

Stereoselective Asymmetric Diels-Alder Reaction of Chiral Lactone Derived from D-Glucose with Cyclopentadiene

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In a previous paper, we reported versatile chiral norbornenes as leukotriene D4 antagonists.¹⁾ The chiral norbornenes were synthesized through asymmetric Diels-Alder reaction between cyclopentadiene and chiral dienophile derived from D-glucose. The chiral dienophile used in the previous study had *trans* double bond, which eventually gave only *trans* relative configurations at newly formed chiral carbons in norbornenes products (A-D) (Fig. 1).

Therefore, it is desirable to synthesize chiral dienophile with *cis*-configuration to expand the diversities of chiral norbornenes (E-H) which would show improved activities as

leukotriene D4 antagonism (Fig. 2).²⁻⁶⁾

In this communication, facial synthesis of *cis* lactone as a chiral dienophile from D-glucose and its application in asymmetric Diels-Alder reaction in high stereoselective manner will be discussed.

Chiral aldehyde **1** is conveniently prepared by a simple three-step reaction from D-glucose.¹⁾ When Wittig reaction was applied to chiral aldehyde **1** with $\text{Ph}_3\text{P}=\text{CHCO}_2\text{Me}$ at room temperature, crude mixture of *trans* ($J = 15.8$ Hz) and *cis* ($J = 11.6$ Hz) dienophile (**2**) was obtained as a *trans* major product (*trans* : *cis* = 4 : 1). However, Horner-Emmons alkenation of chiral aldehyde **1** with bis(2,2,2-trifluoroethyl) (methoxycarbonylmethyl)phosphonate⁷⁾ gave dienophile with *cis* major (*trans* : *cis* = 1 : 4). The *cis-trans* mixture **2** was treated with silica gel in MeOH solution, which smoothly transformed *cis* isomer into lactone **3**. Due to difficulty in separation of the *cis* isomer from *cis-trans* mixture of **2**, the resulting chiral lactone made the separation very convenient.

Diels-Alder reaction between chiral lactone **3** and cyclopentadiene under thermal conditions gave only two *Si*-face attacked stereoisomeric norbonenes with isomeric ratio (major : minor = 11 : 1) out of four possible stereoisomers. The relative configurations at the newly formed carbons at C-5 and C-6 were assigned by ¹H COSY NMR.

Discrimination between *endo* products (**4** or **5**) and *exo* products (**6** or **7**) is relatively easy. The *endo* products (**4** or **5**) have two protons (H5 and H6) in *exo* mode, which show relatively low-field chemical shifts in ¹H-NMR spectra.⁸⁾ The

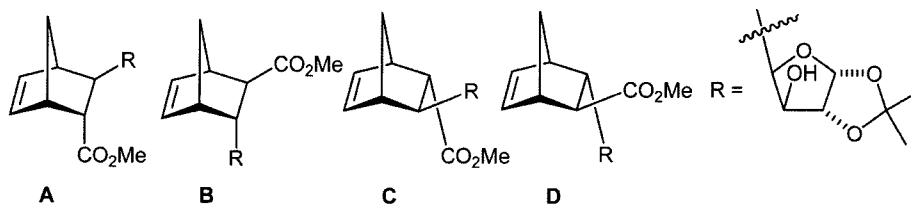


Fig. 1. Enantiomerically pure norbornenes with *trans* relationship substituents from D- glucose.¹⁾

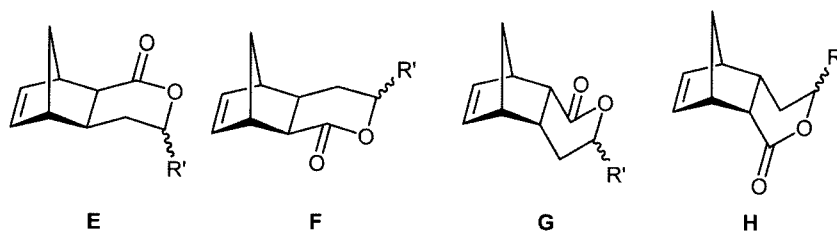


Fig. 2. Enantiomerically pure norbornenes with *cis* relationship substituents.

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Abbreviations: NMR, nuclear magnetic resonance; NOE, nuclear Overhauser enhancement; NOESY, nuclear Overhauser and exchange spectroscopy; TMS, tetramethylsilane.

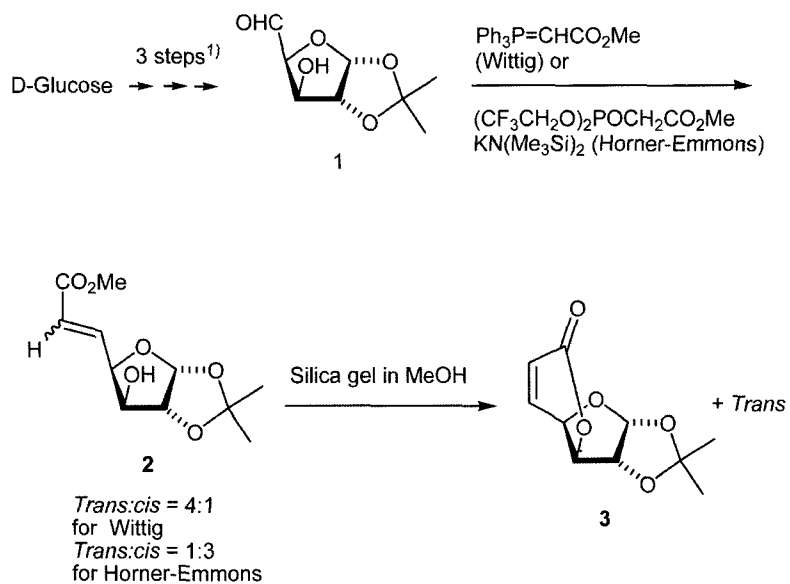


Fig. 3. Synthetic methods for chiral *cis* dienophile **3** from D-glucose.

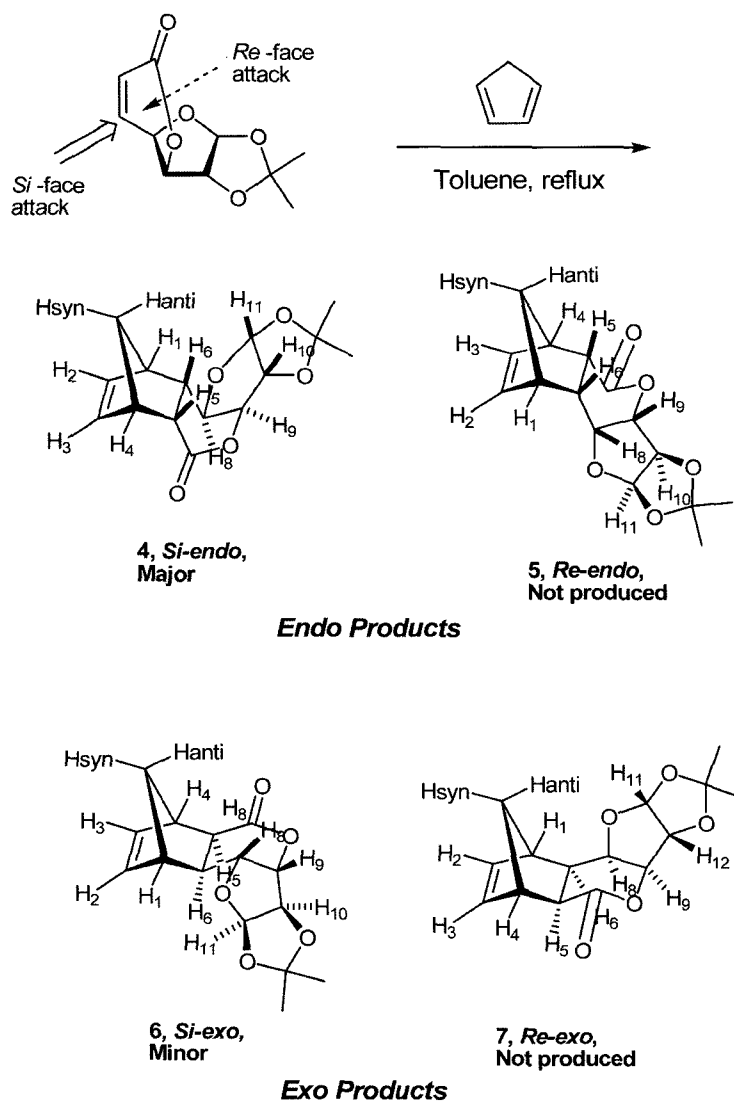


Fig. 4. Asymmetric Diels-Alder reaction between chiral *cis* dienophile **3** and cyclopentadiene.

major stereoisomeric norbornene shows chemical shifts of H-5 at 3.09 ppm and H-6 at 2.91 ppm, indicating it is one of the *endo* norbornenes (**4** or **5**). On the other hand, the minor stereoisomeric norbornene shows relatively high-field chemical shifts of H-5 at 2.46 ppm and H-6 at 2.22 ppm, indicating it is one of the *exo* norbornenes (**6** or **7**). Additional supporting evidence that the major isomer is an *endo* product was obtained from NOESY spectrum, in which strong NOE signals of H-5/ H-7_{anti} and H-6/ H-7_{anti} were observed.

The absolute configuration of the major product **4**, differentiating it from the other possible *endo* isomer **5**, could also be established through NOESY spectrum. Weak NOE signals were shown between H-2, H-3 (both protons have the same chemical shift at 6.28 ppm) and H-9 (4.38 ppm), H-8 (4.28 ppm), which are relatively close to each other in three dimensional model. No NOE between H-6 and H-9 was detected in NOESY spectrum of major product **4**, which indicates that H-5 and H-6 are in opposite direction to H-8 and H-9 in the lactone plane. If major *endo* isomer were **5**, it would show strong NOE between H-6 and H-9 and insensible NOE signals between H-2, H-3 and H-8, H-9. A similar discrimination between two possible minor *exo* isomers **6** and **7** was made through NOESY spectrum. Two strong NOE signals (H-2/H-6 and H-3/H-5) indicate the minor product is an *exo* isomer. A large NOE signal between H6 and H11 rules out the possibility of structure **7**, thus permitting assignment of the minor product **6**.

It is noteworthy that the large coupling constant between H-5 and H-6, which are attached to the newly formed carbons C-5 and C-6 respectively ($J_{5,6} = 9.2$ Hz in major **4**, $J_{5,6} = 8.6$ Hz in major **6**) shows that H5 and H6 are in the *cis* relationship ($J_{5,6} = 4.4$ -5.2 Hz in the *trans* isomers **A-D**),¹⁾ confirming that *cis*-relationship between H-5 and H-6 was maintained during the Diels-Alder reaction.

These two assignments of absolute configurations revealed that the diene attacks the dienophile **3** exclusively from the *si*-face. *Re* face of the dienophile's δ -lactone ring is strongly shielded by two sugar rings. On this steric ground it is clear that favored attack of diene would be exclusively from the *si*-face of the dienophile, resulting in a very high diastereofacial selectivity.

Dienophile 3.; ¹H-NMR (400 MHz, CDCl₃); δ 1.33 (s, 3H, CH₃), 1.54 (s, 3H, CH₃), 4.63 (m, 1H, H₃), 4.82 (m, 2H, H₄, H₅), 6.02 (d, 1H, $J_{3,7}$ 3.7 Hz, H₆), 6.23 (d, 1H, $J_{9,7}$ 9.7 Hz, H₁), 6.96 (dd, 1H, $J_{9,7}$ 9.7, 5.6 Hz, H₃), ¹³C-NMR (100 MHz); δ . 160.8 (C=O), 138.6 (C3), 125.4 (C2), 112.6 (C7), 105.3 (C8), 83.4 (C4), 82.4 (C5), 67.6 (C6), 26.7 (CH₃), 26.2 (CH₃).

4; ¹H-NMR (400 MHz, CDCl₃); δ 1.31 (s, 3H, CH₃), 1.48 (d, 1H, $J_{7syn,7anti}$ 8.7 Hz, H_{7syn}), 1.52 (s, 3H, CH₃), 1.58 (dd, 1H, $J_{1,7}$ 1.7, 8.7 Hz, H_{7anti}), 2.91 (d, 1H, $J_{5,6}$ 9.2 Hz, 3.4 Hz, H₆), 3.09

(dd, 1H, $J_{4,2}$ 4.2, 9.2 Hz, H₅), 3.16 (bs, 1H, H₁), 3.40 (bs, 1H, H₄), 4.28 (bs, 1H, H₈), 4.38 (d, 1H, $J_{2,3}$ 3.8 Hz, H₁₀), 5.78 (d, 1H, $J_{10,11}$ 3.8 Hz, H₁₀), 6.28 (m, 2H, H_{2,3}), ¹³C-NMR (100 MHz); δ . 171.2 (C=O), 136.2 (C2), 135.9 (C3), 111.9 (C12), 103.2 (C11), 84.3 (C10), 81.6 (C9), 73.7 (C8), 50.7 (C7), 48.8 (C4), 46.6 (C1), 40.0 (C5), 39.9 (C6), 26.4 (C14), 26.0 (C13).

6; ¹H-NMR (400 MHz, CDCl₃); δ 1.33 (s, 3H, CH₃), 1.40 (d, 1H, $J_{7syn,7anti}$ 9.6 Hz, H_{7syn}), 1.46 (dd, 1H, $J_{5,6}$ 1.5, 9.6 Hz, H_{7anti}), 1.54 (s, 3H, CH₃), 2.22 (d, 1H, $J_{5,6}$ 8.6 Hz, H₆), 2.46 (ddd, 1H, $J_{1,2}$ 0.8, 1.5, 8.6 Hz, H₅), 2.89 (bs, 1H, H₁), 3.30 (bs, 1H, H₄), 4.41 (t, 1H, $J_{8,9}$ 1.9 Hz, H₈), 4.74 (d, 1H, $J_{10,11}$ 3.8 Hz, H₁₀), 4.67 (d, 1H, $J_{2,3}$ 3.8 Hz, H₉), 5.83 (d, 1H, $J_{11,10}$ 3.8 Hz, H₁₁), 6.25 (dd, 1H, $J_{2,9}$ 2.9, 5.5 Hz, H₃), 6.36 (dd, 1H, $J_{3,1}$ 3.1, 5.5 Hz, H₂), ¹³C-NMR (100 MHz); δ . 171.9 (C=O), 139.0 (C2), 136.5 (C3), 112.1 (C12), 103.4 (C11), 84.4 (C10), 81.6 (C9), 74.7 (C8), 50.7 (C4), 45.9 (C1), 44.5 (C7), 40.0 (C5), 39.3 (C6), 26.4 (C14), 26.1 (C13).

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