid precursor protein and mutant presenilin 1 transgenes. *Neurobiol. Dis.* **6**, pp.231-244.
- Mishkin M. (1978). Memory in monkeys severely impaired by combined but not by separate removal of amygdala and hippocampus. *Nature.* **273**, pp. 297-298.
- Mori T. (2001). Cholesterol accumulates in senile plaques of Alzheimer disease patients and in transgenic APP(SW) mice. *J. Neuropathol. Exp. Neurol.* **60**, pp. 778-785.
- Othman G, Brian L, Matthew S, Mary MH. (2006). High cholesterol content in neurons increases BACE, β -amyloid, and phosphorylated tau levels in rabbit hippocampus. *Exp Neurol.* **200**, pp. 460-467.
- Racchi M, Baetta R, Salvietti N, Ianna P, Franceschini G, Paoletti R, Fumagalli R, Govoni S, Trabucchi M, Soma M. (1997). Secretory processing of amyloid precursor protein is inhibited by increase in cellular cholesterol content. *J. Biochem.* **322**, pp 893-898.
- Refolo LM, Malester B, LaFrancois J, Bryant-Thomas T, Wang R, Tint GS, Sambamurti K, Duff K, Pappolla MA. (2000). Hypercholesterolemia accelerates the Alzheimer's amyloid pathology in a transgenic mouse model. *Neurobiol. Dis.* **7**, pp. 321-331.
- Refolo LM, Pappolla MA, LaFrancosi J. (2001). A cholesterol-lowering drug reduces beta-amyloid pathology in a transgenic mouse model of Alzheimer's disease. *Neurobiol. Dis.* **8**, pp. 890-899.
- Simons M, Keller P, De Strooper B. (1998). Cholesterol depletion inhibits the generation of beta-amyloid in hippocampal neuron. *Proc Natl Acad Sci USA.* **95**, pp. 6460-6464.
- Shie FS, Jin LW, Cook DG, Leverenz JB, LeBoeuf RG. (2002). Diet-induced hypercholesterolemia enhances brain A beta accumulation in transgenic mice. *Neuroreport.* **13**, pp. 455-459.
- Sparks DL, Scheff SW, Hunsaker III JC, Liu H, Landers T, Gross DR. (1994) Induction of Alzheimer-like beta-amyloid immunoreactivity in the brains of rabbits with dietary cholesterol. *Exp. Neurol.* **126**, pp. 88-94
- Sparks DL, Kuo YM, Roher A, Martin T, Lukas RJ. (2000). Alterations of Alzheimer's disease in the cholesterol-fed rabbit, including vascular inflammation: preliminary observation. *Ann N Y Acad Sci.* **903**, pp. 335-344.
- Squire LR, Zola-Morgan S. The medial temporal lobe memory system. *Science* 1991, **253**, pp. 1380-1386.
- Van Hoesen GW, Hyman BT, Damasio AR. (1991). Entorhinal cortex pathology in Alzheimer's disease. *Hippocampus.* **1**, pp. 1-8.
- Vargha-Khadem F, Gadian DG, Watkins KE, Connelly A, Van Paesschen W, Mishkin M. (1997). Differential effects of early hippocampal pathology on episodic and semantic memory. *Science.* **277**, pp. 376-380.
- Wahrle S, Das P, Nyborg AC. (2002). Cholesterol-dependent gamma-secretase activity in buoyant cholesterol-rich membrane microdomains. *Neurobiol. Dis.* **9**, pp. 11-23.
- Witter MP, Amaral DG. (1991). Entorhinal cortex of the monkey. V. Projections to the dentate gyrus, hippocampus, and subicular complex. *J Comp Neurol.* **307**, pp. 437-459
- Yang LB, Lindholm K, Yan R, Citron M, Xia W, Yang XL, Beach T, Sue L, Wong P, Price D, Li R, Shen Y. (2003). Elevated beta-secretase expression and enzymatic activity detected in sporadic Alzheimer disease. *Nat. Med.* **9**, pp. 3-4.