Prevalence and Transmission of Seed-Borne Fungi of Maize Grown in a Farm of Korea

A. B. Basak^{1*} and Min Woong Lee

Department of Applied Biology, Dongguk University, Seoul 100-715, Korea Department of Botany, University of Chittagong, Chittagong 4331, Bangladesh (Received February 7, 2002)

Seed-borne fungi of some maize cultivars/lines grown during the months from May to September of 2001, collected from Dongguk University farm, Go Young City, IL Sang Gu, Korea were detected by blotter method. In all six fungi namely Alternaria alternata (Fr.) Keissler, Aspergillus niger Van Tiegh, Fusarium moniliforme Sheldon, Fusarium sp., Penicillium sp. and Ustilago zeae Unger. were found to associated with maize seeds. Prevalence of seed-borne fungi also varied. The highest percentages of seed-borne fungi were recorded with Fusarium moniliforme and the lowest in Penicillium sp. Transmission of all seed-borne pathogens from seeds to seedlings were also detected by test tube seedling symptom test. Among the seedborne fungi, Alternaria alternata, Fusarium moniliforme and Fusarium sp. produced distinct seed rot and seedling infection symptoms. All the transmitted seed-borne fungi might be caused primary source of infection to the maize crop.

KEYWORDS: Maize, Seed-borne fungi, Transmission

Maize, the important cereal crop of Korea is grown extensively in different glass houses and in field conditions. The crop is affected with as many as 85 diseases (KSPP, 1998; Nyvall, 1989; Rangaswamy, 1979) of which 31 diseases were caused by fungi. Some of the fungal diseases are known to seed-borne and transmitted through seeds (Anderegg and Guthrie 1981; Chang and Kommedahl, 1972; Foley 1962; Futrell and Kilgore, 1969; King 1981; Nyvall 1989; Richardson 1979; Korean Society of Plant Pathology, 1998). From Korea, Kim et al. (1984) tested seven seed samples of corn obtained from Kangweon Provincial Office of Rural Development and found that all the samples were infected with Fusarium moniliforme to an extent of 6.0~79.5%. Nyvall (1989) recorded eight diseases namely-anthracnose leaf blight, black bundle diseases, black kernel rot, ergot, eye spot or brown spot, Fusarium stalk rot, Pythium seed and seedling blight and seed and seedling blight caused by Colletotrichum graminicola (Ces.) G. W. Wilson; Cephalosporium acremonium Cda,; Botryodiplodia theobromae Pat.: Claviceps gigantea Fruentes, de la Isle Ullstrup & Rodriguez; Kabatiella zeae Narita & Hiratsuka; Fusarium moniliforme Sheldon and F. subglutinans (Wollenw & Reink) Nelson, Toussoun & Marasas Comb nov.; Pythium spp.; Diplodia maydis (Earle) Sutton, Gibberella zeae (Schw.) Petch, F. moniliforme Sheldon, Rhizoctonia solani Kühn respectively and found all these fungal pathogens were transmitted by seeds. During the present study, a severe infection on maize kernel was observed at Dongguk University Farm, Go Young City, IL San Gu, during March-

October 2001. Literature review shows that no extensive work on the prevalence of seed-borne fungi and their transmission from seed to seedling of maize seeds has done in Korea. Moreover, every year seed-borne fungi cause heavy yield loss of the crop and some fungi act as a primary source of infection.

Considering the seriousness and common occurrence of kernel rot of maize in a farm of Korea and inadequate information regarding the seed-borne fungi and their transmission from seeds to seedlings this study was undertaken.

Materials and Methods

Isolation of seed-borne fungi. Ten samples of maize seeds of different cutivars/lines grown during June-October, 2001 were collected directly from the experimental field of Dongguk University farm, Go Young City, IL San Gu in the months of November. During collection, seeds from infected maize kernels (Fig. 1A) were isolated separately and preserved in paper bags. Before detection of seed borne fungi from the seeds all the samples were preserved in refrigerator. The mycoflora of the collected seeds samples were detected by blotter method as recommended by International Seed Testing Association (ISTA 1976). The fungi from the seeds were identified by observing their growth characteristics on the incubated seeds in the blotter following the keys offered by different authors (Booth, 1971; Ellis, 1971; Ramnath et al., 1970) and by examining under stereo-binocular microscope whenever necessary.

Test tube-seedling symptom test. The test tube seedling symptom test developed by Khare *et al.* (1977) was

^{*}Corresponding author <E-mail: mwlee@dgu.ac.kr>

48 Basak and Lee

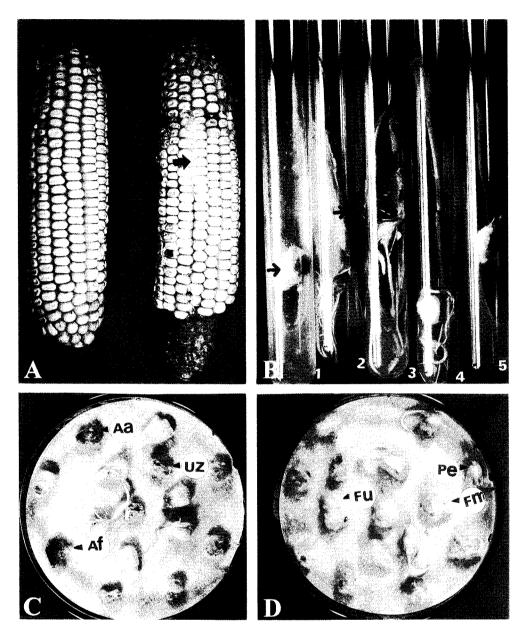


Fig. 1. A. Maize kernel showing infection of seed-borne fungi. B. Transmission of seed borne fungi from seeds to seedlings as determined by the test tube seedling symptom test: (B1) Non germination of seeds or seed rot caused by *Fusarium moniliforme*; (B2) Seedling infection caused by *Alternaria alternata*, (B3), *Fusarium* sp. (B4) and *F. moniliforme* and (B5) A Healthy seedling without fungal infection. C & D. Different seed borne fungi as detected by blotter method: *A. alternata* (Aa), *A. flavus* (Af), *F. moniliforme* (Fm), *Fusarium* sp. (Fu), *Penicillium* sp. (Pe) and *Ustilago zeae* (Uz).

used for this study. Test tube slants were prepared by pouring 6 ml of 2.0% water agar and sterilized in autoclaved for 10 minutes and 15 lb pressure at 121°C. In all two samples having highest percentage of seed infection were employed in this experiment. Two hundred seeds for each sample were used at the rate of one seed per test tube. The test tubes with the seeds were then incubated in the laboratory desk at room temperature $(20 \pm 2^{\circ}\text{C})$. The mouths of the test tubes were properly plugged with cotton and the test tubes were placed on the wooden test incubation. The germination seeds and seedlings in the

test tube examined for the presence of visible symptoms (seed rot, germination failure and infection or death of emerged seedlings) caused by the pathogens present in the seed. The symptoms produced on the germinating seeds and seedlings by the associated pathogen were confirmed by examining the seeds under stereo- binocular microscope.

Results and Discussion

Prevalence of seed-borne fungi. The occurrence of

Table 1. Percentage occurrence of seed borne fungi of selected seed samples of maize grown in a farm of Korea during May-September, 2001

Isolated fungi	Number of seeds with fungi out of 200 seeds	Percentage of		
		Occurrence of fungi ^a	Fungi associated with seeds	
Alternaria alternata	125	8.5	6.3	
Aspergillus flavus	57	3.9	2.9	
Fusarium moniliforme	693	47.0	34.7	
Fusarium sp.	551	37.4	27.6	
Penicillium sp.	26	1.8	1.3	
Ustilago zeae	23	1.6	1.1	

Out of total number of fungal infection of 1475.

individual fungi varied widely. From the total population of fungi encountered in this study, only 5 genera were identified. These were Alternaria alternata (Fr.) Keissler, Aspergillus flavus Lk. Ex Fr., Fusarium moniliforme Sheldon, Fusarium sp., Penicillium sp., and Ustilago zeae Unger. The most predominant fungi detected, in order of prevalence were Fusarium moniliforme, Fusarium sp., Alternaria alternata and Aspergillus flavus Table 1 (Fig 1C and 1D). The highest severity of infection of individual fungus differed greatly. Maximum fungal association was recorded with Fusarium moniliforme and the minimum with Ustilago zeae.

Transmission of seed-borne fungi. Results of transmission of three seed-borne fungi of maize from seed to germinating seeds and seedlings as determined by test tube seedling test are presented in Table 2 (Fig. 1B). In all three kernel rot pathogens namely A. alternata, F. moniliforme and Fusarium sp. were found to transmit to the germinating seeds causing pre-emergence and post emergence death. The rate of transmission of the test kernel rot pathogens from seed to germinating seeds causing pre emergence death or seed rot were always higher than that of transmission to seedling infection or seedling mortality. The highest percentage of seed borne infection 49%, pre emergence death or seed rot (15.5%), seedling infection (11.2%), post emergence death (8.32%) and total disease development (35%) were recorded from the seedlings transmitted from Fusarium moniliforme infected seeds and the lowest from Alternaria alternata.

A. alternata first produced brownish black fluffy col-

ony on the seeds and around the base of seedlings. The fungus infected the cotyledons; as a result the cotyledonary leaves could not open. Sometimes infected seedlings collapsed and finally died (Fig. 1B2). On the total seedborne infection (9%) of *A. alternata*, 3.0% could cause seedling infection or death of seedlings. On the other hand 4.0% seeds could not germinate at all. In field, this pathogen produces Alternaria leaf blight disease. The disease appears as chlorotic streaks that later become necrotic forms on leaves of all ages. Leaves may be killed in one weak if heavy and prolonged dews occur.

Fusarium moniliforme produced orange to dark violet fluffy colony on the seeds and around the base of seedlings (Fig. 1B1). From the colony the mycelia of the fungus were found to infect emerge out seedling. The cotyledonary leaves could not open. Such infected seedlings collapsed and finally died. In an average 11.2% of the seed borne infection of F. moniliforme were capable of causing seedling infection (Fig 1B4) and 8.3% of post emergence death. Moreover 15.5% seeds failed to germinate.

In case of *Fusarium* sp., it produced whitish fluffy colony on the seeds and around the base of seedlings. In the total seed borne infection (26.0%), on an average could not rise to 8.5% seedling infection (Fig. 1B3) and 7.5% post emergence death In this case 41.0% could not germinate

The fungi, which have been detected during the study are known to be transmitted with maize seeds (Richardson, 1979). Most of the fungi encountered in the seeds of maize grown during months from May to September of

Table 2. Transmission of seed borne fungi of maize from seeds to germinating seeds and seedlings as determined by test tube seedling test

Seed-borne fungi	% of seed-borne infection _	% of seed-borne fungi carried			
		Pre-emergence death (%)		Post-emergence	Total disease
		seed rot	seedling	death	development
Alternaria alternata	12.0	4.0	3.0	2.0	9.0
Fusarium moniliforme	49.0	15.5	11.2	8.3	35.0
Fusarium sp.	35.0	10.0	8.5	7.5	26.0

Percent of seed-borne infection of different fungal species was calculated on the basis of 2000 seeds.

50 Basak and Lee

2001 in the present investigation are recorded by different research workers (Chang and Kommedhal 1972; Kim et al., 1984; King and Scott 1981; Windels et al., 1976), but occurrence of Alternaria alternata, Fusarium sp. and Ustilago zeae seems to be new record for Korea. In 1984, Kim et al. detected Fusarium moniliforme as a seed-borne fungus of maize grown in Korea.

The fungal pathogens namely A. alternata, A. flavus, F. moniliforme and U. zeae causing Alternaria leaf blight, Aspergillus ear rot, Fusarium kernel rot and Common smut respectively with maize seeds may provide initial inoculum in the field through root soils of the seedlings. In test tube seedling symptom test, three pathogens namely A. alternata, F. moniliforme and Fusarium sp. were found responsible for transmission from seed to geminating seeds and seedlings. All these fungi under favourable environmental conditions can infect the growing plants and can also serve as source of inoculum for field crop. So, the seed-borne fungi namely- Alternaria, Fusarium should be eliminated to explore the possibility of seed transmission of these fungi under Korean field conditions. Moreover, no infected kernel or seeds from infected kernel should be preserved for raising seedling for the next year. Healthy kernel with seeds should always be preferred for seed preservation purposes.

References

- Anderegg, J. and Guthrie, J. W. 1981. Seed borne Fusarium moniliforme and seedling infection if hybrid sweet corn. Phytopathology 71: 1196-1198.
- Korean Society of Plant Pathology. 1998. List of plant diseases in Korea. 3rd Edition. p. 436.
- Booth, C. 1971. The Genus Fusarium. Commonwealth Mycol. Inst. Kew, Surrey, London, UK. p. 236.

- Chang, M. I. P. and Kommedahl, T. 1972. Interaction among microorganisms occurring naturally and applied to pericarps of corn kernels. Plant Dis. Rep. 56: 861-862.
- Ellis, M. B. 1971. Dematiceous Hyphomycetes. CMI Publication, Surrey, Kew, London, UK. 608 pp.
- Foley, D. C. 1962. Systematic infection of corn by *Fusarium moniliforme*. *Phytopathology* **52**: 870-872.
- Futrell, M. C. and Kilgore, M. 1969. Poor strands of corn and reduction of root growth caused by *Fusarium moniliforme*. *Plant Dis. Rep.* 53: 213-215.
- ISTA. 1976. International rules for seed testing. Seed Science and Technology 4: 3-49.
- Khare, M. N., Mathur, S. B. and Neergaard, P. 1977. A seedling symptoms for detection of *Septoria rodorum* in wheat. *Seed Science and Technoloogy* 5: 613-617.
- Kim, W. G., Oh, I. S., Yu, S. H. and Park, J. S. 1984. Fusarium moniliforme detected in seeds of corn and its pathological significance. Korean J. Mycol. 12: 105-110.
- King, S. B. 1981. Time and infection of maize kernels by Fusarium moniliforme and Cephalosporium acromonium. Phytopathology 71: 796-799.
- and Scott, G. E. 1981. Genotypic differences in maize to kernel infection by *Fusarium moniliforme*. *Phytopathology* **71**: 1245-1247.
- Nyvall, R. F. 1989. Field crop diseases hand book. 2nd edition. Van Nosstrand Reinhold, New York, U.S.A. pp. 91-169.
- Ramnath, Neergaard, P. and Mathur, S. B. 1970. Identification of Fusarium species on seeds as they occur in blotter test. Proc. Int. Seed Test. Ass. 35: 121-144.
- Rangaswamy, G. 1979. Diseases of crop plants in India. 2rd Edition. Prentice Hall of India (Private) limited, New Delhi, 110001, 520 pp.
- Richardson, M. J. 1979. An annotated list of seed borne diseases. 3rd Edition. CAB publications, CMI, Kew, Surrey, UK, pp. 1-320.
- Windels, C. E., Windels, M. B. and Kommedahl, T. 1976. Association of *Fusarium* species with picnic beetles on corn ears. *Phytopathology* **66**: 328-331.