A Convenient and Versatile Synthesis of 2' (and 3')-Amino (and azido)-2' (and 3')-deoxyadenosine as Diverse Synthetic Precursors of Cyclic Adenosine Diphosphate Ribose (cADPR)

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As diverse synthetic precursors of cyclic adenosine diphosphate ribose (cADPR), several adenosine derivatives in which azido or amino group is introduced at 2'- or 3'-position of the sugar moiety of adenosine were prepared from readily available adenosine via conventional protocols. These synthetic sequence employs very efficient reactions conditions that proceed at or below ambient temperature with actual yields of \geq 80% for each individual step.

Key Words : Cyclic adenosine diphosphate ribose (cADPR), 2' (and 3')-Amino (and azido)- 2' (and 3')-deoxyadenosine

Introduction

Cyclic adenosine diphosphate ribose (cADPR, 1) is a naturally occurring cyclic nucleotide and a general mediator involved in Ca^{2-} signaling in a variety of mammalian and invertebrate tissues.¹ Also, recent studies reveal the involvement of cADPR in glucose-dependant insulin secretion in pancreatic cells, proliferation of human T-lymphocytes, and arrhythmogenic oscillations of intracelluar Ca^{2+} in heart cells etc.²⁻¹ Those findings suggest the therapeutic potential of cADPR itself or its derivatives against diabetes, immune and cardiovascular diseases. Therefore, considerable progress has been made in the synthesis of cADPR analogues and the evaluation of their biological effects. In general, the synthetic strategy to prepare cADPR analogues includes intracycli-

zation of NAD⁺ analogues (Figure 1), which can be prepared from the appropriately designed nucleosides/nucleotides.⁵ However, the reported precedents have limitation for the facile preparation of various nucleosides/nucleotides from readily available starting materials.¹⁵ To our knowledge, analogues of cADPR have been prepared from commercially available nucleosides/nucleotides as starting materials in most cases. So, there are still urgent needs for the development of new and diverse nucleosides with more convenient chemical routes.

Most of structural modifications occurred on the adenosine moiety of cADPR, namely, purine and sugar. In particular, a few previous researches show that the structural modification in sugar, especially at 2' and/or 3'-position gives rise to remarkable changes in their activities, namely, considerable

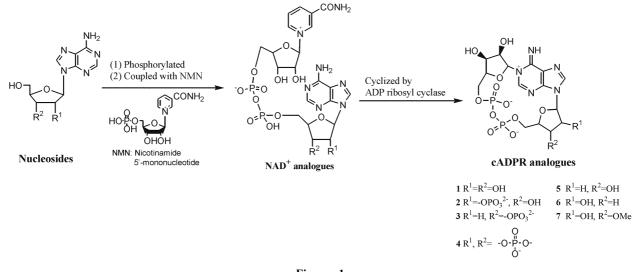
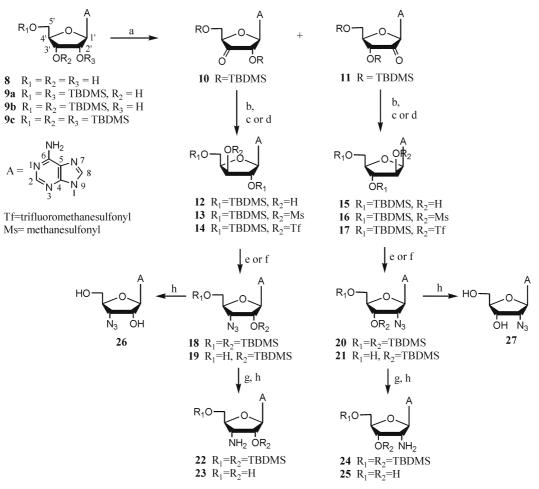


Figure 1

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Scheme 1. (a) CrO₃/Py/Acetic anhydride, CH₂Cl₂, RT; (b) NaBH₄/Acetic acid, THF, 0 °C; (c) Methanesulfonyl chloride, DMAP, CH₂Cl₂, 0 °C; (d) Trifluorosulfonyl chloride, DMAP, CH₂Cl₂, 0 °C; (e) NaN₃, Li₂CO₃, DMF, 120 °C; (f) NaN₃, DMF, RT; (g) Pd/C, H₂, MeOH, RT; (h) NH₄F, MeOH, 60 °C.

change in agonistic activity (2, 5, 6), showing no activities (3, 4), and even reversed activity (7). (Figure 1)^{1a} As mentioned above, however, as the lack of sufficient analogues obtained to date, the relationship between the functionality at 2' /and or 3'-position in sugar moiety and their Ca²⁻ modulating capacities has not been fully understood.

In this point of view, as a part of our synthetic effort to synthesize nucleosides having diverse functionality at 2' or 3'-position of sugar moiety, we designed aminosugar (and azidosugar) nucleosides (23, 25, 26 and 27, Scheme 1) in which 2'- or 3'-hydroxy group in ribose moiety is replaced with primary amine group and azido group. Since amino group is similar to hydroxy group in size and in hydrogenbonding character, and azido group shows interesting biological features when it is introduced to sugar moiety as shown in the example of AZT,⁶ it is considered to be interesting to us to investigate the activity variation of cADPR with amine and azido functionality at sugar moiety. Although there are precedences for the preparation of 2' (and 3')-aminoadenosine analogues,7.9 their synthetic routes are fairly laborious and furthermore each compound should be prepared in separate ways. So, our synthetic strategy was designed in a way that these target compounds (2' and 3'-

substituted products) can be prepared very efficiently using the same reaction steps starting from the disilylated adenosine analogs (**9a** and **9b**) obtained simultaneously in first step (Scheme 1). We now describe the synthesis of 2' (and 3')deoxy-2' (and 3')-azido-adenosine (**27, 26**) and 2' (and 3')deoxy-2' (and 3')-amino-adenosine (**25, 23**) starting from adenosine in very efficient manner by employing conventional protocols with actual yields of >80% for each individual reaction steps (Scheme 1).

Results and Discussion

Treatment of adenosine with *tert*-butyldimethylsilyl (TBDMS) chloride gave silyl-protected compounds, 2',5'bis-*O*-(*tert*-butyldimethylsilyl)adenosine (**9a**) and 3'5'-bis-*O*-(*tert*-butyldimethylsilyl)adenosine (**9b**) in 47% and 30% isolated yield, respectively, together with small amount of 2',3,5'-tris-*O*-(*tert*-butyldimethylsilyl)adenosine (**9c**).¹⁰ Oxidation (chromium trioxide/pyridine/acetic anhydride) of these 2',5'- and 3',5'-*O*-TBDMS-protected adenosine gave the corresponding 3'-keto- and 2'-ketoadenosines (**10** and **11**) in about 90% yield.¹¹ These individual ketonucleotides are known to be useful intermediates for the synthesis of a variety of sugar-modified nucleosides and have been employed to change configuration at C2' and C3' via oxidation-reduction sequence.¹² Although the oxidation of the protected alcohols 9a and 9b to their ketoadenosine 10 and 11, occurred successfully eventually, the chemical properties of compound 11 deserve some mention. Thus, treatment of 9a with the oxidizing agents gave ketoadenosine 10 very nicely with no incidence, which is easily identified by its clear TLC pattern.^{11a} In the case of 2'-ketoadenosine (11), however, there were some problems in separation due to the broad TLC pattern (tailing), which caused difficulty in purification. The ¹H-NMR analysis of the crude compound 11 exhibited the line broadening and doubling of purine peaks, which is consistent with the results reported by F. Hansske et al_{\perp}^{11b} This is likely caused by the formation of Cr complex and its existence as equilibrium mixture of ketone and ketone hydrate.¹⁰ In fact, isolated 2'-ketoadenosine 11 showed very low solubility in ethyl acetate and methylene chloride and the yield was not realized as reported. However, this low yield problem was overcome by using more polar elution solvent (EtOAc-MeOH = 9 : 1. v/v) during chromatography work, and the identity of 2'-ketoadenosine undefined on TLC was confirmed through consecutive reduction which gave arabinofuranosyl derivative (15). In other words, 2'ketoadenosine 11 can be utilized directly for the next step without purification, although it seems to be impure by TLC after the oxidation.

2' (and 3')-Ketonucleosides 11 and 10 were converted to the corresponding arabino (15) and xylofuranosyl nucleoside (12) with high stereoselectivity according to the modified method of M. J. Robins.^{11a} In original method, excess sodium triacetoxyborohydride was used rather than NaBH₄ as a reducing agent. However, it's not convenient to prepare sodium triacetoxyborohydride and a long reaction time (48 hours) and strict temperature (13 °C) control was required for their case. In contrast, in our modified method, THF was used as a reaction solvent in the preparation of sodium triacetoxyborohydride (generated in situ by adding excess AcOH to the suspension of NaBH₄ in THF at 0 °C) and reaction was performed at 0 °C. To our surprise, the reaction time became much more shorter (less than 4 hours) with the consistent high yields (> 90%).

Next we moved on to introduce azide group at 2' position of arabinofuranosyl nucleoside 15 and 3' position of xylofuranosyl nucleoside 12. The first step to mesylate compounds 15 and 12 occurred with no incidence to produce compound 16 and 13 in 93% and 92% yield, respectively. Treatment of 16 and 13 with NaN₃ or LiN₃ according to known procedure.^{6,13} however, produced the desired azido compound 20 and 18 in very poor yields (10-40%) along with 5'-deprotectd azido compounds 21 and 19 (6-15% yield). These problems, however, was easily overcome by utilizing triflate 17 and 14. Thus, the azido group was quantitatively introduced into the 2' (and 3')-position of corresponding triflates (17 and 14) using NaN₃ at room temperature¹⁴ to produce 20 and 18.

With these silvl protected azide compounds 20 and 18 in

hand. several chemical transformations to produce our key intermediates were carried out. First, simple treatment of **20** and **18** with NH₄F/MeOH at 60 °C gave silyl groupdeprotected azido compounds **27** and **26** quantitatively.¹⁵ Second. the azido group of compounds **20** and **18** was reduced with H₂ in MeOH in the presence of Pd/C without loss of the TBDMS protecting groups to produce silyl group-protected amino compounds **24** and **22** in 90% and 92% yield. respectively. Third, the silyl group of amino compounds **24** and **22** can be deprotected under the same condition to give silyl group-deprotected amino compounds **25** and **23** in 87% and 85% purified yield. respectively. The desilylation step is apparently occurred in two-steps on the analysis of TLC, and so the selective desilylation of our particular compounds is possible upon needed.

In summary, these convenient and efficient sequence for the preparation of adenosine analogues (23, 25, 26, 27), which are crucial intermediate for the structural modification of cADPR, were described. In particular, the improvement in the reduction procedure using THF as a co-solvent for the preparation of xylo- and arbinofuranosyl nucleoside (12 and 15) is noticeable in terms of efficiency and convenience. More importantly, compared to other precedent procedures, our synthetic strategy is convenient and versatile as well, from the points of view that several target compounds can be prepared using same starting material via continuous and consecutive processes. With these compounds in hand, the synthetic work for the preparation of cADPR analogs and further modification at adenine moiety is now in progress.

Experimental Section

Melting points were recorded on electrothermal melting point apparatus and are uncorrected. Mass spectra were recorded on a JEOL JMS-DX 303 mass spectrometer (3 KV). ¹H-NMR and ¹³C-NMR data were obtained from Jeol 400 MHz spectrometer and chemical shifts (δ) were reported in ppm in relation to tetramethylsilane (δ 0.00) and CDCl₃ (δ 77.0) for ¹H and ¹³C NMR, respectively; J values are in hertz (Hz). Thin layer chromatography was performed on precoated silica gel 60 F-254 (layer thickness 0.2 mm, Merck). The Flash Column Chromatography was performed on Merck silica gel type 60 (230-400 mesh). Unless specified otherwise the solvent system for all chromatography was EtOAc/Hexane (7 : 3, v/v). The organic solvents and chemicals were obtained from Aldrich. Co. and purified by the appropriate methods before use.

Silylation of Adenosine. TBDMSC1 (8.46 g. 56.13 mmol) was added to a suspension of 8 (5.0 g. 18.7 mmol) in pyridine (35 mL) and stirred at ambient temperature for 48 h under Ar atmosphere. The solvent was evaporated and the residue partitioned (ice-cold 5% HC1/H₂O//CH₂Cl₂). The organic phase was washed (saturated NaHCO₂/H₂O and brine). dried (MgSO₄), and evaporated. The white solid was suspended in EtOAc and applied to a column and separated by Yamazen MPLC system (flow rate: 20 mL/min, detected at 254 nm) to give 2'.5'-bis-O-(tert-butyldimethylsilyl)-

adenosine (9a, 47%), 3'.5'-bis-O-(*tert*-butyldimethylsilyl)adenosine (9b, 30%), 2'.3'.5'-tris-O-(*tert*-butyldimethylsilyl)adenosine (9c, 4%) as white powders with expected spectral properties.¹⁰

9-[2',5'-Bis-O-(*tert*-butyldimethylsilyl)-1- β -D-erythropentofuran-3'-ulosyl]adenine (10). Procedure A. pyridine (2.5 mL, 30.7 mmol) and Ac₂O (1.45 mL, 15.4 mmol) were added consecutively to an ice-cold suspension of CrO₃ (1.54 g, 15.4 mmol) in CH₂Cl₂ (50 mL) and stirred until homogeneous (10-15 min) at ambient temperature under Ar atmosphere. A solution of 9a (3.81 g, 7.67 mmol) in CH₂Cl₂ (10 mL) was added. stirring was continued for 2 h. The reaction mixture was poured into cold EtOAc (1 L) and filtered (glass microfiber filter, GF/A). The filtrate was concentrated and purification (short column chromatography. EtOAc) gave 10 (3.37 g, 89%) as a colorless powder with reported properties.¹¹

9-[3',5'-Bis-O-(*tert*-butyldimethylsilyl)-1- β -D-erythropentofuran-3'-ulosyl]adenine (11). Oxidation of 9b (1.50 g, 3.02 mmol) by procedure A [CrO₃ (0.61 g. 6.04 mmol)/ pyridine (0.98 mL, 12.1 mmol)/Ac₂O (0.57 mL, 6.04 mmol)/ CH₂Cl₂ (25 mL)] gave 11 (1.32 g. 89%) as a pale yellow powder with reported properties.¹¹ The purification was performed on the short column chromatography eluted with EtOAc-MeOH (9 : 1).

9-[2',5'-Bis-O-(*tert*-butyldimethylsilyl)-1- β -D-xylofuranosyl]adenine (12). Procedure B. Ketone 10 (1.0 g. 2.03 mmol) was added to a mixture of NaBH(OAc)₃/THF/AcOH [generated in situ by addition of AcOH (7 mL) to a suspension of NaBH₄ (0.92 g, 24.3 mmol) in THF (50 mL) and then stirred for 1.5 h at 0 °C under Ar atmosphere]. The reaction was stirred for 4 h at 0 °C, and volatiles were evaporated. The residue was partitioned (EtOAc/sat. aqueous NaHCO₃), and the aqueous layer was extracted with EtOAc. The combined organic phase was washed (H₂O, brine). dried over MgSO₄, filtered, and evaporated. The oily residue was applied to flash column chromatography to give 12 (0.90 g. 90%) as a colorless powder with reported properties. ^{11a}

9-[3',5'-Bis-*O*-(*tert*-butyldimethylsilyl)-**1-** β **-D**-arabinofuranosyl]adenine (15). Treatment of ketone **11** (1.30 g. 2.63 mmol) by procedure **B** [NaBH₄ (1.20 g. 31.6 mmol) in THF (60 mL)/AcOH (9 mL)] gave **15** (1.02 g. 79%) as a colorless powder with reported properties. ^{Ua}

9-[3'-O-(Trifluormethanesulfonyl)-2',5'-Bis-O-(*tert*-butyldimethylsilyl)-1- β -D-xylofuranosyl]adenine (14). Procedure C. To a solution of 12 (2.26 g. 4.56mmol) and DMAP (1.68 g. 13.68 mmol) in CH₂Cl₂ (15 mL) was added fresh trifluoromethanesulfonyl chloride (0.53 mL, 5.01 mmol) at 0 °C under Ar atmosphere. The reaction was stirred for 15 min at this temperature and then checked by TLC. A second portion of trifluoromethanesulfonyl chloride (0.1 mL, 0.94 mmol) was added and stirring was continued for 15 min at 0 °C. The reaction was partitioned (ice-cold 1% aqueous AcOH/CH₂Cl₂), and the aqueous layer was extracted with CH₂Cl₂. The combined organic phase was washed (ice-cold saturated aqueous NaHCO₃, ice-cold brine). dried over MgSO₄, filtered, and evaporated. Flash chromatography gave colorless powder of **13** (2.23 g. 78%): mp: 48-52 °C (softening): ¹H-NMR, CDCl₃ δ 8.20 (s, 1H. H8), 7.91 (s, 1H. H2). 6.43 (br, 2H. NH₂). 5.95 (d. 1H. *J* = 2.0 Hz, H1'). 5.01 (m, 1H, H2'), 4.82 (m, 1H. H3'), 4.42 (m, 1H, H4'). 3.99 (m. 2H. H5'a, H5'b). 0.79 (s, 9H, *t*-butyl). 0.75 (s, 9H, *t*-butyl) 0.00 (s. 6H. methyl × 2), -0.02 (s. 3H. methyl). -0.10 (s. 3H. methyl): ¹³C-NMR δ 155.63, 152.93, 149.48. 137.79. 119.60. 89.74, 88.38, 80.46. 79.06, 60.07, 25.46. 25.33. 18.24, 17.65, -5.16, -5.34, -5.59, -5.60.

9-[2'-O-(Trifluormethanesulfonyl)-3',5'-bis-O-(*tert*-butyldimethylsilyl)-1-**β**-D-arabinofuranosyl]adenine (17). Treatment of 15 (1.39 g, 2.80 mmol) by procedure C [DMAP (1.03 g, 8.41 mmol) in CH₂Cl₂ (10 mL)/trifluoromethanesulfonyl chloride (0.33 mL, 3.08 mmol and 0.07 mL, 0.66 mmol)] gave 16 (1.45 g, 82%) as a colorless powder: mp: 116-118 °C: ¹H-NMR, CDCl₃ δ 8.25 (s. 1H, H8). 7.89 (s. 1H, H2). 6.48 (d, 1H. J = 4.0 Hz. H1'). 5.59 (br, 2H. NH₂). 5.10 (m. 1H, H2'). 4.67 (m. 1H, H3'), 3.93 (m, 1H. H4'). 3.80 (dd. 2H. J = 4.4, 10.8 Hz. H5'a), 3.72 (dd. 1H, J = 6.4. 10.8 Hz. H5'b). 0.83 (s. 9H. *t*-butyl), 0.82 (s. 9H, *t*-butyl) 0.07 (s, 6H. methyl × 2). 0.00 (s, 6H. methyl × 2).

9-[3'-O-(Methanesulfonyl)-2',5'-bis-O-(tert-butyldimethylsilyl)-1-**β-D-**xylofuranosyl]adenine (13). Procedure D. To a solution of 12 (1.50 g, 3.02 mmol) and DMAP (1.11 g, 9.08 mmol) in CH₂Cl₂ (10 mL) was added fresh methanesulfonyl chloride (0.28 mL, 3.63 mmol) at 0 °C under Ar atmosphere. The reaction was stirred for 3 h at this temperature and then checked by TLC. A second portion of methanesulfonyl chloride (0.1mL, 1.29 mmol) was added and stirring was continued for 1 h at 0 °C. The reaction was partitioned between ice-cold 1% aqueous AcOH and CH₂Cl₂, and the aqueous layer was extracted once more with CH₂Cl₂. The combined organic phase was washed (ice-cold sat. aqueous NaHCO₃, ice-cold brine), dried over MgSO₄, filtered, and evaporated to give 13 (1.59 g. 92%) as a colorless solid: mp: 57-58 °C: ¹H-NMR, CDCl₃ δ 8.27 (s, 1H. H8), 8.05 (s, 1H. H2), 6.20 (br, 2H, NH₂), 6.04 (d, 1H. J = 2.0 Hz, H1'). 4.88 (m, 1H, H2'), 4.76 (m. 1H, H3'). 4.45 (dd, 1H, J = 4.4, 10.8 Hz, H4'), 3.96 (dd, 1H, J = 4.8, 10.8)Hz. H5'a), 3.91 (dd, 1H, J = 6.8, 10.8 Hz, H5'b). 2.92 (s, 3H, -SO₂CH₃), 0.86 (s. 9H, *t*-butyl), 0.82 (s, 9H, *t*-butyl), 0.07 (s, 3H. methyl), 0.06 (s. 3H, methyl), 0.00 (s, 6H, methyl \times 2); 13 C-NMR δ 155.54, 153.12, 149.67, 138.33, 119.46, 89.71, 80.68, 79.62, 60.22, 38.08, 25.80, 25.46, 18.26, 17.72, -5.08, -5.10. -5.34. -5.42.

9-[2'-O-(Methanesulfonyl)-3',5'-bis-*O*-(*tert*-butyldimethylsilyl)-1-**β**-D-arabinofuranosyl]adenine (16) Treatment 15 (1.07 g. 2.16 mmol) by procedure D [DMAP (0.79 g. 6.48 mmol) in CH₂Cl₂ (20 mL)/methanesulfonyl chloride (0.2 mL, 2.59 mmol/0.1 mL. 1.29 mmol)] gave 16 (1.15 g, 93%) as a colorless solid: mp: 62-64 °C; ¹H-NMR. CDCl₃ δ 8.25 (s, 1H. H8), 7.98 (s, 1H. H2), 6.45 (d. 1H, J = 4.4 Hz, H1'), 6.14 (br. 2H. NH₂). 5.01 (t. 1H. J = 4.4 Hz. H2'), 4.64 (t, 1H. J = 4.0 Hz. H3'), 3.91 (m, 1H. H4'), 3.78 (m. 1H. H5'a, H5'b). 2.45 (s. 3H, -SO₂CH₃), 0.82 (s, 18H, *t*-butyl × 2), 0.08 (s, 3H. methyl), 0.06 (s, 3H. methyl), 0.01 (s, 3H. methyl), 0.00 (s. 3H. methyl): ¹³C-NMR δ 155.60, 153.21. 149.41, 139.63, 119.10, 84.81, 82.63, 82.45, 74.94, 61.77, 37.85, 25.89, 25.60, 18.38, 17.81, -4.62, -4.97, -5.39, -5.42.

3'-Azido-3'-deoxy-2',5'-bis-O-(tert-butyldimethylsilyl)adenosine (18). Procedure E. NaN₃ (0.11 g, 1.71 mmol) was added to a solution of 14 (0.22 g, 0.34 mmol) in anhydrous DMF (4 mL) and the reaction was stirred at ambient temperature for 3 h under Ar atmosphere. The reaction was diluted with H2O (20 mL) and then extracted with EtOAc (20 mL \times 2). The combined organic phase was washed with brine, dried over MgSO4, filtered, and evaporated to give 18 (0.19 g, 99%) as a colorless powder: mp: 106-109 °C: IR (KBr) cm⁻¹ 2107 (strong, -N₃); ¹H-NMR, CDCl₃ δ 8.17 (s. 1H, H8), 8.01 (s. 1H, H2), 5.96 (br. 2H, NH2), 5.88 (d. 1H, J = 4.0 Hz, H1'), 4.71 (t. 1H, J = 4.0Hz, H2'), 4.07 (m. 1H. H3'), 3.92 (m, 1H, H4'). 3.91 (m. 1H. H5'a). 3.68 (dd, 1H, J = 2.0, 11.6 Hz, H5'b), 0.79 (s, 9H. t-butyl), 0.71 (s, 9H, t-butyl), 0.00 (s, 3H, methyl), -0.01 (s, 3H, methyl), -0.08 (s. 3H, methyl), -0.19 (s, 3H, methyl); ¹³C-NMR δ 155.53, 153.01, 149.61, 138.84, 119.92, 88.88. 81.99, 76.96, 62.39, 60.88, 25.94, 25.55, 18.43, 17.90, -5.08, -5.14, -5.36, -5.51; MS (FAB) [M+H]⁺ 521.

2'-Azido-2'-deoxy-3',5'-bis-*O*-(*tert*-butyldimethylsilyl)adenosine (20). Treatment of 17 (0.80 g, 1.27 mmol) by procedure E [NaN₃ (0.42 g, 6.37 mmol)/DMF (15 mL)] gave 20 (0.66 g, 99%) as a colorless powder: mp: 132-135 °C; ¹H-NMR, CDCl₃ δ 8.27 (s. 1H, H8). 8.01 (s. 1H, H2), 6.06 (br. 2H, NH₂), 6.05 (d, 1H, *J* = 5.2 Hz, H1'), 4.63 (t. 1H, *J* = 5.2 Hz, H2'), 4.41 (t. 1H, *J* = 4.8 Hz, H3'). 4.04 (m. 1H, H4'). 3.88 (dd, 1H, *J* = 3.6, 11.2 Hz, H5'a). 3.68 (d. 1H, *J* = 11.2 Hz, H5'b), 0.88 (s, 9H, *t*-butyl). 0.81 (s. 9H, *t*-butyl) 0.10 (s. 3H, methyl). 0.08 (s. 3H, methyl). 0.00 (s. 3H, methyl). -0.01 (s, 3H, methyl): ¹³C-NMR δ 155.70, 153.13, 149.58. 139.08, 120.00, 86.03, 85.50, 72.30, 65.12, 61.86, 25.89. 25.85, 18.35, 18.03, -4.72, -4.98, -5.36, -5.47; MS (FAB) [M+H]⁺ 521.

Azidation of compound 13. Procedure F. To the mixture of NaN₃ (0.13 g, 1.93 mmol) and Li₂CO₃ (0.142 g, 1.93 mmol) in anhydrous DMF (4 mL) pre-heated at 120 °C for 3 hours was added 13 (0.22 g. 0.39 mmol) and stirred at 120 °C and was stirred for 3 days. The reaction was diluted with H_2O (50 mL) and then extracted with EtOAc (50 mL \times 3). The combined organic phase was washed with brine, dried over MgSO₄, filtered, and evaporated. The residue was applied to flash column chromatography to give 18 (0.08 g. 43%) and 19 (0.01 g. 6%) as a white powder: 19: mp: 190-192 °C: ¹H-NMR, CDCl₃ δ 8.43 (s. 1H, H8), 7.91 (s. 1H, H2), 7.36 (d. 1H. J = 12.0 Hz. -OH. exch), 6.07 (br. 2H. NH₂). 5.84 (d. 1H, J = 7.2 Hz, H1'), 5.46 (dd, 1H, J = 7.2, 8.0 Hz, H2'), 4.36 (d. 1H, J = 6.0 Hz, H3'), 4.24 (m, 1H, H4'). 4.04 (d, 1H, J = 12.4 Hz, H5'a). 3.79 (m, 1H, H5'b). 0.88 (s. 9H, t-butyl). 0.00 (s. 3H, methyl). -0.35 (s, 3H. methyl): ¹³C-NMR δ 156.03, 152.56, 148.48, 140.81, 121.22, 90.74, 85.48, 74.61, 63.95, 63.46, 25.48, 17.76, 18.03, -5.16, -5.98.

Azidation of Compound 16. Treatment of 16 by procedure F (0.37 g. 0.65 mmol) [NaN₃ (0.21 g. 3.24 mmol)/Li₂CO₃ (0.24 g. 3.24 mmol) for 4 days] gave 20 (0.05

g. 15%) and **21** (0.05 g. 15%) as a white powder: **21**: mp: 193-195 °C: ¹H-NMR, CDCl₃ δ 8.10 (s. 1H. H8), 8.02 (s, 1H. H2). 6.89 (br. 2H. NH₂), 6.13 (d, 1H, J = 4.4 Hz. -OH, exch), 5.89 (d. 1H. J = 3.2 Hz, H1'). 4.52 (m. 1H. H2'), 4.30 (dd, 1H. J = 4.8. 10.4 Hz. H3'). 4.15 (m. 1H, H4'). 3.80 (m, 2H. H5'a, H5'b). 0.80 (s, 9H, *t*-butyl). 0.00 (s, 6H. methyl × 2); ¹³C-NMR δ 155.67, 155.61, 152.47, 149.13, 118.75, 88.19, 78.29. 66.28. 61.29. 25.45. 17.79. -5.78. -5.92.

3'-Amino-3'-deoxy-2',5'-bis-O-(tert-butyldimethylsilyl)adenosine (22). Procedure G. A solution of 18 (0.10 g, 0.19 mmol) in MeOH (10 mL) was hydrogenated at ambient pressure and temperature in the presence of 10% Pd/C (20 mg) for 16 hours. The mixture was filtered with the aid of celite, and the filtrate was evaporated. The residue was applied to flash column chromatography to give 22 (0.09 g. 92%) as a colorless powder: mp: 167-169 °C; ¹H-NMR. CDCl₃ δ 8.22 (s, 1H, H8), 8.20 (s, 1H, H2), 5.91 (s, 1H, H1'), 5.79 (br. 2H. NH₂), 4.25 (t, 1H, J = 3.6 Hz. H2'). 3.98 (d. 1H, J = 9.6 Hz, H5'a). 3.80 (m, 1H, H4'). 3.79 (m, 1H. H5'b). 3.49 (dd, J = 4.4, 8.0 Hz, H4'), 0.83 (s. 9H, *t*-butyl), 0.82 (s. 9H, t-butyl) 0.09 (s, 3H, methyl), 0.03 (s. 3H, methyl). 0.02 (s. 3H. methyl). 0.00 (s. 3H. methyl); ¹³C-NMR δ 155.30, 152.76, 149.35, 139.07, 119.95, 89.74, 84.93, 77.82, 61.91, 52.19, 26.02, 25.77, 18.56, 18.03, -4.50, -5.04. -5.30. -5.44: MS (FAB) [M+H]⁺ 495.

2'-Amino-2'-deoxy-3',5'-bis-*O*-(*tert*-butyldimethylsilyl)adenosine (24). Hydrogenation of **20** (0.5 g. 0.96 mmol) by procedure **G** gave **24** (0.43 g, 90%) as a colorless powder: mp: 145-147 °C: ¹H-NMR, DMSO-d6 δ 8.15 (s, 1H. H8). 7.97 (s, 1H. H2), 7.13 (br, 2H. NH₂). 5.53 (d, 1H. *J* = 7.6 Hz. H1'), 4.08 (d. 1H. *J* = 4.8 Hz, H3'). 4.01 (dd. 1H. *J* = 4.8, 7.6. H2'), 3.79 (m, 1H. H4'). 3.73 (dd, 1H. *J* = 4.4. 11.2. H5'a). 3.52 (dd, *J* = 4.0. 11.2 Hz. H5'b), 1.59 (br, 1.5H, H2'-NH₂, exch), 0.78 (s. 9H. *t*-butyl), 0.73 (s. 9H. *t*-butyl). 0.00 (s. 6H, methyl × 2), -0.09 (s. 6H, methyl × 2); ¹³C-NMR δ 156.05, 152.48, 149.72, 139.80, 119.22. 87.94, 85.64, 73.80. 62.86, 56.20, 25.78, 25.76. 18.00. 17.89. -4.72, -4.78. -5.49, -5.51; MS (FAB) [M+H]⁻ 495.

3'-Amino-3'-deoxyadenosine (23). Procedure H. A solution of **22** (0.1 g. 0.20 mmol) and NH₄F (0.09 g, 1.76 mmol) in MeOH (10 mL) was stirred in oil bath at 60 °C for 12 hours. The solvent was evaporated and the resulting solid was partitioned between H₂O/EtOAc. The aqueous phase was concentrated to 3 mL and applied to ion-exchange resin column chromatography (Dowex 1×20 -200. OH- form. 2×18 cm). The column was washed with H₂O and eluted with 20-50% MeOH. The chromatographically identical fractions (BuOH-AcOH-H₂O = 60 : 15 : 25) were collected, and the solvent evaporated to give **23** (0.05 g, 85%) with reported properties.⁷

2'-Amino-2'-deoxyadenosine (25). Treatment of **24** (0.39 g. 0.78 mmol) by procedure **H** [NH₄F (0.29 g. 7.88 mmol) in MeOH (15 mL)] gave **25** (0.18 g. 87%) as a colorless powder with reported properties.⁸

3'-Azido-3'-deoxyadenosine (26). Treatment of **18** (0.50 g. 0.90 mmol) by procedure H [NH₄F (0. 50 g, 12.63 mmol) in MeOH (15 mL)] gave **26** (0.28 g. 99%) as a colorless

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powder with reported properties.⁹

2'-Azido-2'-deoxyadenosine (27). Treatment of 20 (0.60 g, 1.15 mmol) by procedure H [NH₄F (0.60 g, 16.13 mmol) in MeOH (15 mL)] gave 27 (0.34 g, 99%) as a colorless powder with reported properties.⁹

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