Antifungal Activity of Lactic Acid Bacteria Isolated from Kimchi Against *Aspergillus fumigatus*

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More than 120 isolates of lactic acid bacteria obtained from Kimchi was screened for antifungal activity against Aspergillus fumigatus. Approximately 10% of the isolates showed inhibitory activity and only 4.16% (five isolates) exhibited strong activity against the indicator fungus A. fumigatus. The five isolates showed a wide rang of antifungal activity against A. flavus, Fusarium moniliforme, Penicillium commune, and Rhizopus oryzae. They were identified by 16S rDNA sequencing as Lactobacillus cruvatus, L. lactis subsp. lactis, L. casei, L. pentosus, and L. sakei. The effect of Lactobacillus on mycelial growth and fungal biomass as well as its ability to produce toxic compounds were determined. The results indicate that the three species, Lactobacillus casei, L. lactis subsp. lactis, and L. pentosus, are active against A. fumigatus.

KEYWORDS: Antifungal activity, Aspergillus fumigatus, Lactic acid bacteria

In storage of foods and feed stocks, a major problem is spoilage and poisoning by fungi, Aspergillus, Fusarium, Penicillium (Nickelson and Jakobson, 1997) that cause great economic losses worldwide (Magnusson et al., 2003). Furthermore, these fungi produce allergenic spores and mycotoxins that cause serious potential health hazards (Nielson and Rios, 2000). Adequate control measures to prevent fungal growth in grains, foodstuffs, foods and feed production and storage are primary importance to avoid contaminating foods and minimizing public health hazards. During the last few years, there has been a growing interest in biopreservation, i.e., the application of microorganisms and/or their metabolites to prevent spoilage and to extend the preservation time of foods (Stiles, 1996). Lactic acid bacteria are becoming interest in biopreservation of food and feed because of the production of antimicrobial compounds such as lactic acid, acetic acid, and bacteriocin produced (Lavermicocca et al., 2000; Lindgern and Dobrogosz, 1990). While many studies focused on antibacterial activities of lactic acid bacteria (Dodd and Gasson, 1994; Lindgren and Dobrogosz, 1990; Stiles, 1996), there are comparatively few reports on antifungal effects. El-Gendy and Marth (1981) reported that Lactobacillus casei inhibited the growth and the aflatoxin production of Aspergillus parasiticus. Early research (Lavermicocca et al., 2000) demonstrated that the antifungal compounds such as phenyllactic acid and 4-hydrophenyllactic acid were produced by Lactobacillus plantarum. In addition, bacteriocin-like substances and other low molecular mass compounds were produced by L. pentosus and L. coryniformis (Okkers et al., 1999; Magnusson et al., 2003).

The objective of this study were to screen and identify

lactic acid bacteria, which effectively inhibited the growth of the fungal species including *Aspergillus fumigatus*, *A. flavus*, *Fusarium moniliforme*, *Penicillium commune* and *Rhizopus oryzae*, and to determine the antifungal spectra of the five isolates of lactic acid bacteria.

Materials and Methods

Isolation of Lactobacillus from Kimchi. Household Kimchi were collected from various sources. The broth of Kimchi was diluted at 1/10⁵ to 1/10⁶. To isolate lactic acid bacteria, the diluted samples were spread on Man-Rogasa-Sharpe (MRS) agar medium containing 2% CaCO₃ and allowed to grow at 37°C for 48 h in anaerobic jars. The plates were then overlaid with malt extract soft agar $(0.05\% \text{ malt extract and } 1\% \text{ agar}) 1 \times 10^6 \text{ spores of}$ Aspergillus fumigatus per ml. After aerobic incubation at 37°C for 48 h, colonies showing inhibition zone were isolated. The isolated strains were transferred onto the modified Lactobacillus selection (Yuki et al., 1999; LBS; Becton Dickinson and Company, Cockeysville, MD, USA) medium containing glucose instead of lactitol as a carbon source and vancomycin as a selective antibiotic (Table 1) and the plates were incubated at 37°C for 48 h in order to isolate Lactobacillus strains from lactic acid bacteria. The isolated Lactobacillus strains were stored at -70°C in MRS broth containing 50% glycerol.

Preparation of spore suspension of *A. fumigatus. A. fumigatus* was kept on potato-dextrose agar (PDA, Difco) plates at 30°C. Spores of fungus were harvested from 10-day-old PDA plate cultures in sterile water containing 0.05% Tween 80, and loosened by 15 min shaking. The suspension was adjusted to 1×10^6 spores/ml.

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Table 1. The composition of the modified LBS agar medium

Trypticase peptone	10 g
Yeast extract	5 g
KH ₂ PO ₄	6 g
tri-Ammonium Citrate	2 g
Tween 80	1 g
Sbdium Acetate Hydrate (3H ₂ O)	25 g
Salts solution	5 <i>ml</i>
Glucose	20 g
Agar	15 g
Vancomycin ^(a)	$10 \ \mu \text{g/ml}$
Distilled water	$1000 \ ml$
Salts solution	
MgSO ₄ ·7H ₂ O	11.5g
FeSO ₄ ·7H ₂ O	0.68 g
MnSO ₄ ·7H ₂ O	3.4 g
Distilled water	100 ml

*Solution of vancomycin is sterilized by filtration and added after autoclaving the medium at 121°C for 15 min.

Influence of Lactobacillus on fungal biomass production in liquid MRS medium. Fungal cultures containing 0.2 ml working culture or spore suspension and 15 ml of MRS in 20×150 mm culture tubes were incubated at 30°C for various times (12, 24, 48 and 72 h incubation). The Lactobacillus and fungus were associated and the mixed culture incubation was continued for 2 weeks at 30°C. Subsequently the mycelium was separated from the medium by filtration on filter paper (No 1. Whatmann), thoroughly washed with ethyl acetate, dried in an oven at 95°C until constant weight, and the dry mass was determined. Cultures of the fungus grown without other microorganisms were used as the control.

Determination of microbial interaction on solid PDA medium. Cultures containing 0.2 ml working culture and 15 ml of PDA medium were transferred into petridish with 9 cm in diameter and incubated at 30°C. After 12, 24, 48 and 72 h incubation, PDA disks with 5 mm-diameter of three-days-old mycelium of the pathogen were plated in the center of the petridish and incubated in the dark at 30°C. Linear growth was estimated by measuring the diameter of the colonies for 2 weeks.

Determination of antifungal spectra. More than 120 isolates were screened against the indicator fungus A. fumigatus, and among these, five isolates were selected for this study. The dual culture overlay assay (Magnusson and Schnürer, 2001) described above in was used to estimate inhibitory activities of the five selected Lactobacillus against four fungi, A. flavus, Fusarium moniliforme, Penicillium commune and Rhizopus oryzae at 30°C. Bacteria were inoculated in 2-cm line on MRS agar plates and incubated at 30°C for 48 h in anaerobic jars. The plates were then overlaid with malt extract soft agar

(0.05% malt extract and 1% agar) containing 1×10^6 spores of each fungus per ml. After 48 h of aerobic incubation at 30°C, the inhibition zone was measured. The inhibition was graded by involving the inhibited growth area per inoculation streak to total area of the petridish (Magnusson $et\ al.$, 2003). The inhibition area was also related to the variation in length of the bacterial streak. The following scale was used: –, no visible inhibition; +, no fungal growth on $0.1\sim3.0\%$ of plate area/bacterial streak; +++, no fungal growth on >8% of plate area/bacterial streak. Inhibition test were carried out in duplicate.

Identification of lactic acid bacteria. All strains were identified by sequence analysis of 16S rDNA. Bacterial DNA was isolated from bacteria grown in MRS broth using DNeasy Tissue Kit (Qiagen). 16S rDNA was amplified by PCR using two primers according to Stackebrandt and Liesack (1993), 5'-GAGTTTGATCCTGGCTCAG-3' and 5'-AGAAAGGAGGTGATCCAGCC-3'. A PCR was run for 35 cycles in a DNA thermal cycler, Genetic analyzer 377 (Perkin-Elmer, Boston, USA), employing the thermal profile according to Yoon et al. (1997). The 16S rDNA sequences of the five isolates of Lactobacillus were aligned using CLUSTAL W software (Nigam et al., 2000). The evolutionary distance matrices were calculated with the DNADIST program within the PHYLIP package (Felsenstein, 1993). The sequence of representative species of the genus Lactobacillus and related taxa were cited using the GenBank Database. The values of 16S rDNA similarity were calculated from the alignment, while the evolutionary distances were calculated using a Kimura two-parameter correction. A phylogenetic tree was constructed using the neighbor-joining method (Saitou and Nei, 1987) based on the calculated distance matrix.

Results and Discussion

Isolation of Lactobacillus spp. One hundred twenty lactic acid bacteria were isolated from various Kimchi, and approximately 10% of the isolated lactic acid bacteria showed antifungal activity against A. fumigatus (data not shown). About 4.16% of the total number of the isolated bacteria had strong activity (+++). For this study, the five isolates, KC-116, KC-304, KC-324, KC-817, and KC-1004, of Lactobacillus with strong (+++) activity were selected. Some researcher (Magnusson et al., 2003; Stiles et al., 1999) reported that only 4~5% of the isolated microorganism showed antagonistic activity against fungi. Antimicrobial substances especially with antibacterial effect from lactic acid bacteria have been well studied (Harris et al., 1989; Kim et al., 1998, 2003; Klaenhammer, 1988; Lee et al., 1999; Niku-Paavola et al., 1999). However, there are relatively few reports on antifungal activity of Lactobacillus spp. (El-Gendy and Marth, 1981; Gourama

Table 2. Inhibitory effect of Lactobacillus isolates against pathogenic fungi

Strain	Species	A. fumigatus ^a	A. flavus	F. moniliforme	P. commune	R. oryzae
KC-116	Lactobacillus curvatus	+++	+++	++	++	+++
KC-304	Lactobacillus lactis subsp. lactis	+++	++	+++	++	+++
KC-324	Lactobacillus casei	+++	+++	++	+	++
KC-817	Lactobacillus pentosus	+++	++	++	++	+
KC-1004	Lactobacillus sakei	+++	++	+++	+++	+++

*All isolates were initially screened against *A. fumigatus*, with degree of inhibition that showed at the initial activity. Symbols are: (+), no fungal growth on 0.1~3.0% of plate area/bacterial streak; (+++), no fungal growth 3~8% of plate area/bacterial streak; (+++), no fungal growth on > 8% of plate area/bacterial streak.

and Bullerman, 1995; Lavermicocca et al., 2000).

Identification of *Lactobacillus* **spp.** Approximately 1,000 bp of the 16S rDNA sequence were determined for the five isolates selected in this study. The sequences derived from the five isolates were used to search databases from the highest similarity rank to determine the species identify of the five isolates in GenBank. Each isolate was identified based on their sequence homology of 16S rDNA. The sequences of the isolates, KC-116, KC-304, KC-324, KC-817 and KC-1004, showed higher homology to the sequence of Lactobacillus species; 99.5% to Lactobacillus curvatus, 99.8% to L. lactis subsp. lactis, 99.7% to L. casei, 99.5% to L. pentosus, and 99.6% to L. sakei, respectively. It suggests that the five isolates belong to the genus Lactobacillus. According to the reports on the correlation between the DNA-DNA homology and 16S rDNA sequence homology (Guha and Jaffe. 1996; Kobayashi and Ritmann 1982), strains reveal DNA-DNA homology higher than 70% and show sequence homology higher than 99.5% when strains were identified. Consequently, the five isolates were identified as *Lactobacillus*, and tentatively named as *Lactobacillus curvatus* KC-116, *L. lactis* subsp. *lactis* KC-304, *L. casei* KC-324, *L. pentosus* KC-817, and *L. sakei* KC-1004, respectively.

Antifungal spectra. The five *Lactobacillus* species inhibited the growth of the *A. fumigatus* in dual culture overlay tests as described by Magnusson and Schnürer (2001), although the inhibition zones varied with the lactic acid bacteria as well as with the fungal strain tested. All the five isolates of *Lactobacillus* inhibited the growth of *A. flavus*, *F. moniliforme*, *P. commune*, and *R. oryzae* (Table 2). The inhibition zone was reduced when the culture supernatants were used (data not shown), which could have been due to a low concentration of antifungal compounds. However, the inhibition of the growth of all the tested fungal species was quite pronounced with the strains KC-116, KC-304, and KC-1004.

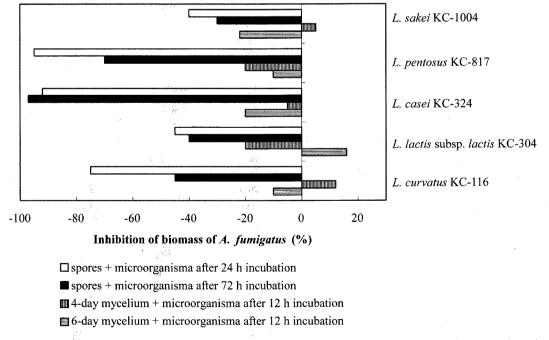


Fig. 1. Inhibitory effects of lactic acid bacteria against mycelial growth of pathogenic fungus, *A. fumigatus*, in paired cultures. Positive values indicate increase of fungal biomass. Individual culture of *A. fumigatus* in MRS medium is used as control.

Influence of Lactobacillus on fungal biomass production in liquid MRS medium. The influence of the five isolated lactic acid bacteria in different growth phases on both spores and mycelia of A. fumigatus was studied. The fungal growth on liquid MRS medium in paired cultures. with lactic acid bacteria expressed as the percentage of decrease or increase of fungal biomass in comparison to the control (individual culture of fungus without bacteria) is presented Fig. 1. The biomass increase after 2 weeks of paired cultures was lower than that of the control in all the samples, when the fungal spores were inoculated into the 24-h old lactic acid bacteria cultures (Fig. 1). The decreasing percentages in paired cultures were 75% in L. curvatus KC-116, 45% in L. lactis subsp. lactis KC-304, 92% in L. casei KC-324, 95% in L. pentosus KC-817, and 40% in L. sakei KC-1004. When the fungal spores were inoculated into the 72-h cultured lactic acid bacteria, the decrease ranges of mycelial biomass were 30~ 97% (Fig. 1). Exactly, the reducing fungal growth was 97% in the case of L. casei KC-324 and 30% in L. sakei KC-1004. The aflatoxin concentration was assayed in post-culture medium (Kumeda et al., 2003). There is no toxin of that kind present with accompanying mycelial biomass decreases in the culture in which fungal spores were inoculated in 24-h old L. curvatus KC-116, L. lactis subsp. lactis KC-304, L. casei KC-324, L. pentosus KC-817, L. sakei KC-1004 cultures as well as in 72-h old all the Lactobacillus cultures (data not shown). Coallier-Ascah and Idziak (1985) reported that the aflatoxins released from A. flavus in the culture broth were decreased in the presence of L. lactis. The bacterium degraded the toxic compounds by inhibitors such as lactic acid and nisin produced and subsequently released into medium before the toxins releasing from fungal body. The mycelial biomass of 4-day and 6-day old fungus paired with 12-h old bacterial cultures (Fig. 1) decreased to 10~20% with the exception of L. lactis subsp. lactis KC-304, L. curvatus KC-1164, and L. sakei KC-1004. These lactic acid bacteria paired culture with A. fumigatus somewhat stimulated the fungal mycelial growth in 4-day or 6-day old fungal mycelia cultures.

Determination of microbial interaction on solid PDA medium. The study on the linear growth of 3-days-old fungal mycelia in the paired culture with 3-days-old lactic acid bacteria in PDA medium was shown in Fig 2. The lowest increase of fungal biomass was shown at the presence of *L. sakei* KC-1004 and *L. casei* KC-324. The activity of these bacteria against the fungus *A. fumigatus* was dependent on their growth phase (Figs. 2 and 3). All the investigated microorganisms in paired culture showed the capacity for fungal mycelial growth inhibition in liquid MRS and solid PDA media. Cabo *et al.* (2002) reported that antifungal activity of lactic acid bacteria is due to a synergistic effect of lactic acid produced. After

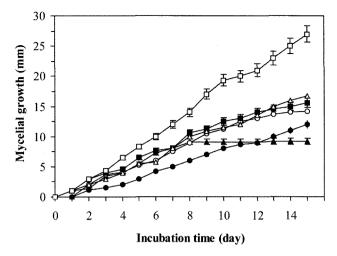


Fig. 2. Mycelial growth of *A. fumigatus* on PDA in mixed-culture with *L. sakei* KC-1004. Symbols: (□), Control; (▲) 6 h culture; (○), 12 h culture; (■), 24 h culture; (△) 48 h culture and (●) 72 h culture of *L. sakei* KC-1004.

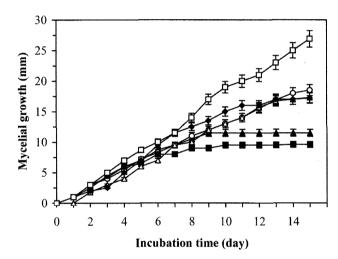


Fig. 3. Mycelial growth of *A. fumigatus* on PDA in mixed-culture with *L. casei* KC-304. Symbols: (□), Control; (♠), 6h culture; (○), 12h culture; (■), 24h culture; (△), 48h culture and (●), 72h culture of *L. casei* KC-324.

HPCL analysis for supernatants of the lactic acid bacteria showing antifungal activities, Magnusson *et al.* (2003) found that the production of lactic acid was the same or even higher in the tested isolates with non-antifungal activity. However, the HPCL analysis of lactic acid in the culture supernatants from these lactic acid bacterial strains gave on explanation for the varying degrees of inhibition of fungi. This varying activity is due to the production of other antifungal compounds. Ström *et al.* (2002) and Magnusson *et al.* (2003) purified three antifungal substances, cyclo(_L-Phe-_L-Pro), cyclo(_L-Phe-*trans*-4-OH-_L-Pro) and phenylacetic acid, from *L. plantarum*, *L. pentosaceus*, and *L. sakei*, respectively. This indicates that other

antifungal compounds are widely distributed among different species of lactic acid bacteria. In this study, the five *Lactobacillus* strains isolated from Kimchi exhibited antifungal activity against several fungi and some different compounds may cause inhibitory activity. Further investigations on the nature of inhibiting compounds and their mechanisms will be able to provide a great potential for the control of *A. fumigatus*.

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References

- Cabo, M. L., Braber, A. F. and Könrrad, P. 2002. Apparent antifungal activity of several lactic acid bacteria against *Penicillium discolor* is due to acetic acid in the medium. *J. Food Protect.* 65: 1309-1316.
- Coallier-Ascah, J. and Idziak, E. S. 1985. Interaction between Streptococcus lactis and Aspergillus flavus on production of aflatoxin. Appl. Environ. Microbiol. 49: 163-167.
- Dodd, H. M. and Gasson, M. J. 1994. Bacteriocins of lactic acid bacteria.
 Pp. 211-251. *In*: Gasson, M. J. and De Vos, W. M. Eds. Genetics and Biotechnology of Lactic Acid Bacteria.
 Blackie Academic and Professional London, London.
- El-Gendy, S. M. and Marth, E. H. 1981. Growth and aflatoxin production by *Aspergillus parasticus* in the presence of *Lactobacillus casei*. *J. Food Protect*. **44**: 211-212
- Felsenstein, J. 1993. PHYLIP: Phylogenetic Inference Package. Version 3.5. Seattle, University of Washington, Washington, USA.
- Gourama, H. and Bullerman, L. B. 1995. Inhibition of growth and aflatoxin production of *Aspergillus flavus* by *Lactobacillus* species. *J. Food Prot.* **58**: 1249-1256.
- Guha, S. and Jaffe, P. R. 1996. Biodegradation kinetics of phenanthrene partitioned into the micellar phase of nonionic surfactants. *Environ. Sci. Technol.* 30: 605-611.
- Harris, L. J., Daeschel, M. A., Stiles, M. E. and Klaenhammuer, T. R. 1989. Antimicrobiol activity of lactic acid bacteria against *Listeria monocytogenes*. J. Food Protect. 52: 384-387.
- Kim, H. T., Park, J. Y., Lee, G. G. and Kim, J. H. 2003. Isolation of a bacteriocin-producing *Lactobacillus plantarum* strain from Kimchi. *Food Sci. Biotechnol.* 12: 166-170.
- Kim, S. K., Lee, E. J., Park, K. Y. and Jun, H. K. 1998. Bacteriocin produced by *Lactobacillus curvatus* SE1 isolated from Kimchi. *J. Microbiol. Biotechnol.* 8: 588-594.
- Klaenhammer, T. R. 1988. Bactericins of lactic acid bacteria. Biochimie 70: 337-349.
- Kobayashi, H. and Ritmann, B. E. 1982. Microbial removal of hazardous organic compounds. Environ. Sci. Technol. 16: 170-183.
- Kumeda, Y., Asao, T., Takahashi, H. and Ichinoe, M. 2003. High prevalence of B and G aflatoxin-producing fungi in sugarcane field soil in Japan: heteroduplex panel analysis identifies a new genotype within *Aspergillus* section *Flavi* and *Aspergillus* nomius. FEMS Microbiol Ecol. 45: 229-238.
- Lavermicocca, P., Valerio, F., Evidente, A., Lazzaroni, S., Corsetti, A. and Gobetti, M. 2000. Purification and characterization of novel antifungal compounds from the sourdough

- Lactobacillus plantarum strain 21B. Appl. Environ. Microbiol. 66: 4084-4090.
- Lee, H. J., Park, C. S., Joo, Y. J., Kim, S. H., Yoon, J. H., Park, Y. H., Hwang, I. K., Ahn, J. S. and Mheen, T. I. 1999. Identification and characterization of bacteriocin-producing lactic acid bacteria isolated from Kimchi. *J. Microbiol. Biotechnol.* 9: 282-291.
- Lindgren, S. E. and Dobrogosz, W. J. 1990. Antagonistic activities of lactic acid bacteria in food and feed fermentations. FEMS Microbiol. Rev. 87: 149-164.
- Magnusson, J. and Schnürer, J. 2001. Lactobacillus coryniformis subsp. coryniformis strain SI3 produces a broad-spectrum proteinaceous antifungal compound. Appl. Environ. Microbiol. 67: 1-5.
- Ström, K., Roos, S., Sjögren, J. and Schnürer, J. 2003. Broad and complex antifungal activity among environmental isolates of lactic acid bacteria. *FEMS Microbiol. Lett.* 219: 129-135.
- Nielson, P. V. and Rios, R. 2000. Inhibition of fungal growth on bread by volatile components from species and herbs, and the possible application in active package, with special emphasis on mustard essential oil. *Int. J. Food. Microbiol.* **60**: 219-229.
- Nigam, P., Armour, G., Banat, I. M., Singh, D. and Marchant, R. 2000. Physical removal of textile dyes and solid-state fermentation of dye-adsorbed agricultural residues. *Bioresour. Tech*nol. 72: 219-226.
- Nikelson, L. and Kakobson, M. 1997. Quantitative risk analysis of aflatoxin toxicity for the consumers of "KenKey"-a fermented maize product. *Food Control* 3: 149-159.
- Niku-Paavola, M. L., Laitila, A., Mattila-Sandholm, T. and Haikara, A. 1999. New types of antimicrobial compounds produced by *Lactobacillus plantarum*. J. Appl. Microbiol. 86: 29-35.
- Okkers, D. J., Dicks, L. M. T., Silvester, M., Joubert, J. J. and Odendaal, H. J. 1999. Characterization of pentocin TV35b acteriocin-like peptide isolate from *Lactobacillus pentosus* with fungistic effect on *Candida albicans*. *J. Appl. Microbiol.* 87: 726-734.
- Pitt, J. I. and. Hocking, A. D. 1999. Fungi and food spoilage. Chapman & Hall, New York, N.Y.
- Saitou, N. and Nei, M. 1987. The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Mol. Biol. Evol.* 4: 406-425.
- Stiles, J., Plockova, M., Toth, V. and Chumchalova, V. 1999. Inhibition of *Fusarium* sp. DMF by *Lactobacillus* strains grown in MRS and Elliker broth. *Adv. Food Sci.* **21**: 117-121.
- Stiles, M. E. 1996. Biopreservation by lactic acid bacteria. *Antonie van Leeuwenhock* **70**: 331-345.
- Ström, K., Sjögren, J., Broberg, A. and Schnürer, J. 2002. *Lactoba-cillus plantarum* MiLAB 393 produces the antifungal cyclic dipeptides cyclo(_t-Phe-_t-Pro), and cyclo(_t-Phe-*trans*-4-OH-_t-Pro) and phenylacetic acid. *Appl. Environ. Microbiol.* **68**: 4322-4327.
- Yoon, J.-H., Lee, S.-T., Kim, S.-B., Kim, W.-Y., Goodfellow, M. and Park, Y.-H. 1997. Restriction fragment length polymorphisms analysis of PCR-amplified 16S ribosomal DNA for rapid identification of *Saccharomonospora* strains. *Int. J. Syst. Bacteriol.* 47: 111-114.
- Yuki, N., Watanabe, K., Mike, A., Togami, Y., Tanaka, R., Ohwaki, M. and Morotomi, M. 1999. Survival of a probiotic, *Lactobacillus casei* strain Shirota, in the gastrointestinal tract: Selective isolation from faces and identification using monoclonal antibodies. *Int. J. Food. Microbiol.* 48: 51-57.