

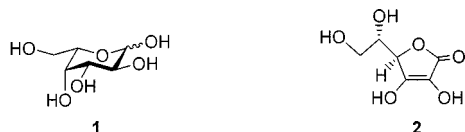
A New Efficient Method for the Synthesis of *L*-Galactose<sup>†</sup>Kwan Soo Kim,<sup>\*</sup> Bong Hwan Cho, and Injac Shin

Department of Chemistry, Yonsei University, Seoul 120-749, Korea

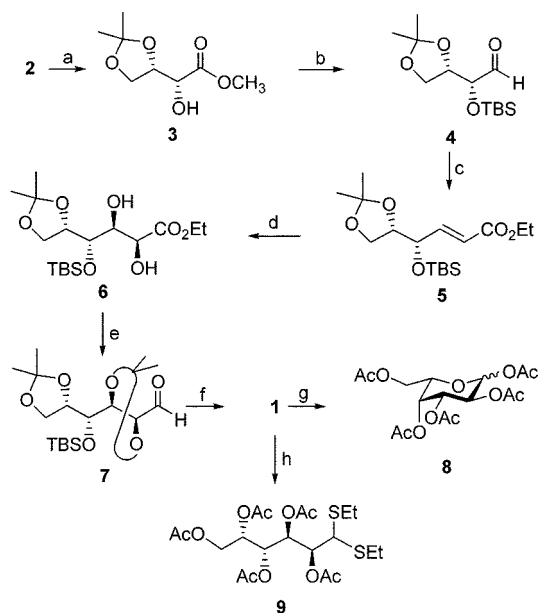
Received May 21, 2002

**Key Words :** *L*-Galactose, *L*-Ascorbic acid, *L*-Sugars

There is a growing need for the synthesis of non-natural *L*-sugars and naturally occurring rare *L*-sugars because of the medicinal potential of *L*-carbohydrates and related nucleosides due to their potent biological activity and lower toxicity compared to their *D*-counterparts.<sup>1</sup> *L*-Sugars are also used as the building block for the synthesis of *L*-oligo-nucleotides and enantio-DNA (DNA having *L*-sugar), which are valuable tools for studying protein-DNA interactions and are promising antisense agents.<sup>2</sup> Although certain *L*-sugars such as *L*-fucose, *L*-rhamnose, and *L*-arabinose are quite abundant in nature, *L*-galactose is a rare sugar and occurs as a minor component in agar-agar, chagual gum, red algae, flaxseed mucilage and a snail galactan.<sup>3</sup> There have been reports for the synthesis of *L*-galactose: (i) a synthesis by reduction of *L*-galactono-1,4-lactone,<sup>4</sup> (ii) a method based on the repeated asymmetric epoxidation starting from achiral 2-butene-1,4-diol,<sup>5</sup> (iii) a synthesis employing the Pummerer rearrangement starting from 6-*S*-phenyl-6-thio-*D*-galactose,<sup>6</sup> and (iv) an enzymatic synthesis by galactose oxidase-catalyzed oxidation of galactitol.<sup>7</sup> These methods have some limitations such as the lengthy synthesis, the carefully controlled reaction in certain steps, and/or the low yield of the product. Herein we report an efficient new method for the synthesis of *L*-galactose (**1**) starting from readily available inexpensive *L*-ascorbic acid (**2**).



The synthesis commenced with transformation of *L*-ascorbic acid (**2**) into the methyl threonate **3** in 74% yield by the known procedure.<sup>8</sup> The hydroxyl group of the compound **3** was protected with *t*-butyldimethylsilyl (TBS) chloride (Scheme 1). The resulting TBS ether was subjected to reduction with DIBAL-H at 78 °C to give the aldehyde **4** in 87% yield. Wittig reaction of the aldehyde **4** with Ph<sub>3</sub>P = CHCO<sub>2</sub>Et in the presence of a catalytic amount of benzoic acid provided the (E)- $\alpha,\beta$ -unsaturated ester **5** in 93% yield along with a small amount of (Z)-isomer (E/Z = 20 : 1). Dihydroxylation<sup>9</sup> of the compound **5** utilizing AD-mix- $\beta$  in the presence of MeSO<sub>2</sub>NH<sub>2</sub> in *t*-BuOH/H<sub>2</sub>O afforded exclusively the diol **6** in 93% yield. Protection of the diol **6** with 2,2-dimethoxypropane followed by reduction of the



**Scheme 1. Reagents and conditions:** (a) see reference 8, 74% in 3 steps; (b) (i) TBSCl, imidazole, DMF, rt, 12 h, 98%; (ii) DIBAL-H, CH<sub>2</sub>Cl<sub>2</sub>, -78 °C, 1 h, 87%; (c) Ph<sub>3</sub>PCHCO<sub>2</sub>Et, benzoic acid (cat.), CH<sub>2</sub>Cl<sub>2</sub>, rt, 4 h, 93%; (d) AD-mix- $\beta$ , MeSO<sub>2</sub>NH<sub>2</sub>, *t*-BuOH-H<sub>2</sub>O, rt, 30 min, then **5**, 0 °C, 12 h, 93%; (e) (i) 2,2-dimethoxypropane, TsOH (cat.), acetone, 4 h, 96%; (ii) DIBAL-H, CH<sub>2</sub>Cl<sub>2</sub>, -78 °C, 1 h, 89%; (f) *c*-HCl, CH<sub>3</sub>CN-H<sub>2</sub>O, rt, 1 h; (g) Ac<sub>2</sub>O, DMAP (cat.), pyridine, 0 °C to rt, 5 h, 80% in 2 steps from **7**; (h) (i) EtSH, *c*-HCl, rt, 10 min; (ii) Ac<sub>2</sub>O, DMAP (cat.), pyridine 0 °C to rt, 4 h, 91% in 3 steps from **7**.

resultant di-*O*-isopropylidene ester with DIBAL-H at 78 °C gave the protected *L*-galactose **7**.<sup>10</sup> Hydrolysis of the purified **7** with *c*-HCl in acetonitrile provided *L*-galactose (**1**), of which acetylation with acetic anhydride in the presence of a catalytic amount of DMAP in pyridine gave the *L*-galactose pentaacetate **8** in 80% yield in two steps. The crude aldehyde **7** could be used without purification for the subsequent hydrolysis and acetylation steps. <sup>1</sup>H and <sup>13</sup>C NMR spectra of the compound **8** were identical with those of *D*-galactose pentaacetate, which we prepared from *D*-galactose. For the purpose of further identification, *L*-galactose (**1**) was treated with EtSH in the presence of *c*-HCl to afford the acyclic *L*-galactose dithioacetal as white solid, of which acetylation with acetic anhydride gave the pentaacetyl-*L*-galactose dithioacetal **9** {[ $\alpha$ ]<sub>D</sub> -10.7 (c 3.4, CHCl<sub>3</sub>)}.<sup>11</sup> <sup>1</sup>H and <sup>13</sup>C NMR spectra of the compound **9** was identical with those of its enantiomer, pentaacetyl-*D*-galactose dithioacetal {[ $\alpha$ ]<sub>D</sub> +10.5 (c 3.4, CHCl<sub>3</sub>) (lit<sup>11</sup>: [ $\alpha$ ]<sub>D</sub> +9.8, CHCl<sub>3</sub>) (lit<sup>12</sup>: [ $\alpha$ ]<sub>D</sub> +11.31, c

<sup>†</sup>This paper is dedicated to the late Professor Sang Chul Shim.

2.2.  $\text{CHCl}_3$ }, which we prepared from *D*-galactose. Thus, the conversion of *L*-ascorbic acid to the *L*-galactose pentaacetate **8** was accomplished in 37% overall yield.

**Acknowledgment.** This work was supported by Korea Research Foundation Grant (KRF-2001-042-D00049).

### References

- (a) Chu, C. K.; Ma, T. W.; Shanmuganathan, K.; Wang, C. G.; Xiang, Y. J.; Pai, S. B.; Yao, G. Q.; Sommadossi, J.-P.; Cheng, Y.-C. *Antimicrob. Agents Chemother.* **1995**, *39*, 979. (b) Schinazi, R. F.; Gosselein, G.; Paraj, A.; Korba, B. E.; Liotta, D. C.; Chu, C. K.; Mathe, C.; Imbach, J.-L.; Sommadossi, J.-P. *Antimicrob. Agents Chemother.* **1994**, *38*, 2172. (c) Gosselein, G.; Schinazi, R. F.; Sommadossi, J.-P.; Mathe, C.; Bergogne, M.-C.; Aubertin, A.-M.; Kim, A.; Imbach, J.-L. *Antimicrob. Agents Chemother.* **1994**, *38*, 1292. (d) Chang, C.-N.; Skalski, V.; Zhou, J. H.; Cheng, Y.-C. *J. Biol. Chem.* **1992**, *267*, 22414. (e) Schinazi, R. F.; McMillan, A.; Cannon, D.; Mathis, R.; Lloyd, R. M.; Peck, A.; Sommadossi, J.-P.; St. Clair, M.; Wilson, J.; Furman, P. A.; Painter, G.; Choi, W. B.; Liotta, D. C. *Antimicrob. Agents Chemother.* **1992**, *36*, 2413. (f) Spadari, S.; Maga, G.; Foehner, F.; Ciarrocchi, G.; Manservigi, R.; Arcamone, F.; Capobianco, M.; Carcuro, A.; Colonna, E.; Iotti, S.; Garbesi, A. *J. Med. Chem.* **1992**, *35*, 4214.
- (a) Goodchild, J. *Bioconjugate Chem.* **1990**, *1*, 16. (b) Uhlmann, E.; Peyman, A. *Chem. Rev.* **1990**, *90*, 543. (c) Beaucage, S. L.; Iyer, R. P. *Tetrahedron* **1993**, *49*, 6123.
- Schafer, A. In *The Carbohydrates: Chemistry and Biochemistry*, 2<sup>nd</sup> ed.; Pigman, W.; Horton, D., Eds.; Academic Press: New York, 1972; Vol. IA, pp 85-86.
- Frush, H. L.; Isbell, H. S. *Methods Carbohydr. Chem.* **1962**, *1*, 127.
- Ko, S. Y.; Sharpless, K. B. *Science* **1983**, *220*, 949.
- Gonzalez, F. S.; Baer, H. H. *Carbohydr. Res.* **1990**, *202*, 33.
- Root, R. L.; Durrwachter, J. R.; Wong, C.-H. *J. Am. Chem. Soc.* **1985**, *107*, 2997.
- Kim, K. S.; Cho, I. H.; Ahn, Y. H.; Park, J. I. *J. Chem. Soc., Perkin Trans. 1* **1995**, 1783.
- Harmuth, C. K.; VanNeieuwenhze, M. S.; Sharpless, K. B. *Chem. Rev.* **1994**, *94*, 2483.
- Compound **7**: <sup>1</sup>H NMR (250 MHz, CDCl<sub>3</sub>) δ 0.12 (s, 6H), 0.92 (s, 9H), 1.31 (s, 3H), 1.33 (s, 3H), 1.41 (s, 3H), 1.50 (s, 3H), 3.67-3.75 (m, 1H), 3.89-3.93 (m, 1H), 3.96-4.08 (m, 3H), 4.57 (dd, *J* = 6.5, 1.1 Hz, 1H), 9.81 (d, *J* = 1.1 Hz, 1H); <sup>13</sup>C NMR (63 MHz, CDCl<sub>3</sub>) δ -4.11, -3.96, 18.44, 25.62, 26.07, 26.44, 26.48, 65.93, 73.37, 77.86, 78.31, 80.38, 109.12, 110.62, 201.46.
- Wolfrom, M. L. *J. Am. Chem. Soc.* **1930**, *52*, 2464.
- Araki, C.; Hirase, S. *Bull. Chem. Soc. Jpn.* **1953**, *26*, 463.