Studies on Viability of Frozen Sperm and Pregnancy Rates after AI with Frozen-Thawed Canine Semen

B. K. Lee, Y. H. Chung and S. K. Kim^{1†}

Dept. of Companion Animal & Animal Resource Science, Joong-Bu University

개 동결 정액의 생존성과 AI 후 임신율에 관한 연구

이봉구·정영호·김상근^{1†} 중부대학교 애완동물자원학과

SUMMARY

본 연구는 개 채취 정액의 동결후의 생존성과 신선 및 동결 정자의 capacitation, acrosome reaction과 생존성을 조사하고, 아울러 신선 및 동결 정액을 자연 발정 또는 발정 유기 암캐에게 인공수정 후 임신율을 조사하였다.

개 채취 정액의 동결 융해 후의 생존성은 64.5±2.30%로서 신선 정액의 생존성에 비해 유의하게 낮게 나타났다(p<0.05). 신선 및 동결 정액의 capacitation, acrosome reaction 및 생존성은 각각 52.5± 4.5%, 9.5±0.6%, 68.8±4.5% 및 16.2±3.2%, 3.2±0.5%, 24.5±2.5%로 나타났다. 신선 및 동결 정액을 자연 발정 또는 발정을 유기한 암캐에 인공수정했을 때 임신율은 각각50.0% 및 33.3%로서 동결 정액을 이용했을 때 임신율이 신선 정액에 비해 낮은 임신율을 나타냈다.

(Key words: canine oocytes, IVM/IVF, stages of reproductive cycle)

INTRODUCTION

In male dogs the number of sperm in ejaculate significantly decreased and female dogs have shown disturbances of reproductive cycle and reproductive diseases (Gunzel, 1986; Kim, 2001).

It was reported that the pregnancy rate was not decreased when artificial inseminated with fresh semen of the 2nd fraction of canine semen, but that the viability was decreased when the sperm was preserved *in vitro* (Maule, 1960; Arthur, 1975). Seager and Fletcher (1973) reported that when ferti-

lized with semen preserved for 1~4 days, the pregnancy rate was 53%. Province et al. (1984), Davis et al. (1963) and Foote (1964) reported that the viability rate was maintained at 50% when the canine semen was cryopreserved with 20% egg yolk extender for 2~4 days or 4~8 days. Harrop (1962) reported that the survival rate of post-thaw sperm was 40~50%. Seager et al. (1975) reported the first success in pregnancy and parturition of canine by AI with frozen semen. Dog epididymal sperm collected after euthanasia has been frozen and successfully used after storage for 3.5 months, although the sperm

¹ 충남대학교 수의과대학 (College of Veterinary Medicine, Chungnam National University)

^{*}Correspondence: E-mail: kskkim@cnu.ac.kr

recovery was poor, the study involved only one animal and the apparent fertility was low (Marks et al., 1994). Harrop (1962) reported a 45~50% survival rate after thawing, which showed the possibility of long term freezing of canine semen. Seager and Fletcher (1973) reported pregnancy rate of 53% when inseminated with semen preserved for 1~4 days. Seager et al. (1975) reported the first success in pregnancy and parturition by using canine frozen-thawed semen.

The studies was carried out to investigate the viability of fresh and frozen sperm and the percentages of capacitated sperm and acrosome-reacted sperm and the pregnancy rates after artificial insemination with frozen-thawed semen.

MATERIALS AND METHODS

1. Collection and Freezing of Semen

Twenty male dogs and 60 female dogs were used in this study and they internal parasites and vaccinated prior to use. Semen was collected by hand massage method. The highest concentration of semen was used.

A 1.0 ml volume of saline was added to sperm suspension and then a 4.0 mL extender buffer modified with cryoprotectants was mixed with the sperm-saline suspension. The ejaculated semen were diluted with physiological saline and tris-buffer (1:3), centrifuged for 6 min with 700 g and the supernatant was removed. Semen was supplemented with 1.5 ml extender-1 at room temperature and the diluted solution was cooled for 45 min at 4°C. Following cooling for 25 min, 1.5 ml extender-2 and extender-2' were supplemented at 4°C and then mixed. Freezing was carried out by the method of Kim (2001).

2. Viability of Frozen Sperm

Straw semen cryopreserved for 1 month was placed for 3 min at room temperature and then

thawed in a water bath at 37°C. In order to remove cryoprotectants, frozen semen was shaken upside down and tris-buffer were removed and dissolved. The dissolved semen samples were moved to a slide glass and observed by microscope and the viability, survival rate, and morphological test was examined by using sperm analyzer imaging system (SAIS Si-100).

3. Al and Pregnancy Diagnosis

Non-estrus dogs were treated with 20 mg of PGF_2a and then with 2,000 IU of PMSG and 2,000 IU of HCG. Estrus, induced dogs and the dogs with natural estrous symptoms were inseminated artificially. Frozen straw semen (0.5 ml) was kept at room temperature for 30 sec thawed in waterbath at 37 °C. Artificial insemination was carried out with transcervical catheter and thawed semen was deposited non-surgically into the uterine body or into a uterine horn at 1~4 days before and after ovulation.

After artificial insemination, the pregnant dogs did not show and had expanded abdomen symptom. At day 20 to 30 after AI. pregnancy was tested by ultra-sonography.

4. Statistical Analysis

For comparison of means, Duncan's multiple range test was performed using SAS package of General Linears Model (GLM) procedures (SAS Institute, 1996).

RESULTS AND DISCUSSION

1. Viability of Frozen Semen

The canine semen was diluted with tris-buffer solution and centrifuged to remove seminal plasma was suspended with extender-1, extender-2 solutions and cooled by vitrification.

Table 1 presents the viability of fresh and frozen semen thawed after freezing. The viability rates of frozen semen by freezing were $64.5\pm2.30\%$. These rates were lower than that of the control group ($84.2\pm2.55\%$). The above results of frozenthawed semen were higher than that of Takeishi *et al.* (1975), who reported that the survival rate of whole semen after cryopreservation was $40\sim65\%$.

2. Sperm Category before and after Freezing

The acrosome status these capacitated, acrosomereated and live sperm was is shown in Table 2.

The percentages of capacitated sperm and acrosome-reacted sperm was 52.5±4.5%, 9.5±0.6% fresh semen and 16.2±3.2%, 3.2±0.5% in frozen semen and live sperm of fresh and frozen semen were 68.8±4.5%, 24.5±2.5%, respectively. Province *et al.* (1984), Davis *et al.* (1963) and Foote (1964) who

Table 1. Viability of frozen-thawed semen following slow and rapid freezing

Treatment	Motility of semen		
	Live (%)	Dead (%)	
Fresh semen ^a	84.2±2.55	15.8±2.25	
Frozen semen ^b	64.5±2.30	35.5±3.10	

^{*} a,b: Values with different letters within same columns differ significantly (p<0.05).

Table 2. Rate of capacitation and acrosome reaction prior to culture for fresh and frozen-thawed ejaculated sperm

Parameter	Fresh semen	Frozen semen
Capacitated sperm ^a	52.5±4.5	16.2±3.2
Acrosome-reacted sperm ^b	9.5±0.6	3.2±0.5
Live sperm ^a	68.8±4.5	24.5±2.5

^{*} a,b : Values with different letters within same columns differ significantly (p<0.05).

re-ported survival rate of 50% when the semen was cryopreserved with 20% egg yolk extender for 2 to 8 days. Harrop (1962) also reported the survival rate of 45~50% in freezing-thawed canine semen. The sperm capacitation rate *in vitro* in dog have been investigated by several researchers (Hewitt and England, 1997; Hewitt and England, 1998; Mahi and Yanagamachi, 1978: Nickson *et al.*, 1993; Shimazu *et al.*, 1992; Yamada *et al.*, 1993).

3. Pregnancy Rate after Al

Table 3 presents the pregnancy rates after with fresh and frozen semen. The pregnancy rate of fresh semen and frozen semen were 65.0% and 40.0%. This results was significantly lower than that of fresh semen (65.0%).

The pregnancy rate was lower with result (81%) reported by Tsutsui and Shimizu (1973) in while AI was performed with fresh semen in the period of 54~108 hrs before and after ovulation. Generally, the pregnancy rate increased when artificial insemination was executed 1~2 days at the 1st fraction and 3~4 days at the 2nd fraction after estrus. There was no respective comparison here (Maule, 1960; Arthur, 1975). This result was similar a or a little lower than that of Seager and Fletcher (1973), who reported pregnancy rate of 53% when inseminated with semen preserved for 1~4 days.

Table 3. Pregnancy rates of frozen-thawed ejaculation and epididymal semen

Treatment	No. of dogs (%)*		
	Examined	ΑI	Pregnant
Fresh semen	20	20	13/20 (65.0) ^a
Frozen semen	30	25	10/25 (40.0) ^a

^{*} a,b: Values with different letters within same columns differ significantly (p<0.05).

^{**} Mean±S.D.

^{**} Mean±S.D.

^{**} Mean±S.D.

^{**} Within 20~30 days after AI. Pregnancy was determined by ultrasound test.

CONCLUSION

The studies were carried out to investigate the viability of fresh and frozen sperm and the percentages of capacitated sperm and acrosome-reacted sperm and the pregnancy rates after artificial insemination with frozen-thawed semen.

- 1. The viability rates of frozen semen by freezing were 64.5±2.30%. These rates were lower than that of the control group (84.2±2.55%).
- 2. The percentages of capacitated sperm and acrosome-reacted sperm was 52.5±4.5%, 9.5±0.6% fresh semen and 16.2±3.2%, 3.2±0.5% in frozen semen and live sperm of fresh and frozen semen were 68.8±4.5%, 24.5±2.5%, respectively.
- 3. The pregnancy rate of fresh semen and frozen semen were 65.0% and 40.0%. This results was significantly lower than that of fresh semen (65.0%).

REFERENCES

- Arthur GH. 1975. Veterinary Reproduction and Obstetrics, 4th ed. The English Language Book Society and Bailliere Tindall. England, pp. 335-337.
- Davis IS, Brattion RW and Foote RH. 1963. The livability of spermatozoa at 5, 25 and 85 ℃ in tris-buffered and citrate-buffered yolk-glycerol extenders. J. Dairy Sci., 46:333-336.
- Foote RH. 1964. The effects of electrolytes, sugars, glycerol and catalase on survival of dog sperm stored buffered-yolk mediums. Am. J. Vet. Res., 25:32-36.
- Gunzel AR. 1986. Semen collection, evaluation, preservation and artificial insemination in the dog. Tierarzti Prax., 14:275-282.
- Harrop AE. 1962. Artificial insemination in the dog.

 In the semen of animals and artificial insemi-

- nation. J.P. Maule, ed. Commonwealth Agri. Bureaux, Farnham Royal, England, pp. 186-189.
- Hewitt DA and England GCW. 1997. The canine oocyte penetration assay its use as an indicator of dog spermatozoal performance *in vitro*. Anim. Reprod. Sci., 50:123-139.
- Hewitt DA and England GCW. 1998. An investigation of capacitation and the acrosome reaction in dog spermatozoa using a dual fluorescent staining technique. Anim. Reprod. Sci., 51:321-332.
- Kim SK. 2001. Studies on the viability of frozen removed seminal plasma by RSP-S and RSP-T semen of small species dogs. Korean J. Anim. Reprod., 25(3):269-275.
- Mahi CA and Yanagamachi R. 1978. Capacitation, acrosome reaction and reaction and egg penetration by canine spermatozoa in a simple defined medium. Gamete Res., 1:102-109.
- Marks SL, Mickelsen WD, Memon MA and Platz CC. 1994. Conception by use of postmortem epididymal semen extraction in dog. J. Am. Vet. Med. Asso., 204:1639-1640.
- Maule JP. 1960. The Semen of animals and artificial insemination, common wealth. Agri. Bur., Farnham. Royal, England, pp. 287-285.
- Nickson DA, Boyd J, Eckersall PD, Ferguson JM, Harvey MJA and Renton JP. 1993. Molecular biological methods for monitoring oocyte maturation and *in vitro* fertilization in bitches. J. Reprod. Fert. (Suppl.), 47:231-240.
- Province CA, Ammann RP, Pickett BW and Squires EL. 1984. Extenders for preservation of canine and equine spermatozoa at 5℃. Theriogenology, 22:409-415.
- Seager SWJ and Fletcher WS. 1973. Progress on the use of frozen semen in the dog. Vet. Rec., 92:6-10.
- Seager SWJ, Platz CC and Fletcher WS. 1975. Conception rates and related data using frozen dog semen. J. Reprod. Fertil., 45:189-192.

Shimazu Y, Yamada S, Kawano Y, Kawayi H, Nakazawa M, Naito K and Toyoda Y. 1992. *In vitro* capacitation of canine spermatozoa. J. Reprod. Dev., 38:67-71.

Takeishi M, Mikami T, Godama Y, Tsunekane T and Iwaki T. 1975. Studies on the physiology of reproduction in the dog W. Artificial insemination using the frozen semen. Japan J. Anim. Reprod., 22:28-33.

Tsutsui T and Simizu T. 1973. Studies on the physiology of reproduction in the dog. I Duration of estrus. Japan J. Anim. Reprod., 18:132-135. Yamada S, Shimazu Y, Kawao Y, Nakazawa M, Naito K and Toyoda Y. 1993. *In vitro* maturation and fertilization of pre-ovulatory canine oocytes. J. Reprod. Fert (Suppl.), 47:227-229.

(접수일: 2005. 12. 5 / 채택일: 2006. 2. 25)