

Low-Dose Gamma Irradiation as Means of Isolating Carotenoid-Hyperproducing Yeast Mutant

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Received: August 17, 2002 Accepted: November 6, 2002

Abstract In order to isolate carotenoid-hyperproducing yeast. low-dose gamma irradiation was used as means of mutagenesis. Phaffia rhodozyma was treated by gamma irradiation of less than 10 kGy, which is considered to be a wholesome irradiation condition established by the Food and Drug Administration. Through repeated rounds of gamma irradiation and visual screening, mutant 3A4-8 was obtained. It produced a 3,824 µg carotenoid/g yeast, 69% higher content than 2,265 µg/g yeast of the unirradiated one. This result indicates that low-dose gamma irradiation could be used as means of mutagenesis to obtain carotenoid-hyperproducing strain of *Phaffia rhodozyma*, since only carotenoid-hyperproducing yeast survived gamma irradiation by scavenging oxygen radicals generated by radiolysis of water.

Key words: Carotenoid, γ-irradiation, mutant

Astaxanthin (3,3-dihydroxy- β -, β -carotene-4,4-dione) is not only an important pigment used in aquaculture industry, but also a strong antioxidant [3, 4, 7, 11, 14]. Astaxanthin is known to play a role in delaying or preventing degenerative diseases [1, 20]. Phaffia rhodozyma is the only yeast that produces astaxanthin, and there has been considerable interest in its commercial use [2, 13, 16, 20]. However, since wildtype strain produces only low quantity of astaxanthin [3, 20], there have been many studies on strain improvement, using antimycin, nitrosoguanidine (NTG), UV, H₂O₂, βionone, and duroquinone [4, 6, 9, 10, 18, 19, 20].

Astaxanthin has been known to protect *P. rhodozyma* against reactive singlet oxygen molecules generated in the natural environment [1, 18, 19]. Thus, a carotenoidhyperproducing strain can survive longer than a wildtype strain under singlet oxygen-rich environment. Gammairradiation produces oxygen radicals generated by radiolysis

of water [5, 17, 21] and can induce mutation of P. rhodozyma through a chromosomal rearrangement [13, 15]. Therefore, the objective of this study was to use γ -irradiation as a selection method of carotenoid-hyperproducing strain.

P. rhodozyma strains used were wildtype 67-385 and antimycin-NTG induced mutant 2A2N [1, 4]. Yeasts were grown as described elsewhere [1, 4, 18]. When P. rhodozyma was grown to log phase, the cells were harvested and resuspended in distilled water and washed twice. The initial cell concentration of strain 67-385 and 2A2N were 5.3×10⁷ and 4.4×10^7 , respectively. The suspension was transferred to a sterile tube and irradiated at room temperature with 0, 1, 2, 3, 3.5, 4, 4.5, 5, 5.5, 6, and 7 kGy, using ⁶⁰Co gamma ray irradiator Type IR-79 (Nordion International Inc., Ontario, Canada) under air. Preliminary experiments were performed with up to 10 kGy. However, the yeast did not grow at 10 kGy. The irradiated cells were diluted with distilled water and plated on YM agar and incubated for 20

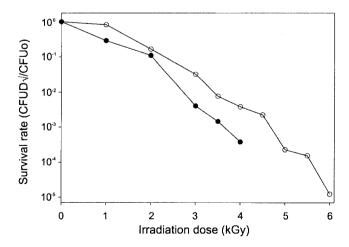


Fig. 1. Survival rate of *P. rhodozyma* by γ -irradiation. CFUy means colony-forming unit after irradiation and CFUo means colony-forming unit without irradiation. —, 67-385; —, 2A2N.

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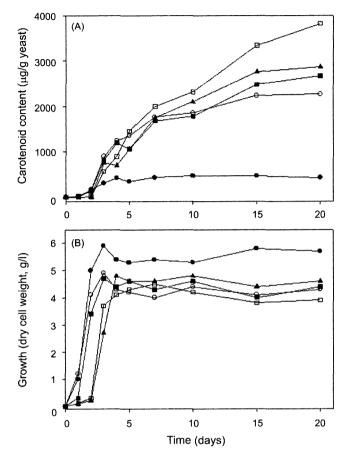
1 ible 1. Carotenoid content of *P. rhodozyma*.

Strains	Carotenoid content (µg/g yeast)
67-385	428
2A2N	2,265
3A	2,668
3A4-8	3,824
3A5-3	2,866

c 1ys. Colonies grown on the plate were counted to c termine survival rate. The number of colonies of strain 6 7-385 decreased more significantly with increasing irradiation c 1938, compared with that of 2A2N (Fig. 1). Strain 2A2N survived up to 6 kGy irradiation, while strain 67-385 did r 1931 survive above 4.5 kGy. This result indicates that the r utant producing more carotenoid is more resistant to copyen radicals than the wildtype strain by scavenging copyen radicals. After the irradiation, the mutant 3A, I wing deep red pigment, was selected and irradiated again to the condition identical to the first irradiation. Two colonies having deep red pigment, 3A4-8 and 3A5-3, were finally selected and their carotenoid content was measured by the method described in the literature [1, 4].

The 3A4-8 and 3A5-3 mutants produced considerably rare carotenoid content than the wildtype strain or the ratant 2A2N (Table 1). In particular, the mutant 3A4-8 raduced the highest carotenoid content, 3,824 μ g/g yeast, after 20 days of culture, compared with 2,265 μ g/g yeast for strain 2A2N. The pattern of carotenoid production in ratant 3A4-8 during growth was different from that of strain 2A2N (Fig. 2). Production of carotenoid in strain 2A2N occurred during the early stationary phase and stopped after 15 days. However, mutant 3A4-8 still showed a significant increase in carotenoid production after 15 days, inplying that mutant 3A4-8 kept on synthesizing caotenoid even after the growth of the mutant had ceased.

A primary function of carotenoid in P. rhodozyma is t) protect cells from singlet oxygen, and it has been cemonstrated to quench singlet oxygen [18]. Carotenoid ontent in the yeast depends on the level of intracellular activated oxygen species, which may induce or activate tie enzymes involved in carotenoid biosynthesis [19]. a staxanthin protects cells by removing reactive oxygen species and rapid synthesis of carotenoid through relief of end-product inhibition in the biosynthesis pathway [18]. chroeder and Johnson [20] also suggested that singlet exygen might induce carotenoid synthesis in P. rhodozyma ly gene activation. Oxygen radicals have been known to cause changes in the molecular properties of proteins as vell as enzyme activities [8, 12, 21]. Therefore, in this tudy, oxygen radicals generated by γ-irradiation might l ave modified the pathway in astaxanthin biosynthesis of .! rhodozyma, and increased carotenoid production of the nutant screened by γ-irradiation.



In conclusion, γ -irradiation could be an efficient method to obtain a carotenoid-hyperproducing strain of *P. rhodozyma* by providing a selection process due to oxygen radicals generated by γ -irradiation. The mutant 3A4-8, screened by consecutive low dose γ -irradiation, produced 69% more carotenoid than the parent strain. To elucidate the mechanism of carotenoid overproduction in the mutant, measurement of the activities of oxygen radical scavenging enzymes such as superoxide dismutase, catalase, and the enzymes involved in carotenoid biosynthesis are in need.

Acknowledgment

This study was supported by a grant from the Korean Ministry of Health and Welfare.

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