

Solvatochromic Fluorescence Behavior of 8-Aminoquinoline-Benzothiazole: A Sensitive Probe for Water Composition in Binary Aqueous Solutions

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Received August 10, 2004

Solvatochromic fluorescence behavior of 8-aminoquinoline based benzothiazole derivative in varying solvent systems has been investigated. Benzothiazole appended 8-aminoquinoline **3** showed distinctive fluorescence color changes depending upon the solvent polarities and the fluorescence color changes occurred over relatively wide span in visible region from 486 nm to 598 nm which can be detected with naked eye. Compound **3** also exhibited significant spectral shifts in λ_{em} as a function of water composition in binary aqueous solvent systems. The changes are due to the specific interaction of **3** by hydrogen bonding with water as well as general solvent effect. The observed solvatochromic fluorescence characteristics of **3** could be used as a new probe for the micro-environmental polarity changes as well as a sensitive sensor for the determination of water composition in binary aqueous solutions.

Key Words : Fluorescence solvatochromism, 8-Aminoquinoline, Solvent polarity, Water composition, Binary aqueous solution

Introduction

In recent years considerable efforts have been given to the design and synthesis of functional molecules that could serve as sensitive sensors for the analytical detection of chemically and biologically important ionic species.¹ For this purpose, the advantages of fluorescence signaling in high selectivity and sensitivity have encouraged the development of a variety of interesting and practically usable fluorescence probes.² Along with intense research efforts for the development of sensors towards ionic species, solvatochromic or solvatofluorochromic methods for the characterization of micro-environment polarity have also been attracted much interest.³⁻⁵ The polarity concept can be applied for the analysis of the nature of important local sites in many of macromolecular systems including synthetic polymers,⁶ proteins,⁷ and nucleic acids.⁸ On the other hand, application of solvatochromic methods to mixed solvent systems, especially to mixed aqueous solutions,^{9,10} is particularly more useful since aqueous solvent systems are widely used in reverse phase liquid chromatography and capillary electrophoresis.¹¹

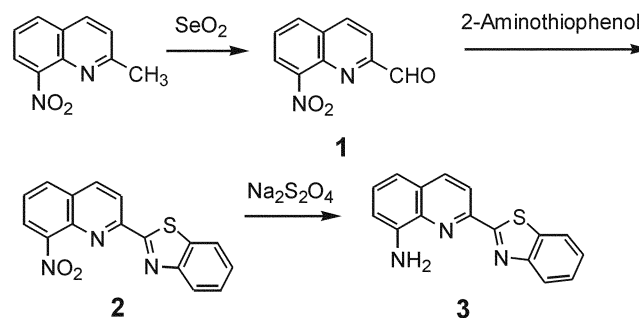
A large number of quinoline derivatives have been synthesized continually because the quinoline moiety has well-defined and attractive ionophoric properties toward a variety of important metal ions.¹² We have reported that the 8-aminoquinoline derivative **3** having appended benzothiazole function has Hg²⁺-selective fluoroionophoric behavior.¹³ During the search for the optimized condition of the Hg²⁺-selective fluorogenic behavior of **3** we have observed a significant dependence of fluorescence behavior upon the polarity of the employed solvents. In this paper, we report that the 8-aminoquinoline derivative having appended benzothiazole function has pronounced solvatochromic fluorescence behavior in aqueous solvent systems. The

compound can be used as a sensitive probe for measuring polarity of the micro-environments in chemical and biological systems as well as for the assessment of the water composition in binary aqueous solutions.

Results and Discussion

The desired compound, benzothiazole derivative of 8-aminoquinoline **3**, was synthesized in three steps¹⁴⁻¹⁶ from 8-nitro-2-methylquinoline as preliminarily reported earlier (Scheme 1).¹³

Preliminary absorption properties of **3** were investigated by UV-Vis spectroscopy. In acetonitrile solution, compound **3** (2.0×10^{-5} M) showed an intense absorption band (λ_{max}) at 310 nm. The absorption spectra of **3** in other solvents of varying polarity were similar in appearance with no appreciable characteristic changes. On the other hand, the ionophore **3** (2.0×10^{-5} M) showed an intense bright yellow fluorescence around 580 nm ($\lambda_{ex} = 310$ nm) in acetonitrile and significant changes in fluorescence spectra were observed in varying solvent systems. Therefore, we have measured fluorescence spectra of **3** in a variety of common organic



Scheme 1

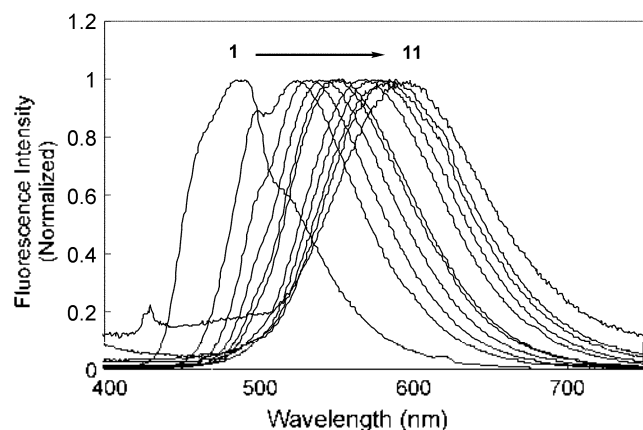


Figure 1. Normalized fluorescence spectra of **3** in varying solvents. Solvents: from left to right, 1, hexane; 2, toluene; 3, ethyl ether; 4, dioxane; 5, ethyl acetate; 6, THF; 7, acetone; 8, acetonitrile; 9, *n*-propanol; 10, ethanol; 11, methanol.

solvents including some of their aqueous solutions. Compound **3** revealed a significant dependence of fluorescence spectral behavior upon the solvent employed and, in general, the fluorescence maximum λ_{em} of **3** was gradually red-shifted as the solvent polarity increased. That is the λ_{em} of **3** was 486 nm in non-polar solvent of *n*-hexane, and was progressively red-shifted to 510 nm in toluene, 541 nm in dioxane, 566 nm in acetone, and eventually to 598 nm in polar solvent of methanol (Figure 1). The fluorescence color changes of **3** in these solvents were distinctive and the blue color of the fluorescence **3** was dramatically changed to green, yellow, orange, and finally red in responding to the solvent polarities from *n*-hexane to methanol. In this case, the span of the spectral changes was relatively wide from 486 nm to 598 nm in visible region and the color changes were distinctive enough to be observable with naked eye.¹⁷

Along with the general solvatochromic fluorescence behavior of **3**, a specific solvent effect has also been observed for the compound in various protic solvents. From the solvent dependent spectral changes, we have made a Lippert plot¹⁸ between Stokes' shift ($\Delta\nu$ = the frequency shift (in cm^{-1}) between absorption and emission) and orientation polarizability which is expressed as $\Delta f = (\epsilon - 1)/(2\epsilon + 1) - (n^2 - 1)/(2n^2 + 1)$ (Figure 2). Stokes' shifts were approximately proportional to the orientation polarizability for the surveyed solvents except for the relatively large deviation of toluene and dioxane, and moderate scattering for protic solvents.¹⁸ The rationale for the significant deviations for the relatively nonpolar solvents of toluene and dioxane is not clear at the moment. However, the excess shifts for protic solvents could be explained by the specific interaction of the hydrogen bonding between the amino group of the fluorophore and the employed solvent molecules. These spectral behaviors suggest that the fluorescence characteristics of **3** are sensitive to both general solvent effects and specific solvent effect in protic solvent systems.

The effect of solvents on the fluorescence of **3** was further investigated systematically by using aqueous binary solutions

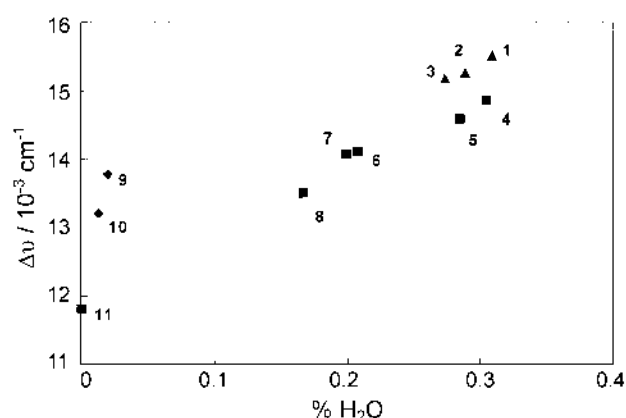


Figure 2. Effects of solvent polarity on the Stokes shift in λ_{em} of **3**. The numbers refer to the following solvents: 1, methanol; 2, ethanol; 3, *n*-propanol; 4, acetonitrile; 5, acetone; 6, THF; 7, ethyl acetate; 8, diethyl ether; 9, dioxane; 10, toluene; 11, hexane.

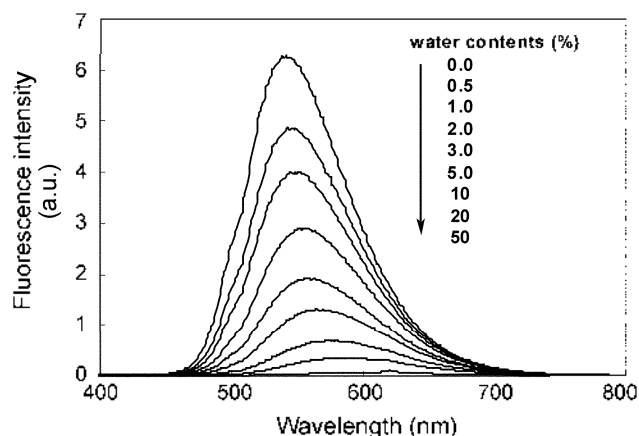


Figure 3. Fluorescence spectra of **3** as a function of water composition in aqueous 1,4-dioxane. $[\mathbf{3}] = 2.0 \times 10^{-5}$ M. $\lambda_{ex} = 310$ nm.

as medium. The compound **3** exhibited a strong fluorescence at 541 nm in pure 1,4-dioxane. However, as the percentage of water increases in aqueous dioxane solution, a gradual decrease in fluorescence intensity and a concomitant red-shift in fluorescence maximum were observed (Figure 3). The red-shifts of fluorescence maximum and the decrease in fluorescence intensity were also visually recognized by the changes in fluorescence color of **3** from yellow to red with increases in water contents of solution. In this case, the absorption spectra of **3** were not affected significantly in aqueous organic solutions of varying water compositions.

The effects of water contents in binary aqueous solvent systems were further investigated in other water miscible aprotic solvents such as acetonitrile, acetone, and THF (Figure 4). Generally, adding small amount of water (up to 5%) induced relatively large shifts in fluorescence maximum. After that, the changes became less significant. That is a typical of specific solvent effect and possibly due to the formation of strong hydrogen bonding between water and the amino group or nitrogen atoms of heterocyclic moieties of compound **3**. This observation of sensitive spectral shifts

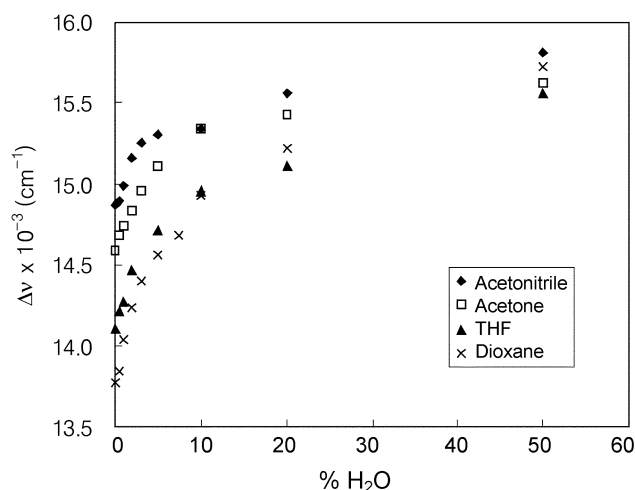


Figure 4. Effects of water composition in aprotic organic solvents on the fluorescence maximum of **3**. [**3**] = 2.0×10^{-5} M. $\lambda_{em} = 310$ nm.

as a function of water composition suggests that the shift in λ_{em} can be applied for the detection of water contents,¹⁹ especially in the range of up to 5% of water composition, in many aqueous organic solvent systems.

Conclusions

In conclusion, we have characterized a new sensitive fluorescent probe for solvent polarity of 8-aminoquinoline appended with benzothiazole function. The quinoline derivative exhibited a wide spectral shift in fluorescence emissions in visible region in response to the polarity of the medium. The compound also showed highly sensitive responses toward changes in water composition in aqueous solvent systems, which can be utilized as a new probe for the determination of water composition in aqueous binary solvent systems. The sensitive solvatochromic fluorescence behavior can also be used for the assessment of polarity characteristics in a variety of micro-environments in chemical and biological systems.

Experimental Section

General. All manipulations were carried out under an inert atmosphere. All solvents used for spectroscopic measurements were purchased from Aldrich Chemical Co. as 'anhydrous' or 'spectroscopic grade'. 2-Methyl-8-nitroquinoline and 2-aminothiophenol were purchased from Aldrich Chemical Co. ¹H and ¹³C NMR spectra were measured on a Varian Gemini-2000 spectrometer. HRMS spectra were obtained with a Micromass Autospec Mass Spectrometer. UV-Vis spectra were measured using a JASCO V-550 spectrophotometer.

Preparation of 8-nitro-2-quinolinecarboxaldehyde 1. A mixture of 2-methyl-8-nitroquinoline (1.00 g, 5.3 mmol) and SeO₂ (1.04 g, 9.4 mmol) in dioxane¹⁴ (30 mL) was heated at 90 °C for 2 h. The reaction mixture was evaporated and the

residue was dissolved in CH₂Cl₂, then filtered to remove undissolved solids. The filtrate was evaporated and redissolved in MeOH. The solution was filtered and the filtrate was evaporated. Crystallization of the crude product from hexane gave pure compound **1** (0.60 g) in 56% yield. ¹H NMR (300 MHz, CDCl₃) δ 10.20 (s, 1H), 8.55 (d, $J = 7.8$ Hz, 1H), 8.19-8.13 (m, 3H), 7.79 (t, $J = 8.1$ Hz, 1H). ¹³C NMR (75 MHz, CDCl₃): δ 193.3, 193.2, 154.2, 139.5, 138.0, 132.1, 130.7, 128.1, 124.7, 119.2. HRMS (EI): Calcd for C₁₀H₆N₂O₃ (M⁺) 202.0378. Found 203.0386.

Preparation of 8-nitroquinoline benzothiazole 2. To a solution of **1** (0.50 g, 2.5 mmol) in toluene (14 mL) was added 2-aminothiophenol (0.27 mL, 2.5 mmol) and acetic acid¹⁵ (0.14 mL). The mixture was refluxed for 3 h, cooled to room temperature and washed with dilute HCl and NaHCO₃ (10 mL). The combined organic layer was dried over Mg₂SO₄ and evaporated. Crystallization from MeOH afforded pure compound **2** (0.40 g) in 52% yield. ¹H NMR (300 MHz, CDCl₃) δ 8.64 (d, $J = 8.7$ Hz, 1H), 8.40 (d, $J = 8.7$ Hz, 1H), 8.15-8.07 (m, 3H), 7.99 (d, $J = 7.8$ Hz, 1H), 7.66 (t, $J = 7.8$ Hz, 1H), 7.54-7.46 (m, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 167.0, 154.6, 153.7, 148.3, 139.5, 137.3, 132.2, 129.8, 126.7, 126.6, 126.4, 125.0, 124.3, 122.5, 120.2. HRMS (EI): Calcd for C₁₆H₆N₃O₂S (M⁺), 307.0415. Found 307.0407.

Preparation of 8-aminoquinoline benzothiazole 3. To a suspension of **2** (0.50 g, 1.63 mmol) in toluene (33 mL) and water (3.8 mL) at 85 °C were added sodium hydrosulfite¹⁶ (1.8 g) and NaHCO₃ (0.5 g) in portion-wise over 2 h. The reaction mixture was washed with 2% NaHCO₃ (10 mL) and the separated organic phase was dried over MgSO₄ and evaporated. Crystallization from CH₂Cl₂ yielded **3** (0.35 g, 78%) as a brown solid. ¹H NMR (300 MHz, CDCl₃) δ 8.42 (d, $J = 8.5$ Hz, 1H), 8.19 (d, $J = 8.5$ Hz, 1H), 8.12 (d, $J = 8.7$ Hz, 1H), 7.96 (d, $J = 8.7$ Hz, 1H), 7.51 (t, $J = 7.4$ Hz, 1H), 7.42 (t, $J = 7.4$ Hz, 1H), 7.37 (t, $J = 7.4$ Hz, 1H), 7.18 (d, $J = 8.0$ Hz, 1H), 6.96 (d, $J = 7.4$ Hz, 1H), 5.11 (s, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 169.9, 154.4, 148.3, 144.1, 137.6, 136.9, 136.3, 129.6, 128.8, 126.2, 125.7, 123.6, 121.8, 118.3, 115.9, 110.6. HRMS (EI): Calcd for C₁₆H₁₁N₃S (M⁺) 277.0674. Found 277.0655.

Fluorescence titration experiments. Fluorescence measurements were performed using an Aminco-Bowman Series 2 Spectrometer. All the solvents used were spectroscopic grade. Incremental amounts of water were added to the stock solution of **3** (2.0×10^{-4} M) in organic solvents by micropipette. After this, the solution was diluted with calculated amounts of water or organic solvents to make the required probe concentration as well as the solvent compositions.

Acknowledgement. This work was supported by Chung-Ang University (2004) and gratefully acknowledged.

References

- (a) *Chemosensors of Ion and Molecule Recognition*; Desvergne, J. P.; Czarnik, A. W., Eds.; Kluwer: Dordrecht, 1997. (b) *Fluorescent Chemosensors for Ion and Molecule Recognition*; Czarnik, A. W.,

- Ed.: American Chemical Society: Washington, DC, 1993.
- Haugland, R. P. *Handbook of Fluorescent Probes and Research Products*. 8th Ed.; Molecular Probes: Eugene, 2001. See also <http://www.probes.com/handbook/>.
 - Reichardt, C. *Solvents and Solvent Effects in Organic Chemistry*. 2nd ed.; VCH: Weinheim, 1988.
 - (a) Yam, V.; Wong, K.; Zhu, N. *J. Am. Chem. Soc.* **2002**, *124*, 6506. (b) Ercelen, S.; Klymchenko, A.; Demchenko, A. *Anal. Chim. Acta* **2002**, *464*, 273. (c) Valle, J.; Catalan, J. *Chem. Phys.* **2001**, *270*, 1.
 - (a) Shin, E. J.; Lee, S. H. *Bull. Korean Chem. Soc.* **2002**, *23*, 1309. (b) Shin, E. J. *Bull. Korean Chem. Soc.* **2004**, *25*, 907.
 - Forlani, F.; Tritto, I.; Piemontesi, F. *Macromol. Chem. Phys.* **2000**, *201*, 401.
 - Cupane, A. L. M.; Fronticelli, C. *J. Biol. Chem.* **1997**, *272*, 26271.
 - Rachofsky, E. L.; Osman, R.; Ross, J. B. *Biochemistry* **2001**, *40*, 946.
 - Hiramoto, H.; Tohma, H.; Yamada, T.; Yamauchi, K.-I.; Siswanta, D.; Yoshioka, N.; Suzuki, K. *Anal. Chim. Acta* **1998**, *373*, 271.
 - Tada, E. B.; Silva, P. L.; El Seoud, O. A. *J. Phys. Org. Chem.* **2003**, *16*, 691.
 - Peyrin, F. X.; Auillaume, Y. C. *Anal. Chem.* **1999**, *71*, 2708.
 - (a) Pearce, D. A.; Jotterand, N.; Carrico, I. S.; Imperiali, B. *J. Am. Chem. Soc.* **2001**, *123*, 5160. (b) Jiang, P.; Chen, L.; Lin, J.; Liu, Q.; Ding, J.; Gao, X.; Guo, Z. *Chem. Commun.* **2002**, 1424. (c) Bagatin, I.; Souza, E.; Ito, A.; Toma, H. *Inorg. Chem. Commun.* **2003**, *6*, 288. (d) Youk, J.-S.; Kim, Y. H.; Kim, E.-J.; Youn, N. J.; Chang, S.-K. *Bull. Korean Chem. Soc.* **2004**, *25*, 869.
 - Youk, J.-S.; Kim, Y. H.; Moon, S. Y.; Choe, J. I.; Chang, S.-K. *Chem. Lett.* **2004**, 702.
 - Hata, T.; Uno, T. *Bull. Chem. Soc. Jpn.* **1972**, *45*, 477.
 - Iwamoto, K.; Araki, K.; Fujishima, H.; Shinkai, S. *J. Chem. Soc. Perkin Trans. 1* **1992**, 1885.
 - Pratt, Y. T.; Drake, N. L. *J. Am. Chem. Soc.* **1960**, *82*, 1155.
 - (a) Miyaji, H.; Sato, W.; Sessler, J. *Angew. Chem. Int. Ed.* **2000**, *39*, 1777. (b) Han, M. S.; Kim, D. H. *Angew. Chem. Int. Ed.* **2002**, *41*, 3809.
 - Lakowicz, J. R. *Principles of Fluorescence Spectroscopy*. 2nd ed.; Kluwer: New York, 1999; p 196.
 - Liu, W.; Wang, Y.; Jin, W.; Shen, G.; Yu, R. *Anal. Chim. Acta* **1999**, *383*, 299.
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