

New Cyclic Peroxides from a Sponge, Plakortis sp.

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Two new cyclic peroxides were isolated from an undescribed sponge of the genus *Plakortis* sp. collected at Discovery Bay, Jamaica. The molecular structures were elucidated by interpreting 1D and 2D NMR and HRMS data. The cyclic peroxides, Compound 1 and 2, exhibited significant antimicrobial activity against pathogenic bacteria and fungi with IC_{50} values of 0.9-5.0 μ g/mL and 0.7-8.0 μ g/mL, respectively.

Key words: Peroxide, Antifungal, Antibacterial, Plakortis

Introduction

Marine sponges are a source of a number of biosynthetically diverse natural products (Stierle and Faulkner, 1980; Davidson, 1991; Compagnone et al., 1998; Perry et al., 2001). Specially, cyclic peroxides from the genus *Plakortis* exhibited a diverse range of bioactivities, including anticancer (Fontana et al., 1998), antibacterial (Chen et al., 2002), and antifungal (Gunasekera et al., 1990) activities. In addition, they affect Ca²⁺ uptake by the cardiac sarcoplasmic reticulum (Patil et al., 1996a).

In the recent years, we have focused in part on peroxide containing marine natural products due to their activity against the phethogenic microbes as well as the malarial parasite *Plasmodium falciparum* (Peng, et al., 2002; Perry, et al., 2001). The polyketide-derived cyclic peroxides have been identified primarily from the genus *Plakortis* of the family Plakinidae.

Here we discussed to isolation and structural elucidation for two new cyclic peroxides that have activities against bacteria and funges from a Jamaican sponge, *Plakortis* sp.

Materials and Methods

Sponge collection and identification

The sponge was collected from inside caves at a depth of 37.5 m off the mouth of Rio Bueno,

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Discovery Bay, Jamaica, on June 2000. The sponge forms a moderately thick encrustation that is very soft, liver-like and very easily torn. It is avocado green in life, emits a bright deep blue watery exudates on exposure to air, and emits a very strong, characteristic sweet Oil of Wintergreen odor. The spicules are mesosclere diods averaging 190 μ m long and extremely rare triods. The sponge is thus an undescribed species of *Plakortis* (Subclass Homoscleromorpha, Order Homosclerophorida, Family Plakinidae). A voucher specimen was deposited at the Natural History Museum, London, United Kingdom.

Extraction and Isolation

The frozen sponge (1.3 kg) was extracted with ethanol (3×2 L). The extract was subjected to silica flash column chromatography using a gradient of hexane to ethyl acetate and finally methanol. We obtained 10 fractions, and fractions 2-6 contained active compounds. The fractions were chromatographed using a gel permeation chromatography on Sephadex LH-20 column (4.9×76 cm) in methanol, and the resulting fractions were purified further using reverse-phase HPLC (Phenomenex, C_{18} 5 μ M ODS 3100A, 10×250 mm) and eluting with a linear gradient of CH₃CN-H₂O (flow rate, 10 mL/min UV detection at 230 nm). This yielded Compound 1 (235 mg) and Compound 2 (163 mg).

Structure analysis

IR and UV spectra were obtained using an AATI

Mattson Genesis Series FTIR and a Perkin-Elmer Lambda 3B UV/Vis spectrophotometer. Optical rotations were measured on an AUTOPOL IV autopolarimeter. Both 1D and 2D NMR spectra were recorded on a Bruker Avance DRX-400 spectrometer. Chemical shift (δ) values were expressed in ppm and were referenced to the residual solvent signals with resonances at $\delta_{\rm H}/\delta_{\rm C}$ 7.26/77.0 (CDCl₃). The samples were processed on an ESI-FTMS 30es ion cyclotron HR HPLC-FT spectrometer with direct injection onto an eletrospray interface and positive or negative mode. The silica gel (230-400 mesh) and Sephadex LH-20 (Pharmacia) used for column chromatography were obtained from Natland International Corporation and Sigma Chemical Co. (USA), respectively.

Results and Discussion

Compound 1 was obtained as a colorless oil $\left[\alpha\right]_{D}^{25}$ of -62° (c 0.18, CHCl₃), and its molecular formula was determined to be $C_{20}H_{34}O_5$, by using positive ion HR-ESIMS [M+H] 355.2406 (calcd. 355.2398). The IR and UV spectra indicated that it was an α , β -unsaturated ketone [IR: 1689, 1647/cm; UV λ_{max} ($\log \varepsilon$) nm: 238 (4.25)] with a hydroxy band (3440 /cm). The ¹H, ¹³C, DEPT, and HMQC NMR experiments allowed the assignment of six methine, seven methylene, and four methyl groups (Table 1). The remaining quaternary centers consisted of a carbonyl (177.7), and oxygenated (82.7) and double-bond oxygenated carbon signals (199.4). The chemical shift values of the two oxygenated carbons suggested a peroxide functionality. A series of downfield ¹³C NMR resonances were assigned a disubstituted olefins [131.5(CH) and 153.0 (CH)]. The proton-carbon connectivity was determined from the HMQC and HMBC experiments and allowed the connection of spin systems to establish the complete structure of Compound 1. In the HMBC spectrum, one 2J and two 3J correlations were observed between H-11 at δ 6.54 and the methine carbon C-10, the ketone carbonyl C-13, and the methylene C-9 signals at 45.0, 199.4, and 35.5, respectively. Two 2J and 3J correlations were observed between the proton H-3 (δ 4.44) and the methine C-4, methylene C-2, ketone carbonyl C-1, and methylene C-5 signals at 34.8, 31.7, 177.7, and 33.2, respectively. The 2*J*, 3*J*, and 4J correlations were observed between the proton H-12 (δ 6.03) and the ketone carbonyl C-13, methine C-10, and the methylene C-19 signals at 199.4, 45.0

Table 1. ¹³C NMR data for compounds 1 and 2 (CDCl₃)

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No.	Compound 1	Compound 2
1	177.7 (s)	178.2 (s)
2	31.7 (d)	31.9 (t)
3	78.7 (d)	79.1 (d)
4	34.8 (d)	35.3 (d)
5	33.2 (t)	33.1 (t)
6	82.7 (s)	84.0 (s)
7	32.2 (t)	133.3 (d)
8	21.1 (t)	130.4 (d)
9	35.0 (t)	38.1 (t)
10	45.0 (d)	44.9 (d)
11	153.0 (d)	133.3 (d)
12	131.5 (d)	132.5 (d)
13	199.4 (s)	26.0 (t)
14	27.5 (q)	14.5 (q)
15	25.5 (t)	25.3 (t)
16	12.1 (q)	11.3 (q)
17	29.8 (t)	33.5 (t)
18	7.5 (q)	7.5 (q)
19	27.6 (t)	27.9 (t)
20	11.1 (q)	12.0 (q)

Compound 1

and 27.6, respectively.

The ¹H NMR (400 MHz, CDCl₃) spectrum (Table 2) showed signals indicating a -CH=CH-moiety [δ 6.54 (1H, dd, J=15.8, 9.0 Hz) and 6.03 (1H, d, J=16.0 Hz)], terminal methyl groups [δ 0.86 (3H, t, J=16.4 Hz) and δ 2.22 (3H, s)], and two isolated moieties at δ 1.48 (1H, m), 1.32 (1H, m), 0.85 (3H, m) and at δ 1.53 (1H, m), 1.35 (1H, m), 0.93 (3H, m), respectively. Five proton spin systems were included based on the analysis of the ¹H-¹H COSY data and include a -CH=CH- (H-11, H-12), CH₃CH₂-[(H-15, H-16), (H-17, H-18), and (H-19, H-20)], and CH₃CH₂CH(CH₂-)CHCH₂-(H-2, H-3, H-4, H-5, H-15, H-16).

Compound 2 was isolated as an oil and its molecular formula was determined to be $C_{20}H_{34}O_4$ by HR-ESIMS[M-H] 337.1356 (calcd. 337.3641). Comparison of the IR and UV spectra of Compound 2 with those of Compound 1 indicated the absence of the α , β -unsaturated ketone group.

The ESI (positive mode) spectrum of Compound

Table 2. ¹H NMR data for compounds 1 and 2 (CDCl₃)

No.	Compound 1	d 1 Compound 2			
2	2.99 (1H, dd, 14.8, 9.6)	3.01 (1H, dd, 16.0, 9.6)			
	2.38 (1H, d, 15.2)	2.39 (1H, br d, 16.0)			
3	4.44 (1H, br m)	4.41 (1H, dt, 5.6, 4.2)			
4	2.10 (1H, m)	2.12 (1H, m)			
5	1.52 (1H, m)	1.75 (1H, dd, 13.2, 4.0)			
	1.25 (1H, m)	1.25 (1H, dd, 13.2, 12.8)			
7	2.01 (1H, t, 11.8)	5.48 (1H, m)			
	1.46 (1H, m)				
8	1.32 (2H, m)	5.48 (1H, m)			
9	2.15 (1H, m)	2.14 (1H, d, 5.6)			
	1.30 (1H, d, 7.6)	2.02 (1H, m)			
10	2.01 (1H, br m)	1.88 (1H, m)			
11	6.54 (1H, dd, 15.8, 9.0)	5.12 (1H, dd, 15.2, 8.4)			
12	6.03 (1H, d, 16.0)	5.37 (1H, 15.2, 6.2)			
13		2.05 (1H, dd, 13.4, 3.4)			
		1.90 (1H, m)			
14	2.22 (3H, s)	0.94 (3H, t, 7.6)			
15	1.25 (1H, m)	1.20 (1H, m)			
	1.16 (1H, m)	1.12 (1H, m)			
16	0.86 (3H, t, 16.4)	0.89 (3H, t, 6.8)			
17	1.48 (1H, m)	1.40 (2H, m)			
	1.32 (1H, m)				
18	0.85 (3H, m)	0.80 (3H, m)			
19	1.53 (1H, m)	1.36 (1H, m)			
	1.35 (1H, m)	1.20 (1H, m)			
20	0.93 (3H, m)	0.82 (3H, t, 7.2)			

Compound 2

2 had pseudo-molecular ion peaks at 356.1237 and 694.9215 ($[M+NH_4]^+$) and ($[2M+NH_4]^+$), respectively, which is comparable to Compound 1. However, the molecular formula of Compound 2 was established to be $C_{20}H_{34}O_4$ by a HR-ESIMS signals at m/z 356.1237 ($[M+NH_4]^+$) and 694.9215 ($[2M+NH_4]^+$).

The ¹³C NMR spectrum contained 20 signals, which were assigned, using the DEPT experiment, as four methyl, seven methylene, and seven methine catbons, leaving two quaternary carbons (Table 1). Comparison of the ¹H and ¹³C NMR spectral data showed a plakotin-type cyclic peroxide ring system with signals at δ 3.01 (1H, dd, J=16.0, 9.6 Hz, H-2e) and 2.39 (1H, br d, J=16.0, H-2a), 4.41 (1H, m, H-3), 2.12 (1H, m, H-4), 1.75 (1H, dd, J=13.2, 4.0, H-5e) and 1.25 (1H, m, H-5a), which was supported by two typical signals for carbon atoms bearing oxygen at 79.1 (CH, C-3) and 84.0 (C, C-6) in the ¹³C NMR.

The E-configuration of the double bond in the side

chain was readily confirmed by the large coupling constant (J=15.2 Hz). The relative stereochemistry of Compound 1 was deduced from the detailed analysis of the coupling constant and NOESY correlations in the combination with molecular modeling. The vicinal coupling constants of the C-5 proton signals at 1.25 (1H, dd, J=13.2, 12.8 Hz) and 1.75 (1H, dd, 13.2, 4.0 Hz) indicated that the C-4 proton must be axial. Significant NOE correlations (Fig. 1) were observed between H-2 and H-5a, H-3e and H-2, H-3e and H-4a, H-4e and H-15, H-4e and H-16, H-5e and H-7, H-8 and H-17, H-8 and H-19, H-8 and H-10, H-10 and H-12, and H-11 and H-19, as well as H-12 and H-19, which required a 6α (equatorial)-ethyl configuration. Such a configuration at C-6 in polyketide-derived cyclic peroxides has previously been reported from sponges in the genus Plakortis (Patil et al., 1996b; Hu et al., 2001).

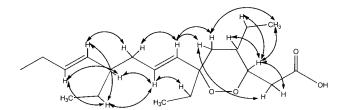


Fig. 1. Significant NOE correlations found for Compound.

The carbon skeleton of Compound 2 was almost similar to that of Compound 1 with exception the carbon signals of the olefinic carbon C-7 (133.3), C-8 (130.4), and methylene C-13 (26.0). The ¹H NMR spectrum of Compound 2 revealed two doublet methyl groups at δ 0.94 (3H, t, J=7.6) and 0.82 (3H, t, J=7.2). In addition to two proton spin systems that are comparable to compound 1[CH₃CH₂CH(CH₂-) CHCH₂-(H-2, H-3, H-4, H-5, H-15, H-16) and CH₃CH₂- (H-17, H-18)]; a third spin system [-CH=CH-CH₂CH(CH₂ CH₃)CH=CHCH₂CH₃ (H-7, H-8, H-9, H-10, H-19, H-20, H-11, H-12, H-13 and H-14)] observed in the ¹H-¹H COSY spectra required a terminal isopropyl group in the side chain. The position of the double bond was confirmed in the HMOC and HMBC experiments. The two olefinic proton signals (H-7 and H-8) overlapped at δ 5.48 (2H, m), whereas another two olefinic signals (H-11 and H-12) did not overlap at δ 5.12 (1H, dd, J=15.2, 8.4) and δ 2.37 (1H, dd, J=15.2, 6.2) in the ¹H NMR spectrum.

Compound 1 and Compound 2 were assayed for antifungal and antibacterial activities against *Candida*

5.0

8.0

Table 3. Data of	antininci Obiai	activities by compound	us i anu z		
Compounds	C. albicans	C. neoformans	S. aureus	MRSA	M. intracellulare
1	n 9*	0.9	0.7	1.6	2.5

2.5

Table 3. Data of antimicrobial activities by compounds 1 and 2

5.0

2

albicans ATCC 90028, Cryptococcus neoformans ATCC 90113, Staphylococcus aureus ATCC 29213, methicillin-resistant Staphylococcus aureus (MRSA) ATCC 90906, and Mycobacterium intracellulare ATCC 23068. The two compounds exhibited significant antifungal and antibacterial activities in vitro with IC₅₀ values of 0.9-5.0 μg/mL and 0.7-8.0 μg/mL, respectively (Table 3).

Compound 1-Colorless oil, $[\alpha]_D^{25}$ =-62 (*c* 0.18, CHCl₃); IR ν_{max} 3440, 2963, 2935, 2877, 1689, 1647, 1460/cm; UV λ_{max} 238 nm (log ε), HR-ESIMS m/z [M+H] 355.2406 (calcd. 355.2398), ¹³C and ¹H and NMR data see Table 1 and 2.

Compound 2-Colorless oil, $[\alpha]_D^{25}$ =-163 (*c* 0.19, CHCl3); IR ν_{max} 2962, 2932, 2876, 1443, 1290, 967/cm; UV λ_{max} 236 nm (log ε), HR-ESIMS m/z [M+NH₄] 356.1237 (calcd. for C₂₀H₃₄O₄ 338.2457), ¹³C and ¹H and NMR data see Table 1 and 2.

References

Chen, Y., J.M. Peter, K.H. Dedra, S.O. Rebecca, C. Katherine, S. Claude, A.P. Shirley and E.W. Amy. 2002. New bioactive peroxides from marine sponges of the family Plakiniidae. J. Nat. Prod., 65, 1509-1512.

Compagnone, R.S., I.C. Pina, H.R. Rangel, F. Dagger, A.I. Suarez, M.V.R. Reddy and D.J. Faulkner. 1998. Antileishamanial cyclic peroxides from the Palauan sponge *Plakortis* aff. *angulospiculatus*. Tetrahedron, 54, 3057-3068.

Davidson, B.S. 1991. Cytotoxic five-membered cyclic peroxides from a *Plakortis* sponge. J. Org. Chem., 56, 6722-6724.

Fontana, A., M. Ishibashi and J. Kobayashi. 1998. New cyclic polyketide peroxides from Okinawan marine

sponge *Plakortis* sp. Tetrahedron, 54, 2041-2048.

8.0

Gunasekera, S.P., M. Gunasekera, G.P. Gunawardana, P. McCarthy and N. Burrers. 1990. Two new bioactive cyclic peroxides from the marine sponge *Plakortis angulospiculatus*. J. Nat. Prod., 53, 669-674.

Hu, J.F., H.F. Gao, M. Kelly and M.T. Hamann. 2001. Plakortides I-L, four new cyclic peroxides from an undescribed Jamaican sponge *Plakortis* sp. (Homosclerophorida, Plakinidae). Tetrahedron, 57, 9379-9383.

Patil, A.D., A.J. Freyer, M.F. Bean, B.K. Carte, J.W. Westly and R.K. Johnson. 1996a. The plakortones, novel bicyclic lactones from the sponge *Plakortis halichondrioides*: Activators of cardiac SR-Ca²⁺-pumping ATPase. Tetrahedron, 52, 377-394.

Patil, A.D., A.J. Freyer, B.K. Carte, R.K. Johnson and P. Lahouratate. 1996b. Plakortides, novel cyclic peroxides from the sponge *Plakortis halichondrioides*: Activators of cardiac SR-Ca²⁺-pumping ATPase. J. Nat. Prod., 59, 219-223.

Peng, J., K. Walsh, V. Weedman, J.D. Bergthold, J. Lynch, K.L. Lieu, I.A. Braude, M. Kelly and M.T. Hamann. 2002. The new bioactive diterpenes cyanthiwigins E-AA from the Jamaican sponge *Myrmekioderma* styx. Tetrahedron, 58, 7809-7819.

Perry, T.L., A. Dickerson, A.A. Khan, R.K. Kondru, D.N. Beratan, P. Wipf, M. Kelly and M.T. Hamann. 2001. New peroxylactones from the Jamaican sponge *Plakinastrella onkodes*, with inhibitory activity against the AIDS opportunistic parasitic infectious *Toxoplasma gondii*. Tetrahedron, 57, 1483-1487.

Stierle, D.B. and D.J. Faulkner. 1980. Metabolites of three marine sponges of the genus *Plakortis*. J. Org. Chem., 45, 3396-3401.

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^{*}Minimum inhibitory concentration (MIC, µg/mL)