

불임여성에서 NAT2, GSTM1, CYP1A1 유전자 다형성과 자궁내막증의 상관관계에 관한 연구

성균관대학교 의과대학 삼성제일병원 산부인과¹, 생식생물학 및 불임연구실²

1 . 2 . 2 . 1 . 1 . 1 . 2

Association between Endometriosis and Polymorphisms of *N-acetyl Transferase 2 (NAT2)*, *Glutathione S-transferase M1 (GSTM1)* and *Cytochrome P450 (CYP) 1A1* Genes in Korean Infertile Patients

Hyun Jeong Song¹, Jin Hyun Jun², Hye Won Choi², Gurl Hur¹, Inn Soo Kang¹,
Mi Kyoung Koong¹, Hyoung-Song Lee²

¹Department of Obstetrics and Gynecology, ²Laboratory of Reproductive Biology and Infertility,
Samsung Cheil Hospital and Women's Healthcare Center, Sungkyunkwan University School
of Medicine, Seoul, Korea

Objective: To investigate the association between endometriosis and polymorphisms of *N-acetyl transferase 2 (NAT2)*, *glutathione S-transferase M1 (GSTM1)*, and *cytochrome P450 (CYP) 1A1* genes in Korean infertile patients.

Materials and Methods: A total of 303 infertile patients who had undertaken diagnostic laparoscopy during January, 2001 through December, 2003 at Samsung Cheil Hospital enrolled in this study. The patients were grouped according to laparoscopic findings: minimal to mild endometriosis (group I: n=147), moderate to severe endometriosis (group II: n=57), normal pelvic cavity (n=99). Peripheral blood was obtained and genomic DNA was extracted. The genotypes of each genes were analyzed using polymerase chain reaction (PCR) and restriction fragment length polymorphism (RFLP). For *NAT2*, RFLP was used to detect the wild type (wt) and mutant (mt) alleles, enabling classification into slow (mt/mt) or fast (wt/wt or wt/mt) acetylation genotypes. For *GSTM1*, PCR was used to distinguish active (+/- or +/+) from null (-/-) genotypes. For *CYP1A1*, *MspI* digestion was used to detect the wild type (A1A1), heterozygote (A1A2) or mutant (A2A2) genotypes.

Result: The genotype frequencies of *NAT2* slow acetylator was 12.8%, 10.9%, 12.8% in group I, group II and control, respectively. The genotype frequencies of *GSTM1* null mutation was 55.3%, 41.8%, 53.2% in group I, group II and control, respectively. The genotype frequencies of *CYP1A1 MspI* polymorphism was 16.3%, 9.1%, 18.1% in group I, group II and control, respectively. No significant difference was observed between endometriosis and normal controls in the genotype frequencies of the *NAT2*, *GSTM1*, *CYP1A1 MspI* polymorphism.

Conclusion: The *NAT2*, *GSTM1*, *CYP1A1* gene polymorphism may not be associated with the

susceptibility of endometriosis in Korean women.

Key Words: Endometriosis, Polymorphism, *NAT2*, *GSTM1*, *CYP1A1*

(stroma) (gland) , *CYP1A1* 15q22-q24 , 가 (*MspI* *Ile-Val*) 가 *MspI* 가 ¹⁰ .

가 (familial study) (twin study) *GSTM1* *GSTT1* , II *GSTM1* 1p13 (null allele) *GSTM1* 가 ^{11,12} .

가 N-acetyl transferase (NAT) II , *NAT1* *NAT2* 가 polymorphic gene *NAT2* .

Rier ⁶ Yang ⁷ 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD or dioxin) 8p22 wild-type *NAT2* allele (*4) 가 fast acetylators , variant alleles (*5, *6, *7) 가 slow acetylators ¹³ Slow acetylator ¹⁴ 가 , fast acetylator ¹⁵ 가 . *NAT2* ,

⁸ procarcinogen , estradiol *GSTM1*, *CYP1A1* 가 , proinflammatory growth factor , remodeling enzyme 가

가 ⁹ Arylhydrocarbon receptor (AhR) 1. aryl hydrocarbon receptor nuclear translocator (ARNT) 2001 1 2003 12

() 가 303 ⁹ minimal-mild (group I: n=147), moderate-severe (group II: n=57), (n=99) 가

가 Cyochrome P450 (CYP) 56 , I , revised American Fertility Society (1985)

Table 1. Sequences of oligonucleotide primers and PCR conditions for *NAT2*, *GSTM1* and *CYP1A1*

		Sequences (5' 3')	Anneling temp ()	Products sizes (bp)
<i>NAT2</i>	Forward	GCT GGG TCT GGA AGC TCC TC	62	547
	Reverse	TTG GGT GAT ACA TAC ACA AGG G		
<i>GSTM1</i>	Foreward	CTG CCC TAC TTG ATT GAT GG	64	218
	Reverse	CTG GAT TGT AGC AGA TCA TGC		
<i>GYP1A1</i>	Forward	CAG TGA AGA GGT GTA GCC GCT	62	340
	Reverse	TAG GGA GTC TTG TCT CAT GCC T		

가 .

2. Genomic DNA

Blood Kit (QIAGEN Inc, Chatsworth, CA, USA) genomic DNA

QIAamp genomic DNA

4 -20

3. (Polymerase chain reaction)

NAT2, *GSTM1* *CYP1A1* polymorphism

PCR (Table 1). PCR total 20 μ l

10 mM Tris-HCl (pH 8.3), 50 mM KCl, 1.5 mM MgCl₂, 0.2 mM dNTPs, 0.5 units of *Taq* DNA polymerase (Roche Diagnostics GmbH, Mannheim, Germany), 50~100 ng genomic DNA

10 pmol . PCR

denaturation 94 2

40 35 cycles

94 40 , 58 , 60 , 62 1 ,

72 1 cycle

extension 10

4.

NAT2 polymorphism wild 12.8%

allele (*4) mutant allele (*5, *6, *7)

PCR *Bam*HI gel UV

(Figure 1). *GSTM1* (+/+ +/ -) null (-/-) mutation

CYP1A1 *Msp*I genotype

5.

SPSS version 10.0 Chi-square test

. *p* 0.05

31.59 \pm 3.38 (Mean \pm STD)

24~42

31.78 \pm 3.54 (Mean \pm STD)

25~40

NAT2 minimal-mild acetylator 12.8% (group I) slow

(group II) 10.9%, moderate-severe

가 (Table 2).

GSTM1

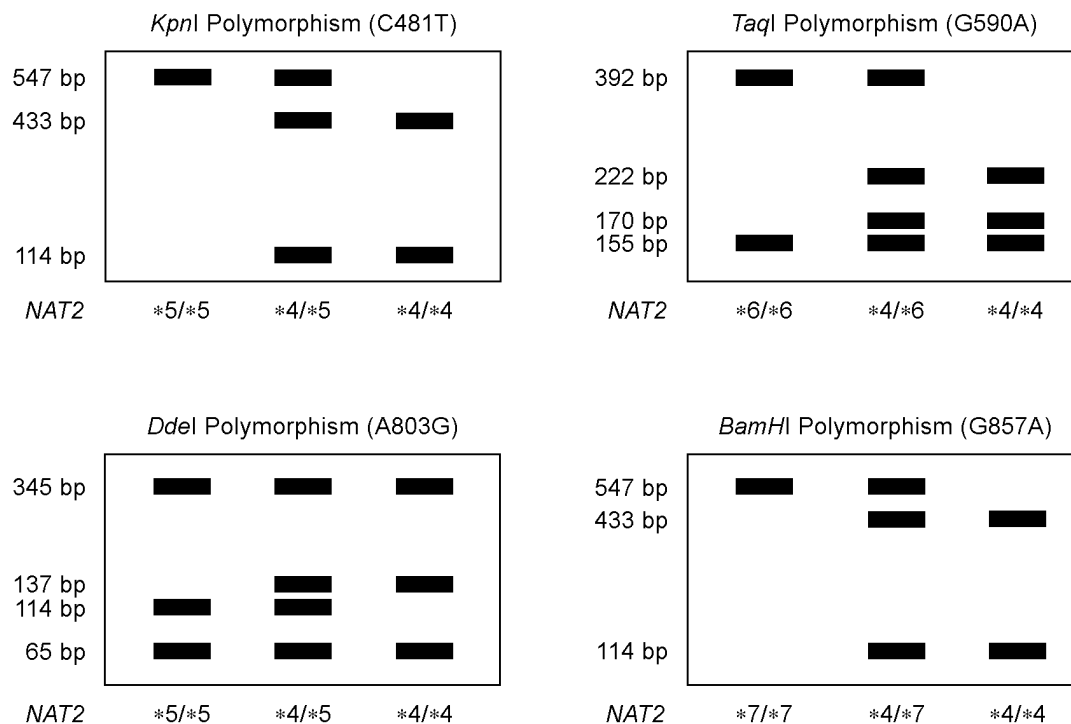


Figure 1. Genotype determination by restriction fragment length polymorphism analysis of the *NAT2* gene PCR products. Following PCR amplification, separate digestions of each PCR product were carried out with the restriction enzymes *KpnI*, *DdeI*, *TaqI* and *BamHI* to detect the substitutions C481T, A803G, G590A, and G857A, respectively. The sizes of the digested products for each restriction enzyme which allow the individual's genotype to be determined are shown diagrammatically (quoted from Nakago *et al.*, Mol Hum Reprod, 2001).

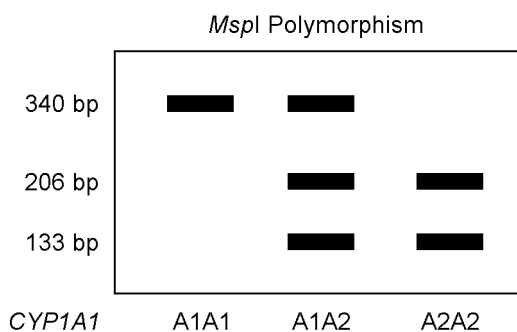


Figure 2. Genotype determination by restriction enzyme digestion of the *CYP1A1* gene PCR products. Following PCR amplification, digestions of each PCR product were carried out with the restriction enzymes *MspI*. A1A1, wild genotype; A1A2, heterozygous genotype; A2A2, homozygous genotype for polymorphism.

가 (Table 2).
CYP1A1
CYP1A1 MspI mutant
 group I 16.3%, group II 9.1%,
 18.1% 가
 (Table 2).

GSTM1 group I 55.3%,
 group II 41.8%, 53.2%

CYP1A1 Hadfield 16 가

Table 2. Frequencies of observed *NAT2*, *GSTM1* and *CYP1A1* genotypes between endometriosis patients and controls

Gene	Polymorphisms	Control	Group I ^a	Group II ^a	p value
<i>NAT2</i>	Slow ^b (mt/mt)	12.8% (12/94)	12.8% (18/141)	10.9% (6/55)	p>0.05
	Fast ^b (wt/mt, wt/wt)	87.2% (82/94)	87.2% (123/141)	89.1% (49/55)	p>0.05
<i>GSTM1</i>	Null (-/-)	53.2% (50/94)	55.3% (78/141)	41.8% (23/55)	p>0.05
	Active (+/+, +/-)	46.8% (44/94)	44.7% (63/141)	58.2% (32/55)	p>0.05
<i>CYP1A1</i>	Mutant (mt/mt)	18.1% (17/94)	16.3% (23/141)	9.1% (5/55)	p>0.05
	Wild (wt/wt, wt/mt)	81.9% (77/94)	83.7% (118/141)	90.9% (50/55)	p>0.05

^a Group I, minimal-mild endometriosis; Group II, moderate-severe endometriosis (rAFS, 1985)

^b Slow, only two variant alleles; Fast, presence of at least one wild-type (*4) allele

21%, 18%, 24% *CYP1A1* *MspI* polymorphism 가 . *GSTM1* 49.2%, 61.5% 가 *CYP1A1* *MspI* polymorphism 가 가 . II, 55.3%, 41.8%, 53.2% 가 group I, 17 가 . *NAT2* Baranova ¹⁸ (wt/wt) 45.1%, slow 34.3% 가 . acetylators 60.0% 38.9% *CYP1A1* mutant type (mt/mt, wt/mt) group I, II, 16.3%, 9.1%, 18.1% 가 3, 4 48.3% 가 *GSTM1* Baranova ¹⁸ 3, 4 (45.8%) *GSTM1* fast acetylator 가 57.4% (75.6%) minimal-mild 33.3% moderate-severe (79.3%) 32.3% *GSTM1* 가 21 , 16 *GSTM1* 가 , Hadfield 45%, slow acetylator 12.2%, 10.2% 52%, 45% Baxer ¹⁹ 가 group I, II, 가 . *GSTM1* 가 가 . 47.6%, 48.9% 17 , *GSTM1* 33.3%, 3, 4 56.9% *GSTM1* 가 ,

- 가
- ,
- 가
- 가
1. Moen NH, Magnus P. The familial risk of endometriosis. *Acta Obstet Gynecol Scand* 1993; 72: 560-4.
2. Kennedy SH, Mardon HJ, Barlow DH. Familial endometriosis. *J Assist Reprod Genet* 1995; 12: 32-4.
3. Treloar SA, O'Connor DJ, O'Connor VM, Martin NG. Genetic influences on endometriosis in Australian twin sample. *Fertil Steril* 1999; 71: 701-10.
4. Simpson JL, Elias S, Malinak LR, Buttram Jr VC. Heritable aspects of endometriosis. I. Genetic studies. *Am J Obstet Gynecol* 1980; 137: 327-31.
5. Lamb KRN, Hoffmann RG, Nichols TR. Family trait analysis: a case-control study of 43 women with endometriosis and their best friends. *Am J Obstet Gynecol* 1986; 154: 596-601.
6. Rier SE, Martin DC, Bowman RE, Dmowski WP, Becker JL. Endometriosis in rhesus monkeys (*Macaca mulatta*) following chronic exposure to 2,3,7,8-tetrachlorodibenzo-p-dioxin. *Fundam Asspl Toxicol* 1993; 21: 433-41.
7. Yang JZ, Agarwal SK, Foster WG. Subchronic exposure to 2,3,7,8-tetrachlorodibenzo-p-dioxin modulates the pathophysiology of endometriosis in the *Cynomolgus* monkey. *Toxicological Sciences* 2000; 56: 374-81.
8. Eskenazi B, Mocarelli P, Warner M, Samuels S, Vercellini P, Olive D, et al. Serum dioxin concentrations and endometriosis: A cohort study in Seveso, Italy. *Environmental Health Perspectives* 2002; 110: 629-34.
9. Rier S and Foster WG. Environmental dioxins and endometriosis. *Semin Reprod Med* 2003; 21: 145-53.
10. Xu X, Kelsey KT, Wiencke JK, Wain JC, Christiani DC. Cytochrome P450 CYP1A1 MspI polymorphism and lung cancer susceptibility. *Cancer Epidemiol Biomarkers Prev* 1996; 5: 687-92.
11. Zhong S, Wyllie AH, Barnes D, Wolf CR, Spurr NK. Relationship between the GSTM1 genetic polymorphism and susceptibility to bladder, breast and colon cancer. *Carcinogenesis* 1993; 14: 1821-4.
12. Murata M, Shiraishi T, Fukutome K, Watanabe M, Nagao M, Kubota Y, et al. Cytochrome P4501A1 and glutathione S-transferase M1 genotypes as risk factors for prostate cancer. *Jap J Clin Oncol* 1998; 28: 657-60.
13. Nakago S, Hadfield RM, Zondervan KT, Mardon H, Manek S, Weeks DE, et al. Association between endometriosis and N-acetyltransferase 2 polymorphisms in a UK population. *Mol Hum Reprod* 2001; 7: 1079-83.
14. Risch A, Wallace DM, Bathers S, Sim E. Slow N-acetylation genotype is a susceptibility factor in occupational and smoking related bladder cancer. *Hum Mol Genet* 1980; 4: 231-6.
15. Ilett KF, David BM, Detchon P, Castleden WM, Kwa R. Acetylation phenotype in colorectal carcinoma. *Cancer Res* 1987; 47: 1466-9.
16. Hadfield RM, Manek S, Weeks DE, Mardon HJ, Barlow DH, Kennedy SH, et al. Linkage and association studies of the relationship between endometriosis and genes encoding the detoxification enzymes GSTM1, GSTT1 and CYP1A1. *Mol Hum Reprod* 2001; 17: 1073-8.
17. (GSTM1, GSTT1 and CYP1A1)
- 2003; 46: 403-8.
18. Baranova H, Canis M, Lvaschenko T, Albuisson E,

- Bothorishvili R, Baranov V, et al. Possible involvement of arylamine N-acetyltransferase 2, glutathione S-transferases M1 and T1 genes in the development of endometriosis. *Molecular Human Reproduction* 1999; 5: 636-41.
19. Baxter SW, Thomas EJ, Campbell IG. GSTM1 null polymorphism and susceptibility to endometriosis and ovarian cancer. *Carcinogenesis* 2001; 22: 63-5.
20. , , , , , , , . GSTM1, GSTT1 . 2003; 46: 1300-5.
21. , , , , , , . N-acetyl transferase 2 . 2003; 46: 2113-7.
-