

Calcium carbonate fibrin adhesive

2

I.

7,8) , Genius Porites
porous resorbable calcium car -
bonate 98%가 calcium car -
bonate (CaCO₃)⁹⁾,
100 - 200 μm

1-2)

10,11). Guillemin

12)

가 ,

12

가

가

가

Ouhayoun^{13,14)}

porous hydroxyapatite

4

3-6).

cal -

Issahakian¹⁵⁾

cium carbonate, calcium sulfate, tricalcium
phosphate, hydroxyapatite

polymers, hydrox -

yapatite . Hydroxya - patite가

(scaffold)

16).

1909 Bergel fibrin powder .

, 2.

. 1944 1)

Cronkite Tedrick ¹⁷⁾fibrinogen
 thrombin , calcium carbonate I
 tissue adhesive 가 , fibrin adhesive
 fibrin adhesive II , calcium carbonate fibrin adhe -
 sive III

(bone sealing), 2)
 가 . (

²¹⁾ ,) 2mg/kg
²²⁻²⁴⁾ Halothane - O₂

2 calcium epinephrine
 carbonate calcium car - (, ,)
 bonate fibrin adhesive .

fibrin 2, 3, 4
 adhesive가 1 mm fis -
 7
 mm, 5 mm, 3 mm 2

II. No. 1/2 round bur

1. No. 1/4 round bur

15 kg 6 .
 1 3 - 0
 가 Mersilk (Ethicon co, U.S.A.)
 2, 3, 4 .

7 (

4 ,) 500 mg
 1 2
 Calcium carbonate(Biocoral
 450, Inoteb, France)

Fibrin adhe -
 sive(Green Plast , ,) (K - Y gel , Johnson & Johnson, U.S.A.)

2

3)

2, 4, 12

(Figure 2).

가

10%

formalin 1

, 5% nitric acid

(3) II

paraffin

Fibrin adhesive

4 μm

. hematoxylin - eosin

(Figure

3).

4)

(4) III

4 12

1)

, 2)

, 3)

가

, 4)

III.

(Figure 4).

1.

1

I

II

III

. 2, 4, 12

2.

1) 2

(1)

가

가

(Figure

1).

(2) I

2) 4 (1) (4) III 가

(Figure 5A, B). 가 가

(2) I (Figure 8A, B, C).

(Figure 6A, B). 3) 12 (1) 가

(3) II 4

(Figure 9A, B).

(Figure 7A, B). (2) I 가

Table 1. Linear histometrics in bucco - lingual sections for treatment modalities at 4 weeks

	Junctional epithelium	New attachment	
		Connective tissue	Alveolar bone
Control	2.46 ± 0.40	0.52 ± 0.62	1.06 ± 0.48
Group I	2.00 ± 0.42	0.48 ± 0.56	0.60 ± 0.21
Group II	2.10	2.40 ± 0.36	1.50 ± 0.17
Group III	1.75 ± 0.78	1.77 ± 0.67	2.23 ± 0.25

Control : flap debridement

Group I : Calcium carbonate graft only

Group II : Application of Fibrin adhesive only

Group III : Application of Fibrin adhesive after calcium carbonate grafting

Table 2. Linear histometrics in bucco - lingual sections for treatment modalities at 12 weeks

	Junctional epithelium	New attachment	
		Connective tissue	Alveolar bone
Control	2.22 ± 0.79	1.30 ± 0.54	1.73 ± 0.73
Group I	2.68 ± 0.74	1.54 ± 0.47	0.82 ± 0.40
Group II	1.45 ± 0.21	2.10 ± 0.17	1.27 ± 0.50
Group III	1.66 ± 0.62	0.85 ± 0.63	3.30 ± 1.44

Legands are the same as table 1

(Figure 10A, B).

(3) II

가

(Figure 11A, B).

(4) III

가

2/3

가

(Figure 12A, B, C).

3.

2.46mm	12	4	1.75 -
		1.45 - 2.68mm	
		I	가
1.77mm, 12		4	0.48 -
		0.85 - 2.1mm	
		4	0.6 -
2.23mm, 12		0.82 - 3.3mm	
	II	4	12

(Table 1, 2).

IV.

scar"

" healing by
가

Bajpai ¹²⁾

, 가

가

가

가
bonate

calcium car -
Natural Coral 1987

Yukna ¹⁰⁾ carbonate phase

hydroxyapatite

carbonate
calcium carbonate

Ouhayoun ¹¹⁾ calcium carbonate

1

3

가

6

가

가, 12

²⁷⁾

가

²⁸⁾

가

가

가

가

가 .

2 fibrin adhesive 4 , 12 가

가 2 가

fibronectin 가 ,

30) 가

fibrin 17) fibrin adhesive 4 , 12 가

Bosch²⁵⁾ fibrin adhesive가 4 ,

12 가 4

adhesive 가 fibrin 가 12 가

26) 4 가

Fibrin adhesive Fibrin adhesive 2

31) fibrin linkage 가

가 가 29) fibrin

fibrin adhesive adhesive 가

Green plast 가 biological carrier

fibrinogen XIII, fibrin adhesive가

Plasma protein, fibronectin, cold agent

insoluble globulin anti - fibrinolytic agent aprotinin, sustained local deliv -

thrombin, thrombin, ery carrier ,

thrombin fibrinogen fibrin 가 Fibrin adhesive

가 fibrin adhesive

가

V.

2
calcium carbonate

fibrin adhesive

가

6

4

I cal-

cium carbonate

II fibrin adhesive

III

calcium carbonate

fibrin adhesive

2, 4, 12

1. 1
2, 4, 12
가

2. 가
4, 12

3. I 가
. 4, 12

4. II fibrin adhe -
sive가 2

5. III
가
2
. 4

. 12
2/3

2
fibrin adhesive

가

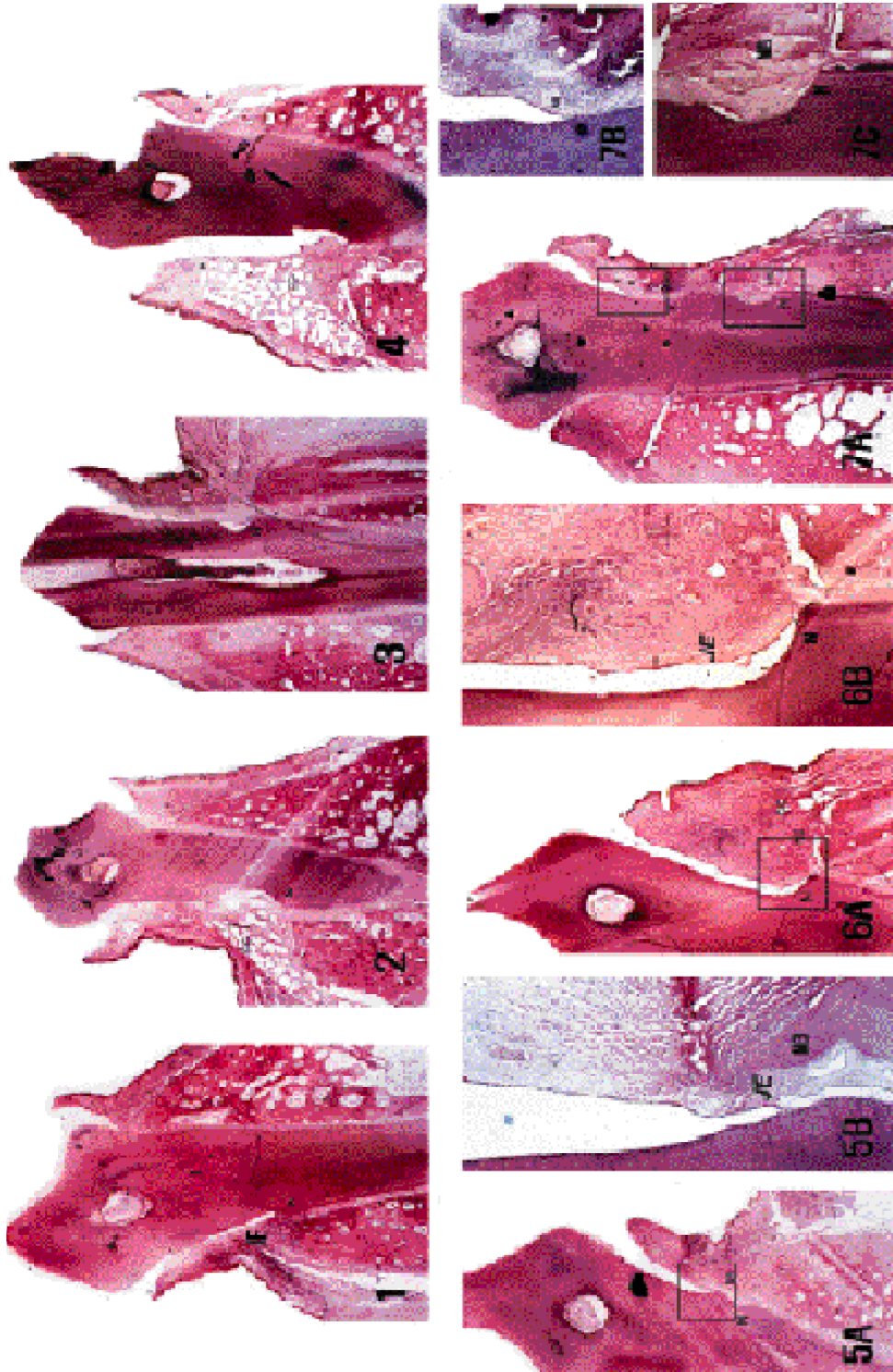
VI.

1. Stahl SS, Froum SJ, Kushner L. : Periodontal healing following open debridement flap procedure. II. Histologic observations. J Periodontol 53 : 15 - 21, 1982.
2. Steiner SR, Crigger M, Egelberg J. : Connective tissue regeneration to periodontally diseased teeth. II. Histologic observations of cases following replaced flap surgery. J Periodont Res 16 : 109 - 116, 1976.
3. Chamberlain ADH, Garrett S, Renvert S, Egelberg J. : Healing after treatment of periodontal intraosseous defects. : IV. Effect of a non - resective versus a partially resective approach. J Clin Periodontol 12 : 525 - 539, 1985.

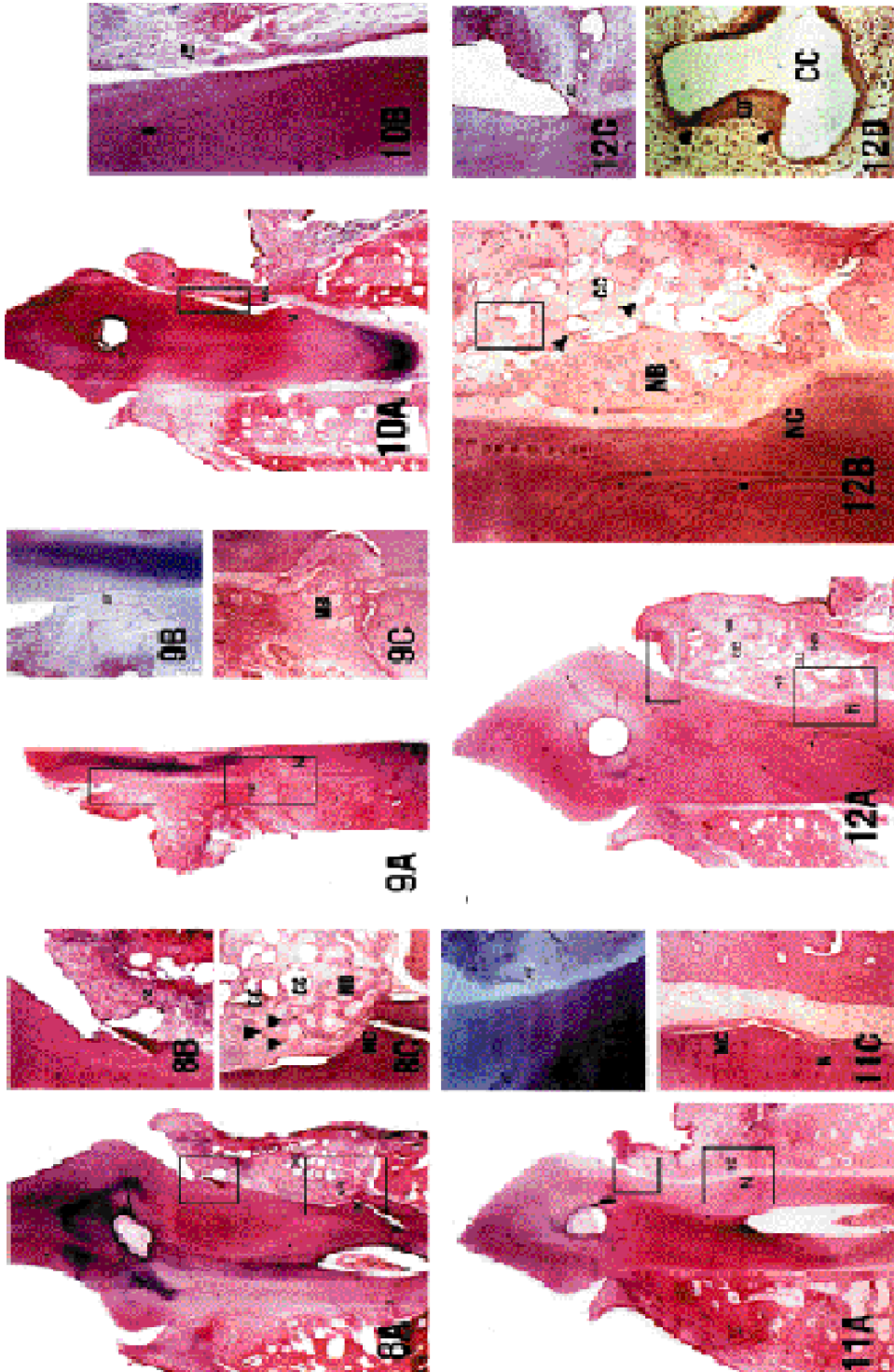
4. Froum SJ, Coran M, Thaller B, Kushner L, Scopp IW, Stahl SS. : Periodontal healing following open debridement flap procedure(I). Clinical assessment of soft tissue and osseous repair. *J Periodontol* 53 : 8 - 14, 1982.
5. Diem CR, Bowers GM, Moffitt WC. : Bone blending : A Technique for osseous implants. *J Periodontol* 43 : 295, 1972.
6. Carranza FA, Kenney EB, Lekovic V, Talamante E. : Histologic study of healing of human periodontal defects after placement of porous hydroxyapatite implants. *J Periodontol* 59 : 682 - 688, 1987.
7. Kenny EB, Lekovic V, Han T, Carranza FA, Dimitrijevic B. : The use of porous hydroxyapatite implant in periodontal defects. Clinical results after six months. *J Periodontol* 56 : 82 - 88, 1985.
8. Kenny EB, Lekovic V, Sa Ferreira JC, Han T, Carranza FA, Dimitrijevic B. : Bone formation with porous hydroxyapatite implants in human periodontal defects. *J Periodontol* 57 : 76 - 83, 1986.
9. Guillemin G, Meunier A, Dallant P, Christel P, Pouliquen JC, Sedel L. : Comparison of coral resorption and bone apposition with two natural corals of different porosities. *J Biomed Mat Res* 23 : 765 - 779, 1989.
10. Yukna RA. : Clinical evaluation of coralline calcium carbonate as bone replacement graft material in human periodontal osseous defects. *J Periodontol* 65 : 177 - 185, 1994.
11. Ouhayoun JP, Shabana AHM, Issahakian S, Patat JL, Guillemin G, Sawaf MH, Forsest N. : Histological evaluation of natural coral skeleton as a grafting material in miniature swine mandible. *J Mat Science in Medicine*. 3 : 222 - 228, 1992.
12. Guillemin G, Patat JL, Fomic J, Chetail M. : The use of coral as a bone graft substitute. *J Biomed Mat Res*. 21 : 557 - 567, 1987.
13. Ouhayoun JP, Issahakian S, Patat JL, Guillemin G. : Influence of biomaterials on the healing pattern of bony defects in miniature pig mandible. *J Dent Res* 68(special issue) : Abst. No. 1244, 1989.
14. Ohgushi H, Okumura M, Yoshikawa T, Inoue K, Senpoku N, Tamai S, Shors EC. : Bone formation process in porous calcium carbonate and hydroxyapatite. *J Biomed Mat Res*. 26 : 885 - 895, 1992.
15. Issahakian S, Ouhayoun JP, Shabana H, Sawaf H. : Evaluation of new biomaterial in periodontal defects : Natural coral. *J Dent Res* 68(special issue) : Abst. No. 274, 1989.
16. Radell BL, Cassingham RJ. : A clinical evaluation of Proplast as a periodontal implant material. *J Periodont* 51 : 110, 1980.
17. Martas H : Fibrin seal : The state of the art. *J Oral Maxillofac Surg*. 43 : 605 - 611, 1985.
18. Pairot T, David AO, Carmela BM,

- Lyn YA. : Autologous fibrin adhesive in mandibular reconstruction with particulate cancellous bone and marrow, *J Oral Maxillofac Surg.* 1994 : 52 : 161 - 165.
19. Wittkamp ARM. : Augmentation of the maxillary alveolar ridge with hydroxyapatite and fibrin glue. *J Oral Maxillofac Surg* 46 : 1019, 1988.
 20. Bonucci E, Marini E, Valdinucci F, Fortunato G : Osteogenic response to hydroxyapatite - fibrin implants in maxillofacial bone defects. *Eur J Oral Sci* 105 : 557 - 561, 1997.
 21. Corrente G, Abundo R, Cardaropoli G, Martuscelli G, Trisi P. : Supracrestal bone regeneration around dental implants using a calcium carbonate and a fibrin - fibronectin sealing system. *Int J Periodont Rest Dent* 17 : 171 - 181, 1997.
 22. Trombelli L, Schincaglia G, Checchi L, Calura G. : Combined guided tissue regeneration, root conditioning, and fibrin - fibronectin system application in the treatment of gingival recession. *J Periodontol* 65 : 796 - 803, 1994.
 23. Pini Prato G, Cortellini P, Clauser C : Fibrin and fibronectin sealing system in a guided tissue regeneration procedure. *J Periodontol* 59 : 679 - 683, 1998.
 24. Warrer K, Karring T : Effect of Tisseel[®] on healing after periodontal flap surgery. *J Clin Periodontol* 19 : 449 - 454, 1992.
 25. Bosch P, Lintner F, Arbes S. : Experimental investigations of the effects of the fibrin adhesive on the kiel heterologous bone graft. *Arch Orthop Trauma Surg* 96 : 177, 1980.
 26. , : 3 가 .
1994 : 6 : 279 - 293.
 27. Gottlow J, Nyman S, Lindhe J, Wennstrom J. : New attachment formation in the human periodontium by guided tissue regeneration. *Case Reports. J Clin Periodont* 13 : 604 - 616, 1983.
 28. Rosenberg MM. : Periodontal and Prosthetic Management for advanced case. 197 - 201, 1988.
 29. Caton JG, Polson AM, Pini Prato G, Bartolucci EG, Clauser C. : Healing after application of tissue - adhesive material to denuded and citric acid - treated root surfaces. *J Periodontol* 57 : 385, 1988.
 30. Knox P, Crooks S, Rimmer CS : Role of fibronectin in the migration of fibroblasts into plasma clots. *J Cell Biol.* 102 : 2318 - 2325, 1986.
 31. Bartolucci E, Pini Prato G : Preliminary observations on the use of a biologic sealing system in periodontal surgery. *J Periodontol* 53 : 731 - 735, 1982.
 32. Cortellini P, Pini Prato GP, Tonetti MS : No detrimental effect of fibrin glue on the regeneration of intrabony defects. *J Clin Periodontol* 22 : 697 - 702, 1995.
 33. , : Oxidized cellulose membrane replemineform hydroxyapatite calcium carbonate .
22(2) 201 - 222, 1992.
 34. , , , : Collagen absorbable hemostat porous resorbable calcium carbonate가 .

(I)



(II)



22(2) 241 - 257, 1992.

35. , : 3

calcium sulfate calcium carbonate
. 26(3) 605 - 624, 1996.

36. , , , , .
.

. : 가
26(4) 907 - 932, 1996.

Figure 1. Control group at 2 weeks. Junctional epithelium(JE) was down growth at the notch(N) area(10X).

Figure 2. Group I at 2 weeks. Calcium Carbonate(CC) were seen only at the notch(N) area(10X).

Figure 3. Group II at 2 weeks. Fibrin adhesive was not seen at the defect(10 X).

Figure 4. Group III at 2 weeks. Calcium Carbonate(CC) were seen at all of the defect(10X).

Figure 5. Control group at 4 weeks. Junctional epithelium(JE) was down growth at the notch(N) area and new bone(NB) was seen at the defect base(A 10X, B 40X).

Figure 6. Group I at 4 weeks. Calcium carbonate(CC) were not seen at the defect and junctional epithelium(JE) was down growth at the notch(N) area(A 10X, B 40X).

Figure 7. Group II at 4 weeks. Junctional epithelium(JE) was not down growth and new bone(NB) was seen around the notch(N) area(A 10X, B 40X).

Figure 8. Group III at 4 weeks. Junctional epithelium(JE) was not down growth and new bone(NB) was seen above the notch(N). the new bone surface were lined with osteoblast(arrow head)(A 10X, B,C 40X).

Figure 9. Control group at 12 weeks. Junctional epithelium(JE) was down growth and new bone(NB)

was seen only at the notch(N) area(A 10X, B 40X).

Figure 10. Group I at 12 weeks. Calcium Carbonate(CC) was not seen at the defect and junctional epithelium(JE) was down growth(A 10X, B 40X).

Figure 11. Group II at 12 weeks. Junctional epithelium(JE) was not down growth and new bone(NB) was seen above the notch(A 10X, B 40X).

Figure 12. Group III at 12 weeks. Calcium Carbonate(CC) was seen at the defect and junctional epithelium(JE) was not down growth. new bone and new cementum(NC) was seen at the defect. the new bone surfaces were lined with osteoid(OT) and osteoblast(arrow head)(A 10X, B,C 40X).

- Abstract -

The Effects of the Combination of Calcium Carbonate and Fibrin Adhesive on the Periodontal Regeneration of Class II Furcation Defect in Dogs

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The purpose of this study was to evaluate the effect of fibrin tissue adhesive and porous resorbable calcium carbonate on the periodontal regeneration of the class II furcation defect in dogs.

Class II furcation defect was surgically created on the second, third, and fourth premolars bilaterally in the mandibles of six mongrel dogs. The experimental sites were divided into four groups according to the treatment modalities: Control - surgical debridement only; Group I - calcium carbonate grafting; Group II - application of fibrin adhesive only; Group III - application of fibrin adhesive after calcium carbonate grafting.

The animals were sacrificed at the 2, 4,

and 12 weeks after periodontal surgery and the decalcified specimens were prepared for histological and histometrical examination.

The results are as follows :

Clinically, there were no inflammatory response in all groups after 2, 4, 12 weeks. In the Control group, junctional epithelium was grown downward to the reference notch. In Group I, graft materials were exfoliated from the defect throughout the experimental periods and new bone was seen in the notch area at 4 and 12 week specimens. In Group II, fibrin adhesive was absorbed at 2 week specimens, and connective tissue attachment increased than that of control group. New cementum and new bone were seen above the notch area. In Group III, the graft material was maintained in the defect throughout the experimental period and inducing the amount of periodontal tissue regeneration was higher than other groups.

These results suggest that the use of fibrin tissue adhesive in conjunction with porous resorbable calcium carbonate would improve the stability of graft material and inhibit the epithelial down growth and make it be a feasible method for periodontal regeneration.