Neurovascular Mechanisms in Stroke, Neurodegeneration and Recovery

Eng H. Lo

Neuroprotection Research Laboratory, Departments of Radiology and Neurology, Massachusetts General Hospital, and Program in Neuroscience, Harvard Medical School, Boston, USA

The emerging concept of the "neurovascular unit" may enable a powerful paradigm shift for neuroscience. Instead of a pure focus on the "neurobiology" of disease, an opportunity now exists to return to a more integrative approach. The neurovascular unit emphasizes that signaling between vascular and neuronal compartments comprise the basis for both function and dysfunction in brain. Hence, brain disorders are not just due to death of neurons, but instead manifested as cell signaling perturbations at the neurovascular interface. In this mini-review, we will examine 3 examples of this hypothesis: neurovascular mechanisms involved in the thrombolytic therapy of stroke, the crosstalk between neurogenesis and angiogenesis, and the link between vascular dysfunction and amyloid pathology in Alzheimer's disease. An understanding of cell-cell and cell-matrix signaling at the neurovascular interface may yield new approaches for targeting CNS disorders.

Key Words: Neurovascular unit, Tissue plasminogen activator, Stroke, Matrix metalloproteinase, Alzheimer's disease, Neurogenesis

INTRODUCTION

Over the past decade, major advances in the neurobiology of CNS disorders were achieved. Fundamental mechanisms underlying neuronal cell death were elucidated, including excitotoxicity, oxidative damage via free radicals, and apoptotic-like events that are triggered after brain injury. Altogether, these advances have revealed a plethora of therapeutic targets for the brain. For example, it was demonstrated that after onset of cerebral ischemia in experimental animal models, interstitial glutamate concentrations rise dramatically to neurotoxic levels (Buchan, 1990; Benveniste, 1991; Lipton & Rosenberg, 1994). Blockade of glutamate receptors of the NMDA or AMPA subtypes all seem to reduce brain infarction in animal stroke models (Hirose & Chan, 1993). Disappointingly, however, a wide range of clinical stroke trials testing glutamate receptor antagonists have failed to show efficacy in patients.

Why have these therapeutic approaches not achieved significant success? Whereas in isolated cell cultures, specific neuronal death pathways were clearly validated, the pharmacologic probes developed based on these paradigms have not translated well into the clinic. There are many reasons why clinical trials of neuroprotection are inherently challenging, and the reader is referred to many other more detailed reviews on this topic (De Keyser et al, 2000; Gladstone et al, 2002). However, an emerging concept that has gained momentum in recent years is the realization that the brain is not a neuron! Brain function and dysfunction

arises from a complex interplay between a network of multiple cell types, including neurons, astrocytes, oligodendrocytes, microglia, and ultimately, the cerebrovasculature that permeates the entire organ.

From a functional perspective, it is the interaction between neuron and astrocyte that mediates neurotransmitter release and reuptake at the synapse. The integrity of the blood-brain barrier depends on cell-cell signaling between the astrocyte and the cerebral endothelium at the microvessel level. And the impressive advances in functional MRI reveal the intricacies of brain function would not be possible without the hemodynamic coupling between neuronal firing and vascular response. A simplified schematic in Fig. 1 summarizes these interactions.

The neurovascular unit provides a conceptual framework that emphasizes cell-cell signaling in the brain (Lo et al, 2004; Hawkins & Davis, 2005; Allan, 2006). Brain disease is therefore manifested as a pdlferturbation in signaling within the cells of the neurovascular unit (Iadecola, 2004; Zlokovic, 2005; Abbott et al, 2006). Even though we have made enormous advances in understanding intra-neuronal mechanisms of cell death, we may have to enlarge our focus to the level of the neurovascular unit if we are to make a difference at the organ level. In this mini-review, we will briefly examine 3 "case studies" of the neurovascular unit: (a) the pleiotropic actions of various proteases during thro-

ABBREVIATIONS: NMDA, N-methyl-D-asparate; AMPA, alpha-amino-3-hydroxy-5-methylisoxazole-4-propionate; CNS, central nervous system; LRP, low density lipoprotein receptor related protein; BBB, blood brain barrier; MMP, matrix metalloproteinase; tPA, tissue plasminogen activator; MRI, magnetic resonance imaging; PET, positron emission tomography.

Corresponding to: Eng H. Lo, Neuroprotection Res Lab, MGH East 149-2401, Charlestown, MA 02129, USA. (Tel) +1-617-726-4043, (Fax) +1-617-726-7830, (E-mail) Lo@helix.mgh.harvard.edu

224 Eng H. Lo

mbolysis that may explain responses in stroke therapy, (b) the interplay between neurogenesis and angiogenesis in brain remodeling after injury, and (c) the cerebrovascular response to amyloid in Alzheimer's disease.

Tissue Plasminogen Activator: A Pleiotropic Neurovascular Mediator in Stroke

Historically, two phenomenon from animal models suggested that active cell death mechanisms are triggered after cerebral ischemia. In the late 1970's, Astrup, Symon and colleagues demonstrated that the ischemic penumbra comprised transiently viable tissue whereby moderate ischemia resulted in loss of evoked potentials but not resting membrane potentials (Astrup et al, 1977). With a return of blood flow, penumbral evoked potentials could be rescued. But without reperfusion, the penumbra would collapse over time and anoxic depolarization takes place. The second phenomenon involved selective neuronal vulnerability, whereby pyramidal neurons in the CA1 sector of the hippocampus underwent delayed cell death within 2~3 days after a transient global cerebral ischemic insult (Garcia, 1988; Paschen, 1996; Harukuni & Bhardwaj, 2006). Taken together, these observations supported the hypothesis that complex and undefined mechanisms are indeed activated that eventually leads to cell death after stroke.

Whatever these neuronal mechanisms might be, a logical therapeutic approach to cerebral ischemia is to restore blood flow. Therefore, thrombolysis with tissue plasminogen activator (tPA) is a rational therapy, and in properly selected patients, tPA works very well (ECASS Study Group, 1995; NINDS rt-PA Stroke Study Group, 1995). However, many limitations still exist. The treatment time window is exceedingly narrow. Not all patients respond. And there remains an overall risk of edema and hemorrhagic conversion (Hacke et al, 1999). Over the past several years, emerging data from cell and animal model systems now suggest that these caveats with tPA stroke therapy might be due to the fact that tPA is not only a "blood molecule" but also has neuroactive properties (Kaur et al, 2004; Benchenane et al, 2005). In this regard, tPA is perhaps best understood as a neurovascular mediator with pleiotropic actions precisely at the critical neurovascular interface.

The primary goal of using tPA in stroke is straight-forward. By converting plasminogen into active plasmin, fibrin is degraded and the offending embolic clot is dissolved. But tPA may do much more. tPA has been shown to be vasoactive. Depending on the concentrations, tPA can be either vasodilatory or vasoconstrictive (Nassar et al, 2004). tPA may also be a critical neuronal mediator that is released in a calcium dependent manner (Gualadris et al, 1996). The plasminogen protease system may play a key role in modulating extracellular microenvironment during synaptic remodeling. Indeed, tPA knockout mice show perturbations in several paradigms of long term potentiation (Baranes et al, 1998).

Tsirka and colleagues demonstrated that tPA can be deleterious to neurons. After kainic acid injections into the hippocampus, tPA deficient knockout mice were resistant to excitotoxic injury compared with wildtype mice (Tsirka et al, 1995; Tsirka et al, 1996). Subsequently, Tsirka, Strickland and Lipton showed that these neuronal effects of tPA were also implicated in stroke; tPA knockout mice suffered significantly reduced infarctions after focal cerebral ische-

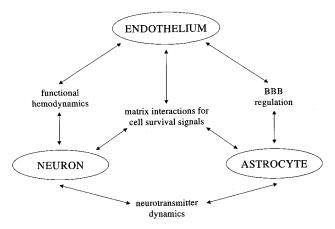


Fig. 1. Schematic of the neurovascular unit, emphasizing the functional aspects of cell-cell and cell-matrix signaling. Note that white matter is not included in this initial simplified schema. A full dissection of signaling between multiple glial cell types should also be considered. Adapted from Lo et al, *Stroke* 2005.

mia (Wang et al, 1998). The underlying mechanisms may involve anoikis since degradation of inter-neuronal laminin seemed to be involved (Chen & Strickland, 1997). Additionally, a linkage between proteolysis and excitotoxicity may also contribute. Vivien and colleagues showed that tPA interacted with the NR1 subunit of the NMDA receptor complex (Nicole et al, 2001). Cleavage of this subunit amplifies calcium currents which may augment excitotoxicity. This novel mechanism may serve to explain why tPA may be neurotoxic under certain conditions. More recently, Tsirka and colleagues have also demonstrated that tPA may behave as a chemokine since sources in damaged brain can activate microglia and trigger downstream neuroinflammation (Rogove et al, 1999; Wang et al, 2003a).

In addition to neuronal processes, tPA can also trigger changes in other protease systems. A major hypothesis is that complications of bleeding and edema after thrombolysis is due to signaling connections between the plasminogen system and the matrix metalloprotease (MMP) system. MMP inhibitors reduce tPA-associated hemorrhage in embolic stroke models (Lapchak et al, 2000; Sumii & Lo, 2002). It was demonstrated that tPA can bind to the LRP lipoprotein receptor and induce a transcriptional upregulation in MMPs (Wang et al, 2003b). tPA knockout mice have reduced MMPs and brain edema after cerebral ischemia (Tsuji et al, 2005). These experimental notions are now supported by accumulating clinical data. Stroke patients with increased MMPs are more likely to have worse outcomes, including increased risk of hemorrhagic conversion (Montaner et al, 2001; Montaner et al, 2003; Alvarez-Sabin et al, 2004). Acute stroke patients that receive tPA seem to have elevated MMP-9 levels (Ning et al, 2006). More recently, elevated plasma MMP-9 levels have been correlated with MMP-9 staining in human stroke brain samples as well (Rosell et al, 2006; Tejima et al, 2006). Taken together, these data raise the possibility that MMP inhibition should be considered as part of a combination stroke therapy together with tPA thrombolysis.

Recent neuroimaging data may further support this idea. Warach, Latour and Kidwell et al showed that leakage of gadolinium contrast can be detected with FLAIR MRI during the hyperacute to acute progression of ischemic stroke

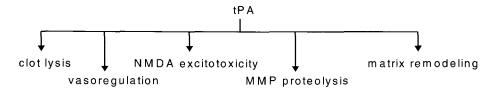


Fig. 2. Multiple actions of tPA at the neurovascular interface. In addition to its intended role as a clot-lysing agent, tPA can also be vasoactive, neuromodulatory, and by activating the MMP protease system, also mediate acute BBB damage or chronic neurovascular remodeling. Adapted from Wang et al, *Stroke* 2004.

(Warach & Latour, 2004). They have termed these subtle enhancement signals HARM or Hyperacute Reperfusion Marker. HARM seems to be correlated with negative outcomes including hemorrhage and edema observed with increased T2 at later times. Interestingly, when tPA reperfusion was compared with reperfusion using mechanical clot remover devices, HARM signals were increased. These initial data suggest that reperfusion with tPA is biologically different from reperfusion via mechanical clot removal.

Altogether, these data indicate that tPA may have multiple actions in brain (Fig. 2). By understanding these complex neurovascular interactions that are affected by tPA, we may be able to better modulate its actions, and hopefully design combination stroke therapies that target the entire neurovascular unit in addition to lysing the clot.

The Neurovascular Niche for Neurogenesis and Brain Remodeling

For many decades, a dogma in neurobiology stated that no new neurons are born in adult mammalian brain. However, beginning in the 1960's and early 1970's, challenges to this dogma began to surface, with initial data coming from the radiobiology literature (Lewis, 1968; Hopewell & Cavanagh, 1972; Privat & Leblond, 1972). DNA labeling studies indicated that in fact, there was significant cell turnover in selected sites of mouse and rat brain, specifically in the hippocampus and subventricular areas. The initial thrust of these early studies focused on the basic idea that radiation injury primarily induced a "reproductive death", i.e. DNA damage was typically not severe enough to induce outright cytotoxicity, and DNA misrepair problems only surfaced when each cell attempted to divide. The surprising radiosensitivity of adult brain suggested that cell division remained intact.

The wider implications of these observations were not fully appreciated until the late 1980's and early 1990's. Snyder, Cepko and colleagues discovered that neuroblast-like precursor cells could be isolated from mammalian brain (Ryder et al, 1990). The multipotent nature of these precursor cells were demonstrated by Weiss and colleagues; isolated cultures could be differentially induced to mature into either new neurons or astrocytes (Reynolds & Weiss, 1992). Taken together, these data led to the idea that pockets of ongoing neurogenesis (and gliogenesis) may persist in adult brain.

The concept of a vascular niche for neurogenesis was first proposed by Palmer, Gage and Goldman. It was discovered that endothelial production of brain-derived neurotrophic factor significantly enhanced neural precursor turn-over in cell cultures (Leventhal et al, 1999). In vivo, detailed micro-

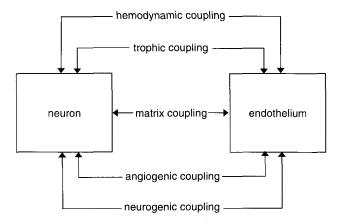


Fig. 3. Schematic emphasizing that neurovascular coupling can be manifested at multiple levels. Traditional hemodynamic couping links neuronal activity with blood flow responses that occur at millisecond timescales. Trophic-matrix coupling at longer distances and timescales as diffusible mediators may provide signals to coordinate neurogenesis and angiogenesis. Based on fundamental concepts first outlined in Park et al, Biochem Biophys Res Commun 2003, and Ward and LaManna, Neurol Res 2004.

scopic analysis revealed that there were close spatial relationships between sites of neurogenesis and active microvasculature, suggesting a functional interdependence between the two mechanisms (Palmer et al, 2000). Mechanistically, it was ultimately demonstrated that conditioned media from cerebral endothelial cells directly induced neuroblast proliferation (Shen et al, 2004). From an evolutionary perspective, similarities between the morphologic development of the nervous system have long been recognized to parallel the intricate branching profiles of the vascular network (Carmeliet, 2003; Eichmann et al, 2005). Increasingly, a molecular dissection of the signals reveal mechanistic overlaps between angiogenic and neurogenic mediators, further supporting the fundamental concept that one cannot examine neurogenesis and angiogenesis separately, but instead one has to consider neurovascular development as a coordinated phenomenon in itself. In adult brain, these trophic and signaling connections may also be vital for homeostasis, as proposed by LaManna et al and Kim and colleagues (Park et al, 2003; Ward & Lamanna, 2004) (Fig. 3).

From a clinical standpoint, interactions between neurogenesis and angiogenesis not only contribute to brain development, but also influence what happens during recovery from brain injury. As discussed earlier, disruptions in neurovascular proteases such as MMPs mediate acute damage. However, because these neurovascular proteases mediate

226 Eng H. Lo

angiogenesis, vasculogenesis and neurogenesis in developing brain, it is likely that they may also play key roles during plasticity and remodeling.

Several labs simultaneously showed that neurogenesis was upregulated after cerebral ischemia. Liu, Sharp and colleagues reported that rates of neuroblast turnover were increased in the hippocampus after transient global cerebral ischemia (Liu et al, 1998). Chopp and colleagues showed that subventricular zone cell kinetics responded sharply after focal ischemia (Zhang et al, 2001). And subsequently, the Lindwall group and the Parent lab both obtained powerful evidence in rat stroke models that increased neuroblast migration was diverted away from the baseline rostral migratory stream toward damaged striatum (Arvidsson et al, 2002; Parent et al, 2002). How do these neuroblasts move? Is it possible that MMPs may also be involved? A recent study showed that doublecortin-positive neuroblasts co-localize with MMP-9 staining (Lee et al, 2006). And broad spectrum inhibition of MMPs significantly thwarted the migratory response of these neuroblasts. Ultimately, the ability of using growth factor supplementation to amplify these neurogenic and presumably angiogenic responses provides promise that therapies for acute stroke and trauma might eventually move into the chronic phase, where one might even speculate about regrowing brain.

Besides the remote migration of newborn cells in damaged Brain, it is also recognized that important neurovascular responses may occur in peri-infarct cortex. Stroke recovery may be based in part on these morphologic substrates, as detected with functional MRI or PET (Dijkhuizen et al, 2003; Kim et al, 2005; Kim et al, 2006). Once again, a biphasic role for neurovascular proteolysis may occur. Zhao et al showed that many weeks after stroke, the peri-infarct cortex remains a dynamic and highly malleable territory (Zhao et al, 2006). Secondary elevations in MMPs are readily apparent, and these signals co-localize with surrogate markers of dendritic and microvessel regrowth. Consequently, inhibition of these MMPs during the delayed phase after stroke made things worse. Markers of neurovascular remodeling were suppressed, and infarcts and cavitations became larger. Furthermore, a significant number of animals showed signs of the development of abnormal and hemorrhagic blood vessels.

The intimate connections between neurogenesis and angiogenesis are critical not only during brain development, but also play a role in remodeling as the brain tries to heal itself after injury, neurodegenerative disease and perhaps even aging. In part, initial data suggest that neurovascular proteases such as MMPs may be involved. Thus, MMP inhibition might reduce hemorrhage during acute brain injury, a delicate promotion of endogenous MMP activities may be required for functional recovery and the matrix integration between neurogenesis and angiogenesis.

Vascular Correlates of Amyloid Neuropathology

Alzheimer's disease is a major cause of dementia in aging populations. Currently, the major pathogenic theories of Alzheimer's are focused on amyloid accumulation and altered tau processing in neurons. A full review of these molecular mechanisms that underlie Alzheimer's is outside the scope of this mini-review, and the reader is referred to many excellent reviews on this subject (Selkoe, 1999; Bossy-Wetzel et al, 2004; Mattson, 2004). However, an

emerging set of ideas over the past 5~6 years now propose that in addition to a pure neuronal disease, Alzheimer's may also have key vascular correlates that must be considered if we are to find the most efficacious treatments for this devastating disorder (Iadecola, 2004; Zlokovic, 2005; Park et al, 2006).

From a purely epidemiological perspective, it is useful to note that major risk factors for sporadic Alzheimer's are mostly cardiovascular in nature (de la Torre, 2002). The prominent Rotterdam study, Honolulu study and others have documented that increased Alzheimer's risk was correlated with hypertension, atrial fibrillation, elevated homocysteine, diabetes, smoking, thrombosis, and atherosclerosis (Breteler, 2000). Increasingly, the research community is acknowledging that there are blurred boundaries and many overlaps between "true" Alzheimer's dementia and vascular-related dementia. PET imaging studies have long demonstrated that early perturbations in cerebral blood flow and metabolism, especially in frontal and temperoparietal cortex, were hallmarks of Alzheimer's and neurodegeneration (de la Torre, 2002). And ultimately, the cognitive deficit profiles in vascular dementia, Alzheimer's and mild cognitive impairment all suggest a spectrum of dysfunction rather than strict and clear-cut categorical differences per se.

A classic example might perhaps be found in analysis of transgenic mice that overexpress amyloid precursor protein (McGowan et al, 2006). As expected, plaques accumulate at later stages of life, corresponding with behavioral defects and neuronal dysfunction. However, a closer look at these brains suggest that vascular changes may occur even earlier. Markers of oxidative and nitrosative stress, as indicated by nitrotyrosine formation, became apparent even before extracellular amyloid deposits occurred (Park et al, 2004). Furthermore, it was shown that critical problems with neurovascular coupling were present in these mice. Facial whisker stimulation is an established paradigm for activating whisker barrel cortex in rodent brains. Realtime measurements of cerebral blood flow using laser Doppler techniques demonstrated that the hemodynamic response coupled with neuronal activation seemed to be significantly suppressed in Alzheimer transgenic mice (Niwa et al, 2002a; Niwa et al, 2002b). The involvement of free radicals was validated when functional rescue of vascular deficits could be achieved by upregulating superoxide dismutase (Iadecola et al, 1999). Taken together, these data strongly suggest that problems in neurovascular coupling may play a central role in the pathology of progressive Alzheimer's disease.

The importance of the cerebrovasculature can also be interpreted in terms of amyloid clearance and kinetics between blood and brain. A rigorous review of these vital concept may be found in a review by Zlokovic (Zlokovic, 2005). Briefly, the amyloid content of brain is not static. Instead there is a dyamic equilibrium and exchange between parenchymal and vascular compartments. This process can be demonstrated by the passive immunization strategy, whereby blood-borne depletion of amyloid may help create gradients so that brain amyloid can be cleared as well. In aging and diseased brain, the receptors responsible for mediating amyloid transport may be altered; these include lipoprotein receptors and RAGE (Davis et al, 2004; Deane et al, 2004). Ultimately, increased deposition in concert with decreased clearance may accelerate the progression of disease and neurodegeneration (Silverberg et al, 2003; Donahue et al, 2006).

In terms of amyloid exposure per se, it may be useful

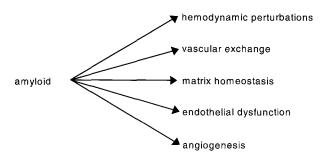


Fig. 4. Hypothesized vascular effects of amyloid. Besides a neurocentric focus, these interactions at the level of the entire neurovascular unit must also be addressed in the search for Alzheimer therapies. Based in part on fundamental concepts outlined by Zlokovic, Trends Neurosci 2005, and Iadecola, Nat Rev Neurosci 2004.

to note that vascular deposits may also be critical. The close association between Alzheimer's dementia and cerebral amyloid angiopathy may yet provide another model system for testing our neurovascular hypotheses (Greenberg, 2002; Zhang-Nunes et al, 2006). Amyloid can disrupt regulated blood flow (Niwa et al, 2002a; Niwa et al, 2002b), and higher levels can trigger apoptotic pathways in cerebral endothelial cells (Xu et al, 2001; Yin et al, 2002; Yin et al, 2005). From a therapeutic perspective, is it possible to hope that targeting endothelium might be more accessible than trying to salvage neurons lying behind the blood-brain barrier?

Once the degenerative process has been initiated, the brain should respond in an effort to restore homeostasis. Therefore, another hypothesis worth pursuing might involve reactions in brain cell turn-over. Baseline neurogenesis is perturbed in Alzheimer brains, although the pro-neurogenic versus anti-neurogenic actions of amyloid remain to be fully clarified (Greenberg & Jin, 2006). More recently, it has also been proposed that amyloid may have effects on angiogenesis (Vagnucci & Li, 2003; Zlokovic, 2005), so these responses in Alzheimer brains may have to be reinterpreted in the context of an integrated milieu of neurovascular remodeling.

It may be interesting to link these notions of neurovascular remodeling to the fact that ApoE isoforms constitute risk factors of Alzheimer's disease (Raber et al, 2004). Although the precise correlations are complex and it is difficult to establish causality, one might simplify the situation to say that ApoE4 isoforms increase the risk of "true" Alzheimer's disease, whereas ApoE2 isoforms may increase the risk of angiopathy and hemorrhage. Recently, it has been proposed that reactive astrocytes may help degrade amyloid, perhaps via MMPs (Deb et al, 2003; Wyss-Coray et al, 2003; Yan et al, 2006). Cell culture studies demonstrate that amyloid induces an upregulation of MMP-9 in cerebral endothelium and astrocytes (Deb et al, 2003; Lee et al, 2003). When astrocytes are exposed to amyloid together with ApoE, it was observed that ApoE4 isoforms tended to suppress MMP-9 levels (Guo et al, 2006). It is attractive to speculate and hypothesize that differential regulation of MMPs by various ApoE isoforms may in fact alter the balance between a beneficial degradation of amyloid versus too much proteolysis that leads to vascular disruption and hemorrhage.

In summary, whereas the majority of resources are now focused on the neurobiology of Alzheimer's disease, it is worthwhile noting that vascular effects may be extremely important as well (Fig. 4). The major risk factors for sporadic Alzheimer's may be mostly vascular in nature. Amyloid perturbs hemodynamic function and blood flow, and triggers oxidative stress and induces cell death in endothelial cells. Alterations in neurovascular transporters may disrupt the delicate balance between deposition and clearance in aging brain. And amyloid may influence neurogenesis and angiogenesis, in part via modulation of neurovascular proteases that regulate degradation and vessel integrity.

Conclusions and Future Directions

For perhaps far too long, the focus in neurological research was on the neurobiology of CNS disorders. However, numerous failures in clinical trials for "pure" neuroprotectants should now be re-interpreted in the context of the neurovascular unit. A prime example can be found in the use of tPA. Besides dissolving the offending clot, tPA may also trigger a broad spectrum of responses within all compartments of the neurovascular unit. These neurovascular responses must be considered in both acute injury as well as delayed remodeling. Similar arguments may be made for neurodegeneration. Many risk factors for Alzheimer's are linked to vascular function. In transgenic models of Alzheimer's, neurovascular coupling seems to be perturbed prior to neuronal dysfunction, and cerebral endothelial markers of oxidative stress and proteolytic pathology often emerge even before parenchymal plaques appear. Indeed, it is not just abnormal amyloid generation, but also abnormal amyloid clearance and neurovascular kinetics that may contribute to Alzheimer's disease. Ultimately, the endogenous brain response to stress and injury may be mediated by a coordinated coupling between neurogenesis and angiogenesis. Experimental models that probe these altered neurovascular phenomenon may now provide new opportunities for basic and translational research.

The cerebral endothelium may not just comprise inert tubes for blood flow. But instead, endothelium may also be active sources of trophic and signaling agents that subserve neuronal survival and function. Ultimately, we hope that investigations will dissect not just neurobiology, but instead the entire neurovascular unit. Is there a change in neurovascular matrix that mediates signaling between the vascular and neuronal compartments? How do alterations in neurovascular homeostasis affect neuronal function? And in turn, how can neuronal defects perturb vascular regulation? The neurovascular unit provides an integrated framework for hypothesis-testing of function and dysfunction. Based on these ideas, we hope that the coming years will reveal new targets and therapeutic approaches for brain injury, neurodegeneration, and recovery.

ACKNOWLEDGEMENT

Supported in part by a stipend from the Korean Society for Pharmacology, and grants from the NIH (R01-NS37073, R01-NS48422, R01-NS53560, R01-NS53548 and P01-NS 10828). The author sincerely thanks Dr. Seong-Ryong Lee from Keimyung University for helpful discussions and advice. The ideas outlined in this mini-review were based on concepts developed by many scientists in the field, and the author apologizes if not all key papers could be cited in the limited context of this short manuscript.

REFERENCES

- Abbott NJ, Ronnback L, Hansson E. Astrocyte-endothelial interactions at the blood-brain barrier. Nat Rev Neurosci 7: 41-53, 2006
- Allan S. The neurovascular unit and the key role of astrocytes in the regulation of cerebral blood flow. *Cerebrovasc Dis* 21: 137—138, 2006
- Alvarez-Sabin J, Delgado P, Abilleira S, Molina CA, Arenillas J, Ribo M, Santamaria E, Quintana M, Monasterio J, Montaner J. Temporal profile of mnatrix metalloproteinases and their inhibitors after spontaneous intracerebral hemorrhage: relationship to clinical and radiological outcome. Stroke 35: 1316-1322, 2004
- Arvidsson A, Collin T, Kirik D, Kokaia Z, Lindvall O. Neuronal replacement from endogenous precursors in the adult brain after stroke. Nat Med 8: 963-970, 2002
- Astrup J, Symon L, Branston NM, Lassen NA. Cortical evoked potential and extracellular K+ and H+ at crttical levels of brain ischemia. Stroke 8: 51-57, 1977
- Baranes D, Lederfein D, Huang YY, Chen M, Bailey C, Kandel E. Tissue plasminogen activator contributes to the late phase of LTP and to synaptic growth in the hippocampal mossy fiber pathway. Neuron 21: 813-825, 1998
- Benchenane K, Berezowski V, Ali C, Fernandez-Monreal M, Lopez-Atalaya JP, Brillault J, Chuquet J, Nouvelot A, MacKenzie ET, Bu G, Cecchelli R, Touzani O, Vivien D. Tissue-type plasminogen activator crosses the intact blood-brain barrier by low-density lipoprotein receptor-related protein-mediated transcytosis. Circulation 111: 2241-2249, 2005
- Benveniste H. The excitotoxin hypothesis in relation to cerebral ischemia. Cerebrovasc Brain Metab Rev 3: 213-220, 1991
- Bossy-Wetzel E, Schwarzenbacher R, Lipton SA. Molecular pathways to neurodegeneration. *Nat Med* 10 Suppl: S2-9, 2004
- Breteler MM. Vascular risk factors for Alzheimer's disease: an epidemiologic perspective. Neurobiol Aging 21: 153-160, 2000
- Buchan AM. Do NMDA antagonists protect against cerebral ischemia: are clinical trials warranted? Cerebrovasc Brain Metab Rev
 2: 1-26, 1990
- Carmeliet P. Blood vessels and nerves: common signals, pathways and diseases. Nat Rev Genet 4: 710-720, 2003
- Chen ZL, Strickland S. Neuronal death in the hippocampus is promoted by plasmin-catalyzed degradation of laminin. *Cell* 91: 917-925, 1997
- Davis J, Xu F, Deane R, Romanov G, Previti M, Zeigler K, Zlokovic BV, Van Nostrand WE. Early-onset and robust cerebral microvascular accumulation of amyloid beta-protein in transgenic mice expressing low levels of a vasculotropic Dutch/ Iowa mutant form of amyloid beta-protein precursor. *J Biol Chem* 279: 20296 20306, 2004
- De Keyser J, Sulter G, Luiten PG. Clinical trials with neuroprotective drugs in acute ischemic stroke: are we doing the right thing? *Trends Neurosci* 22: 535-540, 2000
- de la Torre JC. Alzheimer disease as a vascular disorder. Stroke 33: 1152-1162, 2002
- Deane R, Wu Z, Sagare A, Davis J, Yan S, Hamm K, Xu F, Parisi M, LaRue B, Hu H, Spijkers P, Guo H, Song X, Lenting PJ, Van Nostrand WE, Zlokovic BV. LRP/amyloid beta-peptide interaction mediates differential brain efflux of Abeta isoforms. Neuron 43: 333-344, 2004
- Deb S, Wenjun Zhang J, Gottschall PE. Beta-amyloid induces the production of active, matrix-degrading proteases in cultured rat astrocytes. *Brain Res* 970: 205–213, 2003
- Dijkhuizen RM, Singhal AB, Mandeville J, Wu O, Halpern EF, Finklestein SP, Rosen BR, Lo EH. Correlation between brain reorganization, ischemic damage, and neurologic status after transient focal cerebral ischemia in rats: a functional magnetic resonance imaging study. *J Neurosci* 23: 510-517, 2003
- Donahue JE, Flaherty SL, Johanson CE, Duncan JA 3rd, Silverberg GD, Miller MC, Tavares R, Yang W, Wu Q, Sabo E, Hovanesian V, Stopa EG. RAGE, LRP-1, and amyloid-beta protein in Alzheimer's disease. *Acta Neuropathol (Berl)*, 112: 405-415, 2006
- ECASS Study Group. Intravenous thrombolysis with recombinant tissue plasminogen activator in acute hemispheric stroke. JAMA

- 274: 1017-1025, 1995
- Eichmann A, Le Noble F, Autiero M, Carmeliet P. Guidance of vascular and neural network formation. Curr Opin Neurobiol 15: 108-115, 2005
- Garcia JH. Morphology of global cerebral ischemia. Crit Care Med 16: 979-987, 1988
- Gladstone DJ, Black SE, Hakim AM. Toward wisdom from failure: lessons from neuroprotective stroke trials. Stroke 33: 2123-2136, 2002
- Greenberg DA, Jin K. Neurodegeneration and neurogenesis: focus on Alzheimer's disease. Curr Alzheimer Res 3: 25-28, 2006
- Greenberg SM. Cerebral amyloid angiopathy and dementia: two amyloids are worse than one. Neurology 58: 1587-1588, 2002
- Gualadris A, Jones TE, Strickland S, Tsirka SE. Membrane deplorization induces calcium dependent secretion of tissue plasminogen activator. J Neurosci 16: 2220-2225, 1996
- Guo S, Wang S, Km WJ, Lee S, Froesch MP, Bacskai BJ, Greenberg SM, Lo EH. Effects of ApoE isoforms on beta-amyloid-induced matrix metalloproteinase in rat astrocytes. *Brain Res (in press)*, 2006
- Hacke W, Brott T, Caplan L, Meier D, Fieschi C, von Kummer R,
 Donnan G, Heiss WD, Wahlgren NG, Spranger M, Boysen G,
 Marler JR. Thrombolysis in acute ischemic stroke: controlled trials and clinical experience. Neurology 53 (Suppl 4): s3-s15, 1999
- Harukuni I, Bhardwaj A. Mechanisms of brain injury after global cerebral ischemia. Neurol Clin 24: 1-21, 2006
- Hawkins BT, Davis TP. The blood-brain barrier/neurovascular unit in health and disease. *Pharmacol Rev* 57: 173-185, 2005
- Hirose K, Chan PH. Blockade of glutamate excitotoxicity and its clinical applications. Neurochem Res 18: 479-483, 1993
- Hopewell JW, Cavanagh JB. Effects of X irradiation on the mitotic activity of the subependymal plate of rats. Br J Radiol 45: 461 465. 1972
- Iadecola C. Neurovascular regulation in the normal brain and in Alzheimer's disease. Nat Rev Neurosci 5: 347-360, 2004
- Iadecola C, Zhang F, Niwa K, Eckman C, Turner S, Fischer E, Younkin S, Borchelt DR, Hsiao KK, Carlson GA. SOD1 rescues cerebral endothelial dysfunction in mice overexpressing amyloid precursor protein. Nat Neurosci 2: 157-161, 1999
- Kaur J, Zhao Z, Klein GM, Lo EH, Buchan AM. The neurotoxicity of tissue plasminogen activator? J Cereb Blood Flow Metab 24: 945-963, 2004
- Kim Y, Huang J, Lee SR, Tejima E, Mandeville J, van Meer M, Dai G, Choi YW, Dijkhuizen RM, Lo EH, Rosen BR. Measurements of BOLD/CBV ratios show altered hemodynamics during stroke recovery. *J Cereb Blood Flow Metab* 25: 820-829, 2005
- Kim YR, van Meer MP, Mandeville JB, Tejima E, Dai G, Topalkara K, Qui J, Dijkhuizen RM, Moskowitz MA, Lo EH, Rosen BR. fMRI of delayed albumin treatment during stroke recovery in rats: implication for fast neuronal habituation in recovering brains. J Cereb Blood Flow Metab (in press), 2006
- Lapchak PA, Chapman DF, Zivin JĀ. Metalloproteinase inhibition reduces thrombolytic (Tissue plasminogen Activator)-induced hemorrhage after thromboembolic stroke. Stroke 31: 3034 3040, 2000
- Lee JM, Yin KJ, Hsin I, Chen S, Fryer JD, Holtzman DM, Hsu CY, Xu J. Matrix metalloproteinase-9 and spontaneous hemorrhage in an animal model of cerebral amyloid angiopathy. *Ann Neurol* 54: 379-382, 2003
- Lee SR, Kim HY, Rogowska J, Zhao BQ, Bhide P, Parent JM, Lo EH. Involvement of matrix metalloproteinase in neuroblast cell migration from the subventricular zone after stroke. *J Neurosci* 26: 3491-3495, 2006
- Leventhal C, Rafii S, Rafii D, Shahar A, Goldman SA. Endothelial trophic support of neuronal production and recruitment from the adult mammalian subependyma. *Mol Cell Neurosci* 13: 450 464. 1999
- Lewis PD. Radiosensitivity of the subependymal cell layer of the adult rat brain. Exp Neurol 20: 208-214, 1968
- Lipton SA, Rosenberg PA. Excitatory amino acids as a final common pathway for neurologic disorders. N Engl J Med 330: 613-622, 1994
- Liu J, Solway K, Messing RO, Sharp FR. Increased neurogenesis in the dentate gyrus after transient global ischemia in gerbils.

- J Neurosci 18: 7768-7778, 1998
- Lo EH, Broderick JP, Moskowitz MA. tPA and proteolysis in the neurovascular unit. Stroke 35: 354-356, 2004
- Mattson MP. Pathways towards and away from Alzheimer's disease. Nature 430: 631-639, 2004
- McGowan E, Eriksen J, Hutton M. A decade of modeling Alzheimer's disease in transgenic mice. *Trends Genet* 22: 281–289, 2006
- Montaner J, Alvarez-Sabin J, Molina CA, Angles A, Abilleira S, Arenillas J, Monasterio J. Matrix metalloproteinase expression is related to hemorrhagic transformation after cardioembolic stroke. Stroke 32: 2762-2767, 2001
- Montaner J, Molina CA, Monasterio J, Abilleira S, Arenillas JF, Ribo M, Quintana M, Alvarez-Sabin J. Matrix metalloproteinase-9 pretreatment level predicts intracranial hemorrhagic complications after thrombolysis in human stroke. Circulation 107: 598-603, 2003
- Nassar T, Akkawi S, Shina A, Haj-Yehia A, Bdeir K, Tarshis M, Heyman SN, Higazi AA. In vitro and in vivo effects of tPA and PAI-1 on blood vessel tone. Blood 103: 897-902, 2004
- Nicole O, Docagne F, Ali C, Margaill I, Carmeliet P, MacKenzie ET, Vivien D, Buisson A. The proteolytic activity of tissue plasminogen activator enhances NMDA receptor mediated signaling. Nature Med 7: 59-64, 2001
- NINDS rt-PA Stroke Study Group. Tissue plasminogen activator for acute ischemic stroke. New Engl J Med 333: 1581–1587, 1995
- Ning M, Furie KL, Koroshetz WJ, Lee H, Barron M, Lederer M, Wang X, Zhu M, Sorensen AG, Lo EH, Kelly PJ. Association between tPA therapy and raised early matrix metalloprotein-ase-9 in acute stroke. *Neurology* 66: 1550-1555, 2006
- Niwa K, Kazama K, Younkin SG, Carlson GA, Iadecola C. Alterations in cerebral blood flow and glucose utilization in mice over-expressing the amyloid precursor protein. Neurobiol Dis 9: 61–68, 2002a
- Niwa K, Kazama K, Younkin L, Younkin SG, Carlson GA, Iadecola C. Cerebrovascular autoregulation is profoundly impaired in mice overexpressing amyloid precursor protein. Am J Physiol 283: H315-H323, 2002b
- Palmer TD, Willhoite AR, Gage FH. Vascular niche for adult hippocampal neurogenesis. J Comp Neurol 425: 479-494, 2000
- Parent JM, Vexler ZS, Gong C, Derugin N, Ferriero DM. Rat forebrain neurogenesis and striatal neuron replacement after focal stroke. Ann Neurol 52: 802-813, 2002
- Park JA, Choi KS, Kim SY, Kim KW. Coordinated interaction of the vascular and nervous systems: from molecule- to cell-based approaches. Biochem Biophys Res Commun 311: 247-253, 2003
- Park L, Anrather J, Forster C, Kazama K, Carlson GA, Iadecola C. Abeta-induced vascular oxidative stress and attenuation of functional hyperemia in mouse somatosensory cortex. J Cereb Blood Flow Metab 24: 334-342, 2004
- Paschen W. Glutamate excitotoxicity in transient global cerebral ischemia. Acta Neurobiol Exp (Wars) 56: 313-322, 1996
- Privat A, Leblond CP. The subependymal layer and neighboring region in the brain o the young rat. *J Comp Neurol* 146: 277 302, 1972
- Raber J, Huang Y, Ashford JW. ApoE genotype accounts for the vast majority of AD risk and AD pathology. Neurobiol Aging 25: 641-650, 2004
- Reynolds BA, Weiss S. Generation of neurons and astrocytes from isolated cells of the adult mammalian central nervous system. Science 255: 1707-1710, 1992
- Rogove A, Siao CJ, Keyt B, Strickland S, Tsirka S. Activation of microglia reveals a non-proteolytic cytokine function for tissue plasminogen activator in the central nervous system. J Cell Sci 112: 4007-4016, 1999
- Rosell A, Ortega-Aznar A, Alvarez-Sabin J, Fernandez-Cadenas I, Ribo M, Molina CA, Lo EH, Montaner J. Increased brain expression of matrix metalloproteinase-9 after ischemic and hemorrhagic human stroke. Stroke 37: 1399–1406, 2006
- Ryder EF, Snyder EY, Cepko CL. Establishment and characterization of multipotent neural cell lines using retrovirus vector-mediated oncogene transfer. J Neurobiol 21: 356-375, 1990
- Selkoe DJ. Translating cell biology into the rapeutic advances in Alzheimer's disease. *Nature* 399: A23—A31, 1999
- Shen Q, Goderie S, Jin L, Karanth N, Sun Y, Abramova N, Vincent

- P, Pumiglia K, Temple S. Endothelial cells stimulate self-renewal and expand neurogenesis of neural stem cells. Science $304:\ 1338-1340,\ 2004$
- Silverberg GD, Mayo M, Saul T, Rubenstein E, McGuire D. Alzheimer's disease, normal-pressure hydrocephalus, and senescent changes in CSF circulatory physiology: a hypothesis. *Lancet Neurol* 2: 506-511, 2003
- Sumii T, Lo EH. Involvement of matrix metalloproteinase in thrombolysis-associated hemorrhagic transformation after embolic focal ischemia in rats. Stroke 33: 831-836, 2002
- Tejima E, Zhao BQ, Tsuji K, Rosell A, van Leyen K, Gonzalez RG, Montaner J, Wang X, Lo EH. Astrocytic induction of matrix metalloproteinase-9 and edema in brain hemorrhage. J Cereb Blood Flow Metab, 2006
- Tsirka SE, Gualandris A, Amaral DG, Strickland S. Excitotoxin induced neuronal degenaration and seizures are mediated by TPA. *Nature* 377: 340-344, 1995
- Tsirka SE, Rogove AD, Strickland S. Neuronal cell death and TPA. Nature 384: 123-124, 1996
- Tsuji K, Aoki T, Tejima E, Arai K, Lee SR, Atochin DN, Huang PL, Wang X, Montaner J, Lo EH. Tissue plasminogen activator promotes matrix metalloproteinase-9 upregulation after focal cerebral ischemia. Stroke 36: 1954-1959, 2005
- Vagnucci AH Jr, Li WW. Alzheimer's disease and angiogenesis. Lancet 361: 605-608, 2003
- Wang J, Rogove AD, Tsirka AE, Tsirka SE. Protective role of tuftsin fragment 1-3 in an animal model of intracerebral hemorrhage. Ann Neurol 54: 655-664, 2003a
- Wang X, Arai K, Lee SR, Lee S, Tsuji K, Rebeck GW, Lo EH. Lipoprotein receptor-mediated induction of matrix metalloproteinase-9 in human cerebral endothelial cells. Nature Med 9: 1313-1319, 2003b
- Wang YF, Tsirka SE, Strickland S, Steig PE, Soriano SG, Lipton SA. TPA increases neuronal damage after focal cerebral ischemia in wild type and TPA-deficient mice. Nature Med 4: 228– 231, 1998
- Warach S, Latour LL. Evidence of reperfusion injury, exacerbated by thrombolytic therapy, in human focal brain ischemia using a novel imaging marker of early blood-brain barrier disruption. Stroke 35: 2659-2661, 2004
- Ward NL, Lamanna JC. The neurovascular unit and its growth factors: coordinated response in the vascular and nervous systems. Neurol Res 26: 870-883, 2004
- Wyss-Coray T, Loike JD, Brionne TC, Lu E, Anankov R, Yan F, Silverstein SC, Husemann J. Adult mouse astrocytes degrade amyloid-beta in vitro and in situ. *Nat Med* 9: 453-457, 2003
- Xu J, Chen S, Ku G, Ahmed SH, Xu J, Chen H, Hsu CY. Amyloid beta peptide-induced cerebral endothelial cell death involves mitochondrial dysfunction and caspase activation. J Cereb Blood Flow Metab 21: 702-710, 2001
- Yan P, Hu X, Song H, Yin K, Bateman RJ, Cirrito JR, Xiao Q, Hsu FF, Turk JW, Xu J, Hsu CY, Holtzman DM, Lee JM. Matrix metalloproteinase-9 degrades amyloid-beta fibrils in vitro and compact plaques in situ. *J Biol Chem* 281: 24566-24574, 2006
- Yin KJ, Lee JM, Chen SD, Xu J, Hsu CY. Amyloid-beta induces Smac release via AP-1/Bim activation in cerebral endothelial cells. J Neurosci 22: 9764-9770, 2002
- Yin KJ, Lee JM, Chen H, Xu J, Hsu CY. Abeta25-35 alters Akt activity, resulting in Bad translocation and mitochondrial dysfunction in cerebrovascular endothelial cells. J Cereb Blood Flow Metab 25: 1445-1455, 2005
- Zhang RL, Zhang ZG, Zhang L, Chopp M. Proliferation and differentiation of progenitor cells in the cortex and the subventricular zone in the adult rat after focal cerebral ischemia. Neuroscience 105: 33-41, 2001
- Zhang-Nunes SX, Maat-Schieman ML, van Duinen SG, Roos RA, Frosch MP, Greenberg SM. The cerebral beta-amyloid angiopathies: hereditary and sporadic. *Brain Pathol* 16: 30-39, 2006
- Zhao BQ, Wang S, Kim HY, Storrie H, Rosen BR, Mooney DJ, Wang X, Lo EH. Role of matrix metalloproteinases in delayed cortical responses after stroke. Nat Med 12: 441-445, 2006
- Zlokovic BV. Neurovascular mechanisms of Alzheimer's neurodegeneration. Trends Neurosci 28: 202-208, 2005